



## SITOSTEROL SUCROSIDE FROM THE SUCKERS OF *MENTHA ARVENSIS*\*

RIAZ A. KHAN, ANIL K. SINGH and PAWAN K. AGRAWAL†

Central Institute of Medicinal and Aromatic Plants, Lucknow-226 015, India

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**Key Word Index**—*Mentha arvensis*; Labiatae; suckers; sucrose; sitosterol sucroside.

**Abstract**—Two constituents, isolated from the suckers of *Mentha arvensis*, were identified as 3-*O*- $\beta$ -sitosteryl-glucopyranosyl-(1 $\alpha$   $\rightarrow$  2)-fructofuranoside and sucrose. © 1997 Elsevier Science Ltd. All rights reserved

### INTRODUCTION

*Mentha arvensis* var. *piperascens* is one of the major sources for *l*-menthol and it has been intensively investigated [2, 3]. Four non-volatile constituents, sitosterol- $\beta$ -D-glucoside [4], *l*-menthyl- $\beta$ -D-glucoside, *l*-menthyl-6'-*O*-acetyl- $\beta$ -D-glucoside [5] and lambda-menthyl- $\beta$ -D-glucoside [6] have been isolated from leaves. The present investigation describes the isolation and characterization of two constituents from the ethanolic extracts of the suckers of *M. arvensis*.

### RESULTS AND DISCUSSION

Column chromatography of the chloroform extract of the suckers of *M. arvensis* led to the isolation of compound **1** as an amorphous powder, mp 182°. The <sup>1</sup>H NMR spectrum displayed six methyl signals, distinguishable into two singlets ( $\delta$  0.71 and 1.02), three doublets ( $\delta$  0.84, 0.86 and 0.94), and a triplet ( $\delta$  0.89) in addition to a broad doublet at  $\delta$  5.36 (2H). The <sup>13</sup>C NMR spectrum showed two olefinic carbons at  $\delta$  142.0 (C) and 122.75 (CH) characteristic of  $\Delta^5$  steroidal skeleton [7, 8] and other resonances in the aliphatic region (12–55 ppm) showing significant resemblance with the literature values for sitosterol [9, 10]. The existence of two anomeric carbon resonances at  $\delta$  102.5 (C) and 94.8 (CH), together with 3 methylene resonances at  $\delta$  58.8, 62.8 and 63.7 and 7 oxymethine resonances in the 67–80 ppm region, reflected its diglycosidic nature [11]. A comparative study of the <sup>13</sup>C NMR shielding data for **1** with those observed for sucrose (**2**) (also isolated in the present studies)

[12] showed appreciable similarities. A comparative study of <sup>13</sup>C NMR shielding data for **1** and **2** revealed that the C-3 resonance of the  $\alpha$ -D-glucopyranose was at 5.8 ppm lowerfield position and the C-2 and C-4 resonances were 1.1 ppm upfield in **1** compared with **2**. These glycosylation induced <sup>13</sup>C NMR shifts [13] were consistent with the sitosterol substitution at the C-3 position of the  $\alpha$ -D-glucopyranose. Consequently, the structure of **1** was deduced as 3-*O*- $\beta$ -sitosteryl-glucopyranosyl-(1 $\alpha$   $\rightarrow$  2)-fructofuranoside. To the best of our knowledge, this is the first report of occurrence of any sitosterol sucroside in nature, although the rhamnoglucoside and the rhamnoarabinofuranoside of sitosterol have been reported from *Lindenbergia indica* [14]. The occurrence of sucrose esters has been recently reported from *Nicotiana glauca* [15].

### EXPERIMENTAL

**General.** All mps were determined in open capillaries and uncorr. The NMR measurements were carried out on Bruker WM-400 NMR spectrometer in CD<sub>3</sub>OD for **1** and in D<sub>2</sub>O for **2** [16].

**Plant material and isolation.** The suckers of *Mentha arvensis* were collected from CIMAP field station, Pantnagar (U.P.) and air dried. The ground and defatted suckers (270 g) were extracted with EtOH (3  $\times$  500 ml) and the combined extract was concd to 1/3 its original vol. which was then held at 5° for 48 hr. This led to the deposition of white amorphous material which was filtered and repeatedly washed with MeOH to afford **2**, <sup>13</sup>C NMR (D<sub>2</sub>O): glucopyranose: 91.9 (C-1), 72.1 (C-2), 72.3 (C-3), 68.9 (C-4), 72.8 (C-5), 59.85 (C-6), fructofuranose: 62.06 (C-1), 103.4 (C-2), 81.06 (C-3), 76.2 (C-4), 73.7 (C-5), 61.09 (C-6) identified as sucrose [12].

