

STEROIDAL ALKALOIDS OF *FRITILLARIA MAXIMOWICZII*

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(Received in revised form 22 March 1995)

Key Word Index.—*Fritillaria maximowiczii*; Liliaceae; steroidal alkaloids; 23-isokuroyurinidine; 15,16-*seco*-22 α H,25 β H-solanida-5,14-dien-3 β -ol O-D-glucopyranosyl-(1 \rightarrow 4)- β -D-xylopyranoside; hapepunine 3-O- β -cellobioside.

Abstract.—From the bulbs of *Fritillaria maximowiczii*, in addition to the known jerveratrin alkaloid, kuroyurinidine, three new steroidal alkaloids, 23-isokuroyurinidine, 15,16-*seco*-22 α H,25 β H-solanida-5,14-dien-3 β -ol O- β -D-glucopyranosyl-(1 \rightarrow 4)- β -D-xylopyranoside and hapepunine 3-O- β -cellobioside have been isolated and structurally elucidated.

INTRODUCTION

Fritillaria species are one of the important drug sources often used for depressing coughs, especially in the area of Chain and south-east Asia. In extensive chemical studies, some steroidal alkaloids of the ceveratrum and jerveratrum alkaloid groups [1-5] had been isolated, together with some diterpenes [6]. The bulbs of *F. maximowiczii* (Rinyou-Baimo) grown in north-east China is expected to be a substitute for the bulbs of other *Fritillaria* species, *F. thunbergii* (Setu-Baimo), *cirrhosa*, *unibracteata* and *taipaiensis* (Sen-Baimo), since pharmacological tests of an ethanol extract of this species provided evidence that it also could be used to treat coughs [7]. Therefore, the chemical constituents of this species were investigated.

RESULTS AND DISCUSSION

Fresh bulbs of *F. maximowiczii* were extracted with ethanol to give an extractive, which was partitioned between *n*-butanol and water. The organic layer was shaken with *n*-hexane and 40% methanol, and the aqueous layer was concentrated to give a residue. The residue was subjected to column chromatography on Diaion HP-20 with aqueous methanol to afford several fractions, one of which was subsequently chromatographed on silica gel using chloroform-methanol-water as solvent to provide the four steroidal alkaloids, 1-4.

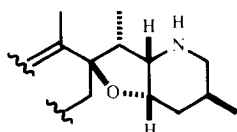
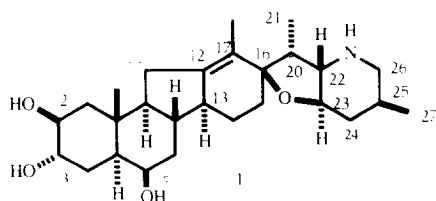
Compound 1 showed in its mass spectrum a $[M]^+$ at m/z 445 and fragment peaks at m/z 125 ($C_8H_{15}N$: base peak) and 114 ($C_6H_{12}O$: 99%) characteristic of the side-chain ring of C-*nor*-D-homo steroids of the jerveratrum group. The 1H and ^{13}C NMR spectra were coincident with those of kuroyurinidine, 2 β ,3 α ,6 β -trihydroxy-5 α -

jerv-12-enine, obtained from *F. camtschatscensis* by Mimaki and Sashida [8].

