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NORNEOLIGNAN AND PHENOLS FROM *CURCULIGO CAPITULATA*

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Key Word Index—*Curculigo capitulata*; Amaryllidaceae; rhizome; 2,4-dichloro-5-methoxy-3-methyl-phenol; curlignan; 4-ethoxy-3-hydroxymethyl-phenol.

Abstract—Nine compounds were isolated from the chloroform-soluble fraction of *Curculigo capitulata*. Of these, 2,4-dichloro-5-methoxy-3-methylphenol, curlignan and 4-ethoxy-3-hydroxymethylphenol are new natural products. Their structures were elucidated mainly based on spectral analysis. © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

Recent chemical investigation of *Curculigo capitulata* (Lour.) O. Kuntze alias *C. recurvata*, has led to the isolation of a series of norlignan glycosides [1–4] and curcapitoside [3], a novel glucosyl-fused phenanthropyran, from the polar fraction. Three of the norlignan glucosides, nyasicoside, 1*S-O*-butyl- and 1*R-O*-butyl-nyasicosides, were found to possess anti-arrhythmic properties [4]. Being interested in such biological activity, we attempted to obtain potential norlignan aglycones from the less polar fraction of this species, which has not been explored yet. Here, we report the outcome of this study.

RESULTS AND DISCUSSION

The chloroform soluble fraction of the ethanol extract of the rhizome was subjected to repeated column chromatography and yielded nine compounds. Six of them were identified as vanillin, 4-hydroxybenzaldehyde [5], ethyl protocatechuate [6], orcinol-1-O- β -D-glucoside [7], 2,6-dimethoxy-benzoic acid [8] and β -sitosterol 3-O- β -D-glucoside [9]. Three of them, 2,4-dichloro-5-methoxy-3-methylphenol (1), curlignan (2) and 4-ethoxy-3-hydroxy-methylphenol (3), were found to be novel natural products. Their structures were elucidated as follows.

Compound 1 was identified as 2,4-dichloro-5-methoxy-3-methylphenol by comparison of its EI

Compound 2, a yellowish viscous liquid, $[\alpha]_D^{25}$ +40°C (c 0.5, MeOH), had a molecular formula C₁₉H₂₀O₇ (HR EI mass spectrometry). It is also phenolic as exemplified by the IR absorption at 3400 cm⁻¹ and the bathochromic shift in the UV spectrum when measured in alkaline conditions. Its ¹H NMR spectrum (Me₂CO-d₆) revealed an ABXpattern and an AB-pattern for the five aromatic protons, three MeO singlets, a double triplet (1H, δ 3.62) coupled to a doublet (1H, δ 5.65) and a multiplet (2H, δ 3.84), as identified in a COSY-45 spectrum. The ¹H NMR spectrum of its acetylated product (2a) revealed two additional singlets for acetyl groups (δ 2.28 and 2.03) and downfield shifted signals for the oxygenated methylene protons from δ 3.84 (2H, m) to δ 4.45 (1H, dd, J = 11.3, 5.4 Hz) and δ 4.30 (1H, dd, J = 11.3, 7.5 Hz), suggesting the presence of a hydoxymethyl group in 2. The ¹³C NMR data revealed that 2 contained two aryl groups (four oxygenated and three

mass and 1 H NMR spectral data [four singlets at δ 6.52 (1H), 5.59 (1H, D₂O exchangeable), 3.83 (3H, OMe) and 2.44 (3H, Ph–Me)] with reported data [10]. It had been reported as the acid hydrolysis product of curculigine A, isolated from the related plant species, *C. orchioides* [10], but it is the first natural occurrence of this compound. Its structure was confirmed by two NOE experiments which indicated that the aryl proton signal (δ 6.52) was enhanced by irradiation at the methoxyl singlet (δ 3.83) but not at the aryl methyl singlet (δ 2.44).

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CH₃ Cl
$$\frac{3}{3}$$
 Cl $\frac{4}{3}$ CH₂OH

1 3

CH₂OH

OCH₃

OCH₃

OCH₃

OCH₃

OCH₃

A R= CO₂CH₃

4 R= CH=CH-CH₂OH, trans

nonoxygenated quaternary carbons and five methines), one conjugated carboxylate (δ 167.0, s), two oxygenated aliphatic carbons (δ 89.5, d; δ 64.2, t), one aliphatic methine (δ 54.1, d) and three methoxy, one of which is an ester methyl (δ 52.0, q). These data pooled together would suggest **2** to be a 2-phenyl benzofuran.

