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PYRROLIZIDINE ALKALOIDS OF *IPOMOEA HEDERIFOLIA* AND RELATED SPECIES*

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Key Word Index—*Ipomoea hederifolia*; *I. neei*; *I. × perigrenium*; *I. quamoclit*; Convolvulaceae; pyrrolizidine alkaloids; platynecine derivatives; ipangulines.

Abstract—The common necine base of the novel pyrrolizidine alkaloids (ipangulines) isolated recently from *Ipomoea hederifolia* is not turneforcidine but its 1β -epimer, platynecine. Three novel ipangulines were isolated from the aerial parts of *I. hederifolia* and identified as 9-*O*-[2-hydroxy-3-(2-acetoxy)-2-methylbutyryl]-7-*O*-salicyloylplatynecine (ipanguline B_2), 9-*O*-[2-hydroxy-3-(2-methylbutyryloxy)-2-methylbutyryl]-platynecine (ipanguline D_{10}) and 9-*O*-salicyloylplatynecine (ipanguline D_{11}). Comprehensive GC-mass spectrometric analysis also revealed a great variety of > 38 platynecine monoesters and diesters. They are classified as ipangulines A (diesters with one phenylacetic acid moiety), ipangulines B (diesters with one salicylic acid moiety), ipangulines C (diesters with two aliphatic moieties) and ipangulines D (either 7- or 9-monoesters). In shoot tips and young leaves, total ipangulines concentrations are up to 0.45% (on dry wt basis). The ipangulines are stored as tertiary alkaloids and not as *N*-oxides. Seeds of different provenance showed similar alkaloid patterns. Taxonomically related species, such as *I. neei* and *I.* × *perigrenium*, also contain platynecine esters in their vegetative tissues. © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

The Convolvulaceae is a large family with almost 2000 species, which are distributed predominantly in the tropical areas of the world. They are known to produce a wide variety of alkaloids, such as ergolines [1, 2], pyrrolidines (e.g., hygrine and cuscohygrine) and the closely related tropanes [3–5]. Some *Ipomoea* species contain unique indolizidine alkaloids of the ipalbidine-type [6] and serotonin-hydroxycinnamic acid conjugates of the ipobscurine-type [7]. Recently, 1-hydroxymethylpyrrolizidines (pyrrolizidine alkaloids, PAs) and 1-aminopyrrolizidines (loline alkaloids) were detected in the tropical bindweed, *Ipomoea hederifolia* [8], and *Argyreia mollis* [9], respectively. These were the first reports of PAs and lolines within the Convolvulaceae.

The PAs found in *I. hederifolia* so far [8], represent a unique type of PAs, i.e. diesters of a necine base esterified to one aliphatic acid and one arylalkylic or arylic acid; they were named ipangulines. Among some 360 naturally occurring PAs, aromatic necic acids are only known from the orchid alkaloids (phalaenopsine-type) [10] but these are exclusive monoesters. To characterise this new type of PA in more detail further alkaloids were isolated from *I. hederifolia* and a comprehensive GC-mass spectrometric analysis was performed to detect the whole variety of structures and minor components.

RESULTS AND DISCUSSION

Identification of platynecine and isolation of novel ipangulines

Analysis of the necine base obtained after hydrolysis of the total PA fraction from *I. hederifolia* revealed a single necine base with an *RI* of 1450, identical with that of authentic platynecine. Since turneforcidine, the 1α -epimer of platynecine, was originally described as

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Fig. 1. Structures of novel alkaloids isolated from *Ipomoea hederifolia*.

the necine base of ipangulines [8], the identity of the necine base was reinvestigated. Platynecine was synthesised by catalytic hydrogenation of retronecine [11]. Comparison of its spectroscopic properties with those of the necine moiety of the ipangulines showed that both compounds were identical.

From dried epigeal part of *I. hederifolia*, three novel PAs could be isolated and their structures elucidated by mass, ${}^{1}H$ NMR and ${}^{1}H$, ${}^{1}H$ COSY spectroscopy. Two of these alkaloids, ipanguline B_2 (32) and ipanguline D_{10} (18), were also detected by GC-mass spectrometry. The third one, ipanguline D_{11} (39), which is the first platynecine monoester with an aryl residue (i.e. salicylic acid) esterified to the C-9 hydroxyl group (Fig. 1), could not be detected by GC-mass spectrometry because we used material from different development stages for isolation.

The EI mass spectrum of 32 showed a [M]⁺ at m/z 435, corresponding to a molecular formula of $C_{22}H_{29}NO_8$. Compared with the values of ipanguline B_1 (28) this suggested an additional acetyl moiety. The ¹H NMR data (Table 1) showed a downfield shift of H-3" (δ 5.09, 1H, q, 6.5 Hz), suggesting that the acetyl moiety is esterified with the hydroxyl at C-3" of the 2,3-dihydroxy-2-methylbutyric acid. The chemical shifts of H-4" and H-5" were in agreement with an erythro-orientation of the hydroxyl groups, as in ipan-

