

Phytochemistry 51 (1999) 337-340

# Diterpenoid and norditerpenoid alkaloids from *Delphinium* carduchorum

Ali H. Meriçli<sup>a,\*</sup>, Filiz Meriçli<sup>a</sup>, Emine Doğru<sup>a</sup>, Hasan Özçelik<sup>b</sup>, Atta-Ur-Rahman<sup>c</sup>, Ayhan Ulubelen<sup>a,\*</sup>

<sup>a</sup>Faculty of Pharmacy, University of Istanbul, 34452 Istanbul, Turkey <sup>b</sup>Faculty of Science and Literature, University of Süleyman Demirel, Isparta, Turkey <sup>c</sup>H.E.J. Research Institute of Chemistry, University of Karachi, Karachi 75270, Pakistan

Received 20 August 1998; received in revised form 2 December 1998

#### Abstract

From the aerial parts of *Delphinium carduchorum*, we have isolated three known norditerpenoid alkaloids (delcaroline, deltatsine, 18-hydroxy-14-*O*-methyl-gadesine) and two new diterpenoid alkaloids, carduchoron and delcarduchol. The structures of the newly isolated alkaloids were established from spectroscopic data. © 1999 Elsevier Science Ltd. All rights reserved.

Keywords: Delphinium carduchorum; Ranunculaceae; Diterpenoid alkaloids; Carduchoron; Delcarduchol

# 1. Introduction

In continuation of our investigations of Turkish Delphinium species (Ulubelen, Meriçli, Meriçli, & Ilarslan, 1992, 1993; Ulubelen, Mericli, Mericli, Ilarslan, & Matlin, 1992; Ulubelen, Meriçli, Meriçli, Ilarslan, & Voelter, 1993; Ulubelen, Meriçli, & Meriçli, 1993), we have now studied the aerial parts of D. carduchorum. We have isolated two new diterpenoid and three known norditerpenoid alkaloids. The known compounds were established as delcaroline (Pelletier, Mody, & Desai, 1981), deltatsine (Joshi et al., 1984) and 18-hydroxy-14-O-methylgadesine (Gonzalez, de la Fuente, Mungia, & Henrick, 1981) by comparison of their NMR data with those of literature values. The structures of the new compounds carduchoron (1) and delcarduchol (2) were derived from 1-D and 2-D NMR techniques.

# 2. Results and discussion

The HR mass spectrum (m/z 339.1827, calc. 339.1834) indicated the molecular formula  $C_{21}H_{25}NO_3$ for carduchoron (1). The NMR spectra indicated the presence of an exo-methylene group [ $\delta_{\rm H}$  4.97 (1H, brs, H-17) and 4.78 (1H, brs, H-17');  $\delta_{\rm C}$  110.1 t, C-17 and 146.8 s, C-16]. The IR and <sup>13</sup>C NMR spectra showed the presence of three carbonyl groups at 1710, 1690 cm<sup>-1</sup> and  $\delta_{\rm C}$  207.6, 209.0 and 177.5, respectively. The presence of an N-methyl group was observed in the NMR spectra [ $\delta_{\rm H}$  2.50, (3H, s);  $\delta_{\rm C}$  42.6 q] and in the IR spectrum, a band 2780 cm<sup>-1</sup> correlated with the presence of this group. The absence of methoxyl groups in the NMR spectra together with the presence of the exo-methylene group indicated that carduchoron (1) is a diterpenoid alkaloid with three carbonyl groups. The location of these groups was accomplished by <sup>1</sup>H and <sup>13</sup>C NMR, <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H-<sup>13</sup>C HETCOR and by COLOC spectra. The chemical shift of C-18 methyl group ( $\delta_{\rm H}$  1.50 (3H, s);  $\delta_{\rm C}$  23.8q) is

<sup>\*</sup> Corresponding authors.

