

THE PRINCIPAL ALKALOID OF *SENECIO SCHWEINFURTHII*

M. H. BENN,\* SIMON MATHENGE,† R. M. MUNAVU‡ and S. O. WERE‡

Chemistry Department, The University of Calgary, Calgary, Alberta, Canada, T2N 1N4; †Botany Department, University of Nairobi, P.O. Box 30197, Nairobi, Kenya; ‡Department of Chemistry and Biochemistry, Egerton University, P.O. Box 536, Njoro, Kenya

(Received 10 April 1995)

**Key Word Index**—*Senecio schweinfurthii*; Compositae; pyrrolizidine alkaloids; 7 $\beta$ -hydroxy-1-methylene-8 $\alpha$ -pyrrolizidine *N*-oxide, 7 $\beta$ -hydroxy-1-methylene-8 $\alpha$ -pyrrolizidine.

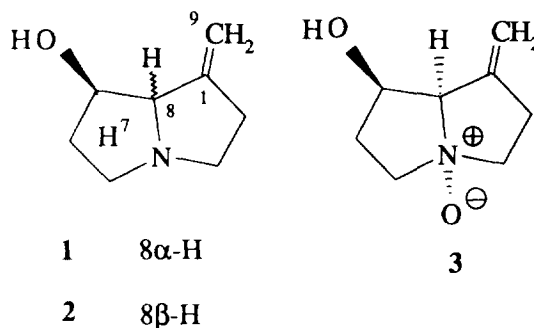
**Abstract**—Air-dried epigeal parts of *Senecio schweinfurthii*, collected from two sites in Kenya, yielded 7 $\beta$ -hydroxy-1-methylene-8 $\alpha$ -pyrrolizidine *N*-oxide, as the predominant alkaloid.

## INTRODUCTION

In continuation of our studies of east African species of *Senecio* [1], we have examined the alkaloids of *S. schweinfurthii* O. Hoffm. (a plant which has extensive synonymy: *S. sera* Schweinf., *S. massaiensis* Muschl., *S. melanophyllus* Muschl., *S. theodori* K. Afzel., *S. roberti-friesii* K. Afzel., *S. roberti-friesii* var. *subcanescens* K. Afzel., *S. roberti-friesii* var. *kilimanjaricus* K. Afzel., and *S. sattimae* K. Afzel. [1]). Our material was first collected on Mt. Kenya and later in the Aberdare Mountains of Kenya.

## RESULTS AND DISCUSSION

Conventional processing of an ethanolic extract of air-dried and pulverized plant material from Mt. Kenya yielded an aqueous extract which gave a strong positive reaction when tested with Mayer's reagent. Most species of *Senecio* contain pyrrolizidine alkaloids [2-4] and these are usually preponderantly present as *N*-oxides [2-6]. Accordingly, the extract was partitioned between  $\text{CHCl}_3$  and dilute aqueous sulphuric acid and a portion of the aqueous extract stirred with Zn dust. After basification and extraction with  $\text{CHCl}_3$ , this yielded alkaloidal material (ca 0.1%) as an oil which crystallized when it was stored at 0°. Both GC-mass spectrometry and NMR data for this product indicated that it was essentially a single substance which was recognized from its spectroscopic properties to be a 7-hydroxy-1-methylenepyrrolizidine. Both 7 $\beta$ -hydroxy-1-methylene-8 $\alpha$ - and 8 $\beta$ -pyrrolizidines (1 and 2, respectively) have been described by Culvenor and Smith who isolated them from *Crotalaria goriensis* [7]. Distillation of our material gave a product whose  $[\alpha]_D$  of  $-153$  agreed with that re-



ported for the 8 $\alpha$ -isomer ( $-150^\circ$ ) and not the 8 $\beta$ -isomer ( $+36^\circ$ ). Additionally, our material had spectroscopic properties identical with those of 1 which we had recently prepared from retronecine [8]. Thus, the *S. schweinfurthii* base is 1.

Culvenor and Smith showed that there was little difference in the amounts of alkaloid isolated from *C. goriensis* seeds with or without reductive work-up, i.e. that there was little *N*-oxide present. However in our case, omission of the reductive step resulted in a large drop in the yield of 1. We therefore examined the unreduced extract for the presence of *N*-oxide and isolated material whose NMR-mass spectrometry data indicated that was 3. Proof for this identification was provided by the oxidation of 1 to 3 which was identical to our isolate. Reductive processing of the *S. schweinfurthii* from our second collection site in the Aberdare Mountains revealed that it too contained 1 as the predominant base.

It is unusual, but preceded [1-4], to find a single pyrrolizidine base in an overwhelming amount; normally several such bases are present in comparable proportions. We also note that the 7-*O*-angelyl derivative of 3 was recently discovered in *S. chrysocoma*, another African species [9]. Together with our findings these appear

\*Author to whom correspondence should be addressed.

to constitute the only reports of the occurrence of methylenepyrrolizidines in the genus *Senecio*.

