

DITERPENOIDS FROM *CALCEOLARIA DENTATA**

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(Received 4 May 1995)

IN HONOUR OF PROFESSOR ANTONIO G. GONZALEZ

Key Word Index—*Calceolaria dentata*; Scrophulariaceae; diterpenes.

Abstract—Five diterpenoids were isolated from the aerial parts of *Calceolaria dentata*. Their structures were elucidated by spectroscopic methods. Three of them are esterified with isovaleric acid.

INTRODUCTION

The large genus *Calceolaria* has been the subject of several chemical studies [1–3]. In the course of our current research on the chemistry of this genus, we have now investigated the aerial parts *Calceolaria dentata* and shown them to contain the new thyrsoflorane derivatives 1–3, the new stemarane derivatives 4 and 5, and the known 17-hydroxy-9-epi-ent-isopimaradiene [4].

RESULTS AND DISCUSSION

Compound 1 was purified and characterized as its methyl ester derivative 1a ($C_{26}H_{40}O_6$, $[M]^+$ at m/z 448). The 1H NMR spectrum of 1a showed the presence of four quaternary methyl groups (δ 0.81, 0.85, 0.93 and 1.02), a singlet at δ 3.38 corresponding to the methylene group of a malonate moiety, a singlet at δ 3.76 (3H, s, OMe) and a singlet at δ 2.03 corresponding to an acetoxyl group. Based on the molecular formula and the 1H NMR and ^{13}C NMR data, it was concluded that 1a was a tetracyclic diterpene of the thyrsoflorane-type with a malonate and an acetyl group.

Treatment of 1a with $LiAlH_4$ in dry diethyl ether gave 3b, whose spectral and physical data were in full accord with those of thyrsoflorin C [5], while treatment of 1a with K_2CO_3 -methanol gave methyl thyrsoflorin B (1b). Therefore, 1a was shown to be methyl thyrsoflorin B acetate [5].

The spectral data for 2, together with the molecular formula $C_{27}H_{44}O_4$ (M^+ at m/z 432), suggested that 2 could be a diterpenoid of the same type as 1. Comparison of the 1H NMR spectrum of 2 with that of 1a showed minor differences in the skeletal proton signals. In particular, the malonyl proton singlet was replaced by

a doublet at δ 0.95 corresponding to two methyl groups and a doublet corresponding to a methylene of an acyl moiety. The ester substituent was assigned to an isovaleryl group based on the typical 1H NMR spectrum (see Experimental), together with the loss of 57 (C_4H_9) and 102 ($C_5H_{10}O_2$) amu in the mass spectra. The ^{13}C NMR spectrum confirmed the presence of this residue (Table 1).

Compound 2 on treatment with K_2CO_3 -methanol gave 2a, whose spectral data were very similar to those of 1b, except that the signals due to the malonate moiety were replaced by those of an isovaleroyl group. Furthermore, treatment of 2 with $LiAlH_4$ in dry ether gave 3b. Therefore, 2 was shown to be 13-isovaleroyl-7-acetoxy-thyrsoflorane.

Compound 3 was purified and characterized as its methyl ester derivative 3a ($C_{29}H_{46}O_6$, $[M]^+$ at m/z 490). Its 1H NMR and ^{13}C NMR spectra were very similar to those of 2 for the skeletal protons, but the acetoxyl ester signals were replaced by those of a malonate ester group. Placement of the malonate group at C-7 was established by means of the ^{13}C NMR data. Therefore, compound 3 was shown to be 13-isovaleryl-7-malonyloxy-thyrsoflorane.

The methyl ester 4a of the new diterpenoid 4 was assigned the molecular formula $C_{24}H_{36}O_4$ ($[M]^+$ at m/z 388). It showed IR absorptions for an ester carbonyl at 1710 and 1730 cm^{-1} and an exomethylene group at 1630 and 880 cm^{-1} . Its 1H NMR spectrum showed a singlet at δ 3.38 characteristic of a malonate group attached to a carbocyclic secondary carbon at δ 4.8 (1H, dt, $J = 5.7, 10.8$ Hz, H-7). The ^{13}C NMR and DEPT spectra showed the presence of three methyl, nine methylene, four methine and four quaternary groups (Table 1) besides the signals of the malonate moiety. From these data, we deduced that 4 was a tetracyclic diterpene with a stemodane or stemarane skeleton, with a malonate group at C-7. However, the presence of three quaternary methyl groups and the malonate attached to a secondary carbon and the exocyclic methylene are only compatible with the stemarane skeleton. This assumption was also supported by the ^{13}C NMR data [6–8].

