

# Alisolide, alisols O and P from the rhizome of *Alisma orientale*

Ming Zhao<sup>a</sup>, Li-Jia Xu<sup>b</sup>, Chun-Tao Che<sup>a,\*</sup>

<sup>a</sup> School of Chinese Medicine, The Chinese University of Hong Kong, Hong Kong

<sup>b</sup> Institute of Medicinal Plant Development, Peking Union Medical College, Chinese Academy of Medical Sciences, Beijing, PR China

Received 11 August 2006; received in revised form 21 April 2007

Available online 26 July 2007

## Abstract

A *nor*-protostane, alisolide (**1**), a rearranged protostane, alisol O (**2**), and a 2,3-*seco*-protostane triterpene, alisol P (**3**), were isolated from the rhizomes of *Alisma orientale*, along with eight known protostane triterpenes. The structures were elucidated to be (17*S*)-3,11-dioxo-23-*nor*-protost-12-en-23(17)-olide, 3-oxo-11 $\beta$ ,23-dihydroxy-24,24-dimethyl-26,27-dinorprotost-13(17)-en-25-oic acid, and (20*R*,23*S*,24*R*)-23,24,25-trihydroxy-2,3-*seco*-protost-13(17)-en-3-oic acid 2,11 $\beta$ -lactone, respectively, by interpretation of spectroscopic data.

© 2007 Elsevier Ltd. All rights reserved.

**Keywords:** *Alisma orientale*; Alismataceae; Protostane triterpenes; Alisolide; Alisol O; Alisol P

## 1. Introduction

*Alisma orientale* (Sam.) Juz. (Alismataceae) is distributed in China, Japan, Mongolia and Russia. The dried rhizome of this plant is a common traditional Chinese medicine known as Rhizoma Alismatis or Zexie, which is a diuretic agent used to “remove dampness and promote water metabolism” in the body according to Chinese medicinal principles (Sun, 1992). Previous studies have shown that it contains protostane triterpenes (Murata and Miyamoto, 1970; Murata et al., 1970; Fukuyama et al., 1988; Geng et al., 1988; Yoshikawa et al., 1993a,b, 1997, 1999; Nakajima et al., 1994; Zeng et al., 1995; Matsuda et al., 1999; Peng and Lou, 2001a; Peng et al., 2002a,b), guaiane sesquiterpenes (Yoshikawa et al., 1994a,b; Peng and Lou, 2001b; Peng et al., 2002c, 2003) and kaurane diterpenes (Nakajima et al., 1994; Yamaguchi et al., 1994; Peng and Lou, 2002). To date, all protostane triterpenes occur only in *Alisma* plants, and they are considered to be chemotaxonomic markers of the genus. In the

present study, alisolide (**1**), alisol O (**2**) and alisol P (**3**) (Fig. 1), were isolated from the rhizome of *A. orientale*, along with eight known protostane triterpenes, alisol A (Yoshikawa et al., 1993b), alisol B, alisol B 23-acetate, 25-*O*-methylalisol A (Nakajima et al., 1994), 25-anhydroalisol A 24-acetate, 25-anhydroalisol A (Peng et al., 2002b), alisol E 23-acetate (Yoshikawa et al., 1993b), as well as 13 $\beta$ ,17 $\beta$ -epoxyalisol A (Peng et al., 2002a).

## 2. Results and discussion

The rhizomes of *Alisma orientale*, collected in Si-Chuan Province, China, were percolated with EtOH–H<sub>2</sub>O (4:1, v/v). After removal of excessive EtOH, the extract separated into an oil and an aqueous layer. Alisolide (**1**), alisols O (**2**) and P (**3**) were purified from the organic portion by repeated column chromatography over silica gel and RP-18 gels.

Alisolide (**1**), C<sub>26</sub>H<sub>36</sub>O<sub>4</sub>, displayed a quasi-molecular ion at *m/z* 413.27096 [M+H]<sup>+</sup> (calculated 413.26864). It showed IR absorption bands at 3447, 1782, 1705, and 1663 cm<sup>−1</sup> assignable to hydroxyl, lactone, ketone, and

\* Corresponding author. Tel.: +852 2609 8130; fax: +852 2603 7203.  
E-mail address: [checht@cuhk.edu.hk](mailto:checht@cuhk.edu.hk) (C.-T. Che).

