

PII: S0031-9422(97)00218-5

24(24¹)[Z]-DEHYDROAMARASTERONE B, A PHYTOECDYSTEROID FROM SEEDS OF *LEUZEA CARTHAMOIDES*

U. A. BALTAYEV, L. DINAN,*† J.-P. GIRAULT‡ and R. LAFONT§

Institute of Petrochemistry and Catalysis, Bashkortostan Republic Academy of Sciences, 450075 Ufa, Prospekt Oktyabrya 141, Russia; * Department of Biological Sciences, University of Exeter, Washington Singer Laboratories, Perry Road, Exeter, Devon EX4 4QG, U.K.; ‡ Université René Descartes, Laboratoire de Chimie et Biochimie Pharmacologiques et Toxicologiques, CNRS-URA 400, 45 Rue Saints-Pères, F-75270 Paris Cedex 06, France; § Ecole Normale Supérieure, Laboratoire de Biochimie, CNRS-EP 119, 46 rue d'Ulm, F-75230 Paris Cedex 05, France

(Received in revised form 29 January 1997)

Key Word Index—Leuzea carthamoides; Rhaponticum carthamoides; Compositae; ecdysteroid; phytoecdysteroid; 24(24¹)[Z]-dehydroamarasterone.

Abstract—A new phytoecdysteroid, $24(24^{\circ})[Z]$ -dehydroamarasterone B, has been isolated from seeds of *Leuzea* (*Rhaponticum*) carthamoides. It has been unambiguously identified by CIMS, ¹³C NMR and ¹H NMR spectroscopy. The biological activity of the ecdysteroid has been determined in the *Drosophila melanogaster* B₁₁ bioassay. The ED₅₀ (5.2×10^{-7} M) is 70-fold higher than that for 20-hydroxyecdysone (7.5×10^{-9} M). © 1997 Elsevier Science Ltd. All rights reserved

INTRODUCTION

Certain species within the Compositae, including Leuzea carthamoides D.C. (syn. Rhaponticum carthamoides [Willd.] Iljin) [1–6], contain very high concentrations of insect steroid hormone analogues (phytoecdysteroids) [7], even though phytoecdysteroids are not of widespread occurrence in the Compositae in general (L. Dinan, unpublished data). While proceeding with a systematic study of the phytoecdysteroids of Leuzea carthamoides, we investigated the seeds for their phytoecdysteroid content [8–10]. We now report the isolation and identification of a new phytoecdysteroid, 24(24¹)[Z]-dehydroamarasterone B (1) from this source.

RESULTS AND DISCUSSION

The CI mass spectrum clearly established the molecular mass of the purified compound 1 as 506, solving for $C_{29}H_{46}O_7$. The UV spectrum was characteristic for an ecdysteroid, as was the fragmentation pattern in the mass spectrum. The ¹H NMR spectrum of 1 (Table 1) was very similar to that of 20-hydroxyecdysone [11] in all regions except for those signals originating from the side-chain (C-22–C-29). However, extra signals with respect to the ¹H NMR spectrum of 20-hydroxyecdysone [11] are observed; a triplet 1H at δ 5.45

