



SESQUITERPENES AND GERANYLGERANYLGLYCEROL FROM THE BROWN ALGAE *TAONIA LACHEANA* AND *TAONIA ATOMARIA* F. *CILIATA*: THEIR CHEMOTAXONOMIC SIGNIFICANCE

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Key Word Index—*Taonia lacheana*; *Taonia atomaria* f. *ciliata*; Dictyotaceae; sesquiterpenes; aromadendranes; geranylgeranylglycerol.

Abstract—Taonia lacheana is a very recent addition to the small Taonia genus. The chemical composition of the new species is characterized by the presence of aromadendrane sesquiterpenes [(+)-spathulenol, (+)-11-epispathulenol and (-)-7,11-dihydroxyaromadendrane] and (-)-(R)-1-geranylgeranylglycerol. A related species from which the last compound had been previously isolated, misidentified as Dilophus fasciola and now recognized to be T. atomaria f. ciliata, has been re-investigated confirming the absence of aromadendrane derivatives. The two Mediterranean Taonia species differ sharply from the Canarian T. atomaria, that is known to elaborate meroditerpenoids.

INTRODUCTION

The small algal genus Taonia includes T. lennebackerae from the Pacific coasts of North America, T. australasica found in the south-eastern Australian waters, and T. atomaria widely distributed in the Atlantic Ocean and in the Mediterranean Sea. A variety of this last species. T. atomaria f. ciliata is common in the Mediterranean and occasionally also found in the Atlantic. Chemical examination of T. atomaria from the Canary Islands has resulted in the isolation of several meroditerpenoids [1-5], exemplified by taondiol (1) [1], while from T. australasica the diterpene taonianone [6] has been isolated. A recent report on the chemistry of T. atomaria from the northern Adriatic Sea [7] describes the identification of the sesquiterpenes 4-cadinene, cadinane-4(14),5-diene, (–)-germacrene D, axenol (–)-cubebol (2) and epi-cubebol, but does not mention any of the meroditerpenoids from the Canarian species.

The recent report from local botanists of a new Taonia species, T. lacheana [8], collected at ca-12 m near Lachea Island (Gulf of Catania) prompted us to study its chemical composition, and in the present paper we report the results obtained. Moreover, we have re-examined a brown alga previously investigated under the name $Dilophus\ fasciola\ [9-12]$, whose classification has been revised into T, $atomaria\ f$. ciliata.

RESULTS AND DISCUSSION

Chromatographic fractionation of the ethylacetate extract from air-dried and ground thalli of *T. lucheana* gave,

along with ubiquitous constituents such as chlorophylls, fats and sterols, two main fractions (A and B), of medium and high polarity, respectively. The main component of fraction A, homogeneous in TLC in different solvent systems, when subjected to ¹³C NMR spectroscopy and GC-mass spectrometry turned out to be a mixture of two isomeric compounds, which were separated by flashchromatography on AgNO₃-impregnated silica gel. The major component, 3, on the basis of its two-dimensional NMR data and $[\alpha]_D$ value, was identified as the sesqui-11β-hydroxy-1β,8α-aromadendrene spathulenol] [13-17]. The isomeric 4 was also a dextrorotatory compound, having ¹H and ¹³C NMR spectra very similar to those of 3 (see Experimental), indicative of the presence of the same carbon skeleton. Moreover, the close similarity of the coupling constants for the cyclopropane protons H-2 and H-4 with the corresponding ones in the spectrum of 3 suggested the identity of the stereochemistry at the bridgehead carbons C-2 and C-4. Thus, the structural differences between 3 and 4 were restricted to the stereochemistry of the chiral centres at C-1, C-4 and C-8. The possibility that 4 could be the known isomer 11β-hydroxy-1β, 8βaromadendrene (allospathulenol) was ruled out on the basis of the differences between the NMR data for 4 and those reported for allospathulenol [18]. Assuming that the stereochemistry of the junction of the five- and seven-membered cycles is identical to that of spathulenol and taking into account the 13C NMR shift of the carbinyl caron (δ 74.7), with respect to the corresponding value for spathulenol ($\delta 81.0$) [17], we suggest for 4 the 828 C. Tringali et al.

structure of $(+)-11\alpha$ -hydroxy- 1β ,8 α -aromadendrene [(+)-11-epispathulenol]. To the best of our knowledge, this is the first reported natural occurrence of this sesquiterpenoid, whose enantiomer has been obtained by synthesis [15].