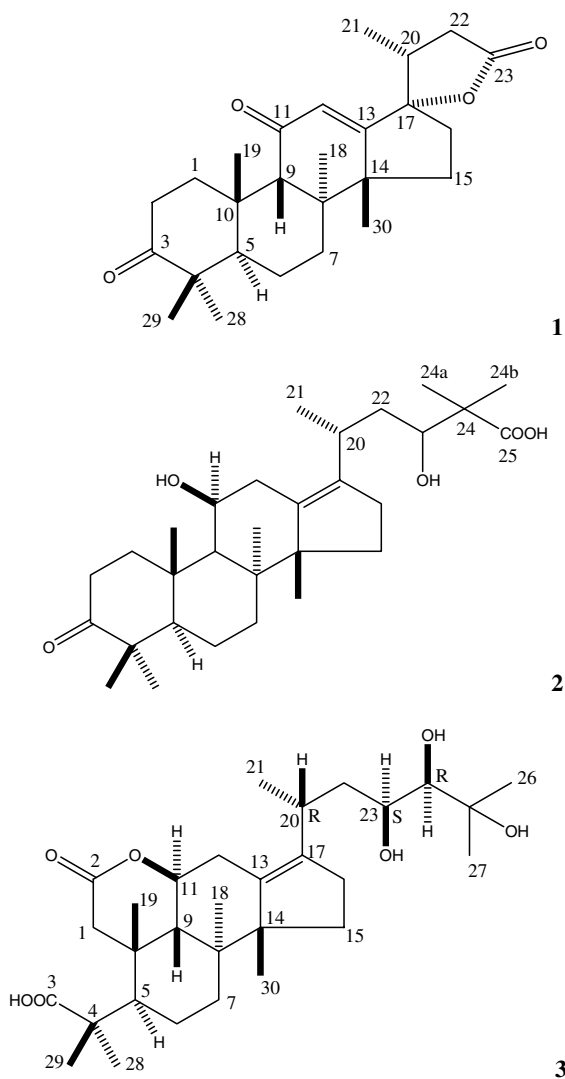


Fig. 1. Chemical structures of 1–3.

enone functional groups, respectively. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopic data of **1** (Table 1) were in agreement with those of the A-, B-, and C-rings of alisol J 23-acetate (Zeng et al., 1995). It was noted, however, that signals for carbons C-24–C-27 were missing from the spectra of **1**. The compound was, therefore, a 23-nor-protostane bearing identical A–C rings as in alisol J 23-acetate. The structure of **1** was further established by interpretation of the COSY, DEPT, HMQC and HMBC data. That is, HMBC correlations were observed between the lactone carbon ( $\delta_{\text{C}}$  174.7) and H-20/H-22. A downfield shift of C-17 ( $\delta_{\text{C}}$  93.8) suggested a C-23(17) lactone structure. To the best of our knowledge, all known structures of protostane triterpenes have a 20*R* stereochemistry, **1** was therefore assumed to have the same stereochemistry at C-20 on the basis of biogenetic ground. The absolute configuration of C-17 was then determined to be *S* based on NOESY results, in particular the correlation observed between H-12 and CH<sub>3</sub>-21 (Fig. 2). Finally, the structure of (17*S*)-3,11-dioxo-23-nor-protost-12-en-23(17)-olide was assigned to alisolide (**1**). In

the literature, only three 23-nor-protostanes have been reported as oxidative reaction products (Murata et al., 1968; Lee et al., 2003). Alisolide (**1**) is therefore the first 23-nor-protostane structure reported from a natural source.

Alisol O (**2**), C<sub>30</sub>H<sub>48</sub>O<sub>5</sub>, displayed a quasi-molecular ion peak at  $m/z$  511.33999 [ $\text{M}+\text{Na}$ ]<sup>+</sup> (calculated 511.33994) in the HRESIMS spectrum and the IR absorption bands at 3436 and 1706 cm<sup>−1</sup> suggested the presence of hydroxyl and carboxyl groups. Analysis of the  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopic data of **2** (Table 1) resulted in structural similarity with alisol B (Nakajima et al., 1994), and the structure of the side-chain was determined by an HMBC analysis. Thus, correlation signals were observed between: H-23 ( $\delta_{\text{H}}$  3.47, *d*,  $J$  = 10.4 Hz) and C-20, C-22, C-24, C-24a, and C-25; CH<sub>3</sub>-24a ( $\delta_{\text{H}}$  1.05) and C-25; CH<sub>3</sub>-24b ( $\delta_{\text{H}}$  1.08) and C-25; CH<sub>3</sub>-21 ( $\delta_{\text{H}}$  0.97, *d*,  $J$  = 5.6 Hz) and C-22, respectively. The stereostructure of **2** was then established by interpretation of the NOESY results (Fig. 2), in which correlations between CH<sub>3</sub>-28 and H-5, CH<sub>3</sub>-18 and H-5, CH<sub>3</sub>-18 and H-11, CH<sub>3</sub>-19 and H-9, as well as between CH<sub>3</sub>-30 and H-9, were observed. The 11-OH was assigned to a  $\beta$  position due to the NOESY correlation observed between H-11 and CH<sub>3</sub>-18. Consequently, alisol O (**2**) was elucidated to be 3-oxo-11 $\beta$ ,23-dihydroxy-24,24-dimethyl-26,27-dinorprotost-13(17)-en-25-oic acid. It is a new rearranged skeleton of protostane, presumably biosynthesized from alisol B (Briggs et al., 1973).

