

A 15 β -HYDROXYWITHANOLIDE FROM *DATURA FEROX*

ADRIANA CIRIGLIANO, ADRIANA S. VELEIRO, JUAN C. OBERTI* and GERARDO BURTON†

Departamento de Química Orgánica, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Pabellón 2, Ciudad Universitaria, (1428) Buenos Aires, Argentina; *Departamento de Química Orgánica and IMBIV (CONICET), Facultad de Ciencias Químicas, Universidad Nacional de Córdoba, Argentina

(Received 10 January 1994)

Key Word Index—*Datura ferox*; Solanaceae; withanolide; 15 β -hydroxynicandrin B.

Abstract—The leaves of *Datura ferox* yielded a new withanolide, 5 α ,12 α ,15 β -trihydroxy-6 α ,7 α -epoxy-1-oxo-22R-witha-2,24-dienolide (15 β -hydroxynicandrin B), in addition to nicandrin B, withanicandrin, withastramonolide and withametelin E previously reported from *Datura* species. The structure and stereochemistry of 15 β -hydroxynicandrin B was established on the basis of spectroscopic evidence and molecular modelling.

INTRODUCTION

Steroidal lactones (C-28) (withanolides) based on the ergostane framework, have been isolated from various genera of the Solanaceae family [1]. To date, six *Datura* species have been reported to contain withanolides, namely *D. quercifolia* [2], *D. stramonium* [2], *D. metel* [1, 3–5], *D. tatura* [6], *D. fastuosa* [7, 8] and *D. ferox* [2].

From leaves of *D. ferox* collected in Argentina we have isolated a new withanolide, 15 β -hydroxynicandrin B (1). The plant also contained, in addition to nicandrin B (withaferoxolide) (2) and withanicandrin (3) previously isolated from its leaves [2], withastramonolide (4) and withametelin E (5) which have been reported from *D. stramonium* [2] and *D. metel* [4], respectively.

RESULTS AND DISCUSSION

15 β -Hydroxynicandrin B (1), C₂₈O₇H₃₈ showed in its FAB-mass spectrum (*m*-nitrobenzylalcohol, KCl) a quasi-molecular ion [M + K]⁺ at *m/z* 525. The ¹H NMR spectrum (Table 1) had two olefinic protons, a double double doublet at δ 6.61 and a double doublet at δ 5.87 corresponding to H-3 and H-2, respectively. The signal at δ 3.05 was assigned to H-6 by analogy with known 5 α -hydroxy-6 α ,7 α -epoxywithanolides [2]. An AB system at δ 2.61 was observed in the five withanolides isolated from this plant, it was assigned to the methylene hydrogens at position C-4 based on the spin-spin couplings and ¹H–¹H correlations with H-2 and H-3 (COSY 45 and DQF COSY).

The presence of two methyl singlets at δ 1.89 and 1.94 indicated a typical α , β -unsaturated δ -lactone side-chain

bearing methyl groups at positions C-24 and C-25. At the high-field end of the spectrum three signals appeared for Me-18 (δ 1.06), Me-21 (δ 1.10) and Me-19 (δ 1.20). The broad signal at δ 3.96 (*W*_{1/2} 6.0 Hz) was characteristic of a 12 α -hydroxywithanolide [2].

The main difference observed in the ¹H NMR spectrum of 1 when compared with the related compounds 2–5, was a downfield shift (*ca* 0.35 ppm) of the signal for H-7, which appeared at δ 3.72, and the presence of an additional signal at δ 4.45 assigned to H-15.

The ¹³C NMR spectrum of 1 (Table 2), complemented by DEPT spectra, was in agreement with the proposed structure. The spectral data were closely related to those of nicandrin B (2) for rings A–C and the side-chain [2, 9], the main differences being in the signals for carbons C-8, C-14, C-15, C-16 and C-18. The signal at δ 69.8 was assigned to C-15, bearing the hydroxyl group.

The presence of a hydroxy group at position 15 β , was established by NOESY experiments and molecular modelling considerations using the PM3 semi-empirical method (Table 3). The NOESY spectrum showed a strong correlation between the signal at δ 4.45 (assigned to H-15) and H-7. Molecular modelling calculations indicated that the H-15/H-7 distance was below 3.0 Å for both the 15 α and 15 β isomers of 1 confirming the presence of the substituent at position C-15. However no NOE was observed between H-15 and Me-18, thus, H-15 must be in the α -orientation.