Flash-chromatography of the more polar fraction (B) from the extract of T. lacheana gave the main component (5) in the pure state. A second subfraction was subjected to acetylation followed by chromatographic fractionation to yield a minor component, 4, and the diacetate 7. Compound 5 was easily identified by spectral analysis as the known (-)-(R)-1-O-geranylgeranylglycerol [9, 19]. The minor component of fraction B, not acetylated under the conditions adopted, showed ¹H and ¹³C NMR resonances strongly resembling those of 3 and 4, the main difference being the absence of the low-field exomethylene signals, which were replaced by the resonances for a second tertiary alcoholic group (1 H: δ 1.16, Me, s; 13 C: δ 74.8, s; 20.0, q). A literature search allowed **6** to be identified as $(-)-7\alpha,11\beta$ -dihydroxy- $1\beta,8\alpha$ -aromadendrane on the basis of IR, NMR and mass spectral data and $[\alpha]_D$ measurement [18, 20-21].

Metabolites 3, 4 and 6 are the first examples of aromadendrane sesquiterpenes isolated from brown algae. Compounds of this class, in particular (+)-spathulenol, occur in many plant genera, while

aromadendrane diols are less frequently encountered [18]. In this connection, mention should be made that the (+)-enantiomer of 6 has been isolated from the soft coral Sinularia mayi [20].

Identification of 5 in the extract of T. lacheana prompted us to reconsider the only other reported occurrence of this compound in a brown alga, D. fasciola [9]. After a botanical re-examination of this seaweed, which led to modification of its classification into T. atomaria f. ciliata, a careful analysis of the ethyl acetate extract was carried out to determine the possible occurrence of aromadendrane sesquiterpenoids. In addition to metabolites identified previously $[\delta$ -cadinene (8), 4,10-epoxymuurolane (9), 1,4-epoxycadinane (10), cubenol (11) and geranylgeranylglycerol (5)] [9, 10, 12], the sesquiterpene 2 [22, 23], identified in an extract of northern Adriatic T. atomaria [15], has been isolated, while no evidence was obtained for the presence of aromadendrane derivatives.

The chemical composition of *T. lacheana*, characterized by aromadendrane sesquiterpenes, differs sharply from that of *T. atomaria* f. *ciliata*, that elaborates sesquiterpenes mostly with the cadinane [10, 12] or germacrane [11] skeleton. However, it is noteworthy that the two species, which occur in the same waters, share the ability to synthesize 5, a metabolite not found until now

in any other alga. The new species also differs chemically from the northern Adriatic *T. atomaria*, again characterized by the presence of sesquiterpenes mostly with cadinane or germacrane skeletons [15]. The chemical composition of the Canarian *T. atomaria* is completely different; several meroditerpenoids have been isolated which are not found in other species belonging to the same genus. The fact that metabolites of this class, for instance taondiol and atomaric acid, have been isolated also from species of the genus *Stypopodium* [24–26], not easily separated morphologically from *Taonia*, suggests the possibility that a botanical misidentification is at the origin of the apparently anomalous chemical composition of the Canarian *T. atomaria*.

EXPERIMENTAL

General. Mps were determined on a Kofler hot stage apparatus. Optical rotations were taken at 25° on a Perkin-Elmer 141 polarimeter in CHCl₃ soln. EIMS spectra were recorded on a Kratos MS 50 instrument, at 70 eV. IR spectra were run on a Perkin-Elmer 684 spectrophotometer in CHCl₃ solution. ¹H NMR spectra were recorded at 200 MHz on an AC-200 Bruker instrument. ¹³C NMR spectra were run at 50.13 MHz on an

AC-200 Bruker instrument equipped with $^1H/^{13}C$ dual probe. All NMR experiments were performed in CDCl₃ with TMS as int. standard. GC-MS analyses were carried out on a Hewlett-Packard gas chromatograph, model 5890, equipped with a MS computerized system, model 5971A, ionization voltage 70 eV, electron multiplier 1700 V, ion source temp. 180°. GC conditions: HP-1 dimethylpolysiloxane capillary column (25 m × 0.2 mm). The oven temp. was held at 60° for 6 min, then programmed from 100° to 250° at 3° min $^{-1}$.