#### EXPERIMENTAL

**Plant material.** A ref. specimen has been deposited in the Herbarium of the University of Calgary.

**Extraction of alkaloids.** Air-dried and pulverized plant material (810 g) was extracted in a Waring blender with 95% EtOH (4 × 2 l). The extracts were then concd under red. pres. to a syrup (128 g). This was partitioned between aq. H<sub>2</sub>SO<sub>4</sub> (1 M, 200 ml) and CHCl<sub>3</sub> (350 ml) with centrifugation to separate the phases. The CHCl<sub>3</sub> layer was then re-extracted with aq. H<sub>2</sub>SO<sub>4</sub> (2 × 75 ml) and the comb. aq. extracts filtered (Celite) and the filtrate (ca 300 ml), which gave a strongly positive reaction when tested with Mayer's reagent [10], was divided into two portions which were processed as follows. (a) *Reductive-ly.* To a portion (ca 200 ml) of the aq. acidic soln was added Zn dust (5 g). The mixt. was stirred overnight at room temp. and then filtered (Celite). The filtrate was brought to pH ca 10 and extracted with CHCl<sub>3</sub> (4 × 75 ml), then EtOAc (1 × 100 ml), and finally with 1-BuOH (100 ml). These organic extracts were separately dried (MgSO<sub>4</sub>) and the solvents removed under red. pres. The CHCl<sub>3</sub> extracts yielded an oil (1.4 g) which was redissolved in aq. H<sub>2</sub>SO<sub>4</sub> (1 M, 100 ml) and this soln was washed with CHCl<sub>3</sub> (2 × 25 ml) before being reacidified to pH ca 10 with NH<sub>4</sub>OH and then extracted with CHCl<sub>3</sub> (4 × 50 ml). The comb., dried (MgSO<sub>4</sub>) CHCl<sub>3</sub> extracts were evapd to give a pale brown oil (610 mg) which crystallized when stored at 0 °C. TLC (CHCl<sub>3</sub>-MeOH-NH<sub>4</sub>OH, 4:1:0.1 and 5:1:0.1), as well as GC-MS, indicated that this material was essentially a single substance. Its <sup>1</sup>H and <sup>13</sup>C NMR spectra were essentially identical with those of the dist. material. Bulb-to-bulb distillation (80 °C at 1 mm) of this oil gave **1** as a colourless oil which crystallized when cooled to 0 °C. Mp 32–33 °C. [α]<sub>D</sub><sup>20</sup> –153 (c 0.5, EtOH), lit. [7] for **1**, mp 35–36 °C, [α]<sub>D</sub><sup>20</sup> –150° (EtOH). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ref. CHCl<sub>3</sub>) δ<sub>H</sub> 7.27 (1H, *q*, *J* = 5.16 and 4.86 (each 1H, *dt*, *J* = 2.1 Hz, H-9A and 9B), 3.89 (1H, *br s*, H-8), 4.14 (1H, *dt*, *J* = 4 and 2 Hz, H-7), 3.07 and 2.65 (each 1H, *m*, H-3A and 3B), 3.12 and 2.78 (each 1H, *m*, H-5A and 5B), 2.51 (2H, *m*, H-2) and 2.01 and 1.92 (each 1H, *m*, H-6A and 6B). <sup>13</sup>C NMR (100 MHz, CHCl<sub>3</sub>, ref. 77.0) 149.0 (C-1), 107.4 (C-9), 73.9 (C-8), 72.2 (C-7), 55.0 (C-3), 52.9 (C-5), 35.6 (C-6) and 34.7 (C-2); these assignments were established by HXCORR spectra. (b) *Without reduction.* A portion of the aq. H<sub>2</sub>SO<sub>4</sub> extract (50 ml) was basified first with Na<sub>2</sub>CO<sub>3</sub>, then NH<sub>4</sub>OH to pH ca 10 and then extracted with CHCl<sub>3</sub> (4 × 50 ml) and then EtOAc (3 × 30 ml). These extracts were dried (MgSO<sub>4</sub>) and the solvents removed under red. pres. to leave a small residue which was redissolved in aq. H<sub>2</sub>SO<sub>4</sub>, washed with CHCl<sub>3</sub> and then recovered by basification with NH<sub>4</sub>OH to pH ca 10 and repeated extraction with CHCl<sub>3</sub>. Removal of solvent from the comb., dried (MgSO<sub>4</sub>) CHCl<sub>3</sub> extracts yielded a brownish oil (58 mg) whose NMR spectra were the same as those of the base obtained by reductive process-