guline B₁ [8, 12]. Thus, 32 is the 3"-acetyl derivative of ipanguline B₁. The other isolated fraction turned out to be a 3:1 mixture of two closely related alkaloids. Ipanguline D_{11} (39) showed a [M]⁺ at m/z 277, corresponding to a molecular formula of C₁₅H₁₉NO₄. The high intensity of the fragment at m/z 95, together with peaks at m/z 139 and 82, suggested the compound to be a 9-monoester of a saturated 7-hydroxy-1-hydroxymethylpyrrolizidine [13]. This was confirmed by the ¹H NMR data (Table 1) showing an upfield shift of H-7 in comparison to the other ipangulines. In the aromatic region, the typical pattern for a salicylic acid moiety could be observed, whereas there were no signals corresponding to a 2,3-dihydroxy-2-methylbutyric acid. Thus, 39 is 9-O-salicyloylplatynecine. Ipanguline D₁₀ (18) exhibited similar NMR data (Table 1) for the necine moiety. Instead of salicylic acid, a 2,3-dihydroxy-2-methylbutyric acid moiety could be deduced from the 'H NMR. Because the q for H-3' (δ 5.16, 1H, 6.5 Hz) showed a downfield shift in comparison to ipanguline A₁, esterification of the C-3' hydroxyl is indicated. In addition, typical signals for a 2-methylbutyric acid could be observed: δ 0.9 $(3H, t, 7.0 \text{ Hz}, H-4''), \delta 1.12 (3H, d, 7.0 \text{ Hz}, H-5''), \delta$ 1.65 (2H, m, $2 \times \text{H-3}''$) and δ 2.30 (1H, m, H-2''). This is in agreement with the mass spectral data. Ipanguline D_{10} is characterised by a [M]⁺ at m/z 357, corresponding to a molecular formula of C₁₈H₃₁NO₆, which is in accordance with 9-O-[2-hydroxy-3-(2methylbutyryloxy)2-methylbutyryl]-platynecine (18).

Analysis of alkaloids in shoots

Shoots of I. hederifolia grown in the greenhouse were separated into stems, shoot tips, including very young leaves, and mature leaves. The three samples were subjected to a rigorous analysis by GC-mass spectrometry [14]. This revealed a great diversity of alkaloids, all derivatives of platynecine (Table 2). Only platynecine derivatives which on EI or CI mass spectrometry show a clear $[M]^+$ or $[M+H]^+$ are included. i.e. a total of 38 compounds from I. hederifolia and an additional eight alkaloids (i.e., 40-47) from I. neei and $I. \times perigrenium cv.$ Cardinal, respectively. The great variety of structures is brought about by esterification of platynecine to various aliphatic or aromatic necic acids and the formation of various combinations of monoesters or diesters. Since the PAs originally isolated from I. hederifolia were named ipangulines [8], we enlarged this classification to distinguish between the various types of compounds (Fig. 2). Ipangulines A comprise diesters esterified with phenylacetic acid at the C-7 hydroxyl and an aliphatic acid at the C-9 hydroxyl; ipangulines B are the respective diesters containing a salicyloyl residue at the C-7 hydroxyl; the ipangulines C are diesters with two aliphatic necic acids and the ipangulines D are either 7- or 9-monoesters. Structures with still ambiguous necic acid moieties were classified as ipanguline X. To identify the structures of the most prominent necic

Table 1. 1 H NMR data of platynecine and compounds 18, 32 and 39 [J (Hz) in parentheses]

	Platynecine	18	32	39
Н	(CD₃OD)	(CDCl ₃)	(CDCl ₃)	(CDCl ₃)
1	2.62 tt (6.0, 12.0)	2.65 m	2.60-2.92 m*	2.81-2.96 m*
2a	2.05 m	1.78–2.10 m*	2.00-2.40 m*	1.78-2.10 m*
2b	2.24 dq-like (9.0, 12.0)	1.78-2.10 m*	2.00-2.40 m*	1.78~2.10 <i>m</i> *
3a	3.34 m*	2.81–2.96 m*	2.60-2.92 m*	2.81-2.96 m*
3b	3.58 dt (7.0, 12.0)	3.18 ddd (3.0, 8.5, 11.0)	3.03 dt (2.0, 8.5)	3.18 ddd (3.0, 8.5, 11.0)
5a	3.34 <i>m</i> *	2.81-2.96 m*	2.60-2.92 m*	2.81-2.96 m*
5b	3.86 ddd (2.0, 8.0, 12.0)	3.35 br t (7.0)	3.16 m	$3.35 \ br \ t \ (7.0)$
6	2.10 m	1.78-2.10 m*	2.00- 2.40 m*	1.78-2.10 m*
7	4.55 br s	4.36 br s	5.60 br s	4.33 br s
8	4.01 m*	3.46 dd (3.5, 8.0)	3.91 dd (3.0, 7.5)	3.49 dd (3.5, 8.0)
9a	3.92 dd (6.0, 11.0)	4.37 dd (8.0, 11.0)	4.36 dd (7.0, 11.0)	4.71 dd (8.0, 11.0)
9b	4.01 m*	4.60 dd (8.0, 11.0)	4.39 dd (7.0, 11.0)	4.86 dd (8.0, 11.0)
3′	and the control of th	5.16 q (6.5)	7.05 dd (1.5, 7.5)	7.00 dd (2.0, 8.0)
1 ′	_	1.32 d(6.5)	7.53 dt (1.5, 7.5)	7.45 dt (2.0, 8.0)
5′	and the second	1.40 s	6.90 dt (1.5, 7.5)	6.89 dt (2.0, 8.0)
5'	_		7.68 dd (1.5, 7.5)	7.83 dd (2.0, 8.0)
2"		2.30 m	· ·	
3″	_	1.65 m	$5.09 \ q \ (6.5)$	1 400 004
4″	_	0.90 t (7.0)	1.21 d(6.5)	
5"		1.12 d(7.0)	1.42 s	_manus
2‴	al addition		2.09 s	

