



A clerodane derivative from *Grangea maderaspatana*

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Abstract

A new clerodane, 15-hydroxy-16-*oxo*-15,16H-hardwickiic acid (15-hydroxycleroda-3,13-dien-15,16-olide-18-oic acid) has been isolated from the aerial parts of *Grangea maderaspatana*, besides the known clerodane diterpenes. © 1999 Elsevier Science Ltd. All rights reserved.

Keywords: *Grangea maderaspatana*; Compositae; Diterpenes; 15-Hydroxy-16-*oxo*-15,16H-hardwickiic acid

1. Introduction

Grangea is a small genus of herbs and distributed in tropical and sub-tropical parts of the Africa and Asia. *G. maderaspatana* is a common weed usually grown in sandy lands and waste places. Previously steroidal constituents (Iyer & Iyer, 1978), hardwickiic acid, the corresponding 1,2-dehydro-derivative, acetylenic compounds (Pandey et al., 1984), eight new clerodane diterpenes including five clerodane, a *nor* clerodane, a *seco*-clerodane and a *nor-seco*-clerodane derivatives along with auranamide (Singh, Jain & Jakupovic, 1988; Singh & Jain, 1990), grangolide and eudesmanolides (Rojatkar, Chiplunkar & Nagasampagi, 1994) have been reported from various parts of this species. Further investigation of its aerial parts afforded a new *trans*-clerodane, named as 15-hydroxy-16-*oxo*-15,16H-hardwickiic acid (**1**) in addition to previously reported compounds.

2. Results and discussion

The compound (**1**) was isolated as colourless gum and purified as its methyl ester. Its structure was established mainly by high field ¹H-NMR and high resolu-

tion mass spectrometry. Its molecular formula C₂₁H₃₀O₅ was followed from accurate mass measurements of peak at *m/z* 344.198 [M-H₂O]⁺ corresponding to molecular composition C₂₁H₂₈O₄. IR spectrum exhibited the presence of, α,β-unsaturated-γ-lactone (1770 cm⁻¹), α,β-unsaturated ester (1715 cm⁻¹) and hydroxyl group (3300 cm⁻¹). The ¹H-NMR spectrum, when recorded in CDCl₃, was very close to that of hardwickiic acid except for lack of the usual furanic proton absorption and the addition of two broad singlets at δ 6.76 (*J* = 1.5, 1.5 Hz) and 5.75 (*J* = 1.5, 1.5 Hz) which could correspond to H-14 and H-15, respectively, in the C-13/C-16 α,β-unsaturated lactone. It was further confirmed from the ¹³C chemical shifts of C-13 (135.0), C-14 (143.5), C-15 (99.5) and C-16 (174.0) carbons that we are dealing with a hydroxy-lactone moiety with β-alkyl substituent. The sequence of protons was determined by spin decoupling experiments. A double doublet at δ 6.60 of H-3 was coupled to H-2α at δ 2.30 and H-2β at δ 2.28 each of which was in turn coupled to H-1α at δ 1.71 and H-1β at δ 1.53. The nature of the lactone group in the side chain followed from the chemical shift of H-14 at δ 6.76 which was coupled vicinally to H-15 at δ 5.75 and allylically to H-12α, β at δ 2.17 and 2.07, respectively. The chemical shifts of H-17, H-19 and H-20 at δ 0.82, 1.27 and 0.77 showed that we are dealing with an A/B ring *trans* fused clerodane. Furthermore, the H-17 signal was doubled. Accordingly the epimeric 15-hydroxy de-

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Table 1
¹H and ¹³C-NMR spectra of compound **1b**

H (400 MHz)		C (100 MHz)	
1	1.71 <i>ddbr</i> (12, 7, 1)	1	18.0
1'	1.53 <i>ddd</i> (12, 11, 5)	2	27.5
2	2.30 <i>m</i>	3	139.0
2'	2.28 <i>m</i>	4	143.0
3	6.60 <i>dd</i> (4, 3)	5	39.0
6	2.32 <i>m</i>	6	37.7
6'	1.15 <i>tbr</i> (12, 3)	7	28.0
7	1.40–150 <i>m</i>	8	37.0
7'	140–150 <i>m</i>	9	39.5
8β	1.63 <i>dq</i> (11, 7)	10	47.0
10β	1.38 <i>dd</i> (12, 1)	11	36.0
11	1.67 <i>tbr</i> (13)	12	20.0
11'	1.50 <i>tbr</i> (13)	13	135.0
12	2.17 <i>tbr</i> (13)	14	143.5
12'	2.07 <i>tbr</i> (13)	15	99.5
14	6.76 <i>dt</i> (1.5, 1.5)	16	174.0
15	5.75 <i>dt</i> (1.5, 1.5)	17	11.0
17	0.82 <i>d</i> , (0.81 <i>d</i>) ^a (7)	18	170.0
19	1.27 <i>s</i>	19	21.0
20	0.77 <i>s</i>	20	19.0
OMe	3.68 <i>s</i>	OMe	51.0

^a Values of epimer.

rivatives of (**1**) were present which could not be separated. Earlier the isolation of corresponding epimeric 15-methoxy derivatives (**2**) from this species (Singh et al., 1988) further ascertained the presence of β-butenolide and proposed structure for (**1**). The detailed assignments of various ¹H- and ¹³C- signals are depicted in Table 1.