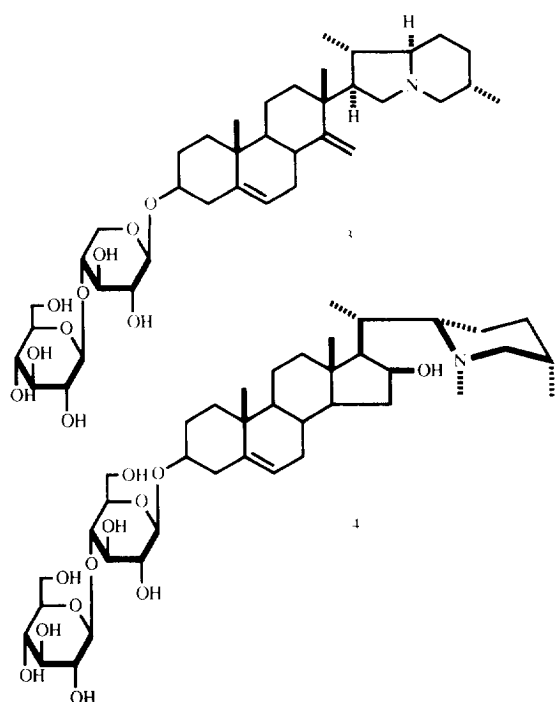
Compound 2 showed a 1H NMR spectrum analogous to that of 1. Therefore, this compound was thought to be a stereoisomer of 1. The 1H - 1H 2D COSY spectrum and the coupling constants revealed the connectivities of the respective protons at 2 α -eq. (*br d*, $J = 2.6$ Hz, at δ 4.58), 3 β -eq (*br d*, $J = 2.9$ Hz at δ 4.66) and 6 α -eq. (*br d*, $J = 2.9$ Hz, at δ 4.25). The steric situation at C-16, -20, -22, -23 and -25 was solved by assignment of the respective signals and coupling constants of the sequence of H₃-21 (*d*, $J = 7.3$ Hz, at δ 1.42), H-20 (*dd*, $J = 7.3, 8.8$ Hz, at δ 2.90), H-22 (*dd*, $J = 8.8, 8.8$ Hz, at δ 3.33), H-23 (*ddd*, $J = 3.7, 8.8, 8.8$ Hz, at δ 3.96), H-26 α -eq. (*dd*, $J = 4.0, 12.6$ Hz, at δ 3.57), H-26 β -ax. (*dd*, $J = 12.6, 12.6$ Hz, at δ 2.84) and the occurrence of NOEs between H₃-18 and H-20, H-20 and H-22 as well as H-22 and H-23. Therefore, the structure of 2 was determined to be 23-isokuroyurinidine. Assignment of the ^{13}C NMR signals for 2 were also attributed without inconsistency.

The mass spectrum of compound 3 showed a $[M]^+$ at m/z 692 (base peak) and fragment ions at m/z 530 [692-hexose] and 398 [530-pentose]. In the 1H NMR spectrum, signals due to two tertiary methyl groups at δ 0.96 and 0.97, two secondary methyl groups at δ 1.09 and 1.18 (each *d*, $J = 6.6$ Hz), one exomethylene group at δ 4.89, 5.06 (each *br s*), one olefinic proton at δ 5.45 (*br d*, $J = 1.1$ Hz) and two anomeric proton signals (each *d*, $J = 7.7$ Hz) were observed. The ^{13}C NMR signals suggested the occurrence of a glucopyranosyl-(1 \rightarrow 4)-xylopyranosyl moiety. Therefore, 3 was hydrolysed with β -glucosidase to give an aglycone, whose electron impact (EI) mass spectrum indicated a $[M + H]^+$ at m/z 398 and a prominent fragment ion at m/z 150 (base peak). The 1H NMR signals were assigned to two tertiary methyl groups (H₃-18 and -19 at δ 0.98 and 1.08), two secondary

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methyl groups (H_3 -21 and -27, each d , $J = 6.6$ Hz, at δ 1.12 and 1.20), one exomethylene group (each 1H, br s , at δ 4.85 and 5.00) and one olefinic proton (H -5, br d , $J = 1.1$ Hz at δ 5.42). The position of the exomethylene group was referred to the ^{13}C NMR data, in which the signals at δ 135.0 (s) and 106.4 (t) could be assigned to C-14 and C-15, respectively, by comparison with those of solanidine. The five proton signals assignable to H_2 -16, H -23 and H_2 -26 adjacent to the nitrogen appeared at δ 2.30–3.35, suggesting that C-16 is not bonded to C-15. On the other hand, the cleaved sugar configurations were determined by GC analysis [9]. Thus, the deduced structure of **3** is 15.16-*seco*-22 α H ,25 β H -solanida-5,14-dien-3 β -ol O - β -D-glucopyranosyl-(1 \rightarrow 4)- β -D-xylopyranoside.

Compound **4** showed in its positive FAB-mass spectrum a $[M + H]^+$ at m/z 754 (75%) and fragment ions at m/z 738 $[M - Me]^+$ (21%) and 722 $[738 - OH]^+$. The 1H NMR spectrum displayed two steroidal angular

methyl groups (δ 0.89 and 1.08), two secondary methyl groups (δ 0.94 and 1.10) and one N -methyl group at δ 2.28, as well as two anomeric protons at δ 5.01 and 5.23 (each 1H, d , $J = 7.7$ Hz). Compound **4** was hydrolysed by β -glucosidase. The EI-mass spectrum of the obtained aglycone showed a $[M]^+$ at m/z 429 and a fragment ion at m/z 112 (base peak) characteristic of an N -methyl-22,26-epimincholestane side-chain. The 1H NMR spectrum exhibited signals due to H -3 (m , δ 3.87), H -16 α (m , δ 4.59) and H -6 (d , $J = 5.1$ Hz, δ 5.43), which were identical to those in hapepunine (22S,25S)- N -methyl-22,26-epimincholest-5-ene-3 β ,16 β -diol [10]. These results and the ^{13}C NMR signals indicated that **4** was hapepunine β -cellobioside.