Alisol P (**3**), C<sub>30</sub>H<sub>48</sub>O<sub>7</sub>, exhibited a quasi-molecular ion peak at  $m/z$  543.32942 [ $\text{M}+\text{Na}$ ]<sup>+</sup> (Calc. 543.32922) in its positive ion HRMS spectrum. The IR spectrum showed absorption bands at 3500–2500, 1748, and 1710 cm<sup>−1</sup>, suggesting the presence of hydroxyl, lactone, and carboxyl functional groups, respectively. The  $^1\text{H}$  NMR spectrum of **3** (Table 1) displayed signals due to seven tertiary methyls ( $\delta_{\text{H}}$  1.03, 1.15, 1.15, 1.22, 1.22, 1.23, 1.28) and a secondary methyl group ( $\delta_{\text{H}}$  1.03). Its  $^{13}\text{C}$  DEPT spectra (Table 1) indicated the presence of thirty signals including two carbonyl carbons, two quaternary olefinic carbons, five sp<sup>3</sup> quaternary carbons (one oxygenated), six sp<sup>3</sup> methines (three oxygenated), seven sp<sup>3</sup> methylenes, and eight methyls. The NMR spectroscopic data of **3** were similar to those of a 2,3-*seco*-protostane-type triterpene such as alismalactone 23-acetate (Yoshikawa et al., 1997), except for the side-chain resonances. Compound **3** was, therefore, likely a 2,3-*seco*-protostane possessing identical A–D rings as for alismalactone 23-acetate. An analysis of the HMBC spectrum supported this assignment. The side-chain was determined to possess 23,24,25-trihydroxy substituents based on interpretation of the HMBC and  $^1\text{H}$ – $^1\text{H}$  COSY spectroscopic data. The stereostructure of **3** was then established on the basis of NOESY results (Fig. 2), in which NOE correlations were observed between: H-5 and CH<sub>3</sub>-18/CH<sub>3</sub>-28; CH<sub>3</sub>-18 and H-11; CH<sub>3</sub>-19 and H-9/CH<sub>3</sub>-29; H-9 and CH<sub>3</sub>-30; H-23 and H-24; H-23 and CH<sub>3</sub>-27, and H-24 and CH<sub>3</sub>-26/CH<sub>3</sub>-27, respectively. Since H-11 exhibited a NOESY correlation with CH<sub>3</sub>-18, it was assigned to

Table 1  
<sup>1</sup>H and <sup>13</sup>C NMR spectroscopic data for alisolide (1), alisols O (2) and P (3)<sup>a</sup>