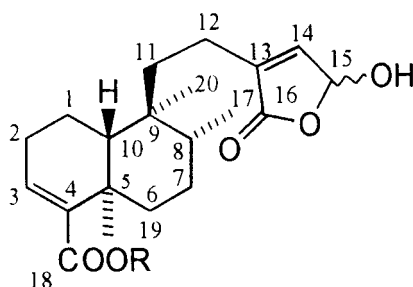
3. Experimental

IR spectra were measured in CCl₄/Nujol and ¹H- and ¹³C-NMR spectra were recorded in CDCl₃. Optical rotations were measured in CHCl₃ and HPLC were performed using MeOH-H₂O mixture in various pro-

portions. Prep TLC performed on TLC glass plates silica gel 60F₂₅₄ precoated layer thickness 0.5 mm. The plant material were obtained from M/S United Chemicals and Allied Products, Calcutta and specimen deposited at RUBL Herbarium, Jaipur.

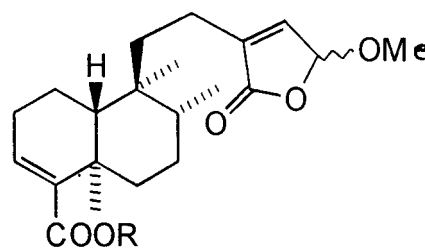
3.1. Extraction and isolation

The air-dried aerial parts (2 kg) of *G. maderaspatana* were extracted with Et₂O–petrol–MeOH (1 : 1 : 1) at room temperature for 24 h. Evaporation of the solvent under vacuo furnished a greenish semisolid mass which was defatted by dissolving in MeOH (250 ml) and left over night in the refrigerator. Filtered, precipitate was rejected and the filtrate was concentrated and column chromatographed over silica gel (60–120 mesh) and following fractions were collected. Fraction-1 (petrol), fraction-2 (petrol–Et₂O, 3 : 1), fraction-3 (petrol–Et₂O, 1 : 1) and fraction-4 (Et₂O). Fraction-1 gave α-humulene (40 mg). Fraction-2 was a yellow oil and separated on TLC (C₆H₆–CH₂Cl₂, 1 : 1) affording 200 mg of phytol, 10 mg of 3-hydroxy-8-acetoxy-pentadeca-1,9,14-trien-4,6-diyne, 40 mg of lupeol and 400 mg mixture of diterpene acids, which on subsequent methylation with ethereal solution of CH₂N₂ and HPLC separation (MeOH–H₂O, 9 : 1) over RP 18 (analytical column) gave 30 mg of methyl 10-epinidoresedate (*R*_t 11.8 min), 100 mg of methyl strictate (*R*_t 12.7 min), 170 mg mixture of methyl esters of centipedic and nidoresedic acids (*R*_t 15.0 min) and 100 mg of methyl ester of hardwickiic acid (*R*_t 16.6 min). Fraction-3 on TLC (C₆H₆–CH₂Cl₂–Et₂O, 4.5 : 4.5 : 1) yielded 6 mg of 3,8-dihydroxy pentadeca-1,9,14-trien-4,6-diyne, 14 mg of *p*-hydroxybenzoic acid and 150 mg complex mixture of diterpene lactones. The latter was treated with CH₂N₂ and subsequent HPLC (MeOH–H₂O, 9 : 1) gave methyl esters of 15-methoxy-16-*oxo*-15,16H-strictic acid (20 mg, *R*_t 6.6 min), 15-methoxy-16-*oxo*-15,16H-hardwickiic acid (**2b**) (30 mg, *R*_t 7.0 min), 15-methoxy-16-*oxo*-nidoresedic acid (16 mg, *R*_t 7.2 min), *nor*-strictic acid (24 mg, *R*_t 7.9 min), *nor*-hardwickiic acid (10 mg, *R*_t 9.4 min) and 2α-acetoxy hardwickiic



1a R = H

1b R = Me



2a R = H

2b R = Me

acid (40 mg, R_f 12.4 min). Fraction-4 was a yellow solid mass and separated on TLC ($C_6H_6-CH_2Cl_2-Et_2O$, 1 : 1 : 1) after treatment with diazomethane affording methyl esters of 16-*oxo*-15,16H-hardwickiic acid (12 mg, R_f 0.30), 16-*oxo*-15,16H-strictic acid (20 mg, R_f 0.28), auranamide (14 mg, R_f 0.25) and compound (**1b**) (30 mg, R_f 0.20).

3.2. 15-Hydroxy-16-oxo-15,16H-hardwickiic acid (15-Hydroxycleroda-3,13-dien-15,16-olide-18-oic acid) (**1a**)

Colourless gum, isolated as its methyl ester, $C_{21}H_{30}O_5$, $[\alpha]_D^{25} -31^\circ$ ($CHCl_3$, c 0.20); IR ν_{max} CCl_4 cm^{-1} : 3300 (OH), 1770 (γ -lactone), 1715 ($C=C-COOR$); MS m/z (rel. int.): 344.198 $[M-H_2O]^+$ (25) (calc for $C_{21}H_{28}O_4$: 344.198), 329 $[344-Me]^+$ (15), 297 $[329-MeOH]^+$ (10); 1H - and ^{13}C -NMR ($CDCl_3$), Table 1.

Acknowledgements

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