To our knowledge, the only other 15 β -hydroxywithanolide reported is the allylic alcohol nicaphysalin C, recently isolated from *Nicandra physaloides* [10].

EXPERIMENTAL

Mps: uncorr. ¹H and ¹³C NMR spectra were measured at 200.13 and 50.32 MHz respectively in CDCl₃ with

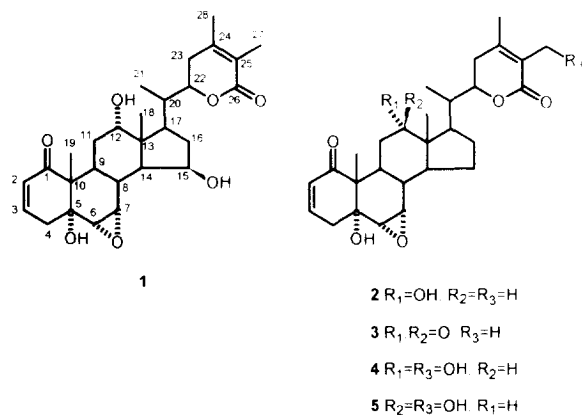
†Author to whom correspondence should be addressed.

Table 1. ^1H NMR spectral data for 15β -hydroxynicantrin B (**1**) in CDCl_3

H	δ	H	δ
2	5.87 <i>dd</i> (2.9, 10.1)	16 α	2.32 <i>ddd</i> (14.1, 13.3, 5.5)
3	6.61 <i>ddd</i> (2.5, 5.0, 10.1)	16 β	1.41 <i>ddd</i> (14.1, 9.0, 1.7)
4 α	2.52 <i>dd</i> (18.0, 5.0)	17	1.92 <i>ddd</i> (13.3, 9.0, 1.7)
4 β	2.70 <i>ddd</i> (18.0, 2.9, 2.5)	18	1.06 <i>s</i>
6	3.05 <i>d</i> (3.8)	19	1.20 <i>s</i>
7	3.72 <i>dd</i> (3.8, 1.7)	20	2.12 <i>m</i>
8	2.21 <i>dt</i> (1.7, 10.8)	21	1.10 <i>d</i> (6.5)
9	2.10 <i>dt</i> (2.7, 10.8)	22	4.35 <i>dt</i> (13.5, 3.5)
11 α	2.85 <i>dt</i> (13.0, 2.7)	23 α	1.99 <i>dd</i> (18.3, 3.5)
11 β	1.61 <i>ddd</i> (13.0, 10.8, 3.5)	23 β	2.51 <i>dd</i> (18.3, 13.5)
12	3.96 <i>brs</i> ($W_{1,2}$ 6.0)	27	1.89 <i>s</i>
14	1.87 <i>dd</i> (10.8, 5.5)	28	1.94 <i>s</i>
15	4.45 <i>dt</i> (1.7, 5.5)		

Chemical shifts in δ from TMS. Coupling constants (in parentheses) in Hz.

Assignments are based on phase-sensitive DQF COSY spectrum and coupling constants.

Table 2. ^{13}C NMR spectral data of **1** in CDCl_3

C	δ	C	δ
1	203.6	15	69.8
2	128.9	16	40.7
3	140.0	17	43.4
4	36.8	18	14.7
5	73.4	19	15.0
6	56.2	20	38.8
7	56.7	21	12.1
8	32.6	22	78.4
9	28.9	23	29.9
10	50.7	24	149.4
11	29.9	25	122.0
12	73.1	26	167.3
13	46.7	27	12.5
14	48.4	28	20.5

TMS as int. standard. Multiplicity determinations (DEPT) and 2D NMR spectra (DQF COSY, NOESY) were obtained using standard Bruker software. FAB-MS were determined on a VG ZAB-BEQQ mass spectrometer. PM3 calculations were performed with AMPAC 4.5 (Semichem, Shawnee, KS).

Plant material and isolation procedure. Whole *Datura ferox* plants were collected in San Antonio de Litín, Córdoba province, Argentina. A voucher specimen is deposited at the Museo Botánico, Universidad Nacional de Córdoba [CORD]. Dried and pulverized leaves (987 g) were extracted with CHCl_3 - Me_2CO - MeOH at room temp. The resulting soln was filtered and evapd to dryness. The residue was partitioned with hexane- MeOH - H_2O (10:3:1), the hexane layer was discarded and the methanolic layer concd and extracted exhaustively with CHCl_3 . The organic layer was evapd to dryness. The residue (3.15 g) was chromatographed on silica gel. Elution with EtOAc-hexane-*iso*-PrOH (30:3:2) afforded four frs containing withanolides which were further purified by flash chromatography using EtOAc-hexane-*iso*-PrOH mixts of increasing polarity.

