



The hydroxylation of the sesquiterpenoid guaioxide by *Mucor plumbeus*

Simone Fontes Arantes, James R. Hanson*

School of Chemistry, Physics and Environmental Science, University of Sussex, Brighton, Sussex, BN1 9QJ, UK

Received 5 January 1999; accepted 17 February 1999

Abstract

Microbiological hydroxylation of guaioxide by *Mucor plumbeus* gave the 7-, 8 α - and 9 β - and 9 α -hydroxy derivatives. © 1999 Elsevier Science Ltd. All rights reserved.

Keywords: *Mucor plumbeus*; Microbiological hydroxylation; Sesquiterpenoid; Guaioxide

1. Introduction

Guaiol **1** is a readily available sesquiterpenoid from guaiac wood oil (Plattner & Lemay, 1940; Plattner & Magyar, 1942). A large number of hydroxylated guaianes are known. Compounds with this azulene carbon skeleton have been obtained from both plants and fungi and a number have interesting biological activity (Fraga, 1998). Their preparation by the microbiological hydroxylation of the more abundant members of this family is an attractive target (Lamare & Furstoss, 1990). The hydroxylation of guaioxide **2** by *Mucor parasiticus* gave (Ishii et al., 1970) the 4 α - and 8 α -monohydroxy- and 4 α ,8 α -dihydroxy derivatives whilst *Streptomyces purpureus* afforded (Ishii et al., 1971) ring A hydroxylation products and 9 α -hydroxy-guaioxide.

2. Results and discussion

Despite numerous attempts, it was not possible to hydroxylate guaiol **1** itself using *Mucor plumbeus*. However, the ether bridge of guaioxide **2**, which was readily prepared from guaiol by treatment with per-

chloric acid in acetic acid (Bates & Slagel, 1962), alters the geometry of the guaiane ring system sufficiently to make it a substrate for *M. plumbeus*.

Incubation of guaioxide **2** with *M. plumbeus* for five days gave four metabolites, which were separated by column chromatography. The first metabolite to be isolated was 7-hydroxyguaioxide **3**. The location of the hydroxyl group at C-7 followed from the appearance of a new tertiary alcohol signal in the ¹³C NMR spectrum at δ_C 79.8 (see Table 1). The methylene signals assigned to C-6 and C-8 showed significant downfield shifts (δ_C 47.0; $\Delta\delta$ 6.2, C-6; δ_C 39.5, $\Delta\delta$ 9.1, C-8) whilst the C-14 and C-15 methyl groups showed γ -gauche shieldings of 3.6 and 1.7 ppm compared to the starting material.

The 8 α - and 8 β -hydroxyguaioxides, **4** (Ishii et al., 1970) and **5**, were isolated as an inseparable mixture (1:1) of secondary alcohols (**4**, H-8, δ_H 3.88, multiplet; **5**, H-8, δ_H 3.62, doublet of triplets, J 11.9 and 4.1 Hz). The mixture was oxidized with chromium trioxide to a single ketone (δ_C 208.15; ν_{max} 1695 cm⁻¹) and hence it was a mixture of epimeric alcohols. In order to confirm the structure, the proton resonances on the seven-membered carbocyclic ring of the ketone were assigned as follows. Irradiation of the geminal methyl group signal at δ_H 1.15 (H-14) gave a nuclear Overhauser effect (nOe) enhancement to a signal at δ_H 1.90 (double-doublet, J 13.7 and 6.4 Hz, H-6 β) and δ_H 2.57

* Corresponding author.