Plant material. Samples of the algae were collected at a depth of, respectively, ca-3 m (T. atomaria f. ciliata) or ca-12 m (T. lacheana) near Lachea Island in April, 1993. Herbarium specimens are deposited at the Dipartimento di Botanica, Università di Catania. Carefully selected and classified thalli of both species were air-dried in the shadow and ground.

Extraction and fractionation. The powdered material of T. lacheana (100 g) was extracted with EtOAc (1.51×3) at room temp. under continuous stirring. Evapn of the solvent under red. pres. afforded a dark-green oil (2.3 g), which was gross-fractionated by open column chromatography (Lichroprep Si-60 Merck, $40-63 \mu m$, increasing amounts of Et₂O in n-hexane as the eluent) to give two main frs (A, B). The less polar frs (A, 180 mg) was

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subjected to flash-chromatography (Lichroprep Si-60, $25-40 \mu m$, eluent i-Pr₂O-hexane from 10 to 60%). The major subfr., homogeneous in TLC in various solvent systems was revealed to be a mixture of two isomeric compounds, 3 and 4 (13C NMR, GC-MS), that were separated by PLC on AgNO3-impregnated silica gel (Lichroprep Si-60, 25-40 μ m; i-Pr₂O-n-hexane, 2:3, as the eluent). Compound 3 (29 mg) was identified as (+)spathulenol basing on 2D NMR analysis (COSY, one bond HETCOR, long-range HETCOR) and $[\alpha]_D$ value $\{+7.2^{\circ} (c \ 1.3 \text{ in CHCl}_{3}); \text{ lit. } +5.7^{\circ} \ [14] \text{ and } -7.5^{\circ} \text{ for }$ (-)-spathulenol [15]. Compound 4 (9 mg) was identified as (+)-11-epispathulenol, mp $102-103^{\circ}$; MS: m/z 220 [M]⁺, 205, 202, 169, 187, 160, 159 (base) 133; IR $v_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3450, 3075, 2920, 1645, 1455, 1375, 1100, 885; $[\alpha]_D = +2.5^{\circ} (c \ 0.5 \text{ in CHCl}_3); [\alpha]_D = +2.0 (c \ 0.7 \text{ in})$ Me_2CO) {lit. -5° in Me_2CO [15]}. ¹³C NMR δ 157.5 (s, C-7); 103.2 (t, C-15); 74.7 (s, C-11); 56.4 (d, C-8 and C-1); 42.3 (*t*, C-10); 38.8 (*t*, C-6); 31.4 (*d*, C-2); 29.7 (*d*, C-4): 29.1 (q, C-12); 28.4 (t, C-9); 28.2 (q, C-13); 25.9 (t, C-5); 19.2 (s, C-3); 16.1 (q, C-14); 1 H NMR: δ 4.74 (1H, bs, H-15a); 4.66 (1H, bs, H-15b); 2.46 (1H, m, H-6a); 2.22 (1H, m, H-8), 2.0-1.0 (7H, overlapped m); 1.23 (3H, s, 12 Me) 1.09 (3H, s, 14 Me); 1.01 (3H, s, 13Me); 0.99 (1H, m, H-5a); 0.62 (1H, ddd, J = 10.5, 9.5, 7 Hz, H-4); 0.32 (1H, t, J = 9.5 Hz, H-2). The more polar fr. (B, 150 mg) was further Lichroprep Si-60 chromatographed (flash) on $(25-40 \mu m)$ eluting with a gradient from 50 to 80% of Et₂O in *n*-hexane and afforded 16 mg of pure (-)-(R)-1-O-geranylgeranylglycerol (5), identity based on MS, ¹H and ¹³C NMR spectra [9, 19] and $[\alpha]_D$ value $\{-2.2^{\circ} (c)\}$ 1.3 in CHCl₃); lit. -2.1° [9], -2.3° [19]}. A second subfr. was exposed overnight to vapours of Ac₂O and pyridine at room temp. and then subjected to flashchromatography (Lichroprep Si-60 25-40 µm, increasing concn of Et₂O in n-hexane as the eluent) to afford 6 mg (-)-7 α ,11 β -dihydroxy-1 α ,8 β -aromadendrane identified via MS, ¹H and ¹³C NMR data [20, 21] and $[\alpha]_D$ value $\{-17.5^{\circ} (c \ 0.2 \text{ in CHCl}_3; \text{ lit. } -17^{\circ} \ [21],$ -22.9° [20]}, and 32 mg of (-)-2(R),3-diacetoxy-1-(R)-O-geranylgeranylglycerol (7) [9], MS: m/z 448 [M]⁺; $MeCO_2$); ¹³C NMR: δ 170.5, 170.2 (s. (s, MeC = CHCH₂O); 135.3, 134.8, 131.1 (s, MeC =):124.3, 124.1, 123.7, 120.3 (d, =CH); 70.3 (t, CH₂O); 67.8 (d, CHOAc); 67.7, 62.9 (t, CH₂O); 39.6 (t, $3 \times \text{CH}_2\text{C} =)$; 26.7, 26.5, 26.3 (t, $CH_2CH =$); 25.6 (q, MeCMe); 20.9, 20.6, 17.6, 16.4 $(q, 2 \times MeCOO \text{ and } 4 \times MeC =);$ ¹H NMR: $\delta 5.30$ (1H, t, J = 7.0 Hz, =CHCH₂O); 5.08 (3H, bt, = CH); 5.18 (1H, m, CHOAc); 4.40–4.10 (4H, m, CHOAc) $2 \times \text{CH}_2\text{O}$); 3.97 (2H, d, J = 7.0 Hz, =CHCH₂O); 2.06 (3H, s, MeCOO); 2.03 (3H, s, MeCOO); 1.98 (12 H, m, $3 \times = CHCH_2CH_2$); 1.65 (6H, bs, $2 \times MeC =$); 1.57 (9H, s, $3 \times MeC =$).

