

## Limonoids from the stem bark of *Khaya grandifoliola*

Hua Zhang<sup>a</sup>, Oluwatoyin A. Odeku<sup>b</sup>, Xiao-Ning Wang<sup>a</sup>, Jian-Min Yue<sup>a,\*</sup>

<sup>a</sup> State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences, 555 Zu Chong Zhi Road, Zhangjiang Hi-Tech Park, Shanghai 201203, People's Republic of China

<sup>b</sup> Department of Pharmaceutics and Industrial Pharmacy, University of Ibadan, Ibadan, Nigeria

Received 10 October 2006; received in revised form 12 March 2007

Available online 25 July 2007

### Abstract

Three limonoids, deacetylkhayanolide E (**1**), 6S-hydroxykhayalactone (**2**), and grandifolide A (**3**), along with three known ones, were isolated from stem bark of the Nigerian medicinal plant *Khaya grandifoliola*. Their structures were characterized on the basis of the application of spectroscopic methods. *In vitro* antimicrobial and cytotoxic activities of the isolates were also tested, but were all found to be inactive.

© 2007 Elsevier Ltd. All rights reserved.

**Keywords:** Meliaceae; Limonoids; Deacetylkhayanolide E; 6S-Hydroxykhayalactone; Grandifolide A; *Khaya grandifoliola*

### 1. Introduction

Some plant species in the Meliaceae and Rubiaceae families, e.g. *Entandrophragma utile*, *Azadirachta indica*, *Morinda lucida*, and *Khaya* species, produce structurally diverse limonoids with important antiparasitic and cytotoxic activities (Obih et al., 1985; Bray et al., 1990; Weenen et al., 1990). There are eight species of *Khaya* growing in tropical regions (Chen et al., 1997), and *K. grandifoliola* has been used for treatment of fever and malaria in Nigeria. In particular, a crude extract of stem bark showed *in vitro* mollusc-killing properties (Makanga and Odyek, 1989), trypanocidal activity against *Trypanosoma brucei* (Owolabi et al., 1990), and antimalarial activity against *Plasmodium bergeri* (Makinde et al., 1988; Awe and Makinde, 1991; Awe et al., 1997). Recent investigations of both the stem bark and the seeds of this plant have led to isolation of eight limonoids and one flavanol (Agbedahunsi and Elujoba, 1998; Tchuendem et al.,

1998; Bickii et al., 2000). In the present paper, we report the isolation and structural elucidation of six additional limonoids, deacetylkhayanolide E (**1**), 6S-hydroxykhayalactone (**2**), grandifolide A (**3**), khayanolide A (**4**, Abdelgaleil et al., 2001), anthothecanolide (**5**, Tchimine et al., 2005), and 3-*O*-acetylanthothecanolide (**6**, Tchimine et al., 2005), of which compounds **1–3** are new. The *in vitro* antimicrobial and cytotoxic activities of compounds **1–6** were also tested, but all tested compounds were inactive.

### 2. Results and discussion

Deacetylkhayanolide E (**1**) had a molecular formula of C<sub>27</sub>H<sub>32</sub>O<sub>10</sub> as deduced from HREIMS at *m/z* 516.1979 [M]<sup>+</sup> (calcd. 516.1995). Its IR spectrum showed the presence of hydroxyl (3498 cm<sup>-1</sup>) and carbonyl (1732 and 1713 cm<sup>-1</sup>) groups. The spectroscopic data of compound **1** (EIMS, IR, and NMR) were very similar to those of khayanolide E (Nakatani et al., 2002). Comparison of the <sup>1</sup>H and <sup>13</sup>C NMR spectroscopic data (Tables 1 and 2) of **1** with those of khayanolide E showed that the only structural difference was the absence of the 1-*O*-acetyl group in **1**, and this was supported by its molecular composition.

\* Corresponding author. Tel./fax: +86 21 5080 6718.

E-mail address: [jmyue@mail.shcnc.ac.cn](mailto:jmyue@mail.shcnc.ac.cn) (J.-M. Yue).

