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Flavonoids from Brosimum acutifolium*

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Abstract

4'-Hydroxy-7,8-[2-(2-hydroxyisopropyl)dihydrofuran]flavan and 4',7-dihydroxy-8-(3,3-dimethylallyl)flavan, together with 10 known plant constituents, were obtained from the trunk bark of *Brosimum acutifolium*. Their structures were elucidated by spectroscopic methods, including 2D NMR spectroscopic techniques. © 2000 Elsevier Science Ltd. All rights reserved.

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1. Introduction

The trunk bark of Brosimum acutifolium, a plant which is distributed mainly in highland forests from east of Pará State (Brazil) to east of Suriname, is used in folk medicine in the Amazon region as an anti-inflammatory and anti-rheumatic agent (van den Berg, 1982). In a previous paper (Torres, Monteiro, Arruda, Muller & Arruda, 1997), we reported the isolation and structure elucidation of two flavans [4'-hydroxy-7,8-(2",2"-dimethylpyran)flavan and 4'-hydroxy-7,8-(3"hydroxy-2",2"-dimethylpyran)flavan] from the trunk bark of B. acutifolium subsp. acutifolium. Further investigation led to the isolation of two new flavans, named brosimine A (1) and brosimine B (2), along with the known compounds 3',7-dihydroxy-4'-methoxyflavan, isobavachin, liquiritigenin, 4-hydroxyloncho-4-hydroxyisocordoin, isoliquiritigenin, carpin, coniferaldehyde, syringaldehyde, sitosterol and stigmasterol. This paper describes the structure elucidation of the two new flavans.

2. Results and discussion

Compound 1 gave a violet spot on a TLC plate with ceric sulphate. The IR spectrum of compound 1 suggested the presence of a hydroxyl group (3436 cm^{-1}) and an aromatic ring (1617 and 1465 cm^{-1}). The HR mass spectrum gave a $[M]^+$ at m/z 326.1514 which corresponds to the molecular formula C₂₀H₂₂O₄ (calcd.: 326.1518). Its 300 MHz ¹H-NMR spectrum (Table 1) showed signals typical of a flavan [δ 5.01 (1H, dd, J = 10.0 and 2.0 Hz, H-2)], 2.01 and 2.16 (1H)each, m, 2H-3), 2.72 (1H, ddd, J = 11.8; 6.4 and 3.2 Hz, H-4 α) and 2.91 (1H, ddd, J = 11.8; 8.5 and 4.3 Hz, H-4β) (Torres et al., 1997; Saini & Ghosal, 1984). The substitution pattern of ring B was proposed with a hydroxyl group at the 4'-position from the ¹H-NMR $[\delta 7.29 \text{ (2H, } d, J = 8.5 \text{Hz}) \text{ and } 6.84 \text{ (2H, } d, J = 8.5 \text{Hz})]$ and EI mass spectra (m/z 120). The presence of a 2-(2hydroxyisopropyl)-dihydrofuran ring was indicated by a set of peaks at δ 4.63 (1H, dd, J = 8.9 and 9.2 Hz), 3.05 (1H, dd, J = 11.8 and 8.9 Hz), 3.12 (1H, dd, J =11.8 and 9.2 Hz), 1.20 and 1.31 (3H each, s), attributed to an oxymethine H-2", two benzylic hydrogens H-1" and a gem-dimethyl group at C-3", respectively. This was confirmed by a COLOC experiment (summarized in Fig. 1) which showed ${}^{3}J_{\text{(CH)}}$ and ${}^{2}J_{\text{(CH)}}$ correlations between the hydrogens Me-4"/Me-5" and

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Table 1 1 H (300 MHz) and 13 C (75 MHz) NMR spectral data for compounds 1 and 2 (CDCl₃, δ)^a

H/C	1		2	
	$\delta_{ m H}$	$\delta_{ m C}$	$\delta_{ m H}$	δ_{C}
2	5.01 (1H, dd, J = 9.9, 1.9)	77.5	4.98 (1H, dd, J = 10.1, 2.4)	77.6
3	2.01 (1H, <i>m</i>); 2.16 (1H, <i>m</i>)	29.7	2.15 (1H, <i>m</i>); 1.98 (1H, <i>m</i>)	30.0
4β	2.91 (1H, ddd, J = 11.8, 8.5, 4.3)	24.6	2.92 (1H, ddd, J = 16.3, 11.0, 5.6)	24.9
4α	2.72 (1H, ddd, J = 11.8, 6.4, 3.2)	24.6	2.72 (1H, ddd, J = 16.3, 5.2, 3.4)	24.9
5	6.84 (1H, d, J = 8.1)	128.7	6.82 (1H, d, J = 8.2)	127.3
6	6.37 (1H, d, J = 8.1)	101.5	6.40 (1H, d, J = 8.2)	108.1
7		159.2 ^b		153.0°
8		113.4		114.6
9		151.7 ^b		153.6°
10		114.1		113.9
1'		134.0		137.5
2',6'	7.29 (2H, d, J = 8.5)	127.4	7.29 (d, J = 8.7)	127.3
3',5'	6.84 (2H, d, J = 8.5)	115.3	6.84 (d, J = 8.7)	115.2
4′		155.3		155.0
1"	3.15 (1H, dd, J = 11.8, 8.9); 3.12 (1H, dd, J = 11.8, 9.2)	28.3	3.41 (d, J = 7.2)	22.4
2"	4.63 (1H, dd, J = 9.2, 8.9)	89.9	5.27 (t, J = 7.2)	122.2
3"		72.1		134.0
4"	1.20 (3H, s)	23.6 ^d	1.74 (3H, s)	25.8
5"	1.31 (3H, s)	26.1 ^d	1.72 (3H, s)	17.8
7-OH			5.20 (1H, s)	

