

PII: S0031-9422(97)00105-2

A PHENOLIC CINNAMATE DIMER FROM PSORALEA PLICATA

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(Received in revised form 24 December 1996)

Key Word Index—*Psoralea plicata*; Leguminosae; sesquiterpenes; tocopherol; phenolic cinnamates; furanocoumarins; plicatin A and B; *p*-coumaric acid derivatives; phenolic dimers; steroids; triterpenes.

Abstract—Caryophyllene oxide, α -tocopherol, Z and E-werneria chromenes, two furanocoumarins, bakuchicin and psoralen, in addition to plicatin-B, lupeol and stigmasterol, have been isolated from the hexane-soluble extract of the aerial parts of *Psoralea plicata*. Plicatin-A, 3-(-3-methyl-2,3-epoxybutyl)-p-coumaric acid methyl ester and a new dimer, α -diplicatin B, were isolated from the ethyl acetate-soluble fraction. From the butanol-soluble matter, roseoside A, daidzin, isopsoralic acid-O-gluco-pyranosyl and isovitexin were isolated. Most of these compounds have been isolated from this species for the first time, although known from other plant sources. © 1997 Elsevier Science Ltd. All rights reserved

INTRODUCTION

The herb, *Psoralea plicata*, known in Arabic as 'Marmid' [1], is widely distributed in the Allaqi area, south of Aswan [2]. The plant is used by Bedouins as a grazing plant and for medicinal uses for different ailments.

Various species of *Psoralea* are reputed in indigenous medicine for their anthelmintic, antipyretic, analgesic, anti-inflammatory, diuretic and diaphoretic properties and are useful in bilious infections, in leprosy and for menstruation disorders [3, 4]. These reports prompted us to carry out systematic phytochemical studies in order to isolate the secondary metabolites found in these important medicinal plants. Previous work has identified the presence of furocoumarins, phenolic cinnamates, coumestans, terpenoids, isoflavones, flavonoids and α-tocopherolquinones [5–22].

RESULTS AND DISCUSSION

A methanol extract of the air-dried aerial parts of *P. plicata* was concentrated and the solvent-free residue exhaustively extracted with hexane, ethyl acetate and *n*-butanol, respectively. The residue obtained from each solvent was chromatographed as detailed in the Experimental section for isolation of its components.

Spectral analysis of compound 1 suggested that it should be β -caryophyllene oxide [23, 24]. The dimethyl protons appeared in the ¹H NMR at δ 0.99 and 1.00, and its ¹³C NMR shifts at δ 21.5 and 29.8, respectively, in agreement with data reported for the *trans*-conformer of caryophyllene oxide [25, 26].

The second oily compound **2** was identified as α -tocopherol, in contrast to tocopherolquinone and its methyl ester reported from the same plant source [11, 27]. Our mass spectrum and fragmentation pattern agreed well with reported data for α -tocopherol [28].

Compounds 3 and 4 were identified as Z- and E-werneria chromenes, previously reported from Werneria species [29], but isolated from P. plicata for the first time. It should be noted that werneria chromene has been prepared semisynthetically by cyclization of plicatin-B, the anti-microbial agent isolated from several Psoralea species [10].

Compound 5 was bakuchicin, an angular furanocoumarin isolated previously from *P. corylifolia* [8]. Compounds 6 and 8 were identified as psoralen and plicatin-A, respectively, being common constituents in most *Psoralea* species [9, 10, 19].

The ¹H NMR spectrum of compound 7 showed two *ortho*-coupled aromatic protons at δ 7.30 and 6.80 (each 1H, d, J = 8.89 Hz) and an isolated proton singlet at δ 7.28. The presence of a *trans*-methyl cinnamate moiety was shown by a pair of doublets of olefinic protons at δ 6.28 and 7.62 (J = 15.93 Hz) and a sharp singlet for a methoxyl group at δ 3.78. In addition, the spectrum showed the presence of a pre-

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nyl group (two methyl singlets at δ 1.76 and 1.78; 1H, multiplet at δ 5.31 and 2H, doublet at δ 3.35, J = 7.16 Hz). The EI mass spectrum of this compound showed a [M]⁺ at m/z 246, giving the molecular formula $C_{15}H_{18}O_3$, with characteristic fragments at m/z 231 [M – Me]⁺, 229 [M – OH]⁺, 215 [M – MeO]⁺ and 191

[M-C₄H₇]⁺. The ¹³C NMR spectrum determined in the DEPT mode, showed the presence of 15 carbon atoms with three methyls (δ 17.6, 25.6 and 51.6), one methylene (δ 28.5), six methines (δ 113.9, 115.7, 121.4, 127.5, 129.9 and 145.7), one carbonyl (δ 168.7) and four quaternary carbon atoms (δ 126.4, 128.0, 133.8

and 156.8). The set spectral data corresponded with plicatin B, an antibacterial compound isolated previously from *P. plicata* and *P. juncea* [9, 10].