^{*} Overlapping signals.

acids, which in most cases was not possible by GCmass spectrometry of the ester alkaloids, the methyl esters of the necic acids were prepared from prepurified total PA extracts of I. hederifolia. This was achieved by transesterification with trimethylsulfoniumhydroxide (TMSH) [15, 16] and direct analysis by GC-mass spectrometry (Table 3). The most characteristic necic acids, i.e. 2-methylbutyric acid, erythro-2,3-dihydroxy-2-methylbutyric acid (e-HMBA) and its threo-epimer (t-HMBA), as well as phenylacetic acid and salicylic acid could be identified unequivocally as methyl esters and, subsequently, were localised as ester components of the PAs (Table 2). The structures of some minor platynecine esters (ipangulines $X_{1.5}$) remain obscure, since their necic acids could not be identified by the methods applied. Erythro- and threo-2,3-dihydroxy-2-methylbutyric acid were found in ca equal amounts in the methyl ester fraction. As platynecine esters, the two epimers represent the respective ipangulines (erythro-HMBA) and isoipangulines (threo-HMBA) [8]. The ipanguline/ isoipanguline pairs are separated by GC. The RI values are very close in case of the unsubstituted diesters (e.g., 16/17, 25/26 and 27/28) but the separation is much better if the HMBA moiety is further acylated (e.g., pairs 9/10, 20/21, 29/30, and 31/32). Separation of the isoipanguline/ipanguline pairs 25/26, 27/28, and 31/32, which in the case of 26, 27/28 and 32 were available as reference compounds. revealed, in all cases, a lower RI for the isoipangulines. From this, we deduce that necine esters with the threoepimer of HMBA leave the column before the respective *erythro*-epimer. Where only one HMBA-ester was detected, differentiation between the HMBA-epimers was not possible (Table 2).

The major PAs found in I. hederifolia are the typical ipangulines A (i.e., 25/26, 29, 33/34, and 36) and B (i.e., 27/28 and 35) (Table 4). Only a few ipangulines C (i.e., 16/17, 20, and 22) and D (i.e., 14) are present in reasonable amounts, the great majority of structures are detectable in trace amounts (<1% of total PAs) only. Differences in the PA patterns exist between stems and leaves; whereas 27/28 are the dominating alkaloids in stems, 29 and 33/34 are major PAs in leaves and shoot tips. The highest total alkaloid concentrations are found in young leaves and shoot tips, being ca two-fold higher than those in mature leaves and more than seven-fold those in stems (Table 4). With PA levels up to 0.45% of dry weight the alkaloid concentrations are within the range of typical PA plants, such as Senecio species [17, 18]. Most interestingly, the ipangulines are largely, if not exclusively, present as free bases and not in the form of their Noxides, as in species of the Asteraceae, Boraginaceae and Fabaceae [10]. This may be related to the fact that the tertiary platynecine derivatives do not necessarily need to be stored as nontoxic N-oxides, since they are not potentially mutagenic and cytotoxic, like the 1,2unsaturated necine derivatives (e.g., retronecine derivatives) [19-21].

Analysis of alkaloids in seeds and of related species

Ipomoea hederifolia seeds of different provenance were compared in their alkaloid patterns (Table 5).

Table 2. GC-MS analysis of the pyrrolizidine alkaloids from shoots of Ipomoea hederifolia (1-38) shoots of I. neei (2, 3, 7, 10, 14-17, 21, 40-43) and roots of Ipomoea × perigrenium (16, 17, 44-47)