Position	1 (CDCl <sub>3</sub> )		2 (CDCl <sub>3</sub> )		3 (CDCl <sub>3</sub> )	
	δ <sub>H</sub>	δ <sub>C</sub>	δ <sub>H</sub>	δ <sub>C</sub>	δ <sub>H</sub>	δ <sub>C</sub>
1	1.92, 2.45	32.4 <i>t</i>	2.09, 2.22	31.0 <i>t</i>	2.56, <i>s</i>	42.9 <i>t</i>
2	2.31, 2.70	33.7 <i>t</i>	2.31, 2.66	33.7 <i>t</i>		173.9 <i>s</i>
3		218.9 <i>s</i>		220.2 <i>s</i>		183.5 <i>s</i>
4		46.9 <i>s</i>		47.0 <sup>b</sup> <i>s</i>		45.2 <i>s</i>
5	2.20, <i>m</i>	48.3 <i>d</i>	2.08	48.4 <i>d</i>	2.67, <i>m</i>	44.8 <i>d</i>
6	1.55, <i>m</i>	20.2 <i>t</i>	1.23, 1.42	20.0 <i>t</i>	1.60, 1.75	18.8 <i>t</i>
7	1.40, 2.18	33.3 <i>t</i>	1.23, 2.01	34.2 <i>t</i>	1.40, 1.58	28.7 <i>t</i>
8		44.4 <i>s</i>		40.5 <i>s</i>		38.7 <i>s</i>
9	2.59, <i>m</i>	55.3 <i>d</i>	1.74, <i>d</i> (10.8)	49.3 <i>d</i>	1.73, <i>d</i> (11.0)	52.3 <i>d</i>
10		37.2 <i>s</i>		37.0 <i>s</i>		38.3 <i>s</i>
11		199.0 <i>s</i>	3.82, <i>m</i>	70.0 <i>d</i>	4.16, <i>dt</i> (5.2, 11.0)	76.7 <i>d</i>
12	5.82, <i>s</i>	124.2 <i>d</i>	2.78, 2.00	33.8 <i>t</i>	2.05, 3.03	30.1 <i>t</i>
13		166.3 <i>s</i>		138.6 <i>s</i>		136.3 <i>s</i>
14		51.3 <i>s</i>		57.0 <i>s</i>		56.6 <i>s</i>
15	1.45, 2.13	30.1 <i>t</i>	1.31, 1.85	30.5 <i>t</i>	1.37, 1.83	30.2 <i>t</i>
16	2.03, 2.17	35.9 <i>t</i>	2.11	28.9 <i>t</i>	2.22, <i>br s</i>	29.0 <i>t</i>
17		93.8 <i>s</i>		134.9 <i>s</i>		139.2 <i>s</i>
18	1.27, <i>s</i>	24.4 <i>q</i>	0.98, <i>s</i>	24.2 <i>q</i>	1.03, <i>s</i>	20.3 <i>q</i>
19	1.07, <i>s</i>	25.1 <i>q</i>	1.01, <i>s</i>	25.6 <i>q</i>	1.15, <i>s</i>	29.2 <i>q</i>
20	2.62, <i>m</i>	37.8 <i>d</i>	2.82, <i>m</i>	28.4 <i>d</i>	2.80, <i>m</i>	28.3 <i>d</i>
21	1.07, <i>d</i> (7.0)	15.8 <i>q</i>	0.97, <i>d</i> (5.6)	20.3 <i>q</i>	1.03, <i>d</i> (7.0)	20.3 <i>q</i>
22	2.84, <i>dd</i> (8.0, 17.6)	36.9 <i>t</i>	1.18, <i>m</i>	36.3 <i>t</i>	1.39, <i>m</i>	39.9 <i>t</i>
	2.41, <i>dd</i> (7.2, 17.6)		1.37, <i>m</i>		1.60, <i>m</i>	
23		174.7 <i>s</i>	3.47, <i>d</i> (10.4)	74.7 <i>d</i>	3.77, <i>dd</i> (3.5, 9.3)	69.6 <i>d</i>
24				46.9 <sup>b</sup> <i>s</i>	3.00, <i>br s</i>	77.4 <i>d</i>
25				181.5 <i>s</i>		74.4 <i>s</i>
26/24a			1.05, <i>s</i>	22.9 <i>q</i>	1.28, <i>s</i>	27.2 <i>q</i>
27/24b			1.08, <i>s</i>	18.2 <i>q</i>	1.22, <i>s</i>	26.2 <i>q</i>
28	1.10, <i>s</i>	29.4 <i>q</i>	1.05, <i>s</i>	29.6 <i>q</i>	1.22, <i>s</i>	29.7 <i>q</i>
29	1.11, <i>s</i>	19.4 <i>q</i>	1.03, <i>s</i>	20.1 <i>q</i>	1.23, <i>s</i>	20.9 <i>q</i>
30	1.29, <i>s</i>	22.4 <i>q</i>	1.13, <i>s</i>	23.1 <i>q</i>	1.15, <i>s</i>	21.2 <i>q</i>

<sup>a</sup> *J* values (Hz) are in parentheses. Multiplicities of carbons were determined by a DEPT experiment. 400 MHz for <sup>1</sup>H and 100 MHz for <sup>13</sup>C NMR.