The filtrate was concd to viscous mass (7.5 g) and extracted with CHCl<sub>3</sub> (3  $\times$  50 ml). The CHCl<sub>3</sub> con-

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† Author to whom correspondence should be addressed.

centrate (3.2 g) was chromatographed over silica gel with  $\text{CHCl}_3$  and increasing polarities of MeOH as eluent to yield **1** (130 mg)  $R_f$  0.6, Solvent system:  $\text{CHCl}_3$ –MeOH– $\text{H}_2\text{O}$ , 3:4:1; in frs eluted with  $\text{CHCl}_3$ –MeOH (3:7).  $^{13}\text{C}$  NMR ( $\text{CD}_3\text{OD}$ ): sitosterol: 38.55 (C-1), 27.2 (C-2), 77.8 (C-3), 41.2 (C-4), 142.0 (C-5), 122.75 (C-6), 33.05 (C-7), 33.7 (C-8), 51.7 (C-9), 37.4 (C-10), 22.7 (C-11), 39.26 (C-12), 41.1 (C-13), 57.4 (C-14), 25.3 (C-15), 30.2 (C-16), 54.69 (C-17), 12.24 (C-18), 19.4 (C-19), 34.9 (C-20), 19.1 (C-21), 35.1 (C-22), 29.3 (C-23), 49.5 (C-24), 25.98 (C-25), 19.8 (C-26), 20.1 (C-27), 23.7 (C-28), 12.3 (C-29). Glucopyranose: 94.89 (C-1), 71.0 (C-2), 78.1 (C-3), 67.8 (C-4), 72.9 (C-5), 58.8 (C-6), fructofuranose: 63.7 (C-1), 102.5 (C-2), 80.0 (C-3), 76.4 (C-4), 74.1 (C-5), 62.8 (C-6). FAB-MS: 644, 599  $[(\text{M} + \text{Na})\text{-Glc}]^+$ , 574, 555, 518, 496, 413, 411, 397, 329, 307, 289, 184, 176, 165.

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#### REFERENCES

1. Khan, R. A., Gupta, A. K., Singh, A. K. and Agrawal, P. K., *Indian Journal of Chemistry*, (in press).
2. Mehra, B. K. in *Mint, Cultivation and Utilization of Medicinal and Aromatic Plants*, ed. C. K. Atal and B. M. Kapoor. CSIR, New Delhi, 1960, p. 173.
3. Duhan, S. P. S., Singh, V. P., Bhattacharya, A. K. and Husain, A., *Perfumer & Flavorist*, 1977, **2**, 57.
4. Shimizu, S., Shibata, H. and Maejima, S., *Journal of Essential Oil Research*, 1990, **2**, 21.
5. Sakata, I. and Mitsui, T., *Agricultural and Biological Chemistry*, 1975, **39**, 1329.
6. Sakata, I. and Koshimizu, Y., *Agricultural and Biological Chemistry*, 1978, **42**, 1959.
7. Agrawal, P. K., Jain, D. C., Gupta, R. K. and Thakur, R. S., *Phytochemistry*, 1985, **24**, 2479.
8. Agrawal, P. K., Jain, D. C. and Pathak, A. K., *Magnetic Resonance Chemistry*, 1995, **33**, 923.
9. Holland, H. L., Diakow, P. R. P. and Taylor, G. J., *Canadian Journal of Chemistry*, 1978, **56**, 3121.
10. Agrawal, P. K. and Bishnoi, V., *Indian Journal of Chemistry*, 1996, **35B**, 86.
11. Agrawal, P. K., *Phytochemistry*, 1992, **31**, 3307.
12. Breitmaier, E. and Voelter, W.,  $^{13}\text{C}$  NMR Spectroscopy. Verlag Chemie, Berlin, 1978, p. 259.
13. Agrawal, P. K. and Pathak, A. K., *Phytochemical Analysis*, 1996, **7**, 113.
14. Tiwari, K. P. and Choudhary, R. N., *Phytochemistry*, 1979, **18**, 2044.
15. Ohya, I., Sinozaki, Y., Tobita, T., Takahashi, H., Motosuzaki, T. and Koiwai, A., *Phytochemistry*, 1994, **37**, 143.
16. Singh, A. K., Pathak, V. and Agrawal, P. K., *Phytochemistry*, 1997, **44**, 555.