In contrast to other *Fritillaria* species, *F. maximowiczii* does not contain ceveratrum-type alkaloids, such as peimine and peiminine.

EXPERIMENTAL

1H and ^{13}C NMR were measured with a JEOL JUM-GX 400 NMR spectrometer and chemical shifts are given in δ (ppm) values with TMS as int. standard. FAB-MS were recorded in a glycerol matrix containing NaI. CC was carried out on Diaion HP-20, Sephadex LH-20 (Pharmacia) and Kieselgel 60 (70–230 and 230–400 mesh, Merck).

Extraction and isolation. Fresh bulbs of *F. maximowiczii* Freyn. (10 kg), collected in Heilongjiang province, China, near Mt. Da-Xing-An-Ling in June, were exhaustively extracted with EtOH. The EtOH extract was concd under red. pres. The viscous concentrate was partitioned between H_2O and 1-BuOH, and then the 1-BuOH portion was shaken with n -hexane and MeOH. The MeOH concentrate was chromatographed on Diaion HP-20, eluting with H_2O to MeOH. The 40% MeOH eluate was repeatedly chromatographed on silica gel with $CHCl_3$ -MeOH (50:1–5:1) and on Sephadex LH-20 with 80% MeOH to give compounds **1** (850 mg), **2** (570 mg), **3** (260 mg) and **4** (440 mg).

Kuroyurinidine (1). Amorphous. $[\alpha]_D^{25} = -10.1^\circ$ (MeOH; c 0.40). EI-MS m/z (rel. int.): 445 (13) $[M]^+$, 430 (18), 332 (72), 125 (100), 124 (89), 114 (99), 110 (56), 1H NMR (pyridine- d_5) δ : 0.82 (3H, d , $J = 6.6$ Hz, H_3 -27), 1.08 (3H, d , $J = 7.4$ Hz, H_3 -21), 1.26 (1H, ddd , $J = 11.3$, 11.3, 9.1 Hz, H_{ax} -24), 1.72 (3H, s , H_3 -18), 1.86 (3H, s , H_3 -19), 2.50 (1H, dq , $J = 9.1$, 7.4 Hz, H -20), 2.79 (1H, dd , $J = 9.1$, 9.1 Hz, H -22), 3.09 (1H, ddd , $J = 13.3$, 13.3, 2.2 Hz, H_{ax} -4), 3.14 (1H, dd , $J = 12.3$, 3.6 Hz, H_{eq} -26), 3.40 (1H, ddd , $J = 9.1$, 9.1, 3.7 Hz, H -23), 4.25 (1H, br d , $J = 2.9$ Hz, H -6), 4.58 (1H, br d , $J = 2.6$ Hz, H -2), 4.66 (1H, br d , $J = 2.9$ Hz, H -3). ^{13}C NMR (pyridine- d_5) δ : 43.5 (C-1), 72.0 (C-2), 72.0 (C-3), 30.5 (C-4), 43.7 (C-5), 72.3 (C-6), 39.6 (C-7), 40.3 (C-8), 57.2 (C-9), 36.9 (C-10), 29.3 (C-11), 126.7 (C-12), 142.6 (C-13), 48.8 (C-14), 25.1 (C-15), 32.4 (C-16), 85.5 (C-17), 13.5 (C-18), 17.6 (C-19), 40.8 (C-20), 11.3 (C-21), 66.9 (C-22), 75.5 (C-23), 40.0 (C-24), 31.4 (C-25), 55.0 (C-26), 19.0 (C-27).