in the ethylenic zone coupled to the multiplet 2H at δ 4.23 in the zone for hydroxymethene and a septuplet 1H (δ 2.87). Moreover, the upfield shift (0.2 ppm) of the 26-Me and 27-Me signals and their splitting to give doublets are indicative of a 25-deoxy compound. 2D COSY and TOCSY [12] experiments lead to correlation of the 26-Me and 27-Me signals with the septuplet 1H at δ 2.87, consequently assigned as H-25. The chemical shift of the H-25 signal and the absence of other correlations than with 26-Me and 27-Me signals, together with the correlation of H-22 with two downfield shifted H-23 at δ 2.41 and 1.95 are in accordance to the presence of an ethylenic double bond between C-24 and C-24¹. The signal at δ 5.45 is similar to one of those observed previously for C-241 in 24(241)-dehydromakisterone A [13]. A phasesensitive NOESY experiment [14] carried out with a mixing time of 500 ms leads to the observation of NOEs between $\{H-25\}H-24^2 \{H-24^2\}26-Me/27-Me$ and between {H-241}H-22 {H-241}H-29 and {H-24¹}H-23 revealing a Z-stereochemistry of the C-24/C-24¹ double bond. The structures of the side-chain and of the steroid nucleus, are confirmed by ¹³C NMR data (Table 2) obtained by PFG-HMQC and PFG-HMBC [15] ¹H-¹³C 2D correlations. HMBC leads to ²J and ³J¹H-¹³C correlations from 26-Me/27-Me; H-25; H-23a; H-23b to the quaternary sp² C at δ 147.4 (assigned as C-24) and from H-25; H-23a; H-23b to the sp² CH at δ 124.3.

Purified 1 possesses agonistic activity in the B_{II} bioassay. The potency (ED₅₀ = 5.2×10^{-7} M) was less

[†] Author to whom correspondence should be addressed.

Table 1. ¹H NMR data of 20-hydroxyecdysone and compound 1

	1	20-Hydroxyecdyson	
l-Hax	1.4 (t, 13)	1.4 (t, 13)	
1-He	1.85	1.85	
2-Hax	$3.99 (m, W_{1/2} = 22, 1H)$	$3.99 (m, W_{1/2} = 22)$	
3-He	$4.07 (m, W_{1/2} = 8, 1H)$	$4.07 (m, W_{1/2} = 8)$	
4-Hax	1.75	1.75	
4-He	1.75	1.75	
5-H	2.34 (1H)	2.36 (t*)	
7- H	5.99 (d, 2.5, 1H)	5.97 (d, 2.5)	
9-Ηα	$3.11 (m, W_{1/2} = 22)$	$3.11 (m, W_{1/2} = 22)$	
11-Hax	1.75	1.75	
11-He	1.85	1.85	
12-Hax	1.73	1.73	
12-He	1.95	1.95	
15-Hα	1.65	1.7	
15-Ηβ	2.05(m)	2.05(m)	
16-Hα	1.95	1.9	
16-Hβ	1.85	1.75	
17-H	2.39	2.34(t)	
22-H	3.72 (d, 10, 1H)	3.43 (d, 10)	
23-Ha	2.41	1.3	
23-Hb	1.95	1.65	
24-Ha		1.75	
24-Hb	_	1.50 (t, d 13, 3, 5)	
25-H	2.87 (sp, 6.6, 1H)	_	
28-H	5.45 (t, 7, X, 1H)	_	
29-Ha/b	4.23 (m, AB, 2H)		
18-Me	0.88 (s, 3H)	0.87(s)	
19-Me	1.00(s, 3H)	1.00(s)	
21-Me	* * *	1.22(s)	
26-Me	* * *	1.24 (s)	
27-Me	1.04(d, 6.6, 3H)	1.24 (s)	

Spectra were obtained in D_2O ; δ in ppm, reference TSPD₄, T = 300 K. ax—axial, e—equatorial.

than that of 20-hydroxyecdysone (ED₅₀ = 7.5×10^{-9} M; [16]), makisterone A $(24\beta$ -methyl-20-hydroxyecdysone; $ED_{50} = 1.3 \times 10^{-8}$ M), 24-epimakisterone A (24α-methyl-20-hydroxyecdysone; $ED_{50} = 2.2 \times 10^{-7}$ [17], M) 24(241)-dehydromakisterone A (ED₅₀ = 7.2×10^{-8} M) or mak- $(24\beta$ -ethyl-20-hydroxyecdysone; isterone C $ED_{50} = 2.0 \times 10^{-7}$ M). The absence of a C-25 hydroxyl group is associated with a significantly higher biological activity (e.g. 20-hydroxyecdysone vs A [25-deoxy-20-hydroxyecdysone; ponasterone $ED_{50} = 3.1 \times 10^{-10}$ M]) [16–18]. By deduction from the bioassay data presented above, it seems probable that the presence of an ethyl group at C-24, the introduction of a 24(24¹)[Z]-double bond and hydroxylation at C-24² each contributes significantly to the 1700-fold lower activity of 1 relative to ponasterone A.