<sup>b</sup> Assignments in same vertical column may be interchangeable.

the α position. The stereochemistry of the side-chain was further assigned as (20*R*, 23*S*, 24*R*) on the basis of the NOESY correlations observed among H-23, H-24, CH<sub>3</sub>-26 and CH<sub>3</sub>-27. Further support of such an assignment was obtained by comparing the NMR spectroscopic data of the side-chain in 3 to those of alisol A (20*R*, 23*S*, 24*R*) (Nakajima et al., 1994) (Table 2). Alisol E, which has a 20*R*, 23*S*, 24*S* stereostructure, exhibits different NMR results (Yoshikawa et al., 1993b) (Table 2). Finally, Alisol P (3) was elucidated to be (20*R*, 23*S*, 24*R*)-23,24,25-trihydroxy-2,3-*seco*-protost-13(17)-en-3-oic acid 2,11β-lactone.

During the course of this study, the X-ray crystallographic data of alisol B 23-acetate (4) (Fig. 3) was also obtained and is now reported for the first time. The compound was previously obtained from *Alisma orientale* as a secondary metabolite in Zexie (Nakajima et al., 1994).

### 3. Concluding remarks

Protostane triterpenes are stereoisomers of dammaranes and characterized by the 8α-CH<sub>3</sub>, 9β-H, 13α-H, and 14β-CH<sub>3</sub> structure. To date, all naturally occurring protostane

triterpenes originate from the *Alisma* genus of plants and they are considered to be chemotaxonomic markers of this taxon. During the course of studying the chemical composition of a Chinese medicinal plant, *A. orientale*, we have obtained several protostane derivatives. Three of these secondary metabolites (1–3) are shown to be new structures, of which alisolide (1) is the first *nor*-protostane reported from a natural source.

## 4. Experimental

### 4.1. General

IR spectra were recorded on a Perkin–Elmer 16PC FT-IR spectrometer, whereas <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopic data were obtained using a JEOL JNM-EX-400 FT-NMR spectrometer at 400 and 100 MHz, respectively, with TMS as internal standard. EI MS were determined on a Finnigan TSQ7000 triple quadrupole mass spectrometer, whereas APCI MS were determined on an Agilent HP 1100 series SL Trap MSD. The X-ray crystallography data were obtained using a Siemens P4-RA diffractometer. CC