*Part 17 in the series "Diterpenoids from *Calceolaria* Species. For part 16 see ref. [1].

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Table 1. ^{13}C NMR spectra data for compounds **1a**, **2**, **2a**, **3a**, **3b**, **4a**, **4b**, **5** and **5a**

C	1a	2	2a	3a	3b	4a	4b	5	5a
1	32.3	32.4	32.4	32.4	32.5	32.1	32.3	47.1	51.5
2	18.6	18.8	18.7	18.8	18.7	18.7	18.7	69.7	65.6
3	41.9	42.0	42.0	42.1	42.0	42.0	42.0	38.4	42.4
4	33.2	33.3	33.1	33.3	33.1	33.2	33.1	34.7	34.9
5	45.9	46.0	46.2	46.1	46.2	36.4	46.6	48.2	48.2
6	27.8	28.1	30.9	28.0	30.7	27.9	30.9	21.7	21.8
7	77.7	76.0	73.7	77.0	73.7	78.5	74.3	39.9	39.9
8	41.7	41.9	45.1	41.8	45.2	43.8	47.1	39.4	39.3
9	53.3	53.4	53.2	53.5	53.1	53.2	53.0	52.6	52.6
10	38.4	38.5	38.6	38.9	38.5	38.6	38.8	40.6	40.8
11	44.5	44.7	44.6	44.7	44.7	39.7	39.7	37.0	37.1
12	43.0	46.2	43.1	43.2	44.2	44.1	44.3	44.4	44.5
13	77.7	77.8	77.9	77.8	76.2	152.3	152.9	153.4	153.6
14	30.4	32.2	32.0	30.6	34.4	30.8	33.3	30.4	30.5
15	32.0	30.7	32.1	32.2	31.8	33.0	32.1	30.7	30.9
16	25.9	26.0	25.9	26.0	26.0	24.7	24.6	23.4	23.4
17	23.0	23.3	23.2	23.1	23.3	104.3	104.0	103.4	103.3
18	22.0	22.6	22.0	22.2	22.1	22.0	22.1	22.6	23.1
19	33.5	33.6	33.5	33.7	33.6	33.5	33.6	33.6	33.7
20	17.5	17.7	17.6	17.7	17.6	17.7	17.8	18.3	18.6

Acetate at δ 21.20 and 170.8.Malonate at δ 53.3, 166.4, 41.9 and 167.Isovaleroyl at δ 172.8, 43.9, 25.5, 22.5 and 20.1.

Reduction of **4a** with LiAlH_4 gave the alcohol **4b**. As expected, the ^1H NMR spectrum of **4b** lacked the resonances for the malonate protons and the H-7 were signals shifted upfield from δ 4.80 to 3.48. On the basis of the above data, and by comparison with the spectral data for similar stemarane diterpenes [7–10], **4b** had to be 13-methylene-7-hydroxy-stemarane and, therefore, **4a** was methyl-7-malonyloxy-13-methylene-stemarane.

The ^1H and ^{13}C NMR spectra of compound **5** showed characteristic signals of a tetracyclic diterpene with a stemarane skeleton [8–10]. The ^1H NMR spectrum showed a signal for an acetyl group (δ 2.03, s) three methyl groups [δ 0.94 (6H) and 1.03 (3H)]. Two doublets at δ 4.40 and 4.44 with a very small coupling constant were attributable to an exocyclic methylene group. The splitting pattern of a signal at δ 5.05 (1H, *tt*, $J = 4.1$, 11.7 Hz), indicated that the ^1H geminal to the acetoxyl group must be between two methylene groups, which meant that it could only be located at C-2, and from the J values this hydrogen must be axial, and in consequence, the acetoxyl must be equatorially oriented. The ^{13}C NMR data were in agreement with the proposed structure (Table 1).

Treatment of **5** with K_2CO_3 –methanol gave **5a**. As expected, the ^1H NMR spectrum of **5a** lacked the signals for the acetyl protons and the H-2 signal was shifted upfield at δ 3.90. Therefore, **5a** was shown to be 2 β -hydroxy-13-methylene-stemarane, and the natural product **5** is 2 β -acetoxyl-13-methylene-stemarane.

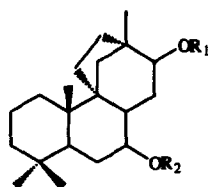
EXPERIMENTAL

Mps: uncorr.; ^1H NMR: 60, 100 and 360 MHz in CDCl_3 with TMS; ^{13}C NMR: 100 MHz. Assignments of ^{13}C NMR chemical shifts were made with the aid of APT and SFORD. IR: film; MS: direct inlet, 70 eV.