Table 1
 ^{13}C NMR data determined in deuteriochloroform at 75 MHz

Carbon	Compound			
	2	3	6	7
1	54.8	53.2	55.6	53.2
2	28.5	28.2	29.0	29.2
3	31.1	30.7	31.6	31.2
4	46.0	46.4	52.0	46.4
5	90.9	87.0	91.4	90.9
6	40.8	47.0	36.4	40.5
7	41.1	79.8	63.0	44.2
8	30.4	39.5	208.1	40.9
9	33.8	32.2	45.9	72.4
10	37.5	37.5	32.9	44.2
11	80.9	81.0	81.0	81.2
12	23.3	23.7	23.1	17.6
13	16.3	17.1	16.5	16.8
14	30.4	26.8	29.8	30.3
15	23.0	21.3	24.3	23.9

(multiplet, H-7). The ^1H : ^{13}C correlation spectrum showed that the signal at δ_{H} 1.90 was linked to a methylene carbon (δ_{C} 36.4) and to a second proton (δ_{H} 2.10, doublet of doublets, J 13.7 and 1.7 Hz). There was also a correlation between a methine resonance at δ_{C} 63.0 and the signal (δ_{H} 2.57) assigned to H-7. Irradiation of H-15 (δ_{H} 1.22) produced an nOe enhancement of δ_{H} 2.57 (H-7, 1.7%) and at δ_{H} 2.61 (1H, m, 3.8%). The ^1H : ^{13}C correlation spectrum showed a correlation between the signal at δ_{H} 2.61, a methylene carbon at δ_{C} 52.0 and a further proton at δ_{H} 2.30 (doublet, J 14.5 Hz, of triplets, J 1.5 Hz). Irradiation of this signal showed that it was coupled to the signal at δ_{H} 2.61 and to a signal at δ_{H} 1.56 which was in turn coupled to the methyl group doublet (δ_{H} 1.00, J 7.0 Hz). This led to the assignment of the signals at δ_{H} 1.56 as H-10, δ_{H} 1.90 as H-6 β , δ_{H} 2.10 as H-6 α , δ_{H} 2.30 as H-9 α , δ_{H} 2.57 as H-7 and δ_{H} 2.61 as H-9 β and to the structure of the ketone as 8-oxo-guaiooxide **6**. Hence the original metabolites were the 8 α - and 8 β -alcohols, **4** and **5**.

The site of hydroxylation (C-9) of the fourth metabolite **7** followed from changes in the ^{13}C NMR spectrum to the position of the signals assigned to C-8 (δ_{C} 40.9; $\Delta\delta$ 10.5 ppm) and C-10 (δ_{C} 44.2, $\Delta\delta$ 6.7 ppm). The ^1H NMR spectrum of 9 α -hydroxyguaiooxide **7** contained a CH(OH) resonance at δ_{H} 3.75 which appeared as a doublet (J 10.8 Hz) of doublets (J 9.8 Hz) of doublets (J 4.0 Hz) (Ishii et al., 1971, δ_{H} 3.83, multiplet). This multiplicity corresponded to a secondary alcohol possessing two diaxial and one axial:equatorial coupling. Irradiation of the H-15 (methyl group) signal (δ_{H} 1.28) gave a nOe enhancement (4.4%) of the CH(OH) signal at δ_{H} 3.75 whilst irradiation of the

methyl group doublet at δ_{H} 0.98 (H-12) also produced nOe enhancements of the H-1 (δ_{H} 1.54, 3.2%) and H-9 (δ_{H} 3.75, 1.1%) signals. Hence the metabolite was assigned the stereochemistry of 9 α -hydroxyguaiooxide **7**.

Guaiooxide as a cyclic ether, is chemically relatively unreactive thus restricting its use as a starting material for partial synthesis. Free radical oxidation with *m*-chloroperbenzoic acid has been shown (Tori et al., 1990) to occur at the tertiary centres C-1, C-4 and C-10 in low yield. However the microbiological hydroxylation by *M. plumbeus* takes place at methylene carbons that are chemically difficult to functionalize. The distribution of metabolites obtained with *M. plumbeus* differed from that found with other organisms.