Table 1  
<sup>1</sup>H NMR spectroscopic data for compounds **1–6** (400 MHz)

Proton	<b>1</b> <sup>a</sup>	<b>2</b> <sup>a</sup>	<b>3</b> <sup>a</sup>	<b>3</b> <sup>b</sup>	<b>4</b> <sup>a</sup>	<b>5</b> <sup>c</sup>	<b>6</b> <sup>a</sup>
2	4.83 <i>d</i> (10.4)						
3		4.01 <i>s</i>	5.74 <i>s</i>	4.74 <i>s</i>	4.25 <i>s</i>	3.60 <i>s</i>	5.73 <i>s</i>
5	4.09 <i>br s</i>	4.22 <i>d</i> (1.6)	3.32 <i>d</i> (7.9)	2.38 <i>d</i> (4.0)	2.61 <i>d</i> (6.5)	2.26 <i>dd</i> (9.6, 7.7)	2.26 <i>dd</i> (9.0, 7.2)
6	4.90 <i>br s</i>	4.92 <i>br s</i>	4.93 <i>d</i> (7.9)	4.38 <i>br s</i>	4.86 <i>d</i> (6.5)	2.70 <i>dd</i> (15.6, 9.6)	2.92 <i>dd</i> (14.2, 9.0)
						2.41 <i>dd</i> (15.6, 7.7)	2.74 <i>dd</i> (14.2, 7.2)
9	3.07 <i>d</i> (8.7)	2.44 <sup>h</sup>	2.78 <i>m</i>	1.61 <sup>l</sup>	2.49 <i>dd</i> (13.2, 5.6)	2.48 <i>br d</i> (12.0)	3.22 <i>br d</i> (11.4)
11	2.16 <sup>d</sup>	2.15 <i>m</i>	2.32 <i>m</i>	1.88 <i>m</i>	1.52 <sup>n</sup>	1.65 <i>m</i>	2.32 <i>m</i>
	1.92 <sup>e</sup>	2.04 <i>m</i>	2.02 <i>m</i>	1.59 <sup>l</sup>	1.33 <i>m</i>	1.39 <sup>o</sup>	2.00 <sup>p</sup>
12	2.22 <sup>d</sup>	1.35 <i>m</i>	1.87 <sup>l</sup>	1.82 <i>br d</i> (14.9)	1.60 <i>m</i>	1.69 <i>m</i>	1.97 <sup>p</sup>
	1.08 <i>br d</i> (11.2)	1.10 <i>dd</i> (14.3, 8.7)	1.37 <i>m</i>	1.33 <i>m</i>	1.47 <sup>n</sup>	1.39 <sup>o</sup>	1.54 <i>br d</i> (14.7)
14		1.89 <sup>i</sup>	2.42 <sup>k</sup>				
15	3.83 <i>d</i> (18.6)	3.60 <i>dd</i> (15.0, 13.0)	3.48 <i>d</i> (19.1)	2.86 <sup>m</sup> (2H)	3.52 <i>d</i> (18.2)	3.35 <i>d</i> (19.2)	3.80 <i>d</i> (18.7)
	3.36 <i>d</i> (18.6)	3.00 <i>dd</i> (15.0, 6.0)	3.02 <i>dd</i> (19.1, 8.0)		3.15 <i>d</i> (18.2)	2.73 <i>d</i> (19.2)	3.15 <i>d</i> (18.7)
17	5.85 <i>s</i>	5.37 <i>s</i>	6.10 <i>s</i>	5.32 <i>s</i>	5.87 <i>s</i>	5.56 <i>s</i>	6.11 <i>s</i>
18	1.39 <i>s</i>	0.87 <i>s</i>	1.02 <i>s</i>	0.98 <i>s</i>	1.18 <i>s</i>	0.98 <i>s</i>	0.75 <i>s</i>
19	1.94 <i>s</i>	1.92 <i>s</i>	2.11 <i>s</i>	1.39 <i>s</i>	1.94 <i>s</i>	4.42 <i>d</i> (11.6)	4.83 <i>d</i> (11.2)
						4.15 <i>d</i> (11.6)	4.59 <i>d</i> (11.2)
21	7.65 <sup>f</sup>	7.65 <i>br s</i>	7.53 <i>br s</i>	7.49 <i>br s</i>	7.35 <i>br s</i>	7.62 <i>br s</i>	7.66 <i>br s</i>
22	6.55 <i>br s</i>	6.51 <i>d</i> (1.2)	6.49 <i>d</i> (1.0)	6.40 <i>d</i> (1.2)	6.45 <i>d</i> (1.2)	6.48 <i>d</i> (1.2)	6.54 <i>dd</i> (1.7, 0.6)
23	7.64 <sup>f</sup>	7.63 <i>t</i> (1.6)	7.61 <i>t</i> (1.5)	7.43 <i>t</i> (1.7)	7.61 <i>t</i> (1.7)	7.52 <i>t</i> (1.8)	7.60 <i>t</i> (1.8)
28	1.72 <i>s</i>	1.89 <i>s</i>	1.23 <i>s</i>	0.95 <i>s</i>	1.66 <i>s</i>	0.94 <i>s</i>	1.18 <i>s</i>
29	2.67 <i>d</i> (12.2)	2.60 <i>d</i> (13.7)	1.83 <i>s</i>	1.30 <i>s</i>	2.79 <i>d</i> (12.1)	1.29 <i>s</i>	1.64 <i>s</i>
	2.17 <i>d</i> (12.2)	2.46 <i>d</i> (13.7)			2.40 <i>d</i> (12.1)		
30	3.49 <i>d</i> (10.4)	4.03 <i>d</i> (12.6)	3.60 <i>d</i> (14.1)	2.53 <i>d</i> (14.3)	3.41 <i>s</i>	2.93 <i>d</i> (13.9)	3.59 <i>d</i> (13.8)
		2.78 <i>d</i> (12.6)	2.41 <sup>k</sup>	1.57 <sup>l</sup>		1.50 <i>dd</i> (13.9, 1.5)	2.40 <i>dd</i> (13.8, 1.4)
–COOMe	3.73 <i>s</i>	3.92 <i>s</i>	3.83 <i>s</i>	3.81 <i>s</i>	3.67 <i>s</i>		
CH <sub>3</sub> COO–			1.74 <i>s</i>	2.15 <i>s</i>			1.74 <i>s</i>
1-OH	6.99 <i>br s</i>	8.05 <sup>A</sup>		3.90 <i>br s</i>			7.23 <i>br s</i>
OH	7.57 <sup>g</sup> (6-OH)	8.04 <sup>A</sup> (3-OH)		4.76 <i>br s</i> (2-OH)			10.08 <i>br s</i> (2-OH)
	7.36 <i>br s</i> (8-OH)	7.23 <i>br s</i> (6-OH)		2.88 <sup>m</sup> (6-OH)			7.08 <i>s</i> (14-OH)