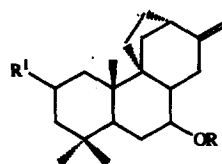
Calceolaria dentata Ret. Pav, collected in Malleco, IX Región, Chile, in November 1992, was identified by Dr. Otto Zoellner, Universidad Católica de Valparaíso. A voucher specimen is deposited at the Universidad Técnica Federico Santa María. The aerial parts of *C. dentata* (1500 g) were extracted at room temp. successively with petrol and CH_2Cl_2 for 48 hr each, affording 80 g syrup. The crude material was chromatographed on a silica gel column (400 g HF_{254} for TLC), and eluted with mixts of petrol and EtOAc of increasing polarity. Frs (125 ml) were combined based on TLC and ^1H NMR (60 MHz) monitoring, and the combined frs. purified by repeated CC on silica gel or silica gel impregnated with AgNO_3 (10%).

The molecular formulae were deduced by a combination of low resolution MS and the ^{13}C NMR spectra.

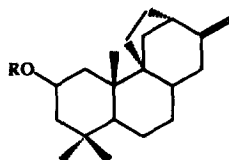
Methyl thyriflorin B acetate (1a). Oil, $[\alpha]_D^{25} - 76.5^\circ$ (c, 1.0 CHCl_3). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 2910–2760, 1720–1700, 1430, 1270, 1190, 1140, 1020, 980; ^1H NMR (300 MHz) δ : 4.77 (1H, *dt*, $J = 5.7$, 10.9 Hz, H-7), 4.60 (1H, *dd*, $J = 6.0$, 10.5 Hz, H-13), 3.76 (3H, s, OMe), 3.38 (2H, s, CH_2 malonyl), 2.03 (3H, s, OCOCH_3), 1.02 (3H, s, Me-17), 0.93 (3H, s, Me-20), 0.85 (3H, s, Me-18), 0.81 (3H, s, Me-19); ^{13}C NMR: Table 1; MS m/z (rel. int.): 448 ($\text{C}_{26}\text{H}_{40}\text{O}_6$) $[\text{M}]^+$ (2.0), 331 $[\text{M} - \text{C}_4\text{H}_5\text{O}_4]^+$ (1.0), 330



	R ₁	R ₂
1	COCH ₂ COOH	COMe
1a	COCH ₂ COOMe	COMe
1b	COCH ₂ COOMe	H
2	COCH ₂ CH(Me) ₂	COMe
2a	COCH ₂ CH(Me) ₂	H
3	COCH ₂ CH(Me) ₂	COCH ₂ COOH
3a	COCH ₂ CH(Me) ₂	COCH ₂ COOMe
3b	H	H



	R
4	COCH ₂ COOH
4a	COCH ₂ COOMe
4b	H



	R
5	COMe
5a	H

[M - C₄H₆O₄]⁺ (3.0), 272 [M - C₄H₅O₄ - C₂H₃O₂]⁺ (2.0), 270 [M - C₄H₆O₄ - C₂H₄O₂]⁺ (3.0), 175 (3.0), 173 (4.0), 171 (4.0), 162 (4.0), 159 (7.0), 157 (6.0), 147 (4.0), 145 (7.0), 133 (6.0), 131 (8.0), 123 (5.0), 118 (9.0), 104 (10.0), 95 (9.0), 93 (10.0), 81 (13), 69 (15), 59 (12), 57 (10), 55 (15), 43 (100), 41 (20).

Treatment of **1a** with K₂CO₃-MeOH gave **1b**, whose physical and spectral characteristics were identical to methyl thyriflorin B [5]. Treatment of **1a** with LiAlH₄ in dry Et₂O gave thyriflorin C (**3b**).

13-Isovaleroyl-7-acetoxy-thyriflorane (2). Oil, [α]_D²⁵ + 22.35 (c 1.2, CHCl₃). IR ν_{max}^{CHCl₃} cm⁻¹: 2920–2780, 1710, 1700, 1455, 1380–1360, 1250, 1200, 1150, 1120, 1040–1020, 1000, 930, 800–730; ¹H NMR (300 MHz) δ: 4.71 (1H, dt, J = 5.5, 11 Hz, H-7), 4.60 (1H, dd, J = 5.9, 10.5 Hz, H-13), 2.15 (2H, d, J = 6.0 Hz, H-2'), 2.09 (3H, s, OCOCH₃), 1.02 (3H, s, Me-17), 0.95 (6H, d, J = 6.4 Hz, Me-4' and Me-5'), 0.92 (3H, s, Me-20), 0.84 and 0.82 (3H each, s, Me-18 and Me-19); ¹³C NMR: Table 1.