						(i:
	Alkaloid	R_1 (at O^7)	R_2 (at O°)	RI	[M] ⁺ , characteristic ions (rel. int.)	CI-MS ^a
_	Ipanguline D ₁	Н	COC,H,	1703	227 (1) 209 (10) 183 (4) 1A0 (7) 05 (05) 60 (2) (1) (1) 209 (1) 183 (4) 1A0 (7) 60 (8)	
7	7-(2-Methylbutyryl)-platynecine	MeBu	Н	1778	22) (1), 201 (10), 103 (4), 140 (7), 23 (80), 62 (100), 71 (5), 53 (8), 43 (13) 241 (0.2), 210 (0.4), 156 (34), 139 (63), 123 (5), 113 (17), 82 (100), 57 (13)	
m .	9-(2-Methylbutyryl)-platynecine	Н	McBu	1803	241 (0.6), 223 (9), 197 (2), 140 (9), 124 (6), 95 (100), 82 (80), 55 (8)	-
4	Ipanguline D ₂ ^b	Н	COC,H ₀ O?	1886		Ī,
v	9-Tigloylplatynecine	Н	Tigloyl	1898	239 (0.2) 221 (15), 195 (3), 140 (5), 123 (6.1), 23 (30), 62 (100), 73 (13), 53 (14)	7
9	Isoipanguline D ₃	H	t-HMBA	2022	273 (=) 255 (3) 212 (3); 158 (26) 140 (5); 14 (14) 05 (45) 03 (100)	Z
_	Ipanguline D ₃	Н	e-HMBA	2037	273 (=), 255 (7), 212 (14), 158 (29), 140 (24), 114 (14), 55 (46), 62 (100)	
20	Ipanguline D_4^{b}	Н	COC,H,10,7	2085	2992 (0.3), 281 (3), 255 (3), 212 (3), 140 (13), 65 (10), 62 (10)	
6	Isotpanguline D,	Н	Ac-t-HMBA	2108	315 (14) 297 (2) 271 (5) 254 (3) 278 (10) 200 (7) 158 (0) (40 (20) 65 (22) 62 (36)	;
9	Ipangulinc D _s	Н	Ac-e-HMBA	2157	315 (2), 297 (2), 271 (9), 254 (2), 228 (13), 200 (7), 158 (9), 140 (29), 95 (72), 82 (100), 43 (23)	- : ź
Ξ	Ipanguline D_{e}	PAA	Н	2168	275 (-), 156 (19) 139 (65) 124 (3) 113 (16) 01 (20), 62 (100), 62 (100), 43 (41)	ı,
12	Ipanguline D,	Н	PAA	2212	275 (1) 257 (5) 231 (8) 140 (5) 95 (6) 91 (19) 92 (100), 52 (100), 53 (1)	 ź :
13	Ipanguline $\mathbf{D}_{\mathrm{s}}^{\mathrm{b}}$	Н	COC,H,O.2	2226	3437 (7) 299 (2) 228 (3) 158 (8) 140 (40) 122 (110), 32 (110), 33 (18)	- ź
7	Ipangulinc D,º	Ή	DiAc-HMBA	2246	357 (1) 339 (1) 313 (8) 228 (6) 140 (40), 122 (11), 30 (69), 53 (62), 62 (100), 71 (40), 43 (33)	;
13	Ipanguline X ₁ ^b	ć.	į	2302	385 (-), 297 (17), 254 (27), 228 (20), 310 (34), 139 (8), 134 (7), 95 (100), 62 (91), 43 (34)	-
					(19), 43 (48)	ī.
91	Isoipanguline C ₁	MeBu	/-HMBA	2308	357 (0.1), 313 (1), 255 (8), 242 (21), 238 (16), 224 (18), 140 (100), 138 (50), 127 (57), 05 (52), 05	- 7
					(33), 82 (67), 57 (29)	- Ż
17	Ipanguline C	MeBu	e-HMBA	2311	357 (=), 312 (0.5), 255 (7), 242 (19), 238 (13), 224 (10), 140 (100), 128 (44), 123 (62), 82 (23), 85	
,	:			•	(37), 82 (79), 57 (30)	
<u>×</u>	Ipanguline D_{10}	H	MeBu-e-HMBA	2327	357 (3), 313 (3), 242 (14), 228 (16), 158 (12), 140 (44), 122 (13), 96 (100), 95 (61), 85 (30), 82	7
5	1	i			(93), 57 (41)	
5	Ipanguine X_2°	Tigloyl?	HMBA?	2376	355? (0.2), 340 (0.2), 311 (2), 255 (12), 240 (25), 238 (23), 222 (19), 140 (100), 138 (91), 122 (65).	
2	Cariffication 1	;			96 (31), 95 (36), 83 (44), 82 (77), 55 (28)	
07	tsotpanguille C2	McBu	Ac-t-HMBA	2391	399 (0.1), 313 (8), 297 (22), 254 (37), 242 (14), 224 (32), 140 (32), 138 (90), 122 (77), 95 (65), 82	z
21	Inanguline C.	Mab	A - ITAKDA	7676	(100), 57 (39), 43 (38)	
	7	100 A	AC-C-IIIIDA	0747	399 (0.1), 313 (6), 297 (18), 254 (46), 242 (13), 224 (32), 140 (32), 138 (88), 122 (76), 95 (65), 82	Ľ
22	Ipanguline C ₃ °	MeBu	DiAc-HMBA	2506	(100), 37 (41), 43 (39) 441 (f) 13 339 (20) 280 (22) 234 (22) 210 (11) 138 (100) 133 (20) 62 (62) 62 (62) 62 (62)	
					(34)	
23	Ipanguline C4°	MeBu	MeBu-HMBA	2606	441 (0.1), 339 (10), 254 (29), 242 (12), 224 (25), 138 (66) 122 (100) 96 (40) 95 (30) 85 (44) 92	- 2
	:				(53), 57 (63)	Į,
5 7	ipanguine X_3	6.	٠.	2667	439 (-), 339 (16), 280 (8), 254 (22), 240 (9), 222 (14), 210 (7), 138 (99), 122 (100), 96 (36), 95	_
					(45), 85 (19), 82 (65), 57 (27), 55 (17)	•
9	Isoipanguine A ₁	PAA	t-HMBA	2702	391 (0.1), 276 (14), 258 (16), 255 (12), 238 (25), 140 (100), 138 (60), 122 (57), 96 (28), 95 (41),	Z
36	Ingamiliae A				91 (38), 82 (95)	
3	ipangunic A ₁	PAA	e-HMBA	2707	391 (0.1), 276 (12), 258 (14), 255 (8), 238 (16), 140 (78), 138 (57), 122 (52), 96 (29), 95 (45), 91	- z
					(34), 82 (100)	