<sup>A</sup> Interchangeable signals.

<sup>a</sup> Recorded in C<sub>5</sub>D<sub>5</sub>N.

<sup>b</sup> Recorded in CDCl<sub>3</sub>.

<sup>c</sup> Recorded in CD<sub>3</sub>OD.

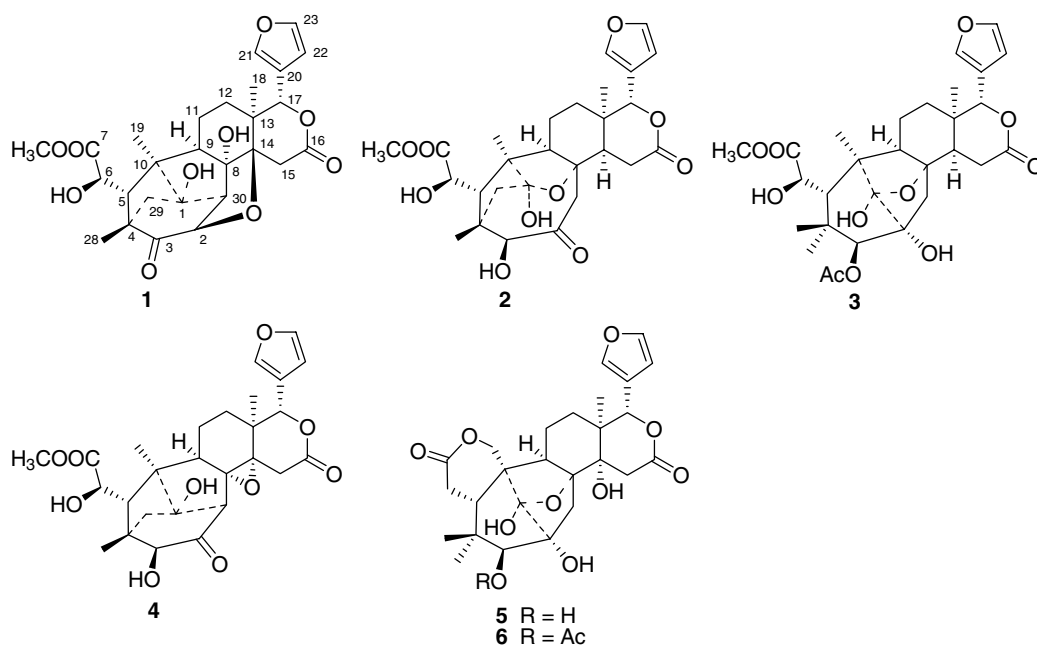
<sup>d–p</sup> Overlapping signals.

