



# ARATICULIN, A *BIS*-TETRAHYDROFURAN POLYKETIDE FROM *ANNONA*CRASSIFLORA SEEDS

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**Abstract**—A new tetrahydroxy adjacent bis-tetrahydrofuran polyketide, araticulin, has been isolated from the cytotoxic ethanolic extract of *Annona crassiflora* seeds. Its structure was established by NMR and mass spectrometry as a *threo-trans-threo* isomer of purpureacin-2, the only previously known bis-tetrahydrofuran polyketide bearing an OH group at C-12.

#### INTRODUCTION

Annona crassiflora, known popularly as 'araticum', is a native Brazilian tree found in the 'cerrado' area. Its fruit is edible and the seeds are traditionally used in folk medicine against snake bites [1]. A number of Annonaceae species of the genus Annona have been shown to contain polyketides with significant cytotoxic, antitumour, pesticidal, antimicrobial and antiparasitic activities [2, 3]. We have recently described a cytotoxic nonadjacent bis-tetrahydrofuran polyketide from the petrol extract of A. crassiflora seeds that we named crassiflorin [4]; this was shown to be identical to cherimolin-2 or bullatanocin [5, 6].

In the present paper, we report the structural determination of a new polyketide, named araticulin (1), which was isolated from the cytotoxic ethanolic extract of *A. crassiflora* seeds.

### RESULTS AND DISCUSSION

A 75% ethanol extract of *A. crassiflora* seeds exhibited significant *in vitro* cytotoxicity to human lung carcinoma (A-549) and melanoma (RPMI 7951) cells.

Compound 1 was isolated by a combination of open column chromatography on silica gel and washing with pH 4 buffer. It was obtained as a whitish wax, mp  $65-67^{\circ}$ . The FAB mass spectrum showed an  $[M + H]^+$  at m/z 639.51, corresponding to the molecular formula  $C_{37}H_{66}O_8$ . Spectral characteristics of 1 and its derivatives, including IR, <sup>1</sup>H NMR, <sup>13</sup>C NMR, HREI and FAB mass spectrometry, suggested that it belonged to the rare class of adjacent bis-tetrahydrofuran (THF) polyketides [2, 3].

On the basis of its spectral data (Table 1), **1** was suggested to be a new polyketide possessing a terminal  $\alpha, \beta$ -unsaturated  $\gamma$ -lactone with a 4-OH group [2, 3].

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Table 1	13C NIMD and	I 'LI NIMD date	for compound	1 and its tetra-acetate	(CDCL)

	1		1	Araticulin tetra-acetate	
C-Atom*	(50 MHz)	H-Atom*	(200 MHz)	(500 MHz)	
1	174.63	3a	2.51 <i>dd</i>	2.52	
2	131.17	3b	2.37 dd	_	
3	33.40	4	3.84 m	5.11 m	
4	69.96	5	1.54 m	1.67	
5	37.33	11	1.63 m	1.67	
6-10	29.3-29.7	12	3.59 m	4.58 m	
11	37.5	13-14	1.63 m		
12	71.67	15	3.39 m	4.85 m	
13	33.53	16	3.84 m	3.98 m	
14	31.89	17-18	1.67-1.96 m	1.97 m	
15	74.04†	19	3.84 m	3.90 m	
16	83.17‡	20	3.84 m	3.90 m	
17-18	28.37-28.96	21-22	1.67-1.96 m	1.97 m	
19	81.74	23	3.84 m	3.98 m	
20	81.85	24	3.39 m	4.85 m	
21-22	25.64-25.47	34	0.86 t	0.87 t	
23	82.84‡	35	7.17 d	7.07 s	
24	74.15†	36	5.05 ddd	5.09 ddd	
25	33.32	37	1.41 <i>d</i>	1.39 d	
26-33	28.37-28.96	4 OAc	_	2.022	
34	14.10	12 OAc	<del>_</del>	2.034	
35	151.79	15 OAc	_	2.074	
36	77.97	24 OAc	_	2.070	
37	19.10	<del></del>	_		

<sup>\*</sup>Biogenetic numbering.

The adjacent bis-THF ring system, with the usual OH groups on each side, was indicated by the <sup>13</sup>C NMR (CDCl<sub>3</sub>) signals of **1** (Table 1) at  $\delta$  74.15, 83.17, 82.84, 81.85, 81.74 and 74.04, and <sup>1</sup>H NMR chemical shifts (Table 1) at  $\delta$  3.39 (H-15, H-24) and 3.84 (m, H-16, H-19, H-20 and H-23). The signals in the <sup>1</sup>H and <sup>13</sup>C NMR spectra of **1** at  $\delta$  3.59 and 71.67 are characteristic of a hydroxyl group in an alkyl chain [5].

The THF rings were located from C-16 to C-20 in the hydrocarbon chain on the basis of typical fragments observed in the EI mass spectrum of 1 and its trimethylsilyl derivative. Allocation of the fourth hydroxyl group at C-12 was based on the occurrence of a fragment ion at m/z 269.1750 ( $C_{15}H_{25}O_4$ , calc. 269.17522) due to the cleavage between C-12 and C-13; this was confirmed by the EI mass spectrum of the corresponding trimethylsilyl derivative by the fragment ion at m/z 413.3.