Compound 9 was obtained as an oil and identified by spectral measurements as 3-(3-methyl-2-3-epoxybutyl-)-p-coumaric acid methyl ester, an epoxide reported from *Baccaris* species [21]. It is the first time that this epoxide has been identified *Psoralea*.

The ¹H NMR spectrum of compound 10 showed three signals for aromatic protons, two doublets, at δ 6.65 and 6.99 (J = 8.9 Hz) and one broad singlet at δ 7.02. In addition, the spectrum showed the presence of a prenyl group (6H singlet at δ 1.88, 1H multiplet at δ 5.29 and 2H doublet at δ 3.20 (J = 7.1 Hz). Furthermore, the presence of two methine protons at δ 3.90 and 4.35, each being a double doublet (A₂B₂ pattern showing eight lines for H-1" and H-2", J cis/J trans 10.4/7.2) [30] and of the A_2B_2 pattern type was characteristic of cyclobutyl protons, since in this case each proton on the cyclobutane ring is cis-coupled with one of its neighbours and trans-coupled with the other; this suggested that compound 10 should be a dimer [31]. The ¹³C NMR of compound 10 showed 15 carbon atoms, the same as that of plicatin-B, but compound 10 contained two methine carbons shifted upfield at δ 40.75 and 47.10; this is also characteristic of methine carbons in a cyclobutyl ring. In addition, there were four methine, one methylene, four quaternary carbons, one methoxyl, two methyls and one carbonyl carbon. The mass spectrum of compound 10 showed only peaks for the monomer but the corresponding spectrum of its diacetate showed a [M]+ at m/z 576, giving the molecular formula $C_{34}H_{40}O_8$. This observation confirmed that compound 10 was a dimer of plicatin-B, given the name α -diplicatin B. Compound 10 is symmetrical, because only half of the total number of protons and carbon atoms are found in its ¹H and ¹³C NMR spectra. Although the finding of a naturally occurring dimer, such as α-diplicatin B, is unusual, an equivalent dimer of cinnamic acid (referred to as gratissimic acid) together with its dimethyl ether (gratissimin) has been previously reported in Ocimum gratissimum [30], together with α dicerptene in Pityrogramma triangularis [31].

From the *n*-butanol soluble extract, four glycosides, namely, roseoside A 11 [32–37], diadzin 12 [38–40], isopsoralic acid-*O*-glucopyranosyl 13 [41] and isovitexin 14 [42, 43] were isolated, respectively. The HC-correlations based on HMBC for the acetates of compounds 11 and 13 are presented in Figs 1 and 2.

The ¹H NMR, spectrum of compound 11, showed a signal for one olefinic proton at δ 5.88 (1H, br s), which revealed in the correlation spectrum (HMBC), a cross-peak with the methyl group at δ 1.88 (3H, d, J=1.8 Hz) on a quaternary carbon. The methylene protons at δ 2.43 and 2.25 (each 1H, d, J=17.0 Hz, H-2 α and β), showed cross-peaks with two geminal methyl groups at δ 1.01 and 1.07 (each, 3H, s). The UV spectrum revealed that compound 11 contained a conjugated chain, which was confirmed by ¹³C NMR

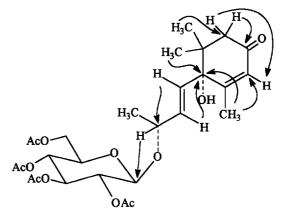


Fig. 1. ¹H-¹³C correlation based on HMBC of roseoside A tetraacetate.

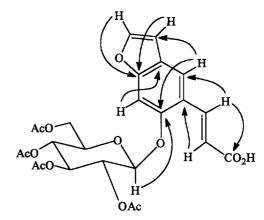


Fig. 2. ¹H-¹³C correlation based on HMBC of isopsoralic acid-6-*O*-glucoside tetraacetate.

showing the characteristic shift for a ketonic carbon at δ 197.7. The ¹H NMR spectrum also showed two other olefinic protons at δ 5.77 (1H, d, J = 10.3 Hz) and δ 5.78 (1H, dd, J = 10.3 and 5.5 Hz), with the latter showing a cross-peak with a proton at δ 4.23 (1H, m), which, in turn, showed coupling (HH-COSY)with methyl protons at δ 1.23 (3H, d, J = 6.4 Hz). Compound 11 also showed the anomeric proton for glucose at δ 4.56 (1H, d, J = 7.9 Hz) which showed a cross-peak (CH-COSY) with the proton at δ 4.23, suggesting that the linkage of the sugar is at the OH on the carbon at δ 77.0. The ^{13}C NMR spectrum revealed the presence of one anomeric carbon at δ 100.0 and five other carbons (one methylene and four methines), besides four acetate group carbons. The coupling constant, NOESY and the chemical shift of the anomeric proton, confirmed that the sugar was β linked. Acid hydrolysis of this compound with 5% HCl, gave D-glucose and vomifoliol (blumenol) [32-36]. From the above mentioned data and optical rotation, we found that compound 11 is similar to roseoside-A, previously isolated from Vinca rosea [37].