The structure of compound **1** was further confirmed by 2D NMR spectra, including HSQC, HMBC, and NOESY spectra (Supplementary data). Compound **1** was therefore determined to be deacetylkhayanolide E.

6*S*-Hydroxykhayalactone (**2**) showed a molecular formula of C<sub>27</sub>H<sub>34</sub>O<sub>10</sub> as determined by HREIMS at *m/z* 518.2146 [M]<sup>+</sup> (calcd. 518.2152), which is 16 mass units more than that of khayalactone (Tchuendem et al., 1998). The spectroscopic data (EIMS, IR, and NMR) of **2** were very similar to those of khayalactone, indicating that the structures of both compounds were related, and the only difference is likely the presence of one more hydroxyl group in **1**. The NMR spectroscopic data further indicated that compound **2** possessed one more oxygenated methine (δ<sub>H</sub> 4.92, *br s*; δ<sub>C</sub> 73.4) and one fewer methylene group than khayalactone, supporting the above deduction. In the HMBC, the correlations of proton signal (δ<sub>H</sub> 4.92, *br s*) to C-4 (δ<sub>C</sub> 47.3), C-5 (δ<sub>C</sub> 44.1), C-7 (δ<sub>C</sub> 176.7), and C-10 (δ<sub>C</sub> 56.6) established that the additional hydroxyl group in **2** was located at C-6. Comparison of the chemical shifts of both H-6 and C-6 of **2** with those of compound **1** sug-

gested that the 6-OH had the *S*-configuration. The structure of **2** was thus elucidated, and confirmed by HMQC, HMBC, and NOESY spectra.

Grandifolide A (**3**) was determined to have a molecular formula C<sub>29</sub>H<sub>38</sub>O<sub>11</sub> by HREIMS at *m/z* 562.2392 [M]<sup>+</sup> (calcd. 562.2414). The NMR spectroscopic data (Tables 1 and 2, in C<sub>5</sub>D<sub>5</sub>N) of compound **3** were very similar to those of 3-*O*-acetylanthothecanolide (**6**, Tchimene et al., 2005), indicating that they were structurally related analogs. In the HMBC spectrum, the correlations of OMe (δ<sub>H</sub> 3.83, *s*)/C-7 (δ<sub>C</sub> 175.8) and H<sub>3</sub>-19 (δ<sub>H</sub> 2.11, *s*)/C-10 (δ<sub>C</sub> 46.2) showed that the partial structure of **3** (C-5 to C-7, C-5 to C-10, and C-10 to C-19) is the same as that of compound **2**. The proton signal at δ<sub>H</sub> 2.42 was assignable to H-14 by its HMBC correlations with C-8 (δ<sub>C</sub> 79.2), C-13 (δ<sub>C</sub> 35.7), and C-15 (δ<sub>C</sub> 28.5), indicating that the C-14 was a methine. The relative stereochemistry of compound **3** was confirmed by NOESY spectroscopic analysis (Supplementary data). The complete assignments of the <sup>1</sup>H and <sup>13</sup>C NMR spectroscopic data of **3** were therefore fully achieved by HMQC spectroscopy.



The three known compounds were identified by spectroscopic methods to be khayanolide A (**4**, Abdelgaleil et al., 2001), anthotheanolide (**5**, Tchimine et al., 2005), and

3-*O*-acetylanthotheanolide (**6**, Tchimine et al., 2005). As different deuterated solvents were used in our research for the NMR measurements of known compounds **4–6**, some protons or carbons of these compounds showed different resonances from the literature values. Therefore, the NMR data of **4** and **6** in  $C_5D_5N$ , and **5** in  $CD_3OD$  are reported for the first time in this paper.

Compounds **1–6** were tested in an antimicrobial assay against *Escherichia coli*, *Staphylococcus aureus*, *S. epidermidis*, and *Candida albicans* by the microdilution method (Yang and Yue, 2001). However, they were all inactive with the MIC values  $>50 \mu\text{g/ml}$ . The *in vitro* activity against the P-388 (murine leukemia) and A-549 (human lung adenocarcinoma) cell lines were also evaluated using the MTT (Alley et al., 1988) and SRB (Skehan et al., 1990) methods, respectively, but none proved to be active.