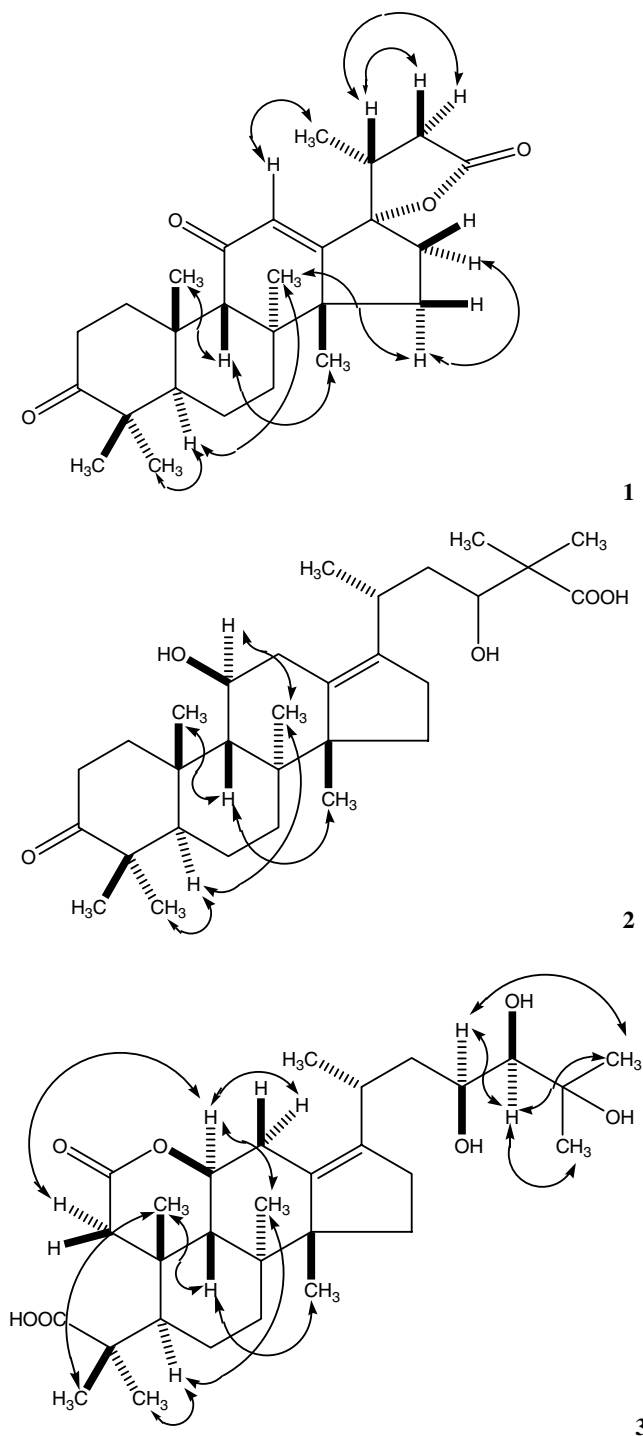


Fig. 2. Key NOESY correlations of 1–3.

was carried out on silica gel (100–200 mesh, 230–400 mesh), RP-18 silica gel (40–63  $\mu\text{m}$ ) and Sephadex LH-20, respectively, and TLC was conducted on silica gel 60 F<sub>254</sub> and RP-18 F<sub>254</sub> plates.

#### 4.2. Plant material

The dried rhizomes of *Alisma orientale* (Sam.) Juzep. were collected in Si-chuan Province, China, in 2002, and

Table 2

Comparison of NMR spectroscopic data of alisol P (3), alisol A and alisol E<sup>a</sup>

	Alisol P (3)	Alisol A <sup>b</sup> (20 <i>R</i> , 23 <i>S</i> , 24 <i>R</i> )	Alisol E <sup>c</sup> (20 <i>R</i> , 23 <i>S</i> , 24 <i>S</i> )
C-20	28.3	28.7	28.0
C-21	20.3	20.3	20.4
C-22	39.9	40.4	38.5
C-23	69.6	69.5	73.2
H-23	3.75 ( <i>dd</i> , <i>J</i> = 3.6, 9.6 Hz)	3.77 ( <i>dd</i> , <i>J</i> = 3.5, 9.3 Hz)	3.47 ( <i>t</i> -like)
C-24	77.5	77.6	79.1
H-24	3.00 ( <i>br s</i> )	3.00 ( <i>br s</i> )	3.27 ( <i>d</i> , <i>J</i> = 6 Hz)
C-25	74.4	74.1	71.6
C-26	27.2	27.6	25.9
C-27	26.2	26.4	26.4

<sup>a</sup> Chemical shifts expressed in ppm.

<sup>b</sup> Data extracted from Nakajima et al. (1994).

<sup>c</sup> Data extracted from Yoshikawa et al. (1993b).

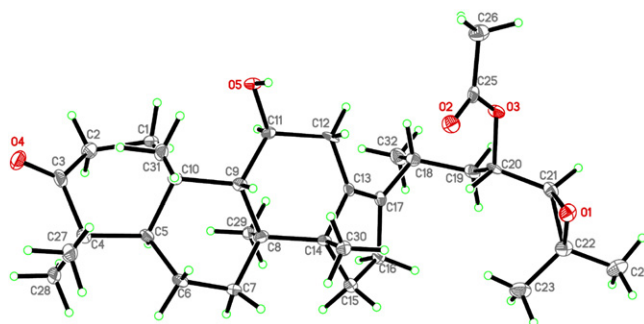


Fig. 3. X-ray structure of alisol B 23-acetate (4).

identified by Prof. Rui-Chao Lin of the Beijing Institute of Control of Biological and Pharmaceutical Products, Bei-jing. Voucher specimens (No. SCM072002) have been deposited in the School of Chinese Medicine, The Chinese University of Hong Kong.

#### 4.3. Extraction and isolation

Dried rhizomes of *Alisma orientale* (2.5 kg) were powdered and percolated with EtOH–H<sub>2</sub>O (4:1, v/v) at room temperature until the percolate became colorless. After removal of excessive EtOH under reduced pressure, the remaining solution was allowed to stand for several hours to separate into an oil and an aqueous layer. The aqueous layer was then further extracted with EtOAc (2 L  $\times$  3 times). The oil layer was subjected to silica gel CC eluted with a petroleum ether–EtOAc gradient (9:1  $\rightarrow$  1:1) to afford three fractions (Fr. I–III). Fr. I was fractionated by silica gel CC using mixtures of petroleum ether–EtOAc (10:0  $\rightarrow$  8:2) as mobile phase to yield thirty fractions (Fr. I<sub>1</sub>–I<sub>30</sub>). Fr. I<sub>15</sub> was subjected to further separation on a RP-18 column eluted with MeOH–H<sub>2</sub>O (4:1, v/v) to yield alisolide (1, 8 mg). Fr. I<sub>20</sub> yielded a crop of alisol B 23-ace-