3. Experimental

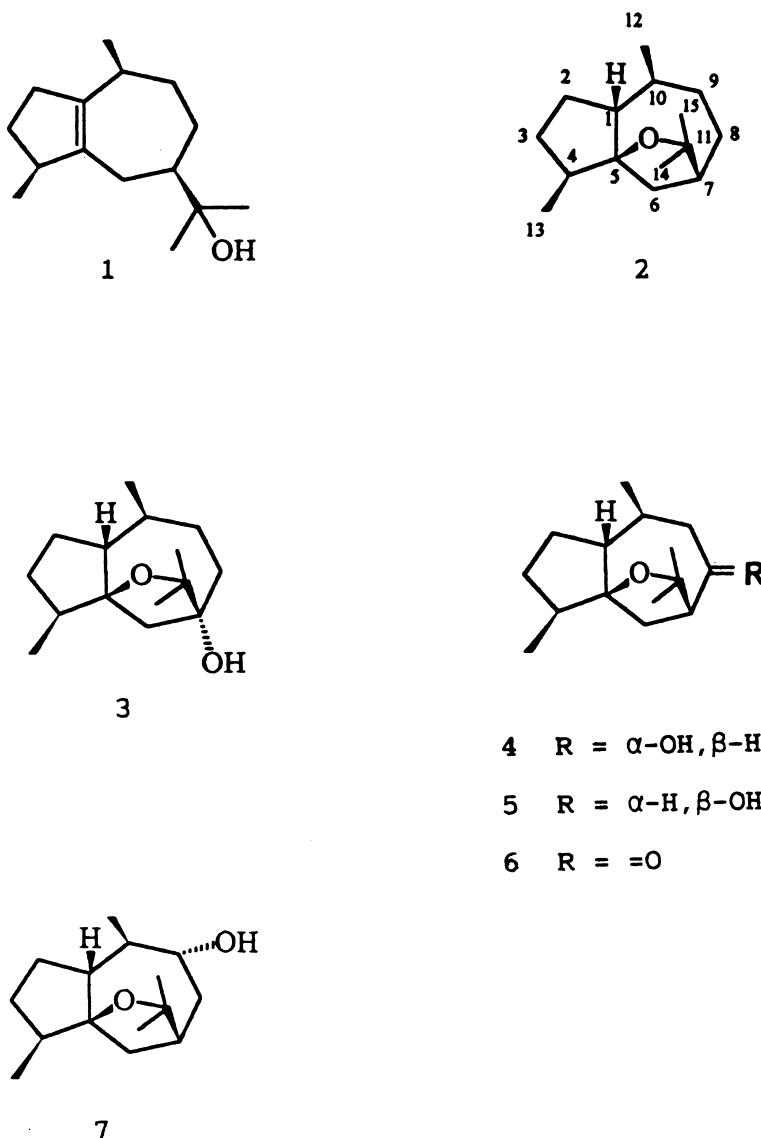
^1H NMR spectra were recorded in CDCl_3 at 300 or 500 MHz. ^{13}C NMR spectra were determined at 75 MHz. IR spectra were recorded as nujol mulls. Chromatography was carried out on silica, Merck 9385. Light petroleum refers to the fraction, b.p. 60–80°. Extracts were dried over anhydrous sodium sulfate. Jones reagent refers to a solution of chromium trioxide (26.72 g) in concentrated sulfuric acid (23 cm³) diluted to 100 cm³ with water.

3.1. Fermentation conditions

M. plumbeus (IMI 116688) was grown on shake culture on a medium comprising (per litre): glucose (30 g), potassium dihydrogen phosphate (2 g), magnesium sulfate (2 g), ammonium tartrate (2 g), yeast extract (1 g), calcium chloride (0.1 g), sodium chloride (1 g), iron(II) ammonium sulfate (0.2 g) and a trace elements solution (2 cm³). The latter contained (per litre): zinc sulfate (1 g), iron(II) sulfate (1 g), cobalt nitrate (1 g), ammonium molybdate (1 g), copper sulfate (0.1 g) and manganese sulfate (0.1 g). The culture was grown in shake culture in 250 cm³ conical flasks each containing 100 cm³ medium for 36 h prior to the addition of the substrate.