EXPERIMENTAL

General. The UV spectrum was obtained in MeOH. CIMS was obtained on a Riber 10-10B apparatus

(Nermag S.A.) in chemical desorption mode with NH_3 as the reagent gas. HPLC sepn was performed on Gilson equipment consisting of two model 303 pumps, a holochrome detector (set at 242 nm) and controlled by the Gradient Manager program.

NMR spectroscopy. The experiments were run at 500 MHz for ¹H, at 300 K, on a Bruker AMX 500 Spectrometer equipped with a Silicon Graphics workstation. Presaturation of the solvent was used for all the 1D and homonuclear 2D ¹H experiments. The sample was lyophilised twice and dissolved in D₂O. The errors on the chemical shifts are 0.01 ppm for ¹H and 0.1 ppm for ¹³C. TSPD4, 3-(trimethylsilyl)[2,2,3,3 d_4 propionic acid (sodium salt), was used as internal reference for the proton and carbon shifts. A degassed sample was used for the ¹H NOESY experiments using a 2D phase-sensitive method (States-TPPI). ¹H NOESY experiments in D₂O were performed using a mixing time of 500 ms. Zero quantum coherence was suppressed by incorporation of a randomly positioned 180° pulse during the mixing time [19]. FIDs were acquired (32 scans) over 5102 Hz into a 2K data block for 256 incremental values of the evolution time and a relaxation delay of 2 s. The raw data were zero filled to a $2K \times 2K$ matrix. Water suppression was

Table 2. ¹³C NMR data of 20-hydroxyecdysone and compound 1

	*			
		20-Hydroxyecdysone	1	
C-1	CH_2	36.4	36.4	
C-2	CH	68.3	68.3	
C-3	CH	68.2	68.2	
C-4	CH_2	32.3	32.3	
C-5	CH	51.4	51.4	
C-6	C	209.3	209.3	
C-7	CH	122.1	122.1	
C-8	C			
C-9	CH	34.9	34.9	
C-10	C	39.1	39.1	
C-11	CH_2	21.0	21.0	
C-12	CH_2	32.0	32.0	
C-13	C	48.3	48.5	
C-14	C	86.1	86.3	
C-15	CH_2	31.2	31.2	
C-16	CH_2	21.0	21.0	
C-17	CH	50.2	50.2	
C-18	CH_3	18.0	18.1	
C-19	CH_3	24.2	24.2	
C-20	CH	79.4	79.0	
C-21	CH_3	20.6	20.6	
C-22	CH	78.4	75.7	
C-23	CH_2	27.0	34.7	
C-24	CH_2	41.6	147.4 (C)	
C-25	C	72.6	30.4 (CH)	
C-26	CH_3	28.3	21.7	
C-27	CH_3	29.3	22.2	
C-28	CH	_	124.3	
C-29	CH_2		58.7	

Spectra were obtained in $D_2O.~\delta$ in ppm, reference $TPSD_4, \\ T=300~K.$

performed by a low power transmitter pulse of presaturation (70 dB) during relaxation delay and mixing time. One half-sinusoid (5% truncated) shape homospoil gradients of 10 G cm⁻¹ were used during mixing time.

Extraction. Air-dried seeds of Leuzea carthamoides (1.2 kg) were milled and then extracted with MeOH. The combined extracts were evapd under vacuum at 40-45° to a vol. of 250 ml and this was then diluted with 375 ml H₂O. After extraction of the hydrophobic compounds by partitioning against hexane, the phytoecdysteroids were extracted into BuOH. The BuOH was removed under vacuum to give 30.4 g of crude material. After the isolation of known ecdysteroids, the frs containing 24(24¹)-dehydroamarasterone B (43.6 mg) were separated by RP-HPLC (Spherisorb ODS-2, 250 × 4.6 mm i.d., eluted with MeOH-H₂O [1:1] at 1 ml min⁻¹). Further purification was achieved on a silicic acid column (30 cm × 10 mm i.d.), eluted with CHCl₃-MeOH (9:1), yielding 5.3 mg of 1 (overall yield = 0.00004%).

