

DIASTEREO- AND ENANTIO-SELECTIVE REDUCTION OF ETHYL 2-METHYL-3-OXOBUTANOATE BY PLANT CELL CULTURES

KAORU NAKAMURA,* HIROCHIKA MIYOSHI,† TOMOAKI SUGIYAMA† and HIROKI HAMADA*†

*Institute for Chemical Research, Kyoto University, Uji, Kyoto 611, Japan; †Department of Applied Science, Okayama University of Science, 1-1 Ridai-cho, Okayama 700, Japan

(Received in revised form 10 May 1995)

Key Word Index Biotransformation; reduction; *Marchantia polymorpha*; *Glycine max*; hydroxy ester; diastereoselective; reduction; enantioselective.

Abstract—Ethyl 2-methyl-3-oxobutanoate was reduced diastereo- and enantio-selectively to the corresponding antiand syn-(S)-hydroxyester by callus of *Marchantia polymorpha* and *Glycine max*, respectively.

INTRODUCTION

Diastereo- and enantio-selective synthesis of 2-alkyl-3-hydroxyalkanoates is a challenging problem in the field of synthetic organic chemistry [1, 2]. Chemical methods have been developed which involve aldol condensation of chiral compounds [1] and reduction of α -alkyl- β -keto esters with BINAP-Ru complexes [2]. The latter method is effective for 2-alkylamino derivatives, but is not effective for 2-alkyl derivatives. Biotransformations such as microbial reductions in aqueous media [3, 4], and microbial reduction in organic media [5] are also effective. Herein, we report on a novel biotransformation for the reduction of ethyl 2-alkyl-3-hydroxyalkanoate by plant cell suspension cultures [(for review of plant cell biotransformations see refs [6-8]).

RESULTS AND DISCUSSION

Due to the β -keto ester structure, racemization of the enantiomers of ethyl 2-methyl-3-oxobutanoate (1), the starting material, is very rapid and the reduction product, 2-methyl-3-hydroxybutanoate (2), is hardly racemized [3]. The reduction would form one stereoisomer selectively out of the four possible stereoisomeric hydroxyl esters if the reduction proceeded diastereo- and enantioselectively (Scheme 1).

Several plant cell cultures were tested for the ability to reduce compound 1, and the enantioselectivity and diastereoselectivity of the reduction were measured (Table 1).

The yield of the hydroxyl ester depends on the species of plant. Thus, Glycine max, Catharanthus roseus and Marchantia polymorpha afforded the hydroxyl ester in high yields while Nicotiana tabacum, Pharaenopsis sp. and Phytolacca americana gave only moderate yields. The diastereoselectivity of the reduction was species dependent. Callus from C. roseus afforded a low anti-

selectivity (anti:syn = 61:39). Similar results were obtained with P. americana (anti:syn = 13:7) and Pharaenopsis sp. (anti:syn = 18:7). Although the diastereoselectivities of these calli were not satisfactory, we found that the anti-isomer could be prepared selectively by using callus of M. polymorpha as the reducing biocatalyst (anti:syn = 48.2:1.7). It is well known that BINAP-Ru catalyst exhibits high enantioselectivity when reducing 1, whereas BINAP catalyst shows no diastereoselectivity towards 2-alkyl-3-oxobutanoate [2]. Thus, biocatalysts are superior to chemical catalysts in terms of diastereoselectivity. Several calli afforded syn-selectivity. Thus, the callus from N. tabacum (anti:syn = 19:31) showed low syn-selectivity while that of G. max (anti:syn = 2:23) gave excellent synselectivity.

The enantioselectivity of the reduction with M. polymorpha was excellent; the stereochemistry of the antiisomer (2S,3S) was 3S and the enantiomeric excess (ee) was over 99%. The ee of the syn-isomer (2R,3S) obtained from G. max was also excellent (97%) (Scheme 2). Thus,

1 2

Scheme 1. Reduction products of 1.