only possible when there is a carbonyl group at C-6, as also observed in spirasine II (Farg, Xiao-tian, & De quan, 1986; Wu, Wu, Niwa, Lu, & Hirata, 1988) hetidine (Pelletier, Aneja, & Gopinath, 1968; Jiang & Pelletier, 1988), thalicsessine (Wu, Wu, Niwa, Lu, & Hirata, 1987). The oxo group at C-6 also caused the chemical shift at C-5 ( $\delta_{\rm C}$  58.9) and C-7 ( $\delta_{\rm C}$  50.9) to move towards higher fields, as also observed in the above three compounds. Twenty-one carbons were observed in the 1-D, <sup>1</sup>H-coupled <sup>13</sup>C (APT) spectrum and the chemical shift patterns indicated two methyl, seven methylene, five methine and seven quaternary carbons. The two other carbonyl groups could be placed at one of the following positions: C-1-C-3, C-7, C-11, C-13, C-15, and C-19. The chemical shifts (Table 1) clearly indicated that no carbonyl group is present at C-1 to C-3, C-7 is seen to be an isolated methylene, with AB doublets at  $\delta_{\rm H}$  2.25 and 2.75 ( $J=18{\rm Hz}$ ) and with  $\delta_{\rm C}$  50.9 t. Since the chemical shift of C-16 is at  $\delta_{\rm C}$ 144.5, there should not be a carbonyl group at C-15; also, the C-15 protons were observed at  $\delta$  2.26 and 2.38 as doublets (J = 14 Hz). This leaves only three positions C-11, C-13, and C-19 for the placement of the two carbonyl groups. One of the carbonyl signals is at  $\delta$  177.5, indicating that the carbonyl should be attached to the nitrogen and, therefore, should be placed at C-19. Due to chemical shifts induced on C-9 ( $\delta_{\rm C}$  74.1) and C-12 ( $\delta_{\rm C}$  61.1), the last carbonyl should be placed at C-11 instead of C-13; COLOC experiments correlated this decision. A survey in the literature showed the presence of a compound thalicsessine (3) (Wu et al., 1987) with an N-CH<sub>2</sub>CH<sub>2</sub>OH group

341.1991) as indicated by its HR mass spectrum. The NMR spectra indicated a diterpenoid alkaloid instead of a norditerpene. The exo-methylene group was observed at  $\delta_{\rm H}$  5.01 (1H, t, J = 1.5 Hz), and 4.97 (1H, t, J = 1.5 Hz);  $\delta_{\rm C}$  155.1 s, C-16 and 110.0 t C-17. The chemical shift of C-16 indicated the presence of a hydroxyl group at C-15 [ $\delta_{\rm H}$  3.95 brs;  $\delta_{\rm C}$  75.0 d H-15  $\alpha$  ]. Twenty-one carbons were observed in the <sup>1</sup>H-coupled <sup>13</sup>C (APT) spectrum of **2**, and indicated two methyl, seven methylene, six methine and six quaternary carbons. Two of the quaternary carbons were keto groups as indicated in the IR (1710 and 1700 cm<sup>-1</sup>) and in the  $^{13}$ C NMR spectra ( $\delta_{\rm C}$  210.5 and 213.0). Since the location of the hydroxyl group was evident, the two keto groups should be placed at one of the following positions: C-1-C-3, C-6, C-7, C-11, C-13 or C-19. The chemical shift of Me-18 being at  $\delta_{\rm H}$  1.00 (3H, s) and  $\delta_{\rm C}$  27.3 clearly showed that there was no carbonyl group at C-6. The presence of an N-methyl group at  $\delta_{\rm H}$  2.37 s,  $\delta_{\rm C}$  41.8 q instead of  $\delta_{\rm H}$  2.50, as in compound 1, ruled out the presence of a keto group at C-19; also the lack of a carbonyl group signal around  $\delta_{\rm C}$ 170-180 correlated with the absence of a carbonyl group at this position. The chemical shift of C-1 to  $\delta_C$ 48.2 t and C-3 to  $\delta_{\rm C}$  52.0 t, and the lack of a triplet signal around  $\delta_{\rm C}$  20.0 for C-2, indicated the presence of a carbonyl group at C-2. The second carbonyl was placed at C-13 due to the chemical shifts of C-12 ( $\delta_{\rm C}$ 53.4 d) and C-14 ( $\delta_{\rm C}$  56.6 d). Both positions were also correlated by COLOC experiments. Table 2 shows the <sup>1</sup>H-<sup>1</sup>H COSY, HETCOR and COLOC results of delcarduchol (2).

and three carbonyl groups at C-6, C-11 and C-19. The <sup>13</sup>C NMR shifts of compound **1** Table 1 are quite similar to those. Thus, carduchoron was assigned the structure (**1**).

The second new compound delcarduchol (2) had the molecular formula  $C_{21}H_{27}NO_3$  (m/z 341.1984, calc.