23-Isokuroyurinidine (2). Amorphous. $[\alpha]_D^{25} = -14.2^\circ$ (MeOH, c 0.40). EI-MS m/z (rel. int.): 445 (12) $[M]^+$, 430

(28), 332 (48), 125 (100), 124 (76), 114 (55), 110 (46). $^1\text{H NMR}$ (pyridine- d_5) δ : 0.84 (3H, J = 6.6 Hz, H_3 -27), 1.42 (3H, d , J = 7.3 Hz, H_3 -21), 1.64 (3H, s , H_3 -18), 1.86 (3H, s , H_3 -19), 2.84 (1H, dd , J = 12.6, 12.6 Hz, H_{ax} -26), 2.90 (1H, dq , J = 8.8, 7.3 Hz, H -20), 3.10 (1H, ddd , J = 13.3, 13.3, 2.2 Hz, H_{ax} -4), 3.33 (1H, dd , J = 8.8, 8.8 Hz, H -22), 3.57 (1H, dd , J = 12.6, 4.0 Hz, H_{eq} -26), 3.96 (1H, ddd , J = 8.8, 8.8, 3.7 Hz, H -23), 4.25 (1H, $br\ d$, J = 2.9 Hz, H -6), 4.58 (1H, $br\ d$, J = 2.6 Hz, H -2), 4.66 (1H, $br\ d$, J = 2.9 Hz, H -3). NOEs were observed between H_3 -18 and H -20; H -20 and H -22; H -22 and H -23. $^{13}\text{C NMR}$ (pyridine- d_5) δ : 43.4 (C-1), 71.8 (C-2), 71.9 (C-3), 30.4 (C-4), 43.6 (C-5), 72.2 (C-6), 39.9 (C-7), 40.7 (C-8), 57.0 (C-9), 36.8 (C-10), 29.3 (C-11), 126.2 (C-12), 144.0 (C-13), 48.6 (C-14), 24.8 (C-15), 31.8 (C-16), 86.4 (C-17), 13.4 (C-18), 17.5 (C-19), 38.6 (C-20), 12.2 (C-21), 64.3 (C-22), 72.9 (C-23), 37.9 (C-24), 28.4 (C-25), 51.5 (C-26), 18.2 (C-27).

15,16-*seco*-22 α H,25 β H-Solanida-5,14-dien-3 β -ol *O*- β -D-glucopyranosyl-(1 \rightarrow 4)- β -D-xylopyranoside (**3**). Amorphous. $[\alpha]_D^{25}$ = -20.5 (MeOH, c 0.40). Pos. FAB-MS m/z (rel. int.): 692 (100) $[\text{M} + \text{H}]^+$, 530 (10) $[692 - \text{glc}]^+$, 398 (16) $[530 - \text{xy}]^+$, 380 (23) $[530 - 150]^+$. $^1\text{H NMR}$ (pyridine- d_5) δ : 0.96, 0.97 (each, 3H, s , H -18, 19), 1.09, 1.18 (each, 3H, d , J = 6.6 Hz, H_3 -21, -27), 4.89, 5.06 (each, $br\ s$, H_2 -15), 4.91, 5.16 (each, 1H, d , J = 7.7 Hz, H -2, xyl, H -1), 5.45 (1H, $br\ d$, J = 1.1 Hz, H -6). $^{13}\text{C NMR}$ (pyridine- d_5) δ : 37.1 (C-1), 30.2 (C-2), 78.2 (C-3), 39.0 (C-4), 141.0 (C-5), 121.0 (C-6), 35.6 (C-7), 33.7 (C-8), 50.9 (C-9), 36.3 (C-10), 20.5 (C-11), 39.0 (C-12), 42.3 (C-13), 135.0 (C-14), 106.7 (C-15), 56.3 (C-16), 51.5 (C-17), 17.6 (C-18), 19.1 (C-19), 38.2 (C-20), 19.0 (C-21), 69.3 (C-22), 30.2 (C-23), 30.8 (C-24), 27.7 (C-25), 58.4 (C-26), 19.9 (C-27). xyl: 103.2 (C-1), 74.7 (C-2), 76.5 (C-3), 78.9 (C-4), 64.7 (C-5). glc: 103.7 (C-1), 74.4 (C-2), 78.2 (C-3), 71.7 (C-4), 78.2 (C-5), 62.7 (C-6).