Bioassay. The biological potency of the purified ecdysteroid was determined with a microplate-based bioassay using the *Drosophila melanogaster* B_{II} cell line [16].

 $24(24^{1})[Z]$ -Dehydroamarasterone B (1). Amorphous UV λ_{max} nm: 243 nm. ^{1}H NMR (D₂O); see Table 1. ^{13}C NMR (D₂O); see Table 2. CIMS m/z: 524 [M+H+NH₃]⁺, 507 [M+H]⁺, 489 [M+H-H₂O]⁺, 471 [M+H-2H₂O]⁺, 453 [M+H-3H₂O]⁺, 380 [524-{C-22-C-24}]⁺, 363 [M+H-{C-22-C-24}]⁺, 345.

Acknowledgements—The Royal Society, London, is thanked for the award of a Kapitza Fellowship to U.A.B. The technical assistance of Pensri Whiting is gratefully acknowledged.

REFERENCES

1. Yakubova, M. R. and Sakharova, N. A., Rastitelnye Resursy, 1980, 16, 98.

- 2. Vereskovskii, V. V., Chekalinskaya, I. I. and Pashina, G. V., Rastitelnye Resursy, 1985, 19, 60.
- 3. Baltaev, U. A. and Abubakirov, N. K., Khimiya Prirodnykh Soedinenii, 1987, 681.
- Girault, J.-P., Lafont, R., Varga, E., Hajdu, Z., Herke, I. and Szendrei, K., *Phytochemistry*, 1988, 27, 737.
- 5. Varga, E., Szendrei, K., Hajdu, Z., Hornok, L. and Csaki, G., Herba Hungarica, 1986, 25, 115.
- Piš, J., Budesinsky, M., Vokac, K., Laudova, V. and Harmatha, J., Phytochemistry, 1994, 37, 707.
- Lafont, R., Bouthier, A. and Wilson, I. D., in Insect Chemical Ecology, ed. I. Hrdy. Academia Prague and SPB Academic Publishers, The Hague, 1991, p. 197.
- 8. Baltaev, U. A., Khimiya Prirodnykh Soedinenii, 1991, 806.
- Baltaev, U. A., Khimiya Prirodnykh Soedinenii, 1992, 231.
- 10. Baltaev, U. A., Phytochemistry, 1995, 38, 799.
- Girault, J. P. and Lafont, R., Journal of Insect Physiology, 1988, 34, 701.
- Bax, A. and Davis, D. G., Journal of Magnetic Resonance, 1985, 65, 355.
- Baltaev, U., Gorovits, M. B., Rashkes Y. V. and Abubakirov, N. K., Khimiya Prirodnykh Soedinenii, 1978, 463.
- Neuhaus, D. and Williamson, M. P., in The Nuclear Overhauser Effect in Structural and Conformational Analysis. VCH, 1989.
- 15. Hurd, R. E. and John, B. K., Journal of Magnetic Resonance, 1991, 91, 648.
- Clément, C. Y., Bradbrook, D. A., Lafont, R. and Dinan, L., Insect Biochemistry and Molecular Biology, 1993, 23, 187.
- 17. Harmatha, J. and Dinan, L., Archives of Insect Biochemistry and Physiology, 1997, 35, 219.
- Dinan, L., in Ecdysone: from Chemistry to Mode of Action, ed. J. Koolman. Thieme, Stuttgart, 1989, p. 345.
- Rance, M., Bodenhausen, G., Wagner, G., Wuthrich, K. and Ernst, R. R., Journal of Magnetic Resonance, 1985, 62, 497.