13-Isovaleroyl-7-hydroxy-thyriflorane (2a). Compound **2** (100 mg) was treated with K₂CO₃-MeOH. After usual work, **2a** was obtained as an oil, [α]_D²⁵ = -14.13° (c 5.0, CHCl₃). IR ν_{max}^{CHCl₃} cm⁻¹: 3520–3460, 2900–2780, 1705, 1450, 1380–1360, 1290, 1260–1200, 1040, 1010, 1000, 980, 930–910, 720; ¹H NMR (300 MHz) δ: 4.65 (1H, dd, J = 6.0, 10 Hz, H-13), 3.39 (1H, dt, J = 5, 10.5 Hz, H-7), 2.16 (2H, d, J = 6.1 Hz, H-2'), 1.42 (1H, m, H-3'), 1.02 (3H, s, Me-17), 0.95 (6H, d, J = 6.4 Hz, Me-4' and Me-5'), 0.92 (3H, s, Me-20), 0.84 and 0.82 (3H each, s,

Me-18 and Me-19); ¹³C NMR: Table 1; MS m/z (rel. int.): 390 (C₂₅H₄₂O₃) [M]⁺ (1.0), 288 [M - C₅H₁₀O₂]⁺ (92.5), 270 [288 - H₂O]⁺ (52.2), 246 (18.9), 203 (13.7), 260 (17.4), 255 (21.1), 231 (12.6), 175 (10.6), 165 (19.5), 164 (59.4), 149 (13.0), 123 (20.3), 109 (18.9), 107 (18.0), 104 (16.3), 95 (21.1), 93 (21.6), 81 (35.5), 79 (20.9), 69 (38.4), 67 (23.9), 57 (96.1), 55 (47.6), 41 (100.0).

Treatment of **2** with LiAlH₄ in dry Et₂O gave **3b**.

13-Isovaleroyl-7-malonyloxy-thyriflorane (3). After addition of ethereal CH₂N₂, **3** was transformed into **3a**, crystals, mp 67–69° (MeOH), [α]_D²⁵ = -10.7° (c 1.3, CHCl₃). IR ν_{max}^{KBr} cm⁻¹: 2920–2830, 1680, 1720, 1250, 1190, 1140, 1120, 1040–1020, 980, 870, 800; ¹H NMR (300 MHz) δ: 4.77 (1H, dt, J = 5.6, 10.9 Hz, H-7), 4.60 (1H, dd, J = 5.9, 10 Hz, H-13), 3.73 (3H, s, OMe), 3.35 (2H, s, CH₂ malonate), 2.16 (2H, d, J = 6.2 Hz, H-2'), 1.01 (3H, s, Me-17), 0.94 (6H, d, J = 6.5 Hz, Me-4' and Me-5'), 0.91 (3H, s, Me-20), 0.83 and 0.80 (3H each, s, Me-18 and Me-19); ¹³C NMR: Table 1; MS m/z (rel. int.): 490 (C₂₉H₄₆O₆) [M]⁺ (2), 388 [M - C₅H₁₀O₂]⁺ (2), 373 [M - OCOCH₂ - COOCH₃]⁺ (6), 372 [M - C₄H₆O₄]⁺ (10), 357 [372 - Me]⁺ (8), 287 [M - C₅H₁₀O₂ - C₄H₅O₃]⁺ (3), 272 [M - C₅H₉O₂ - C₄H₅O₄]⁺ (5), 270 [M - C₅H₁₀O₂ - C₄H₆O₄]⁺ (100), 255 [270 - Me]⁺ (27.5), 242 (16), 159 (11.4), 149 (11.7), 145 (11.8), 133 (13), 118 (17), 109 (15), 107 (12.8), 105 (17.8), 101 [C₅H₉O₂]⁺ (8.5), 95 (14.2), 91 (13.9), 85 (15), 81 (16.2), 69 (13.5), 57 (18.4).

Treatment of **3a** with LiAlH_4 in dry Et_2O gave **3b** whose physical and spectroscopic characteristics were in full agreement with those of thyrsoflorin C [5].