# O Me .... N OH

# 3. Experimental

#### 3.1. General

IR spectra were recorded in CHCl<sub>3</sub>. <sup>1</sup>H NMR spectra were measured at 200 MHz, <sup>13</sup>C NMR at 50 MHz

Table 1 NMR data of carduchorone (1) and <sup>13</sup>C NMR of thalicsessine (3)

Position	<sup>13</sup> C	$^{1}H$	COSY <sup>1</sup> H- <sup>1</sup> H	COLOC <sup>13</sup> C- <sup>1</sup> H	3
1	38.1t	α 2.01 dd (5,12)	1β, 2α, 2β	C-5, C-9, C-20	39.8
_	22.0	β 1.60 brd (12)	$1\alpha$ , $2\alpha$		20.6
2	23.0t	α 1.75m	1α, 2β		20.6
3	35.2t	β 1.40m α 1.85m	$1\alpha$ , $1\beta$ , $2\alpha$		34.2
3	33.21	β 1.40m	2α, 2β, 3β 2β, 3α		34.2
4	46.9s	р 1.40Ш	2β, 3α		46.5
5	58.9s	2.50s		C-6, C-19, C-20	60.0
6	207.6s	2.303		C 0, C 13, C 20	207.6
7	50.9t	α 2.75d (18)	7β		51.5
	50.50	β 2.25d (18)	7α		01.0
8	44.5s	F ==== (==)			43.9
9	74.1d	1.66s		C-6, C-11	75.6
10	43.5s			,	42.9
11	209.0s				208.9
12	61.1d	2.30brs		C-11, C-17	63.7
13	29.7t	α 1.90m	13β, 14		33.3
		β 1.40m	13α, 14		
14	47.4d	1.80m	13α, 13β		47.0
15	48.5t	α 2.26d (14)	15β	C-12, C-16, C-17	51.5
		β 2.38d (14)	15α		
16	144.5s				141.6
17	110.1t	4.97brs		C-12, C-15	111.1
17'		4.78brs			
18	23.8q	1.50s		C-6, C-10	25.5
19	177.5s				177.1
20	53.2d	2.02d (3)		C-5, C-12, C-13	53.9
21	42.6q	2.50s			49.7
22					60.9

in CDCl<sub>3</sub>. HR-MS were measured at 70 eV. Chromatographic separations were carried out on a Chromatotron instrument using rotors coated with 1-mm thick layers of neutral Al<sub>2</sub>O<sub>3</sub>.

#### 3.2. Plant material

Aerial parts of *D. carduchorum* Chowdhuri and Davis were collected in eastern Turkey from the Van-Gevas Mountain at an altitude of 2500 m in August 1993. A voucher specimen is deposited at the University of Süleyman Demirel (Isparta), Faculty of Sciences, under H.Ö. 6326.

#### 3.3. Extraction of alkaloids

Dried and powdered aerial parts were extracted by percolation at room temp., using MeOH. After evap. in vacuo, a residue (7g) was obtained. This was dissolved in EtOH and adjusted to pH 1.5 (5%  $\rm H_2SO_4$ ) and extracted with  $\rm CH_2Cl_2$  (20×100ml) to give a neutral fr. (5.5g). The aq. soln was basified (pH 8–10) and extracted with  $\rm CH_2Cl_2$  (25×100ml) to yield 200 mg of a crude alkaloid mixt. This was separated on a neutral  $\rm Al_2O_3$  rotor of a Chromatotron with petrol–EtOAc–

MeOH systems to yield the alkaloids in the following order: delcaroline (12 mg), deltatsine (8 mg), **1** (22 mg), **2** (15 mg) and 18-hydroxy-14-*O*-methylgadenine (6 mg).

#### 3.4. Carduchoron (1)

IR  $v^{\text{CHCl}_3}$  cm<sup>-1</sup>: 3050, 2980, 2860, 2780, 1710, 1690, 1645, 1370, 1250, 1050, 980, 890.  $^{1}\text{H}$  and  $^{13}\text{C}$  NMR (CDCl<sub>3</sub>) in Table 1. HRMS m/z (rel. int.): 339.1827 [M]<sup>+</sup> (78), 324 [M-15]<sup>+</sup> (14), 310 [M-29]<sup>+</sup> (100), 296 (15), 279 (20), 178 (12), 124 (60), 113 (26), 97 (16), 83 (20), 71 (33), 57 (42).