tate (**4**) (50 mg) upon purification over RP-18 silica gel. Fr. II was subjected to further silica gel CC eluted with petroleum ether–EtOAc (8:2 → 6:4) to afford 25 fractions (Fr. II<sub>1</sub>–II<sub>25</sub>). From Fr. II<sub>6</sub>, alisol B (15 mg) was obtained after chromatography on RP-18 silica gel. Fr. II<sub>12</sub> was further purified on a RP-18 column (MeOH–H<sub>2</sub>O, 5.5:4.5 → 7.5:2.5) followed by silica gel chromatography (CHCl<sub>3</sub>–MeOH, 10:0 → 9.8:0.2) to afford alisol O (**2**, 10 mg). Fr. II<sub>14–16</sub> and Fr. II<sub>18–21</sub> were subjected to silica gel CC eluted with mixture of CHCl<sub>3</sub>–MeOH (10:0 → 9.7:0.3), respectively, to afford alisol A (16 mg), 25-*O*-methylalisol A (11 mg), 25-anhydroalisol A 24-acetate (10 mg), and 25-anhydroalisol A (15 mg). Fr. II<sub>23</sub> was purified over a RP-18 column (MeOH–H<sub>2</sub>O, 6:4 → 6.5:3.5, as mobile phase) to obtain alisol E 23-acetate (12 mg). Lastly, Fr. III and the EtOAc-soluble portion of the aqueous layer were combined and purified over a silica gel column using CHCl<sub>3</sub>–MeOH (10:0 → 7:3) as mobile phase to afford twenty fractions (Fr. III<sub>1–20</sub>). Fr. III<sub>10–15</sub> was applied to a silica gel column (eluted with CHCl<sub>3</sub>–MeOH, 9:1 → 8:2) and then on a RP-18 column (eluted with MeOH–H<sub>2</sub>O, 5.5:4.5 → 6.5:3.5) to afford alisol P (**3**, 15 mg) and 13 $\beta$ ,17 $\beta$ -epoxyalisol A (15 mg), respectively.

#### 4.4. Alisolide (**1**)

Colorless crystal, C<sub>26</sub>H<sub>36</sub>O<sub>4</sub>; IR (KBr)  $\nu_{\max}$ : 3447, 2980, 1782, 1705, and 1663 cm<sup>−1</sup>. For <sup>1</sup>H and <sup>13</sup>C NMR spectroscopic data, see Table 1; APCIMS *m/z* 413 [M+H]<sup>+</sup>; HRMS Calc. for C<sub>26</sub>H<sub>37</sub>O<sub>4</sub>H [M+H]<sup>+</sup> 413.26864, found *m/z* 413.27096.

#### 4.5. Alisol O (**2**)

White powder, C<sub>30</sub>H<sub>48</sub>O<sub>5</sub>; IR (KBr)  $\nu_{\max}$ : 3436, 1706 cm<sup>−1</sup>; For <sup>1</sup>H and <sup>13</sup>C NMR spectroscopic data, see Table 1; HRESIMS Calc. for C<sub>30</sub>H<sub>48</sub>O<sub>5</sub>Na [M+Na]<sup>+</sup> 511.33994, found *m/z* 511.33999.

#### 4.6. Alisol P (**3**)

Colorless crystal, C<sub>30</sub>H<sub>48</sub>O<sub>7</sub>; IR (KBr)  $\nu_{\max}$ : 3500–2500, 1748 and 1710 cm<sup>−1</sup>. For <sup>1</sup>H and <sup>13</sup>C NMR spectroscopic data, see Table 1; APCIMS *m/z* 520 [M]<sup>+</sup>; HRMS Calc. for C<sub>30</sub>H<sub>48</sub>O<sub>7</sub>Na [M+Na]<sup>+</sup> 543.32922, found *m/z* 543.32942.