### 3. Experimental

#### 3.1. General

Optical rotations were determined on a Perkin–Elmer 341 polarimeter, whereas IR spectra were recorded on a Perkin–Elmer 577 spectrometer. NMR spectra were measured on Varian Mercury plus 400 and Bruker AM-400 instruments. EIMS and HREIMS (70 eV) were carried out on a Finnigan MAT 95 mass spectrometer. All solvents used were of analytical grade (Shanghai Chemical Reagents Company, Ltd.). Silica gel (20–40  $\mu\text{m}$ , Qingdao Haiyang Chemical Company, Ltd.) and RP-18 silica gel (150–200 mesh, Merck) were used for cc, with pre-coated silica gel GF254 plates (Yantai Jiangyou Silica Gel Exploitation Company, Ltd.) used for TLC.

Table 2  
 $^{13}\text{C}$  NMR spectroscopic data for compounds **1–6** (100 MHz)

C	<b>1</b> <sup>a</sup>	<b>2</b> <sup>a</sup>	<b>3</b> <sup>a</sup>	<b>3</b> <sup>b</sup>	<b>4</b> <sup>a</sup>	<b>5</b> <sup>c</sup>	<b>6</b> <sup>a</sup>
1	84.8	118.1	110.3	108.3	85.6	109.9	109.0
2	75.8	204.4	80.2	79.5	211.8	83.4	81.9
3	207.3	86.7	84.4	83.7	86.5	82.7	82.8
4	51.8	47.3	38.5	38.1	43.4	39.9	38.1
5	43.1	44.1	46.7	49.1	45.7	38.4	38.0
6	72.7	73.4	71.4	71.0	72.5	31.3	30.7
7	175.9	176.7	175.8	175.6	175.0	178.5	174.5
8	88.5	83.0	79.2	78.2	77.1	82.5	80.3
9	56.8	55.7	64.3	62.9	55.3	55.0	54.1
10	59.4	56.6	46.2	44.1	58.1	46.8	46.2
11	17.1	17.8	22.0	20.4	20.1	22.2	21.2
12	26.6	29.9	36.3	35.4	32.0	32.5	31.3
13	38.4	39.7	35.7	35.2	37.4	42.3	41.1
14	84.3	54.6	45.8	44.7	64.1	73.7	72.4
15	34.2	30.5	28.5	27.6	37.5	38.3	38.3
16	170.4	172.7	170.5	170.4	170.2	173.5	170.9
17	80.7	80.0	78.4	78.1	77.2	79.3	77.7
18	14.6	24.3	22.9	22.5	16.5	16.7	16.3
19	21.0	20.8	22.7	22.3	19.6	75.6	74.4
20	121.9	121.6	122.7	121.2	121.4	123.1	122.2
21	141.7	141.6	141.5	140.7	141.6	143.6	141.7
22	110.9	111.0	110.9	109.9	110.6	112.0	111.0
23	143.4	143.5	143.2	143.0	143.4	145.1	143.2
28	16.2	25.0	25.5	24.8	18.6	25.8	23.8
29	46.5	49.8	24.5	23.9	46.1	23.1	22.7
30	65.1	38.6	41.1	39.7	62.7	43.2	42.9
–COOMe	52.2	52.1	51.8	52.6	51.6		
–OAc			171.1	171.9			170.2
			20.7	21.0			20.6

<sup>a</sup> Recorded in  $C_5D_5N$ .

<sup>b</sup> Recorded in  $CDCl_3$ .

<sup>c</sup> Recorded in  $CD_3OD$ .

### 3.2. Plant material

The stem bark of *K. grandifoliola* (2.5 kg) was collected in August 2005 from the botanical garden of the University of Ibadan, Ibadan, Nigeria, and was identified by Mr. Ayo Owolabi, a botanist of the Department of Botany and Microbiology, University of Ibadan. A voucher specimen (#218) is deposited at the Herbarium of the Department of Pharmacognosy, Faculty of Pharmacy, University of Ibadan, Ibadan, Nigeria.