The ¹H NMR spectrum of compound 13 showed two furan protons at δ 7.60 (1H, d, J = 2.2 Hz) and δ 6.65 (1H, dd, J = 2.2 and 0.5 Hz). Also, there were

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two singlets for two aromatic protons at δ 7.79 and 7.37, in addition to a cinnamic acid group (two transcoupled olefinic protons) at δ 8.04 and 6.94 (J = 16.1Hz). The chemical shifts and splitting patterns of the furan, aromatic and olefinic protons were similar to psoralen, which was confirmed by a CH-COSY spectrum. The correlation spectrum (HMBC), showed a cross-peak between H-3 (δ 6.75), C-4 (δ 129.2) and C-8 (δ 156.5). Furthermore, H-2 (δ 7.60), displayed a cross-peak with C-9 (δ 123.6), that showed a crosspeak with H-7 (δ 7.37). From these correlations, we suggest that compound 13 has a benzofuran cinnamic acid structure. The ¹H NMR and ¹³C NMR spectra showed one proton at δ 5.08 (1H, d, J = 7.8 Hz), its carbon at δ 100.3 showing a cross-peak with the carbon at δ 153.5, revealing that the sugar was β -linked to the carbon at δ 153.5. The ¹H NMR of compound 13 and ¹³C NMR also showed four acetates and a sugar containing five methine carbons and one methylene group, indicating that the sugar was hexose. This was confirmed by acid hydrolysis with 5% HCl, giving glucose (TLC). From the above mentioned data and mass spectrum, we suggest that compound 13 was isopsoralic acid $1 \rightarrow 6$ -O- β -D-glucoside, isolated previously from Coronilla glauca [41].

EXPERIMENTAL

¹H NMR and ¹³C NMR (at 100 MHz), spectra were recorded using a Bruker AM-400 and Gemini-300, respectively, NOESY and HMBC on a Bruker AM-500.

Extraction and isolation of constituents. Air-dried aerial parts (2 kg) of *P. plicata* Del. (Cullen plicatum Delile C. H. Stirt) were powdered and exhaustively extracted with 75% MeOH by maceration. The alcohol extract was concd under red. pres. to a syrupy consistency (179 g). The solvent-free residue (50 g) was mixed with 200 ml H₂O and 100 ml MeOH, transferred to a separatory funnel and partitioned between hexane, EtOAc and *n*-BuOH. Each fr. was dried (Na₂SO₄) and concd to a syrupy residue (10 g hexane residue, 3.6 g EtOAc residue and 5 g *n*-BuOH residue).

Hexane-soluble fraction. Separation of compounds was achieved using repeated flash CC on silica gel, eluting with a hexane-EtOAc gradient. Compounds obtained were 1 (50 mg), 2 (25 mg), 3 (5 mg), 4 (30 mg), 5 (5 mg), 6 (18 mg), 7 (700 mg), lupeol (45 mg) and stigmasterol (10 mg).

Ethyl acetate-soluble fraction. The fr. (3.6 g) was slurried with 7 g of silica gel and transferred to a silica gel column, previously packed by the wet method in hexane–CHCl₃-MeOH (3.5:1.4:0.1), starting elution with the same solvent system to give pure epoxy compound 9 (7 mg) and the new dimer 10 (5 mg). The column was then eluted with MeOH and the MeOH eluate (1 g) subjected to ODS CC, eluting with MeOH–H₂O (4:1) to give another crop of compound 10 (30 mg).

Butanol-soluble fraction. The fr. (5 g) was dissolved

in a small amount of MeOH, transferred to the top of a Sephadex-LH 20 column, previously packed in MeOH, and elution was started with MeOH (each fr. 250 ml) Each fr. obtained was conc. to 5 ml under red. pres. and analysed by TLC using EtOAc–MeOH– $\rm H_2O$ (100:16.5:13.5), where two subfrs, 1 and 2, were obtained.

Separation of compounds 11 and 12 from subfr. 1. Subfr. 1 (200 mg) was dissolved in a small amount of MeOH and transferred to a flash silica gel column previously packed with CHCl₃-MeOH-H₂O (30:20:1) and eluted by the same solvent to give two frs A and B.

Separation of compound 11 by acetylation. Fr A (20 mg) was dissolved in 2 ml pyridine and 1.9 ml Ac₂O and left for 24 hr at room temp. The reaction mixt. was tested by TLC for complete acetylation, then purified by flash silica gel CC eluted with hexane—Me₂CO (3:2), to give pure compound 11 acetate (17 mg).

Separation of compound 12 by acetylation. Fr. B (30 mg) was acetylated in a similar manner to fr. A and the mixt. purified by flash silica gel CC with toluene—MeOH (7:3), to give pure compound 12 acetate (20 mg).

Separation of compounds 13 and 14. Subfr. 2 (500 mg), was dissolved in a small amount of MeOH and transferred to a flash silica gel column, previously packed with CHCl₃–MeOH–H₂O (37:12:1) and eluted with the same solvent system; 100 ml frs were collected Frs 15–25 gave compound 13 (30 mg) and frs 30–45 gave compound 14 (150 mg).

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