27	Isoipanguline B ₁	Sal	t-HMBA	2770	393 (0.1), 278 (16), 260 (6), 255 (18), 238 (25), 140 (100), 138 (70), 122 (47), 121 (29), 96 (20)	
ę	:				95 (26), 82 (62)	
87	Ipanguline \mathbf{B}_{l}	Sal	e-HMBA	2772	393 (0.1), 278 (13), 260 (5), 255 (15), 238 (21), 140 (100), 138 (67), 122 (46), 121 (31), 96 (21), 95 (28), 82 (64)	
29	Isoipanguline A ₂	PAA	Ac-t-HMBA	2782	433 (0.2), 297 (31), 276 (8), 258 (20), 254 (29), 140 (24), 138 (68), 122 (54), 96 (26), 95 (59), 91	ź
ş	A suffine A	ė V	4 03 411	0	(36), 82 (100), 43 (35)	
3	Panganne A2	L A A	Ac-e-HMBA	7810	433 (0.1), 297 (25), 276 (7), 258 (20), 254 (34), 140 (24), 138 (70), 122 (58), 96 (25), 95 (62), 91 (35), 82 (100), 43 (33)	- Ż
<u>.</u>	Isoipanguline B ₂	Sal	Ac-t-HMBA	2837	435 (-), 297 (42), 260 (6), 254 (36), 210 (8), 140 (75), 138 (100), 122 (64), 121 (51), 96 (27), 95	ī Ž
32	Inanguline B.	Sol	Ac a UMABA	1013	(57), 82 (98), 43 (30)	
!	rpungumin 12	J41	AC-C-HIMIDA	6/07	455 (), 297 (34), 260 (7), 254 (42), 210 (8), 140 (74), 138 (100), 122 (67), 121 (49), 96 (26), 95 (57), 82 (99), 43 (32)	_
33	Isoipanguline A,	PAA	DiAc-t-HMBA	2886	475 (0.1), 389 (1), 339 (28), 280 (21), 258 (21), 210 (8), 138 (94), 122 (65), 95 (99), 91 (28), 82	r Ž
7	Inanguline A	DAA	A CANTA A SCI	1000	(100), 43 (35)	
35	Ipanguline B.	Sal	DiAc-e-FIMBA	2691	477 (-), 339 (18), 280 (5), 258 (19), 210 (7), 138 (87), 122 (67), 95 (67), 91 (52), 82 (100), 43 (72)	;
				17/2	777 (77, 553 (24), 260 (17), 200 (5), 210 (7), 140 (52), 158 (100), 122 (60), 121 (40), 93 (69), 82 (70), 43 (21)	Z
æ	Ipanguline A4°	PAA	MeBu-HMBA	2985	475 (0.5), 339 (14), 276 (7), 258 (19), 254 (28), 140 (21), 138 (69), 122 (100), 96 (55), 95 (36), 91	ı Z
;	:				(35). 85 (40), 82 (80), 57 (55)	
31	Ipanguline X_4^8	٠.	6.	3055	497 (0.1), 339 (12), 298 (5), 280 (13), 254 (27), 141 (12), 140 (17), 138 (43), 122 (100), 96 (35),	z
Ş					95 (22), 85 (21), 82 (42), 71 (9), 57 (28)	
\$	Ipanguline X_s°	ć	٠.	3083	499 (0.1), 456 (4), 384 (5), 339 (11), 300 (2), 282 (10), 254 (25), 140 (30), 138 (39), 122 (100), 96	
9	•				(33), 95 (22), 85 (23), 82 (45), 57 (31)	
₹ ;	9-Acetylplatynecine	T,	Ac	1557	199 (1), 181 (5), 155 (5), 140 (3), 95 (56), 82 (100), 55 (10), 43 (19)	
-	Ipanguline X ₆ "	٠.	ç.	2100	315? (), 300 (0.1), 200 (21), 182 (22), 140 (50), 138 (41), 122 (45), 95 (42), 82 (100), 55 (16), 43	
4	Inanonline D. be	Ac UMBA9	11	9616	(47)	
43	ryangamo X b	AC-HIMBA:	Ц.	6717	315 (-), 228 (4), 200 (4), 156 (34), 139 (87), 122 (12), 113 (25), 82 (100), 43 (24)	
}	Ipangumic A;		·. }	/177	312? (), 297 (6), 254 (11), 182 (26), 138 (46), 122 (42), 95 (53), 82 (100), 43 (78)	
1	Ipanguine Dis	lig-HMBA?	I	2358	355 (4), 312 (1), 296 (2), 196 (15), 156 (29), 139 (100), 122 (9), 113 (18), 96 (11), 83 (46), 82	
ţ	\$ \$ 1	;			(79), 55 (27)	
ç	Ipanguline D_{14}^{***}	I	Tig-HMBA?	2375	355 (11), 337 (2), 240 (10), 228 (12), 196 (56), 140 (30), 122 (10), 96 (52), 95 (47), 83 (72), 82	
46	46 Ipanguline D ₁₅ b.c	Н	Tig-HMBA?	2387	(100); 33 (43) 355 (12): 337 (1): 240 (10): 228 (12): 196 (29): 140 (32): 122 (10): 96 (60): 95 (50): 83 (74): 82	
					(100), 55 (43)	
4	47 Ipanguline $D_{16}^{b,c}$	Tig-HMBA?	н	2395	355 (3), 240 (4), 228 (5), 196 (5), 156 (29), 139 (100), 122 (9), 113 (19), 96 (13), 83 (49), 82 (82).	
1					33 (23)	