Methyl-13-methylene-7-malonyloxy-stemaranane (4a). Crystals, mp: $75\text{--}77^\circ$ (M), $[\alpha]_D^{25} = 0^\circ$ (c, 2.0, CHCl_3). IR $\nu_{\text{max}}^{\text{KBr}} \text{cm}^{-1}$: 2900–2820, 1730–1710, 1620, 1410, 1370, 1360, 1240, 1140, 1100, 1080, 1040, 1010, 980, 900, 850; ^1H NMR (300 MHz) δ : 4.80 (1H, *dt*, $J = 6$, 11 Hz, H-7), 4.49 (1H, *brs*, H-17), 4.41 (1H, *brs*, H-17'), 3.76 (3H, *s*, OMe), 3.39 (2H, *s*, CH_2 malonyl), 2.62 (1H, *t*, $J = 4.9$ Hz, H-12), 1.02 (3H, *s*, Me-20), 0.84 and 0.81 (3H each, *s*, Me-18 and Me-19); ^{13}C NMR: Table 1; MS *m/z* (rel. int.): 388 $[\text{M}]^+$ ($\text{C}_{24}\text{H}_{36}\text{O}_4$) (2.3), 271 $[\text{M} - \text{C}_4\text{H}_5\text{O}_4]^+$ (22.0), 270 $[\text{M} - \text{C}_4\text{H}_6\text{O}_4]^+$ (68.8), 255 $[270 - \text{Me}]^+$ (100), 185 (15.7), 159 (18.8), 149 (15.8), 148 (59.5), 147 (288), 131 (22.2), 119 (16.4), 117 (15.1), 107 (15.0), 105 (21.3), 91 (21.5), 79 (11.2).

Treatment of **4a** with LiAlH_4 in dry Et_2O gave 7-hydroxy-13-methylene-stemaranane (**4b**) crystals, mp $166\text{--}168^\circ$ (MeOH). ^1H NMR (300 MHz) δ : 4.54 (1H, *d*, $J = 1.9$ Hz, H-17), 4.50 (1H, *d*, $J = 1.9$ Hz, H-17'), 3.48 (1H, *dt*, $J = 5.4$, 10.6 Hz, H-7), 2.65 (1H, *dd*, $J = 5.0$, 15 Hz, H-12), 1.05 (3H, *s*, Me-20), 0.89 and 0.88 (3H each, *s*, Me-18 and Me-19); ^{13}C NMR: Table 1.

2-Acetoxy-13-methylene-stemaranane (5). Oil, $[\alpha]_D^{25} = -38.07$ (c, 3.0, CHCl_3). IR $\nu_{\text{max}}^{\text{CHCl}_3} \text{cm}^{-1}$: 2940–2780, 1710, 1630, 1450, 1350, 1250, 1200, 1150, 1020, 980, 930, 870, 800; ^1H NMR (300 MHz) δ : 5.05 (1H, *tt*, $J = 4.1$, 11.7 Hz, H-2), 4.44 (1H, *d*, $J = 4.5$ Hz, H-17), 4.40 (1H, *d*, $J = 4.5$ Hz, H-17'), 2.62 (1H, *t*, $J = 5.1$ Hz, H-12), 2.02 (3H, *s*, OAc), 1.09 (3H, *s*, Me-20), 0.90 (6H, *s*, Me-18 and Me-19).

Treatment of **5** with $\text{K}_2\text{CO}_3\text{--MeOH}$ gave 2-hydroxy-13-methylene-stemaranane (**5a**). IR $\nu_{\text{max}}^{\text{CHCl}_3} \text{cm}^{-1}$: 3800–3000, 2940–2900, 2840, 2680, 2660, 2440, 1640, 1590, 1520, 1470, 1420, 1230, 1200, 1020, 940; ^1H NMR (300 MHz) δ : 4.46 (1H, *d*, $J = 4.4$ Hz, H-17), 4.39 (1H, *d*, $J = 4.4$ Hz, H-17'), 3.91 (1H, *tt*, $J = 4.0$, 11 Hz, H-2), 1.03 (3H, *s*, Me-20), 0.90 and 0.88 (3H each, *s*, Me-18 and Me-19).

Acknowledgements—We are grateful to Prof M. Nicoletti (Università La Sapienza, Rome, Italy), A. San Feliciano (Universidad de Salamanca, Spain) and R. Riguera (Universidad de Santiago de Compostela, Spain) for recording the ^1H , ^{13}C NMR and mass spectra, and to Prof. Otto Zoellner (Universidad Católica de Valparaíso, Chile) for identification of the plant material. This research was supported by a grant from FONDECYT N° 1941048.

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