^a Coupling constants (*J* in Hz) are given in parentheses.

carbons C-2" (δ 89.9) and C-3" (δ 72.1). Based on the presence of two *ortho* coupled doublets in ¹H-NMR $[\delta 6.84 \text{ (1H, } d, J = 8.1 \text{ Hz, H-5}) \text{ and } 6.37 \text{ (1H, } d,$ J = 8.1Hz, H-6)] and comparison with ¹H-NMR spectroscopic data of the two flavans isolated previously (Torres et al., 1997), the dihydrofuran ring was placed at the C-7/C-8 positions, which was confirmed by a NOE experiment. Irradiation of H-4 α (δ 2.72) resulted in the enhancement of the H-5 signal (δ 6.84). A possstructure is 4'-hydroxy-7,8-(2",2"ible isomeric dimethylpyran)flavan (1a), previously obtained by us from the same species (Torres et al., 1997); however, its ¹H- and ¹³C-NMR data are not totally superimposable on those of 1, and the signals at $\delta_{\rm H}$ 4.63 (H-2") and $\delta_{\rm C}$ 89.9 (C-2") are characteristics of a dihydrofuran ring bonded to an aromatic ring (Macias, Massanet, Rodrigues-Luis & Salvá, 1989). Therefore, structure 1a for compound 1 was eliminated. The foregoing is consistent with structure 1, for which the name brosimine A is proposed. The ¹³C-NMR signals (Table 1) were assigned using HETCOR and COLOC experiments and by comparison of the observed values with those reported for model compounds (Torres et al., 1997; Macias et al., 1989).

Compound **2** also gave a violet color with ceric sulfate. It was assigned the molecular formula $C_{20}H_{22}O_3$ by HR-MS (m/z 310.1568, [M]⁺, calcd. 310.1569). The ¹H-NMR spectrum (Table 1) of **2** was similar to that of **1**, but with signals assigned to a 3,3-dimethylallyl moiety [δ 5.27 (1H, t, J = 7.2 Hz, H-2"), 3.41 (2H, d, J = 7.2 Hz, H-1"), 1.72 (3H, s, H-4") and 1.74 (3H, s, H-5")] replacing those of the 2-(2-hydro-

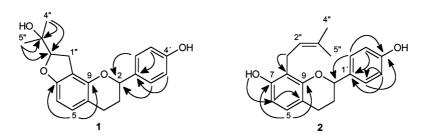


Fig. 1. CH long-range correlations in COLOC spectra of 1 and 2.

b,c Interchangeable signals.

^d Solution in CDCl₃ referenced to CHCl₃ at δ 7.26 ppm (¹H) and δ 77.23 ppm (¹³C).

xvisopropyl)-dihydrofuran ring of 1. The EI mass spectrum showed characteristic retro Diels-Alder fragments at m/z 120 and 190 indicating the presence of a 4'-OH group in the B-ring and of 3,3-dimethylallyl and hydroxyl groups in the A-ring. The prenyl and OH groups were located at the 8 and 7-positions in ring A, respectively, based on the presence of two ortho-coupled aromatic hydrogens [δ 6.82 and 6.40 (d, J = 8.2 Hz)] and by NOE and COLOC (Fig. 1) experiments. A NOE was observed for H-5 (δ 6.82) when H-4 α (δ 2.72) was irradiated, and the OH-7 hydrogen $(\delta 5.20)$ exhibited a cross-peak to the aromatic carbon C-6 (δ 108.1). Thus **2**, for which the trivial name brosimine B is proposed, is 4',7-dihydroxy-8-(3,3-dimethylallyl)flavan. The ¹³C-NMR frequencies (Table 1) of 2 were assigned on the basis of HETCOR and COLOC (Fig. 1) NMR experiments.