15,16-*seco*-22 α H,25 β H-Solanida-5,14-dien-3 β -ol. A suspension of **3** (28 mg) and crude hesperidinase in NaOAc-HOAc was incubated for 2 days at 37°. The reaction mixt. was evapd under red. pres. to give a residue, which was chromatographed over silica gel with *n*-hexane-Me₂CO (4:1) to afford the aglycone (12 mg). EI-MS m/z (rel. int.): 398 (2) $[\text{M} + \text{H}]^+$, 397 (3) $[\text{M}]^+$, 396 (4) $[\text{M} - \text{H}]^+$, 246 (1) $[396 - 150]^+$, 204 (2), 150 (100), 136 (26), 98 (5). $^1\text{H NMR}$ (pyridine- d_5) δ : 0.98, 1.08 (3H, each, s , H_3 -18, 19), 1.12, 1.20 (each, 3H, d , J = 6.6 Hz, H_3 -21, -27), 4.85, 5.00 (1H, each, $br\ s$, H_2 -15), 5.42 (1H, $br\ d$, J = 1.1 Hz, H -6). $^{13}\text{C NMR}$ (pyridine- d_5) δ : 37.4 (C-1), 32.3 (C-2), 71.1 (C-3), 43.1 (C-4), 141.0 (C-5), 121.2 (C-6), 35.6 (C-7), 34.0 (C-8), 50.8 (C-9), 36.3 (C-10), 20.7 (C-11), 38.2 (C-12), 42.5 (C-13), 135.0 (C-14), 106.4 (C-15), 56.9 (C-16), 51.2 (C-17), 18.4 (C-18), 19.1 (C-19), 37.1 (C-20), 18.7 (C-21), 68.3 (C-22), 29.9 (C-23), 31.0 (C-24), 28.0 (C-25), 58.7 (C-26), 19.9 (C-27). The insol. part from the above solvent was converted into the corresponding TMSi ethers of methyl 2-(polyhydroxyalkyl)-thiazoli-

dine-4(*R*)-carboxylates followed by GC analysis. The sugar moiety was composed of D-glucose and D-xylose.

Hapepunine 3-*O*- β -cellobioside (**4**). Amorphous. $[\alpha]_D^{25}$ = -29.1 (MeOH, c 0.43). Pos. FAB-MS m/z (rel. int.): 754 (75) $[\text{M} + \text{H}]^+$, 738 (21) $[\text{M} - \text{Me}]^+$, 722 (100) $[738 - \text{OH}]^+$. $^1\text{H NMR}$ (pyridine- d_5) δ : 0.89, 1.08 (3H each, H_3 -18, -19), 0.94, 1.10 (each, 3H, d , J = 7.0 Hz, H_3 -21, -27), 2.28 (3H, s , *N*-Me), 5.01, 5.23 (each, 1H, d , J = 7.7 Hz, 2 \times glc H -1). $^{13}\text{C NMR}$ (pyridine- d_5) δ : 37.3 (C-1), 30.7 (C-2), 78.4 (C-3), 38.8 (C-4), 141.0 (C-5), 121.7 (C-6), 32.0 (C-7), 31.6 (C-8), 50.2 (C-9), 36.8 (C-10), 21.1 (C-11), 40.5 (C-12), 43.1 (C-13), 54.3 (C-14), 29.9 (C-15), 71.4 (C-16), 50.2 (C-17), 13.3 (C-18), 17.5 (C-19), 40.4 (C-20), 19.0 (C-21), 58.8 (C-22), 30.1 (C-23), 37.2 (C-24), 31.6 (C-25), 62.4 (C-26), 19.3 (C-27), 43.9 (*N*-Me), glc: 102.2 (C-1), 74.8 (C-2), 76.8 (C-3), 81.3 (C-4), 76.4 (C-5), 62.4 (C-6), glc: 104.9 (C-1), 74.8 (C-2), 78.2 (C-3), 71.4 (C-4), 78.2 (C-5), 62.1 (C-6).

Hapepunine. Compound **4** (35 mg) was dissolved in a small amount of DMSO (0.8 ml) and suspended in NaOAc-HOAc buffer and β -glucosidase (from almonds). The mixt. was incubated for 3 days at 37° and was then evapd under red. pres. The residue was chromatographed over silica gel with *n*-hexane-Me₂CO (3:1) to provide the aglycone (8 mg). EI-MS m/z (rel. int.): 429 (1) $[\text{M}]^+$, 428 (2) $[\text{M} - \text{H}]^+$, 414 (2) $[\text{M} - \text{Me}]^+$, 397 (5) $[414 - \text{OH}]^+$, 112 (100). $^1\text{H NMR}$ (pyridine- d_5) δ : 1.07, 1.10 (each, 3H, s , H_3 -18, -19), 1.07, 1.08 (each, 3H, d , J = 7.0 Hz, H_3 -21, -27), 2.28 (3H, s , *N*-Me), 3.87 (1H, m , H -3 α), 4.59 (1H, m , H -16 α), 5.43 (1H, d , J = 5.1 Hz, H -6). The sugar was identified using the method described above.

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