e-HMBA = erythro-2,3-dihydroxy-2-methylbutyryl moiety; t-HMBA = threo-2,3-dihydroxy-2-methylbutyryl; Ac-HMBA = 3-acetoxy-2-hydroxy-2-methylbutyryl; DiAc-HMBA = 2,3-diacetoxy-2-methylbutyryl; MeBu = 2-methylbutyryl; PAA = phenylacetyl; Sal = salicyloyl; Tig = tigloyl.

[&]quot;Analysed by CI-MS, N = NH," I = isobutane.

According to GC-MS, a platynecine ester but identity of necic acids uncertain or tentative.

Orientation of hydroxyl groups of HMBA moiety unclear.

Fig. 2. The four structural types of the ipangulines. R = aliphatic necic acid.

Table 3. GC-MS analysis of necic acid methyl esters prepared from a purified crude extract of 1. hederifolia by transesterification with TMSH

Necic acid methyl ester	RI^a	m/z [M] ⁺	Relative intensity
2-Methylbutyric acid methyl ester	755	116	26
Tiglic acid methyl ester	855	114	2
Hexanoic acid methyl ester	915	130	1
threo-2,3-Dihydroxy-2-methylbutyric acid methyl ester	968	148	7
erythro-2,3-Dihydroxy-2-methylbutyric acid methyl ester	973	148	8
Benzoic acid methyl ester	1063	136	< 1
Octanoic acid methyl ester	1112	158	< 1
Phenylacetic acid methyl ester	1142	150	51
Salicylic acid methyl ester	1163	152	1
O-Methylsalicylic acid methyl ester ^b	1298	166	3

a DB-1 column.

Although there are some differences, the A_1 ipangulines (25/26) are always the dominating PAs. In contrast to the vegetative plant parts (see Table 4), which accumulate preferentially 25/26, further esterified with acetic acid (29, 33/34) or methylbutyric acid (36), seeds are devoid of such esters. Furthermore, the B ipangulines are absent from the seed pattern.

Within the Convolvulaceae, *I. hederifolia* belongs taxonomically to the sectio Quamoclit [22]. Thus, two additional species, i.e. *I. neei* and *I. × perigrenium*, an assumed hybrid of *I. hederifolia* (D. F. Austin, personal communication) and presumably *I. quamoclit*, were included in a preliminary screening for PAs. No PAs were detected in seeds of *I. quamoclit* and *I. neei* but vegetative plant material available from *I. neei* and *I. × perigrenium* were found to contain PAs (Table 6). Like *I. hederifolia*, the two species contain only platynecine esters and, among the various necic acids, HMBA is most prominent. In contrast to *I. hederifolia*, aromatic necic acids (i.e., phenylacetic acid and salicylic acid) are absent from the PAs of these two species which, in turn, produce some PAs not found

in *I. hederifolia*, i.e. **40–47** (Tables 2 and 6). More detailed studies are needed to evaluate whether the presence of ipanguline-type PAs is a general feature and, thus, a chemosystematic marker of the sectio Quamoclit within the Convolvulaceae. However, to date, ipangulines have not been detected in numerous species of the genus *Ipomoea* outside this sectio or of other convolvulaceous genera.

EXPERIMENTAL

General

 1 H NMR (300/400 MHz) and 13 C NMR (75/100 MHz) were recorded on a Bruker AC 300 or AC 400. EIMS: direct inlet system, 70 eV. HRMS: 80 eV. CC: silica gel 60 (70–230 mesh). TLC and prep. TLC: precoated silica gel plates 60 F_{254} (0.25 mm).

Plant material

Seeds of *I. hederifolia* L. (syn.: *I. angulata* Lamk.) were collected near Ella, Sri Lanka; Chiang Mai,

^bO-methylated by TMSH.

Table 4. Relative abundance and total concentrations of pyrrolizidine alkaloids in stems, shoot tips and leaves of greenhousegrown *Ipomoea hederifolia*