The known compounds 3',7-dihydroxy-4'-methoxy-flavan (Masaoud, Ripperger, Porzel & Adam, 1995)

isobavachin (Komatsu, Yokoe & Shirataki, 1978), liquiritigenin (Achenbach, Stocker & Constenla, 1988; Fukai, Wang, Inami & Nomura, 1990), hydroxylonchocarpin (Braz Filho, Gottlieb, Mourão, Rocha & Oliveira, 1975), hydroxyisocordoin (Pisteli, Speka, Flamini, Mele & Morelli, 1996) isoliquiritigenin (Achenbach et al., 1988; Fukai et al., 1990), coniferal-dehyde (Borges-Del-Castilho, Bradley-Delso, Ferrero, Bueno & Rodrigues-Luis, 1983), syringaldehyde (Barakat, Nawwar, Buddrus & Michael, 1987), sitosterol and stigmasterol, were identified by comparison of their spectral and physical data with those reported in the literature.

3. Experimental

3.1. General

Mps uncorrected. IR were recorded in NaCl film.

¹H- and ¹³C-NMR spectra were recorded at 300 and 75 MHz, respectively, in CDCl₃ on a Varian GEMINI 300 instrument. EIMS were obtained by direct probe insertion at 70 eV.

3.2. Plant material

Brosimum acutifolium was collected in Portel county, State of Pará, Brazil and identified by Dr. Maria Elisabeth van den Berg of the Botanic Department of Museum Paraense Emílio Goeldi, Belém, Pará.

3.3. Extration and isolation

The powdered bark (2.3 kg) was extracted with hexane, CH₂Cl₂, EtOAc and MeOH, successively, at room temperature. After evaporation of the solvent the CH₂Cl₂ extract (4 g) was subjected to CC on silica gel (70–230 mesh) using hexane–CH₂Cl₂–MeOH at different ratios of increasing polarity furnishing 40 frs. The frs. were analyzed by silica gel TLC with the solvent system hexane–CH₂Cl₂–MeOH (10:10:1) or hexane–Me₂CO (10:3), and were detected by UV light and by spraying with 10% alcoholic H₂SO₄ or ceric sulfate.

Fr. 2 afforded a mixture of coniferaldehyde and syringaldehyde (35 mg). Frs. 15 and 29 afforded stigmasterol (23 mg) and sitosterol (31 mg), respectively. Frs. 24–28 were rechromatographed on silica gel (70–230 mesh) using hexane–CH₂Cl₂–MeOH in order of increasing polarity, furnishing **2** (8 mg) and hydroxylonchocarpin (20 mg). Frs. 33–34 were rechromatographed on silica gel (70–230 mesh) using hexane–Me₂CO in order of increasing polarity to yield **1** (16 mg), 3-7-dihydroxy-4'-methoxyflavan (16 mg) and hydroxyisocordoin (17 mg).

The EtOAc extract (10.77 g) was subjected to CC on

silica gel (70–230 mesh) eluted with a hexane–Me₂CO gradient to give 22 frs. Frs. 10–12 were further purified by CC on silica gel (230–400 mesh) and eluted with CH₂Cl₂–EtOAc (95:5) to furnish isobavachin (12 mg). Fr. 17 after purification on silica gel using hexane–EtOAc–MeOH gradient yielded liquirtigenin (7 mg) and isoliquiritgenin (4 mg).

3.4. Brosimne A (1)

 $[\alpha]_{D}^{25}$ -7.06° (CHCl₃, c 0.35); IR ν_{max}^{film} (cm⁻¹): 3436, 1617, 1465, 1268, 1232; ¹H- and ¹³C-NMR spectral data: see Table 1, HR-MS: m/z 326.1514 for C₂₀H₂₂O₄ (calcd: 326.1518); EIMS m/z (rel. int. %): 326 [M]⁺ (3), 308 [M - H₂O]⁺ (1), 293 [308 - Me]⁺ (1), 173 [C₁₁H₉O₂]⁺ (10), 120 [C₈H₈O]⁺ (35), 107 [C₇H₇O]⁺ (20), 69 [C₅H₅]⁺ (53), 55 [C₄H₇]⁺ (100).

3.5. Brosimine B (2)

 $[\alpha]_{D}^{25}$ -4.88° (CHCl₃, c 0.5); IR ν_{max}^{film} (cm⁻¹): 3387, 1608, 1514, 1450, 1062, ¹H- and ¹³C-NMR spectral data: see Table 1, HR-MS: m/z 310.1568 for $C_{20}H_{22}O_{3}$ (calcd: 310.1569); EIMS m/z (rel. int. %): 310 [M]⁺ (62), 256 [$C_{16}H_{15}O_{3}$]⁺ (53), 190 [$C_{12}H_{14}O_{2}$]⁺ (36), 135 [$C_{8}H_{7}O_{2}$]⁺ (98), 120 [$C_{8}H_{8}O$]⁺ (100), 107 [$C_{7}H_{7}O$]⁺ (90), 91 [$C_{7}H_{7}$]⁺ (55), 55 [$C_{4}H_{7}$]⁺ (19).

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