The relative stereochemistry around the two THF rings was determined with the aid of empirical  $^{1}$ H and  $^{13}$ C NMR shift rules established with a series of model compounds [7, 8]. Comparison of  $^{1}$ H NMR data for the tetra-acetyl derivative (Table 1) and model compounds [7], indicated that the chemical shifts at  $\delta$  4.85 (H-15, H-24), 3.98 (H-16, H-23), 3.90 (H-19, H-20) and 2.07 (15-OAc, 24-OAc) shown by araticulin tetra-acetate were close to those for the *threo-trans-threo-trans-threo* model, suggesting this relative stereochemistry for the THF moieties (C-15/C-16, C-16/C-19, C-19/C-20, C-20/C-23 and C-23/C-24).

The structure of 1 is closely related to that of purpureacin-2, isolated from *A. purpurea* leaves [9]. These two isomeric compounds are the first representatives of adjacent bis-THF Annonaceous polyketides bearing a hydroxyl group at C-12. However, the minor differences in the <sup>13</sup>C and <sup>1</sup>H NMR chemical shifts suggest that 1 is a new stereoisomer of purpureacin-2 whose relative stereochemistry was not reported by Cepleanu *et al.* [9]. However, the stereochemistry at C-4, C-12 and C-36 in 1 are unknown.

Compound 1 showed significant activities against human tumour cell lines in culture when compared with adriamycin (Table 2).

## **EXPERIMENTAL**

General. Mps: uncorr. IR: KBr. <sup>13</sup>C NMR: 50 MHz, CDCl<sub>3</sub>. <sup>1</sup>H NMR: 200 and 500 MHz, CDCl<sub>3</sub>.

Plant material. Fruits of A. crassiflora were collected in Itatiaiuçu and Curvelo, Minas Gerais, Brazil. Plant material was identified by J. L. Pedersoli, Instituto de Ciências Biológicas, UFMG, Belo Horizonte, Minas Gerais, Brazil. A voucher specimen is deposited at the BHCB, UFMG, Belo Horizonte.

Bioassays. Crude extracts were evaluated for cytotoxicity at the Ohio State University, using standard protocols for A 459 (human lung carcinoma), HT-29 (human colon adenocarcinoma), MCF-7 (human breast carcinoma), RPMI 7951 (melanoma) and U 251 (CNS carcinoma).

<sup>†,‡</sup>Assignments in vertical column interchangeable.

Table 2. Bioactivities of 1 on human tumor cell lines

	ED <sub>50</sub> (μg ml <sup>-1</sup>	1)			
Compound	A-549*	HT-29†	MCF-7‡	RPMI-7951§	U-251
Araticulin (1)	$6 \times 10^{-3}$	6×10 <sup>-1</sup>	$4 \times 10^{-3}$	10 <sup>-3</sup>	10 <sup>-2</sup>
Adriamycin¶	$4 \times 10^{-3}$	$3 \times 10^{-3}$	$2\times10^{-3}$	$5 \times 10^{-3}$	$1 \times 10^{-2}$

<sup>\*</sup>Human lung carcinoma.

Extraction and isolation. Powdered seeds (1.25 kg) were successively extracted in a Soxhlet apparatus with petrol and 75% EtOH. The EtOH extract (91.1 g) was fractionated on a silica gel column with a gradient of hexane-CH<sub>2</sub>Cl<sub>2</sub>-EtOAc-MeOH to give 82 frs. A CH<sub>2</sub>Cl<sub>2</sub> soln of the residue (1.4 g) of the fr. eluted with EtOAc was washed with a buffer soln at pH 4. The residue from the organic layer gave a mixt. of a yellow wax and a white powder. The wax was extracted with EtOAc and the residue recrystallized from EtOAc to give a white amorphous powder (40 mg) of 1.

Araticulin (1).  $C_{37}H_{66}O_8$ . [α]<sub>D</sub> +9.6° (MeOH; c 0.27). IR  $\nu_{max}$  cm<sup>-1</sup>: 3450, 2920, 2840, 1750, 1600, 1450, 1300, 1060, 1020, 920. FABMS (3-NBA), m/z: 639.51 [MH]<sup>+</sup>, 621.56 [MH –  $H_2O$ ]<sup>+</sup>, 603.59 [MH –  $2H_2O$ ]<sup>+</sup>, 585.59 [MH –  $3H_2O$ ]<sup>+</sup>, 567.59 [MH –  $4H_2O$ ]<sup>+</sup>. EI-MS: 467, 449, 397, 379, 369, 309, 241, 269, 171, 141. EI-MS, TMSi derivative m/z: 683, 613, 593, 543, 453, 413, 383, 313, 243, 213. <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>). Table 1. <sup>1</sup>H NMR (200 and 500 MHz, CDCl<sub>3</sub>), Table 1.

Araticulin tetraacetate. FABMS (m-NBA), m/z 807.79 [MH]<sup>+</sup>, 747.69 [MH – AcOH]<sup>+</sup>, 687.54 [MH – 2AcOH]<sup>+</sup>, 627.56 [MH – 3AcOH]<sup>+</sup>, 567.70 [MH – 4AcOH]<sup>+</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>), Table 1.

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<sup>†</sup>Human colon adenocarcinoma.

<sup>‡</sup>Human breast carcinoma.

<sup>§</sup>Melanoma.

Human CNS carcinoma.

Positive control standard.