3.2. Incubation of guaiooxide with *M. plumbeus*

Guaiooxide (710 mg) dissolved in ethanol (30 cm³), was evenly distributed between 29 flasks of *M. plumbeus* and the fermentation continued for a further five days. The mycelium was filtered and the broth was extracted with diethyl ether. The extract was dried and the solvent evaporated to give a gum which was chromatographed on silica. Elution with 5% diethyl ether:light petroleum gave 7 α -hydroxyguaiooxide **3** (23 mg) which crystallized from light petroleum as needles, m.p. 118–120° (found: HREIMS 238.192 C₁₅H₂₆O₂



requires 238.193), ν_{\max} 3388, 1086, 1064 cm^{-1} ; δ_{H} 0.90 (3H, *d*, *J* 6.3 Hz, H-12), 0.95 (3H, *d*, *J* 7.0 Hz, H-13), 0.99 (1H, *m*, H-2 α), 1.12 (3H, *s*, H-14), 1.16 (3H, *s*, H-15), 1.57 (1H, *td*, *J* 13.8, 4.1 Hz, H-8 β), 1.91 (1H, *m*, H-2 β), 1.96 (1H, *dd*, *J* 11.7, 2.0 Hz, H-6 β), 2.12 (1H, *d*, *J* 11.7 Hz, H-6 α). Elution with 7% diethyl ether: light petroleum gave a mixture of alcohols **4** and **5** (25 mg) which were oxidized to 8-oxoguaiaoxide (vide infra). Further elution with 10% diethyl ether:light petroleum gave 9 α -hydroxyguaiaoxide **7** (60 mg) which crystallized from light petroleum as needles, m.p. 117–119° (found: HREIMS 238.193 $\text{C}_{15}\text{H}_{26}\text{O}_2$ requires 238.193). ν_{\max} 3380, 1107 cm^{-1} , δ_{H} 0.95 (3H, *d*, *J* 7.1 Hz, H-13), 0.98 (3H, *d*, *J* 6.5 Hz, H-12), 1.12 (3H, *s*, H-14), 1.28 (3H, *s*, H-15), 1.31 (1H, *m*, H-8 α), 1.54 (1H, *dt*, *J* 10.1, 8.5 Hz, H-1), 2.08 (1H, *m*, H-8 β), 3.75 (1H, *ddd*, *J* 10.8, 9.8, 4.0 Hz, H-9 β).

3.3. Oxidation of the alcohols **4** and **5**

The mixture of alcohols **4** and **5** (25 mg) from the above was dissolved in acetone (5 cm^3) and titrated with the Jones reagent until a strong persistent orange colour was obtained. Methanol was then added, the solvent was evaporated and the products recovered in diethyl ether. The extract was washed with aqueous sodium hydrogen carbonate, water, brine and dried. The solvent was evaporated to give 8-oxoguaiaoxide **6** (24 mg) as an oil, ν_{\max} 1695, 1164, 1045 cm^{-1} , δ_{H} 0.91 (1H, *m*, H-2), 0.93 (3H, *d*, *J* 6.4 Hz, H-13), 1.00 (3H, *d*, *J* 7.0 Hz, H-12), 1.15 (3H, *s*, H-14), 1.22 (3H, *s*, H-15), 1.25 (1H, *m*, H-3), 1.56 (1H, *m*, H-10), 1.63 (1H, *dd*, *J* 10.3, 8.0 Hz, H-1), 1.69 (1H, *m*, H-3), 1.76 (1H, *m*, H-4), 1.90 (1H, *dd*, *J* 13.7, 6.4 Hz, H-6 β), 1.93 (1H, *m*, H-2), 2.10 (1H, *dd*, *J* 13.7, 1.7 Hz, H-6 α), 2.30 (1H,

dt, *J* 14.5, 1.5 Hz, H-9 α), 2.57 (1H, *m*, H-7), 2.61 (1H, *m*, H-9 β).

Acknowledgement

SFA thanks CNPq (Brazil) for financial assistance.

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