ing. (c) The remainder of the aq. H<sub>2</sub>SO<sub>4</sub> extract (50 ml) was basified as before to pH ca 10 and then extracted with 1-BuOH (4 × 50 ml). The comb. BuOH extracts were dried (Na<sub>2</sub>SO<sub>4</sub>) and evapd *in vacuo* to a reddish-brown syrup. This was dild with H<sub>2</sub>O and again evapd under red. pres., to remove traces of BuOH. The residual glass was largely insol. in CDCl<sub>3</sub> but sol. in MeOH. A <sup>1</sup>H NMR (CD<sub>3</sub>OD) revealed, besides signals apparently due to glycosidic materials, relatively intense signals which were attributable to the *exo*-methylene functionality (δ<sub>H</sub> 5.05 and 4.9 each 1H, *br s*), as well as others corresponding to H-7 and H-8 of a pyrrolizidine alkaloid. A portion (88 mg) of the BuOH extract was subjected to vacuum short CC on silica gel 60 (3.3 cm × 3.5 cm) using 1-BuOH-EtOH-H<sub>2</sub>O-NH<sub>4</sub>OH (120:25:15:10) as eluant and collecting frs of 10 ml. After repeated re-evapn under red. pres. with H<sub>2</sub>O added to remove BuOH, the individual frs were examined by <sup>1</sup>H NMR: frs 7 and 8 corresponded to pure **3**. These were comb. to afford the *N*-oxide as a near-colourless oil (20 mg). MS (FAB) *m/z* 156 [*M* + 1]<sup>+</sup>. NMR δ<sub>H</sub> (MeOH-*d*<sub>4</sub>, δ<sub>H</sub> 3.31 as ref.) 5.29 (1H, *q*, *J* = 2 Hz, H-9A), 5.12 (1H, *q*, *J* = 2 Hz, H-9B), 4.54 (1H, *dt*, *J* = 5.4 and 2 Hz, H-7), 4.32 (1H, *br dt*, *J* = 5.4 and 2 Hz, H-8), 3.97 (1H, *ddd*, *J* = 11.9, 9.7 and 7.1 Hz, H-5A), 3.91 (1H, *dt*, *J* = 11.2 and 8.2 Hz, H-3A), 3.64 (1H, *m*, H-5B), 3.59 (1H, *m*, H-3B), 2.99 (1H, *m*, H-2A), 2.80 (1H, *m*, H-2B), 2.73 (1H, *m*, H-6A) and 2.14 (1H, *m*, H-6B). δ<sub>C</sub> (MeOH-*d*<sub>4</sub>, δ<sub>C</sub> 49.0 as ref.) 142.2 (*s*, C-1), 111.9 (*t*, C-9), 92.0 (*d*, C-8), 72.3 (*d*, C-7), 70.0 (*t*, C-3), 68.5 (*t*, C-5), 34.9 (*t*, C-6), 33.4 (*t*, C-2); δ<sub>C</sub> (D<sub>2</sub>O, TSP-*d*<sub>4</sub> δ<sub>C</sub> 0 as ref.) 142.9 (*s*, C-1), 115.1 (*t*, C-9), 92.7 (*d*, C-8), 73.6 (*d*, C-7), 71.2 (*t*, C-3), 69.4 (*t*, C-5), 35.6 (*t*, C-6), 34.5 (*t*, C-2).

**Preparation of 3 by oxidation of 1.** To a soln of **1** (10 mg) in D<sub>2</sub>O (0.8 ml) was added 30% H<sub>2</sub>O<sub>2</sub> (0.1 ml) and the reaction monitored by <sup>13</sup>C NMR. After 2 hr, the spectrum was identical with that of the product isolated from the unreduced *S. schweinfurthii* extract. The reaction mixt. was evapd to dryness under red. pres., taken up in MeOH and again evapd to leave **3** as a colourless glass (10 mg) with spectroscopic properties identical to those of the natural product.

**Acknowledgements**—The authors thank the Natural Sciences and Engineering Research Council of Canada for a grant in aid of research (to MHB) which helped support this work.

#### REFERENCES

1. Jeffrey, C. (1968) *Kew Bull.* **41**, 873.
2. Rizk, A.-F. M. (1990) *Naturally Occurring Pyrrolizidine Alkaloids*. CRC Press, Boca Raton, FL.
3. Mattocks, A. R. (1986) *Chemistry and Toxicology of Pyrrolizidine Alkaloids*. Academic Press, London.
4. Bull, L. B., Culvenor, C. C. J. and Dick, A. T. (1968) *The Pyrrolizidine Alkaloids: Their Chemistry, Pathogenicity and other Biological Properties*. North Holland, Amsterdam.

5. Hartmann, T. and Zimmer, M. (1986) *J. Plant. Physiol.* **122**, 67.
6. Hartmann, T., Ehmke, A., Eilert, U., von Borstel, K. and Theuring, C. (1989) *Planta* **177**, 98.
7. Culvenor, C. C. J. and Smith, L. W. (1961) *Aust. J. Chem.* **14**, 284.
8. Hanselmann, R. and Benn, M. (1993) *Tetrahedron Lett.* **34**, 3511.
9. Liddell, J. R. and Logie, C. G. (1993) *Phytochemistry* **34**, 1198.
10. Cordell, G. A. (1981) *Introduction to Alkaloids: A Biogenic Approach*. John Wiley and Sons, New York.