			PAs	s, relative abur	ndance (%)
	Alkaloid	RI	Stems	Shoot tips	Mature leaves
1	Ipanguline D ₁	1703	tr	<1	<1
2	7-(2-Methylbutyryl)-platynecine	1778	tr	< 1	< 1
3	9-(2-Methylbutyryl)-platynecine	1803	< 1	< 1	< 1
4	Ipanguline D ₂	1886		tr	
5	9-Tigloylplatynecine	1898		tr	
6	Isoipanguline D ₃	2022	< 1	< 1	<1
7	Ipanguline D ₃	2037	<1	<1	
8	Ipanguline D ₄	2085		tr	tr
9	Isoipanguline D ₅	2108	tr	< 1	< 1
0	Ipanguline D ₅	2157	< 1	< 1	< 1
1	Ipanguline D ₆	2168			< 1
2	Ipanguline D ₇	2212	< l	< 1	< 1
3	Ipanguline D ₈	2226	tr	tr	
4	Ipanguline D ₉	2246	4	1	7
5	Ipanguline X ₁	2302	tr	tr	tr
6/17	Isoipanguline/Ipanguline C ₁	2308-11	8	3	6
8	Ipanguline D ₁₀	2327	< l	< 1	<1
9	Ipanguline X ₂	2376	tr	< 1	
0	Isoipanguline C ₂	2391	2	4	7
:1	Ipanguline C ₂	2426	tr	< 1	< 1
2	Ipanguline C ₃	2506	5	3	5
3	Ipanguline C ₄	2606	tr	<1	< 1
4	Ipanguline X ₃	2667	>1	tr	tr
5/26	Isoipanguline/Ipanguline A_1	2702-7	14	13	12
27/28	Isoipanguline/Ipanguline B ₁	2770-2	21	2	1
9	Isoipanguline A ₂	2782	8	25	20
0	Ipanguline A ₂	2810	<1	< 1	< 1
1	Isoipanguline B ₂	2837	<1	1 >	< 1
32	Ipanguline B ₂	2873	tr	< 1	< 1
33/34	Isoipanguline/Ipanguline A ₃	2886-91	15	26	23
35	Ipanguline B ₃	2927	4	tr	2
6	Ipanguline A ₄	2985	5	9	9
37	Ipanguline X ₄	3055	> i	tr	tr
38	Ipanguline X ₅	3083	>1	>1	> 1
	mg g ⁻¹ (dry wt)				
	bases + N-oxides:		0.595	4.458	1.841
Tertiary 1	bases:		0.525	3.618	1.956

Thailand; Guayaquil, Ecuador; Santa Ana, Panama. Seeds of *I. quamoclit* L. were collected near San Lorenzo, Panama, and those of *I. neii* (Spreng.) O'Donell near Boquete, Panama. *Ipomoea* × *perigrenium cv.* Cardinal was purchased from Gartencenter Pluta, Berlin-Dahlem. Plants were grown from seed in the greenhouse and provided roots and shoots for alkaloid extraction. Voucher specimens are deposited at the Institut für Pharmazie II, Freie Universität Berlin.

For detailed GC-MS analysis sterile cultures of differentiated plantlets of *I. hederifolia* (origin: Sri Lanka) were established on MS medium (phytohormones omitted) [23]. Plantlets are kept as stock-cultures and used during the growing season to initiate

greenhouse plants. For PA analysis, the long shoots (1 m) were divided into (a) shoot tips (10 cm) with the youngest leaves; (b) mature leaves; (c) stems without leaves. Harvested material was immediately frozen, lyophilised and kept cold until used.

Semisynthesis of (-)-platynecine

Monocrotaline (300 mg) was refluxed with 800 mg of BaOH in $\rm H_2O$ for 1 h. The mixt. was acidified and extracted with CHCl₃. After evapn of the aq. layer, retronecine was dissolved in EtOH (60°) and hydrogenated for 2 h (Pd/C-catalyst). Evapn gave 20 mg (-)-platynecine. Colourless gum. [α] $_{\rm D}^{20}$ -51° (EtOH, c 0.9). EIMS m/z (rel. int.): 157 [M] $^+$ (13), 113 (24),

Table 5. GC-MS analysis of PA extracts from seeds of Ipomoea hederifolia of different provenances

					Relative ab	undance (%)	
	Alkaloid	RI	$[M]^+ (m/z)$	Sri Lanka	Panama	Ecuador	Thailand
	Anhydroplatynecine	1130	139	2	3	4	5
5	9-Tigloylplatynecine	1898	239		2	< 1	
11	Ipanguline D ₆	2168	275	< 1	2	2	1
12	Ipanguline D ₂	2212	275	< 1	6	6	6
14	Ipanguline D ₉	2246	357	< 1	<1		
17	Ipanguline C ₁	2308	357	< 1	2	9	4
19	Ipanguline X	2376	355?	< 1	31	6	< 1
25/26	Isoipanguline/Ipanguline A ₁ ^a	2702-7	391	97	54	71	80
Total P.	As, mg g ⁻¹ (dry wt)				0.15	0.16	0.28

^a Two epimers poorly resolved.

Table 6. Pyrrolizidine alkaloids in two additional species of the sectio Quamoclit (Convolvulaceae) compared with *Ipomoea hederifolia*

	Alkaloid	RI	$[M]^+$ (m/z)	Ipomoea neei (shoots)	Ipomoea × perigrenium (roots)	Ipomoea hederifolia (whole plant)
40	9-Acetylplatynecine	1557	199	+		
2	7-(2-Methylbutyryl)-platynecine	1778	241	+		+
3	9-(2-Methylbutyryl)-platynecine	1803	241	+		+
7	Ipanguline D ₃	2037	273	+		+
41	Ipanguline X ₆	2100	315?	+		
42	Ipanguline D ₁₂	2125	315	+		
10	Ipanguline D ₅	2157	315?	+		+
43	Ipanguline X ₇	2217	315	+		
14	Ipanguline D ₉	2246	357	+		+
16	Isoipanguline C ₁	2308	357	+	+	+
17	Ipanguline C ₁	2311	357	+	+	+
44	Ipanguline D ₁₃	2358	355		+	
45	Isoipanguline D ₁₄	2375	355		+	
46	Ipanguline X ₈	2387	355		+	
47	Ipanguline D ₁₅	2395	355		+	
21	Ipanguline C	2426	399	+		+

82 (100). ¹H NMR (Table 1). ¹³C NMR (MeOH): δ 28.9 (t, C-2), 37.1 (t, C-6), 44.4 (d, C-1), 55.3 (t, C-5), 56.3 (t, C-3), 60.7 (t, C-9), 72.0 (d, C-7), 74.4 (d, C-8), identical with data in Ref. [24].