### 3.3. Extraction and isolation

The powdered stem bark of *K. grandifoliola* was percolated three times with EtOH–H<sub>2</sub>O (85:15, v/v) (3L for each time) at room temperature. The combined extracts were evaporated to dryness under reduced pressure to give a residue (156 g), which was partitioned between H<sub>2</sub>O (2.0 l) and EtOAc (500 ml × 3). The EtOAc solubles were combined and evaporated to dryness in vacuo to give a dry residue (58 g).

The EtOAc extract was separated on a silica gel column eluted with petroleum ether/acetone (50:1 to pure acetone) to obtain five fractions (F1–F5). F2 (14.2 g) was further separated into four sub-fractions using a silica gel column (CHCl<sub>3</sub>/MeOH 200:1 to 5:1). The four sub-fractions were purified by extensive column chromatography (silica gel and RP-18 silica gel) to afford **1** (16 mg), **2** (6 mg), **3** (21 mg), **4** (153 mg), **5** (10 mg), and **6** (120 mg).

### 3.4. Deacetylkhayanolide E (**1**)

White amorphous powder;  $[\alpha]_D^{20}$  – 30.3 (*c* 0.12, C<sub>5</sub>H<sub>5</sub>N); IR  $\nu_{\text{max}}^{\text{KBr}}$  cm<sup>–1</sup>: 3498, 2951, 1732, 1713, 1464, 1389, 1242, 1148, 1045, 1026, 770; EIMS *m/z* (rel. int.): 516 [M]<sup>+</sup> (82), 420 (13), 402 (41), 378 (100), 361 (9), 343 (10), 198 (35), 165 (57), 125 (34); HREIMS *m/z* 516.1979 [M]<sup>+</sup> (calcd. for C<sub>27</sub>H<sub>32</sub>O<sub>10</sub>, 516.1995). For <sup>1</sup>H and <sup>13</sup>C NMR spectroscopic data, see Tables 1 and 2.

### 3.5. 6S-Hydroxykhayalactone (**2**)

White amorphous powder;  $[\alpha]_D^{20}$  – 60.2 (*c* 0.13, C<sub>5</sub>H<sub>5</sub>N); IR  $\nu_{\text{max}}^{\text{KBr}}$  cm<sup>–1</sup>: 3383, 2949, 2879, 1738, 1699, 1460, 1271, 1252, 1171, 1024, 822; EIMS *m/z* (rel. int.): 518 [M]<sup>+</sup> (15), 500 (6), 475 (56), 457 (100), 439 (18), 411 (13), 379 (9), 364 (11), 229 (29); HREIMS *m/z* 518.2146 [M]<sup>+</sup> (calcd. for C<sub>27</sub>H<sub>34</sub>O<sub>10</sub>, 518.2152). For <sup>1</sup>H and <sup>13</sup>C NMR spectroscopic data, see Tables 1 and 2.

### 3.6. Grandifolide A (**3**)

White amorphous powder;  $[\alpha]_D^{20}$  – 41.5 (*c* 0.67, CHCl<sub>3</sub>); IR  $\nu_{\text{max}}^{\text{KBr}}$  cm<sup>–1</sup>: 3446, 2951, 1732, 1635, 1441, 1375, 1240, 1163, 1063, 1028, 876, 602; EIMS *m/z* (rel. int.): 562 [M]<sup>+</sup> (4), 544 (16), 502 (8), 484 (16), 469 (11), 448 (29), 407 (22), 302 (26), 183 (100), 134 (77), 95 (98); HREIMS

*m/z* 562.2392 [M]<sup>+</sup> (calcd. for C<sub>29</sub>H<sub>38</sub>O<sub>11</sub>, 562.2414). For <sup>1</sup>H and <sup>13</sup>C NMR spectroscopic data, see Tables 1 and 2.

## Acknowledgements

We wish to acknowledge the Chinese Academy of Sciences and The Third World Academy of Sciences for the CAS-TWAS Visiting Scholar Fellowship awarded to O.A. Odeku. The financial support from the Ministry of Science and Technology of China (2002CB512807) is greatly acknowledged. We also thank Mr. Ayo Owolabi for the identification of the plant material.

## Appendix A. Supplementary material

Supplementary data (IR, MS, 1D and 2D NMR spectra of compounds **1–6**). Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.phytochem.2007.06.009.