This proposed skeleton was confirmed by NOE studies of 2 and its peracetate 2a (Fig. 1). Among these, irradiation at the H-2 doublet (δ 5.65) or H-3 double triplet (δ 3.62) caused the enhancement of the corresponding H-2' and H-6' signals. A chemical model study of 2 indicated that these two NOE results could be possible only if H-2 and H-3 are trans-oriented. The absolute stereochemistry at C-2 and C-3 of 2 was determined to 2S,3R by the comparison of its CD curve with that of 2S,3R-(+)dehydro-diconiferyl alcohol (4) [11], both showing one negative and one positive Cotton effect around 230 and 280 nm, respectively. These data established the structure of 2 as depicted in Fig. 1. This novel natural product is named curlignan after its plant origin and skeleton similar to neolignan. Curlignan (2) could be produced biogenetically by an oxidative coupling of two coniferyl alcohol, followed by oxidative degradation of the side-chain at C-5.

The assigned structure for **2** was also confirmed by the analysis of the HMBC spectrum (Table 1). Critical correlations for the ring skeleton include the couplings of H-2 to C-2', C-6' and C-9, and H-3 to C-3a and C-1'. The unambiguous ¹H and ¹³C NMR assignments for **2** are listed in Table 1 from the analysis of NOE, HMQC and HMBC spectra. The ¹H NMR spectrum measured in CDCl₃ was also assigned and is listed in the Experimental

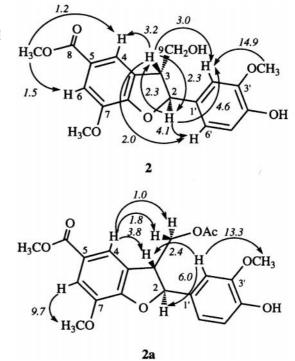


Fig. 1. NOEs (italics, %) of $\mathbf{2}$ (Me₂CO- d_6) and $\mathbf{2a}$ (CDCl₃).

section for reference, although it was not so well-resolved as that measured in Me_2CO-d_6 .

Compound 3, a yellowish viscous liquid, had a molecular formula $C_9H_{12}O_3$ as deduced from EI mass spectrometry and NMR data. It is phenolic as exemplified by the IR absorption at 3300 cm⁻¹ and the bathochromic shift in the UV spectrum when measured in alkaline conditions. Its 1H NMR spectrum revealed an ABX-pattern for the aromatic protons, signals for an ethoxyl group and a two-

Table 1. 13 C and 1 H NMR (δ , mult.) (Me₂CO- d_6) and HMBC data for compound **2**

Position	¹³ C	¹ H (<i>J</i> in Hz)	HMBC ($J = 8 \text{ Hz}$) correlated C (#)
2	89.5 d	5.65 d (6.7)	2′,6′,9
3	54.1 d	3.62 dt (6.1, 6.7)	3a, 1'
3a	130.6 s		
4	119.8 d	7.61 d (1.3)	6, 7a, 8
5	124.1 s		
6	114.6 d	$7.50 \ d \ (1.3)$	4, 7a, 8
7	144.9 s		
7a	153.6 s		
8	167.0 s		
9	64.2 t	$3.84 \ m$	
1'	133.6 s		
2'	110.6 d	$7.03 \ d \ (1.2)$	2, 4', 6'
3'	148.5 s		
4'	147.6 s		
5'	115.7 d	6.81 d (8.1)	1', 3'
6'	120.2 d	6.88 dd (1.2, 8.1)	2, 2', 4'
7-OMe	56.3 q	3.88 s	7
8-OMe	52.0 q	3.83 s	8
3'-Ome	56.4 q	3.81 s	3'

proton singlet (δ 4.60) corresponding to an oxygenated benzylic methylene. These data indicated 3 to be a 1,2,4-trisubstituted benzene. The location of these substituents was determined by NOE experiments. Irradiation at the methylene singlet enhanced the *meta*-coupled aromatic proton (δ 6.50, d, J=3.0 Hz), and the methylene quartet (δ 3.57, J=7.0 Hz). Irradiation at the latter quartet enhanced an *ortho*-coupled aryl proton (d, J=8.6 Hz) and the methyl triplet (δ 1.24, J=7.0 Hz), in addition to the methylene singlet. These data taken together established 3 to be 4-ethoxy-3-hydroxymethylphenol.

Besides these compounds, some cycloartenol related triterpenes, which had been found in other related plants, such as curculigol A from C. orchioides [12] and certain polyunsaturated long chain alkenes having structure similar to β -carotene were also detected. They were not further isolated and characterized because of the very limited amounts obtained.