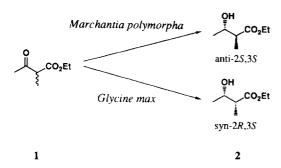
1420

Table 1. Reduction of ethyl 2-methyl-3-oxobutanoate (1) by plant cell calli

Callus	Yield of 2 (%)	Anti(ee %):Syn(ee %)
Marchantia polymorpha	88	96(99*):4(13†)
Pharaenopsis sp	30	72(93*):28
Phytolacca americana	41	65:35
Catharanthus roseus	90	61(99*):39(84*)
Nicotiana tabacum (Bright yellow)	24	56:44
Nicotiana tabacum (Mild cure)	36	38:62
Glycine max	100	8(>95*):92(97*)

^{*}The 3S-hydroxyl ester is obtained preferentially.

[†] The 3R-hydroxyl ester is obtained preferentially.



Scheme 2. Biotransformation of compound 1.

only one stereoisomer out of four possible stereoisomers was obtained by using reduction with plant cell cultures.

EXPERIMENTAL

Analytical. Chemical yields and diastereomeric ratios were determined on a capillary GC column (HR-20M, 0.25 mm × 25 m, 120°, He, 2 ml min⁻¹) using tetradecane as an int. standard. The absolute configurations and enantiomeric excesses were determined on a chiral capillary GC column (Chiraldex G-TA, 0.25 mm × 30 m, 85°, He 2 ml min⁻¹). The standard sample of 2 was prepd according to ref. [3].

Cultivation of cells. Callus tissues were cultured on agarose with the following media: C. roseus, Schenk and Hildebrandt medium [9]; G. max and N. tabacum (Bright yellow and Mild cure), Murashige and Skoog's medium [10]; M. polymorpha, MSK-II medium [11]; Pharaenopsis, VWII medium [12].

Substrate. Compound 1 was purchased from Tokyo Kasei and used without further purification.

Biotransformation of 1. Callus (10 g) was suspended in 30 ml K-Pi buffer (pH 7.0, 0.1 M) and 0.1 mmol 1 was added to the resulting mixt. After 24 hr, 2.5 mg tetradecane (GC int. standard) in 0.5 ml $\rm Et_2O$ was added to the reaction mixt., which was then extracted with EtOAc (10 ml \times 3). Chemical yields and diastereomeric ratios were determined by GC.

REFERENCES

- Masamune, S., Choy, F. A., Kerdesky, D. and Imperiali, B. (1981) J. Am. Chem. Soc. 103, 1566.
- Noyori, R., Ohkuma, T., Kitamura, M., Takaya, H., Sayo, N., Kumobayashi, H. and Akutagawa, S. (1987) J. Am. Chem. Soc. 109, 5856.
- 3. Nakamura, K., Miyai, T., Nagar, A., Oka, S. and Ohno, A. (1989) *Bull. Chem. Soc. Jpn* **62**, 1179.
- Nakamura, K., Kawai, Y., Miyai, T. and Ohno, A. (1990) Tetrahedron Letters 31, 3631.
- 5. Nakamura, K., Takano, S. and Ohno, A. (1993) Tetrahedron Letters 34, 6068.
- Suga, T. and Hirata, T. (1990) Phytochemistry 29, 2393.
- 7. Gotoh, S., Aoki, M., Iwaeda, T., Izumi, S. and Hirata, T. (1994) Chem. Letters 1519.
- 8. Hirata, T., Izumi, S., Akita, K., Yoshida, H. and Gotoh, S. (1993) Tetrahedron: Asymm. 4, 1645.
- Schenk, R. U. and Hildebrandt, A. C. (1972) Can. J. Botany 50, 199.
- Murashige, T. and Skoog, F. (1962) *Physiol. Plant.* 15, 493.
- Katoh, K., Ishikawa, M., Miyake, K., Ohta, Y., Hirose, Y. and Iwamura, T. (1989) Physiol. Plant. 49, 241.
- Vacin E. F. and Went, F. W. (1949) Bot. Gaz. 110, 605.