Extraction and fractionation. The powdered material of T. atomaria f. ciliata (180 g) was extracted with EtOAc (21×3) and the extract (dark-green oil, 11.7 g) was fractionated by open-column chromatography (Lichroprep Si-60 40-63 μ m, increasing amounts of Et₂O in n-hexane as the eluent) to afford six main fractions, A-F, in order of increasing polarity. Fr. A (2.6 g) was subjected to

GC-MS analysis and the known sesquiterpene δ cadinene (8) was identified on the basis of GC retention time, computer matching with NBS library and comparison of fragmentation pattern with that reported in lit. [27]. Fr. B (630 mg) was subjected to flash-chromatography on Lichroprep Si-60 (25-40 µm), with EtOAc-nhexane (0.5:99.5) as the eluent. The following compounds were isolated in order of increasing polarity: 4,10-epoxymuurolane (9) (160 mg), 1,4-epoxycadinane (10) (110 mg) and cubenol (11) (60 mg), all of them identified on the basis of reported spectral data (MS, IR, NMR) [10, 12, 28, 29]. The third fr. (C, 1.5 g), was analysed by GC-MS and three of the major constituents were identified as the known fatty acids myristic, palmitoleic and linolenic. Fr. D (1.3 g) was flash-chromatographed (Lichroprep Si-60, 25-40 μm, gradient from 0 to 5% MeOH in CH₂Cl₂). A subfr. of intermediate polarity was subjected to further flash-chromatography (Lichroprep DIOL, 25-40 µm, gradient from 0 to 12% of EtOAc in nhexane) to afford (-)-cubebol (2) (45 mg), identified via MS, IR, ¹H and ¹³C NMR spectra [7, 23] and $[\alpha]_D$ value $\{-46.1^{\circ} (c \ 1.2 \text{ in CHCl}_3); \text{ lit. } -48.3^{\circ} \ [23] \ -67^{\circ} \ [7] \}.$ Fr. E (4.3 g) was not subjected to further analysis since preliminary TLC showed that chlorophylls and sterols were the main constituents. The more polar fr. (F, 1.2 g) was subjected to flash-chromatography (Lichroprep Si-60, 25-40 µm, elution with a gradient of Et₂O in nhexane from 50 to 80%) and gave geranylgeranylglycerol (55 mg), identical in all respects to compound 5 isolated from T. lacheana.