EXPERIMENTAL

M.p.s: uncorr. ¹H NMR (400.13 MHz) and ¹³C NMR (100.61 MHz): Me₂CO-*d*₆ or CDCl₃ using the solvent peak as int. standard. MS: direct inlet system. UV: MeOH. IR: KBr disc. CD: MeOH.

Plant material

See Ref. [4].

Extraction and isolation

This part of the work is an extension study focused on the isolation of the CHCl3-sol. constituents from the rhizome of C. capitulata. The CHCl₃ fraction (13 g) from dried powdered rhizomes (5.10 kg) [4] was chromatographed on a silica gel column (70-230 mesh, 500 g), eluted with 0 to 15% MeOH in CHCl₃, to give 8 frs. Fr. 1 (36 mg) was recrystallized from CHCl₃ to give 2,4-dichloro-5methoxy-3-methylphenol (1, 5.2 mg) as needle crystals. Fr. 2 (85 mg) was rechromatographed on a silica gel column (230-400 mesh, 20 g), eluted with CHCl₃ to give vanillin (10 mg) as an amorphous powder. Repeated chromatography on silica gel columns (230-400 mesh) of fr. 4 (396 mg), eluted with 0.5% MeOH in CHCl₃, gave amorphous 2 (5.4 mg), 3 (38.2 mg) and 4-hydroxybenzaldehyde (5.0 mg). Fr. 6 (280 mg) yielded ethyl protocatechuate (25.2 mg) after purification on a silica gel column (230-400 mesh, 15 g), eluted with 1.5% MeOH in CHCl₃. Fr. 7 (946 mg) yielded 1-O-β-Dglucosyl-orcinol (30 mg), 2,6-dimethoxybenzoic acid (16 mg) and 3-O- β -D-glucosyl- β -sitosterol (40 mg) via repeated silica gel cc (230-400 mesh), eluted with 2 to 10% MeOH in CHCl₃.

Peracetylation of 2

Compound 2 (2.0 mg) was stirred with py-Ac₂O (0.5 ml, 3:1) overnight. Usual work-up yielded the peracetylated product 2a (2.2 mg).

2,4-Dichloro-5-methoxy-3-methylphenol (1)

Needle crystals, m.p. $131-132^{\circ}$ C. $R_{\rm f}$ 0.49 (MeOH–CHCl₃, 1:19). IR $v_{\rm max}$ cm⁻¹: 3450, 2940, 1600, 1460, 1420, 1345, 1295, 1230, 1160, 1100, 1080, 945, 820. UV $\lambda_{\rm max}$ nm (log ε): 229 (sh, 3.88), 291 (3.40); MeOH + NaOH: 243 (3.91), 303 (3.65). 13 C NMR (CDCl₃): δ 154.6 (s, C-5), 150.4 (s, C-1), 135.1 (s, C-3), 114.9 (s, C-2), 112.4 (s, C-4), 97.8 (d, C-6), 56.3 (q, OMe), 17.9 (q, 3-Me). EIMS m/z (rel. int.): 210 [M + 4]⁺ (10), 208 [M + 2]⁺ (56), 206 [M]⁺ (100), 191 (12), 163 (30), 99 (20), 69 (25).

Curlignan (2)

Amorphous powder. R_f 0.25 (MeOH-CHCl₃ (1:19). $\left[\alpha\right]_{D}^{23} + 40.0$ (MeOH, c 0.5). IR v_{max} cm⁻¹: 3400, 2940, 1710, 1610, 1520, 1435, 1325, 1205, 1180, 1070, 770; UV λ_{max} nm (log ε): 228 (sh, 4.38), 276 (4.19), 298 (sh, 3.83); MeOH + NaOH: 231 (sh, 4.34), 273 (4.26), 303 (sh, 4.10). CD nm ($\Delta \varepsilon$): 317 (0), 293 (+4.82), 280 (+3.96), 271 (+4.88), 245(+0.07), 240 (+0.73), 229 (-5.69), 211 (+10.27). ¹H and ¹³C NMR (Me₂CO-d₆): Table 1. ¹H NMR (CDCl₃): δ 7.57 (1H, br s, H-4), 7.53 (1H, br s, H-6), 6.87 (2H, br s, H-2' and H-6'), 6.86 (1H, br s, H-5'), 5.56 (1H, d, J = 7.1 Hz, H-2), 3.90 (1H, s, 7-OMe), 3.89 (2H, m, H-9a), 3.87 (3H, s, 8-OMe), 3.83 (3H, s, 3'-OMe), 3.65 (1H, dt, J = 7.1, 6.1 Hz, H-3). HR EI MS $[M]^+$ m/z: 360.1212 ($C_{19}H_{20}O_7$ requires 360.1209); EIMS m/z (rel. int.): [M]⁺ 360 (70), 342 (100), 331 (60), 327 (85), 310 (25), 295 (20), 283 (15), 239 (15), 208 (15), 181 (80), 149 (45), 117 (50), 97 (35), 71 (45), 57 (50).