## References

- Abdelgaleil, S.A.M., Okamura, H., Iwagawa, T., Sato, A., Miyahara, I., Doe, M., Nakatani, M., 2001. Khayanolides, rearranged phragmalin limonoid antifeedants from *Khaya senegalensis*. Tetrahedron 57, 119–126.
- Agbedahunsi, J.M., Elujoba, A.A., 1998. Grandifolin from *Khaya grandifoliola* stem bark. Niger. J. Nat. Prod. Med. 2, 34–36.
- Alley, M.C., Scudiero, D.A., Monks, A., Hursey, M.L., Czerwinski, M.J., Fine, D.L., Abbott, B.J., Mayo, J.G., Shoemaker, R.H., Boyd, M.R., 1988. Feasibility of drug screening with panels of human tumor cell lines using a microculture tetrazolium assay. Cancer Res. 48, 589–601.
- Awe, S.O., Makinde, J.M., 1991. Antimalarial effects of the stem bark aqueous extracts of three *Khaya* species. Fitoterapia 62, 467–473.
- Awe, S.O., Olajide, O.A., Adeboye, J.O., Makinde, J.M., 1997. Pharmacological evaluation of *Khaya grandifoliola* methanolic extract. J. Pharm. Res. Dev. 2, 20–23.
- Bickii, J., Njifutie, N., Foyere, J.A., Basco, L.K., Ringwald, P., 2000. In vitro antimalarial activity of limonoids from *Khaya grandifoliola* C.D.C. (Meliaceae). J. Ethnopharmacol. 69, 27–33.
- Bray, D.H., Warhurst, D.C., Connolly, J.D., O'Neill, M.J., Phillipson, J.D., 1990. Plants as sources of antimalarial drugs. Part 7. Activity of some species of Meliaceae and their constituent limonoids. Phytother. Res. 4, 29–35.
- Chen, S.K., Chen, B.Y., Li, H., 1997. In: Chinese Flora (Zhongguo Zhiwu Zhi), vol. 43. Science Press, Beijing, 3, pp. 46.
- Makanga, B., Odyek, O., 1989. Mollusc-killing agents from *Khaya grandifoliola*. Trop. Med. Parasitol. 40, 117–118.
- Makinde, J.M., Awe, S.O., Agbedahunsi, J.M., 1988. Effect of *Khaya grandifoliola* extract on *Plasmodium bergeri bergeri* in mice. Phytother. Res. 2, 30–32.
- Nakatani, M., Abdelgaleil, S.A.M., Kassem, S.M.I., Takezaki, K., Okamura, H., Iwagawa, T., Doe, M., 2002. Three new modified limonoids from *Khaya senegalensis*. J. Nat. Prod. 65, 1219–1221.
- Obih, P.O., Makinde, J.M., Laoye, J., 1985. Investigation of various extracts of *Morinda lucida* for antimalarial actions on *Plasmodium bergeri bergeri* in mice. Afr. J. Med. Sci. 14, 45–49.
- Owolabi, O.A., Makanga, B., Thomas, E.W., Molyneux, D.H., Oliver, R.W., 1990. Trypanocidal potentials of African woody plants: in vitro

- trial of *Khaya grandifoliola* seed extracts against *Trypanosoma brucei*. *J. Ethnopharmacol.* 30, 227–231.
- Skehan, P.A., Storeng, R., Monks, A., McMahon, J., Vistica, D., Warren, J.T., Bokesch, H., Kenney, S., Boyd, M.R., 1990. New colorimetric cytotoxicity assay for anticancer-drug screening. *J. Nat. Can. Inst.* 82, 1107–1112.
- Tchimene, M.K., Tane, P., Ngamga, D., Connolly, J.D., Farrugia, L.J., 2005. Four tetranortriterpenoids from the stem bark of *Khaya anthotheca*. *Phytochemistry* 66, 1088–1093.
- Tchuendem, M.-H.K., Ayafor, J.F., Connolly, J.D., Sterner, O., 1998. Khayalactone, a novel limonoid from *Khaya grandifoliola*. *Tetrahedron Lett.* 39, 719–722.
- Weenen, H., Nkunya, M.H.H., Bray, D.H., Mwasumbi, L.B., Kinabo, L.S., Kilmali, V.A.E.B., 1990. Antimalarial activity of Tanzanian medicinal plants. *Planta Med.* 56, 368–370.
- Yang, S.P., Yue, J.M., 2001. Two novel cytotoxic and antimicrobial triterpenoids from *Pseudolarix kaempferi*. *Bioorg. Med. Chem. Lett.* 11, 3119–3122.