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REFERENCES

- 1. González, A. G. (1971) Tetrahedron Letters 2729.
- González, A. G., Darias, J. and Martín, J. D. (1972) An. Quim. 68, 1187.
- 3. González, A. G., Darias, J., Martín, J. D. and Pascual, C. (1973) Tetrahedron 29, 1605.
- González, A. G., Alvarez, M. A., Martín, J. D., Norte, M., Pérez, C. and Rovirosa, J. (1982) *Tetrahedron* 38, 719.
- González, A. G., Martín, J. D., Pérez, C., Rovirosa, J., Tagle, B. and Clardy, J. (1984) Chem. Letters 1649.
- Murphy, P. T., March, G. and Wells, R. J. (1981) Tetrahedron Letters 22, 1555.
- De Rosa, S., De Giulio, A., Iodice, C. and Zavodnik
 N. (1994) Phytochemistry 37, 1327.
- Cormaci, M., Furnari, G. and Pizzuto, F. (1994) Nord. J. Bot. 14, 463.
- Amico, V., Oriente, G., Piattelli, M., Tringali, C., Fattorusso, E., Magno, S. and Mayol, L. (1977) Experientia 33, 989.

- 10. Amico, V., Oriente, G., Piattelli, M., Tringali, C., Fattorusso, E., Magno, S. and Mayol, L. (1979) Experientia 35, 450.
- 11. Fattorusso, E., Magno, S., Mayol, L., Amico, V., Oriente, G., Piattelli, M. and Tringali, C. (1978) Tetrahedron Letters 43, 4149.
- Fattorusso, E., Magno, S. and Mayol, L. (1979) Gazz. Chim. Ital. 109, 589.
- Bowyer, R. C. and Jefferies, P. R. (1963) Chem. Ind. 1245.
- Juell, S. M., Hansen, R. and Jork, H. (1976) Arch. Pharm. 309, 458.
- Surburg, H. and Mondon, A. (1981) Chem. Ber. 114, 118.
- Inagaki, F. and Abe, A. (1985) J. Chem. Soc. Perkin Trans. II 1773.
- 17. Krebs, H. C., Rakotoarimanga, J. V. and Habermehl, G. G. (1990) Magn. Reson. Chem. 28, 124.
- Gijsen, H. J. M., Wijnberg, J. B. P. A., Stork, G. A. and De Groot, A. (1992) Tetrahedron 48, 2465.
- Zhang, D. and Poulter, C. D. (1993) J. Am. Chem. Soc. 115, 1270.

- Beechan, C. M. and Djerassi, C. (1978) *Tetrahedron* 34, 2503
- Bohlmann, F., Grenz, M., Jakupovic, J., King, R. M. and Robinson, H. (1983) *Phytochemistry* 22, 1213.
- 22. Vonášek, F., Herout, V. and Šorm, F. (1960) Collec. Czech. Chem. Commun. 25, 919.
- 23. Tanaka, A., Tanaka, R., Uda, H. and Yoshikoshi, A. (1972) J. Chem. Soc. Perkin, Trans. I 1721.
- González, A. G., Darias, J., Martín, J. D. and Norte, M. (1974) Tetrahedron Letters, 3951.
- Gerwick, W. H. and Fenical, W. (1981) J. Org. Chem. 46, 22.
- Rovirosa, J., Sepulveda M., Quezada E. and San-Martin, A. (1992) Phytochemistry 31, 2679.
- 27. Eight Peak Index of Mass Spectra, 2nd Edn. Mass Spectrometry Data Centre, AWRE, Reading, U.K. 1974.
- 28. Ohta, Y. and Hirose, Y. (1967) Tetrahedron Letters 2073.
- Nagasampagi, B. A., Yankov, L. and Sukh Dev (1968) Tetrahedron Letters 1913.