# 3.5. Delcarduchol (2)

IR  $v^{\text{CHCl}_3}$  cm<sup>-1</sup>: 3480, 3040, 2960, 2840, 2778, 1710, 1700, 1650, 1420, 1370, 1268, 1150, 1050, 960, 890. <sup>1</sup>H and <sup>13</sup>C NMR (CDCl<sub>3</sub>) in Table 2. HRMS m/z (rel. int.): 341.1984 [M]<sup>+</sup> (76), 326 [M-15]<sup>+</sup> (62), 313 [M-28] <sup>+</sup> (22), 298 (14), 282 (18), 254 (15), 192 (23), 176 (15), 136 (12), 98 (12), 84 (100), 83 (97), 70 (17).

Table 2 NMR data of delcarduchol (2)

Position	<sup>13</sup> C	<sup>1</sup> H	$COSY ^{1}H-^{1}H$	COLOC <sup>13</sup> C <sup>-1</sup> H
1	48.2t	α 3.26 d (13)	1β	C-2, C-5, C-20
		β 1.30 d (13)	1α	
2	213.0s	,		
3	52.0t	α 2.90d (12)	3β	C-2, C-5, C-10
		β 1.90d (12)	3α	
4	41.0s	, , , ,		
5	59.9d	1.86brs		
6	29.3 <sup>a</sup> t	α 2.85m	6β, 7α, 7β	C-4, C-10, C-20
		β 1.60m	6α, 7α, 7β	
7	36.2t	α 2.75m	6α, 6β, 7β	C-15
		β 1.65m	6α, 6β, 7α	
8	40.6s	,	, ,,	
9	46.6d	2.04dd (4,7)	11α, 11β	
10	50.4s	· / /	, ,	
11	23.4 <sup>a</sup> t	α 1.80m	9, 11 β	C-13, C-16, C-17
		β 2.35m	9, 11 α	,,
12	53.4d	2.60brd (3Hz)	,	C-13, C-16, C-17
13	210.5s	, ,		,,
14	56.6d	1.65m		
15	75.0d	α 3.95brs		
16	155.1s			
17	110.0t	5.01t (1.5)	17'	
17'		4.97t (1.5)	17	
18	27.3q	1.00s		
19	57.9t	α 2.16d (13)	19β	
		β 1.98d (13)	19α	
20	67.3d	2.97brs		
21	41.8q	2.37s		

# Acknowledgements

This study was partly supported by the University of Istanbul Research Fund Ö-385 and partly by the Research Fund of Turkish Academy of Sciences (TÜBA), awarded to A.U.

#### References

Farg, S., Xiao-tian, L., & De quan, Yu. (1986). *Heterocycles*, 24, 2105.

Gonzalez, A. G., de la Fuente, G., Mungia, O., & Henrick, K. (1981). *Tetrahedron Lett.*, 22, 4843.

Jiang, Q., & Pelletier, S. W. (1988). Tetrahedron Lett., 29, 1875.

Joshi, B. S., Glinski, J. A., Chokshi, H. P., Chen, S. Y., Srivastava, S. K., & Pelletier, S. W. (1984). Heterocycles, 22, 2037. Pelletier, S. W., Aneja, R., & Gopinath, K. W. (1968). Phytochemistry, 7, 625.

Pelletier, S. W., Mody, N. V., & Desai, R. C. (1981). *Heterocycles*, 16, 747.

Ulubelen, A., Meriçli, A. H., & Meriçli, F. (1993). J. Nat. Prod., 56, 780.

Ulubelen, A., Meriçli, A. H., Meriçli, F., & Ilarslan, R. (1992). *Phytochemistry*, 31, 1019.

Ulubelen, A., Meriçli, A. H., Meriçli, F., & Ilarslan, R. (1993). *Phytochemistry*, 33, 213.

Ulubelen, A., Meriçli, A. H., Meriçli, F., Ilarslan, R., & Matlin, S. A. (1992). *Phytochemistry*, 31, 3239.

Ulubelen, A., Meriçli, A. H., Meriçli, F., Ilarslan, R., & Voelter, W. (1993). *Phytochemistry*, 34, 1165.

Wu, Y. C., Wu, T. S., Niwa, M., Lu, T. S., & Hirata, Y. (1987). Heterocycles, 26, 943.

Wu, Y. C., Wu, T. S., Niwa, M., Lu, S. T., & Hirata, Y. (1988). *Phytochemistry*, 27, 3949.