Early frs of eluate (EtOAc-hexane-*iso*-PrOH 100:10:1) yielded withanicantrin (**3**) (10.6 mg), crystals from EtOAc-hexane, mp 264–266° (lit. [2] 267°). Continued elution with EtOAc-hexane-*iso*-PrOH (50:5:1) furnished a mixt. of two withanolides which were purified by prep. reverse-phase TLC, MeOH - H_2O (7:3) yielding withametelin (**5**) (1.0 mg), crystals from EtOAc-hexane, mp 259–260° (lit. [4] 260–262°) and 5 α ,12 α ,15 β -trihydroxy-6 α ,7 α -epoxy-1-oxo-22*R*-witha-2,24-dienolide (15 β -hydroxynicantrin B) (**1**) (1.5 mg), crystals from EtOAc-hexane, mp 266–267°; ^1H and ^{13}C NMR: Tables 1 and 2; FAB-MS (*m*-nitrobenzylalcohol, KCl) m/z (rel. int.): 525 [$\text{M} + \text{K}$] $^+$ (100), 487 [$\text{M} + 1$] $^+$ (17), 485 [$\text{M} - 1$] $^+$ (15), 469 (10), 451 (12). Further elution with EtOAc-hexane-*iso*-PrOH (50:5:2) afforded withas-tramonolide (**4**) (16 mg), crystals from EtOAc-hexane, mp 271–273° (lit. [2] 269–270°) and nicantrin B (withaferoxolide) (**2**) (34 mg), crystals from

Table 3. Correlations displayed by 15 β -hydroxynicandrin B (**1**) in the NOESY spectrum*

H	NOESY correlations	H-H distance (Å) [†]
7	H-15	2.8
8	H-18, H-19	< 2.4
11 β	H-18, H-19	< 2.2
12	H-18	2.4
20	H-18	2.4

*Interactions between vicinal and geminal hydrogens are not included.

[†]Calculated distances for the most stable conformer of **1** (AMPAC 4.5, PM3).

EtOAc-hexane, mp 271–272 (lit. [2] 278°, [9] 246–248°). ¹H NMR spectra for the four known withanolides (**2**–**5**) were in agreement with the reported data [2, 4, 9].

Acknowledgements—We thank Prof. A. T. Hunziker (Universidad Nacional de Córdoba) for identification of the plant. Financial support by CONICET (Argentina) and Fundación Antorchas is gratefully acknowledged.

REFERENCES

1. Glotter, E. (1991) *Nat. Prod. Rep.* 415.
2. Evans, W. C., Grout, R. J. and Mensah, M. L. K. (1984) *Phytochemistry* **23**, 1717.
3. Gupta, M., Bagchi, A. and Ray, A. B. (1991) *J. Nat. Prod.* **54**, 599.
4. Gupta, M., Manickam, M., Sinha, S. C., Sinha-Bagchi, A. and Ray, A. B. (1992) *Phytochemistry* **31**, 2423.
5. Jahromi, M. A. F., Manickam, M., Gupta, M., Oshima, Y., Hatakeyama, S. and Ray, A. B. (1993) *J. Chem. Res. (S)* 234.
6. Shingu, K., Yahara, S. and Nohara, T. (1990) *Chem. Pharm. Bull.* **38**, 3485.
7. Manickam, M., Sinha-Bagchi, A., Sinha, S. C., Gupta, M. and Ray, A. B. (1993) *Phytochemistry* **34**, 868.
8. Manickam, M., Awasthi, S. B., Oshima, Y., Hisamichi, K., Takeshita, M., Sahai, M. and Ray, A. B. (1994) *J. Chem. Res. (S)* 306.
9. Bagchi, A., Neogi, P., Sahai, M., Ray, A. B., Oshima, Y. and Nikino, H. (1984) *Phytochemistry* **23**, 853.
10. Shingu, K., Yahara, S. and Nohara, T. (1994) *Chem. Pharm. Bull.* **42**, 318.