Isolation of alkaloids

Aerial parts of *I. hederifolia* (2 kg) were extracted with 201 of MeOH (70%). After evapn, the extract was acidified and partitioned between H₂O and organic solvents. The aq. layer was then made alkaline and extracted with CHCl₃ to yield a crude alkaloid mixt., which was fractionated on silica gel [MeOH–CHCl₃–NH₃ (33%)-mixts]. Frs 18–29 were further purified by prep. TLC (CHCl₃–MeOH, 4:1) to give ipanguline B₂ (32). Ipanguline D₁₀ (18) and ipanguline D₁₁ (39) were

isolated from frs 68-80, again using prep. TLC in CHCl₃-MeOH-NH₃ (33%) (75:23:2).

Ipanguline B_2 (32)

EIMS m/z (rel. int.): 435 [M]⁺ (0.1), 391 (4), 320 (8), 297 (60), 254 (34), 210 (7), 140 (47), 138 (70), 122 (47), 95 (48), 82 (100); HRMS, m/z: 435.1891 ($C_{22}H_{29}NO_8$, calc. 435.1893), 391.1634 ($C_{20}H_{25}NO_7$, calc. 391.1631), 297.1575 ($C_{15}H_{23}NO_5$, calc. 297.1576). ¹H NMR (Table 1).

Ipanguline D_{10} (18)

EIMS *m/z* (rel. int.): 357 [M]⁺ (3), 313 (6), 242 (4), 228 (6), 158 (3), 140 (23), 139 (30)*, 96 (34), 95 (86).

^{*} Due to an impurity with a 7-monoester.

82 (100); HRMS, m/z: 357.2153 ($C_{18}H_{31}NO_6$, calc. 357.2151). ¹H NMR (Table 1).

Ipanguline D_{11} (39)

EIMS m/z (rel. int.): 277 [M]⁺ (13), 259 (7), 233 (8), 140 (23), 139 (30)*, 95 (86), 82 (100); HRMS, m/z: 277.1313 (C₁₅H₁₉NO₄, calc. 277.1314). ¹H NMR (Table 1).

Alkaloid extraction for GC and GC-MS [14]

Ground lyophilised plant material (i.e., shoot tips, mature leaves and stems), 0.25 g each, was extracted twice in a mortar with 2 ml 1 M HCl. After centrifugation, a prepurified total alkaloid fr. was obtained from the supernatant by liquid—liquid extraction using silica earth columns (Extrelut, Merck) [14]. Samples were stored in MeOH prior to analysis. In expts where PA N-oxides were expected, the acidic aq. extracts were reduced by adding Zn dust in excess and left to stand for 5 h, prior to liquid—liquid extraction.

Preparation of necic acid methyl esters by transesterification [15, 16]

A 0.2 M soln of trimethylsulfoniumhydroxide (TMSH, Macherey & Nagel, Düren) $(20 \,\mu\text{l})$ was added to $10 \,\mu\text{l}$ MeOH containing $10 \,\mu\text{g}$ alkaloids. The mixt. was directly injected into the GC-MS-instrument.

GC-MS analysis [14]

A GC equipped with a 30 m \times 0.32 mm fused-silica capillary column (DB-1, J & W Scientific) was used. Conditions: injector 250°; split ratio 1:20; temp. programme 70–300°, 6° min⁻¹; carrier gas He 0.5 bar. The capillary column was directly coupled to the quadrupole mass spectrometer. EI-mass spectra were recorded at 40 eV. CI mass spectra were recorded with the same GC-MS system using NH₃ or *iso* butane as reagent gas, at 100 eV. Conditions for GC-MS analysis of the necic acids were the same as those given for the alkaloids but with a temp. programme 30–200°, 6° min⁻¹. Retention indices (*RI*): Kovats indices [10, 25] were calculated with reference to a set of coinjected hydrocarbons.

GC analysis [14]

PAs were separated on a 15 m \times 0.25 mm fusedsilica capillary column (DB-1, J & W Scientific). Conditions: injector 250°, split ratio 1:20, temp. programme 150–300°, 6° min⁻¹, carrier gas He 0.75 bar, detector: dual FID, PND. Acknowledgements—The authors are indebted to Mrs E. Bäumel-Eich (Berlin) for essential support in exploring and collecting the plant material and to Mrs C. Theuring (Braunschweig) for skilful technical assistance. The work was supported by grants from the Deutsche Forschungsgemeinschaft and Fonds der Chemischen Industrie to T.H.

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^{*} Due to an impurity with a 7-monoester.

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