Curlignan peracetate (2a)

¹H NMR (CDCl₃): δ 7.56 (1H, br s, H-4), 7.55 (1H, br s, H-6), 7.00 (1H, d, J = 8.0 Hz, H-5′), 6.95 (1H, d, J = 1.7 Hz, H-2′), 6.92 (1H, dd, J = 8.0, 1.7 Hz, H-6′), 5.58 (1H, d, J = 6.8 Hz, H-2), 4.45 (1H, dd, J = 11.3, 5.4 Hz, H-9a), 4.30 (1H, dd, J = 11.3, 7.5 Hz, H-9b), 3.93 (3H, s, 7-OMe), 3.87 (3H, s, 8-OMe), 3.80 (1H, m, H-3), 3.79 (3H, s, 3′-OMe), 2.28 (3H, s, 4′-OAc), 2.03 (3H, s, 9-OAc). EIMS [M]⁺ m/z: 444.

4-Ethoxy-3-hydroxymethylphenol (3)

Amorphous powder. $R_{\rm f}$ 0.38 (MeOH–CHCl₃, 1:9). IR $\nu_{\rm max}$ cm⁻¹: 3300, 2960, 2910, 1500, 1455, 1350, 1200, 1070, 1000, 820, 780 cm⁻¹. UV $\lambda_{\rm max}$ nm (log ε): 229 (sh, 3.76), 297 (3.55); MeOH + NaOH: 253 (sh, 3.97), 301 (sh, 3.80). ¹H NMR (CDCl₃): δ 6.71 (1H, d, J = 8.6 Hz, H-5), 6.64 (1H, dd, J = 8.6, 2.9 Hz, H-6), 6.50 (1H, d, J = 2.9 Hz, H-2), 4.60 (2H, s, -CH₂OH), 3.57 (2H, q, J = 7.0 Hz, OCH₂CH₃), 1.24 (3H, t, J = 7.0 Hz, OCH₂CH₃).

¹³C NMR (CDCl₃): δ 149.8 (s, C-4), 148.7 (s, C-1), 123.2 (s, C-3), 117.1 (d, C-6), 115.9 (d, C-5), 114.8 (d, C-2), 71.8 (t, OCH₂CH₃), 66.3 (t, -CH₂OH), 15.0 (q, -OCH₂CH₃). EIMS m/z (rel. int.): [M]⁺ 168 (30), 122 (100), 94 (70), 66 (15), 55 (10).

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REFERENCES

- 1. Chifundera, K., Messana, I., Galeffi, C. and De Vicente, Y., *Tetrahedron*, 1991, **47**, 4369.
- Chifurdera, K., Palazzino, G., Messana, I., Ping, L., Galeffi, C. and Cannarsa, G., Phytochemistry, 1994, 35, 1343.
- 3. Lee, S. S., Chang, W. L. and Chen, C. H., *Tetrahedron Lett.*, 1996, **37**, 4405.
- Chang, W. L., Su, M. J. and Lee, S. S., J. Nat. Prod., 1997, 60, 76.

- Taguchi, H., Yosioka, I., Yamasaki, K. and Kim, I. H., Chem. Pharm. Bull., 1981, 29, 55.
- Yahara, S., Satoshiro, M., Nishioka, I., Nagasawa, T. and Oura, H., *Chem. Pharm. Bull.*, 1985, 33, 527.
- Xu, J. P. and Dong, Q. Y., Zhongcaoyao, 1986, 17, 8–38.
- 8. Thong-Ngarm, I. and Chantrapromma, K., Warasan Songkhla Nakkharin, 1983, **5**, 245.
- 9. Lee, S. S., Lin, B. F. and Chen, K. C. S., *Chin. Pharm. J.*, 1995, **47**, 511.
- Xu, J. P. and Dong, Q. Y., *Zhongcaoyao*, 1987,
 18, 194, 222.
- 11. Hirai, N., Okamoto, M., Udagawa, H., Yamamuro, M., Kato, M. and Koshimizu, K., *Biosci. Biotechol. Biochem.*, 1994, **58**, 1679.
- 12. Mirsa, T. N., Singh, R. S., Tripathi, D. M. and Sharma, S. C., *Phytochemistry*, 1990, **29**, 929.