#### 4.7. X-ray data for alisol B 23-acetate (**4**)

Colorless prisms, C<sub>32</sub>H<sub>50</sub>O<sub>5</sub>. Crystal size 0.40 × 0.35 × 0.30 mm<sup>3</sup>; The space group is *P*2<sub>1</sub>2<sub>1</sub>2<sub>1</sub>; The crystals belong to the orthorhombic system with cell parameters *a* = 7.5492(6) Å, *b* = 14.5156(11) Å, *c* = 26.888(2) Å, *V* = 2946.4(4) Å<sup>3</sup>, *Z* = 4, *D*<sub>calcd</sub> = 1.160 mg/m<sup>3</sup>,  $\mu$  = 0.076 mm<sup>−1</sup>, *F*(000) = 1128. The structures were refined by full-matrix least-squares on *F*<sup>2</sup> using SHELXL-97. Final discrepancy indices of *R*<sub>1</sub> = 0.0565,

*wR*<sub>2</sub> = 0.1156 and GOOF = 1.007 for observed data with *I* > 2 $\sigma$ (*I*). The final difference electron density map contains maximum and minimum peak heights of 0.232 and −0.209 e Å<sup>−3</sup>. Crystallographic data (excluding structure factors) for the structural analysis have been deposited with the Cambridge Crystallographic Data Center as supplementary publication numbers CCDC 612128. Copies of the data can be obtained, free of charge, on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK [fax: +44(0)-1223-336033; e-mail: deposit@ccdc.cam.ac.uk].

#### Acknowledgements

The authors are grateful to Dr. Shu-Guang Wang, Dr. Xiao-Qi Zhang, and Dr. Ying Wang, for providing IR, NMR and X-ray crystallographic data.

#### References

- Briggs, L.H., Bartley, J.P., Rutledge, P.S., 1973. Degradation of the lanosterol side chain. *J. Chem. Soc., Perkin Trans. 1: Org. Bioorg. Chem.* 8, 806–809.
- Fukuyama, Y., Geng, P.W., Wang, R., Yamada, T., Nakagawa, K., 1988. 11-Deoxyalisol C and alisol D: new protostane-type triterpenoids from *Alisma plantago-aquatica*. *Planta Med.* 54, 445–447.
- Geng, P.W., Fukuyama, Y., Yamada, T., Wang, R., Bao, J.X., 1988. Triterpenoids from the rhizome of *Alisma plantago-aquatica*. *Phytochemistry* 27, 1161–1164.
- Lee, S.M., Kim, J.H., Zhang, Y., An, R.B., Min, B.S., Joung, H., Lee, H.K., 2003. Anti-complementary activity of protostane-type triterpenes from *Alismatis rhizoma*. *Arch. Pharmacol. Res.* 26, 463–465.
- Matsuda, H., Kageura, T., Toguchida, I., Murakami, T., Kishi, A., Yoshikawa, M., 1999. Effects of sesquiterpenes and triterpenes from the rhizome of *Alisma orientale* on nitric oxide production in lipopolysaccharide-activated macrophages: absolute stereostructures of alismaketones-B 23-acetate and -C 23-acetate. *Bioorg. Med. Chem. Lett.* 9, 3081–3086.
- Murata, T., Miyamoto, M., 1970. Biological-active triterpenes of *Alismatis rhizoma*. II. Structures of alisol A and alisol A monoacetate. *Chem. Pharm. Bull.* 18, 1354–1361.
- Murata, T., Shinohara, M., Hirata, T., Kamiya, K., Nishikawa, M., Miyamoto, M., 1968. New triterpenes of *Alisma plantago-aquatica* var *orientale*. *Tetrahedron Lett.* (1), 103–108.
- Murata, T., Shinohara, M., Miyamoto, M., 1970. Biological-active triterpenes of *Alismatis rhizoma*. IV. Structures of alisol B, alisol B monoacetate and alisol C monoacetate. Some reactions of the  $\alpha$ -hydroxy epoxide of the alisol B derivatives. *Chem. Pharm. Bull.* 18, 1369–1384.
- Nakajima, Y., Satoh, Y., Katsumata, M., Tsujiyama, K., Ida, M., Shoji, J., 1994. Triterpenoids of *Alisma orientale* rhizomes and the crude drug *Alismatis rhizoma*. *Phytochemistry* 36, 119–127.
- Peng, G.P., Lou, F.C., 2001a. Terpenoids of *Alisma orientalis* Juzep. *Nat. Prod. Res. Dev.* 13 (4), 1–4.
- Peng, G.P., Lou, F.C., 2001b. Structure determination of oplopanone by 2D inadequate experiment. *Nat. Prod. Res. Dev.* 13 (6), 9–11.
- Peng, G.P., Lou, F.C., 2002. Isolation and identification of diterpenes from *Alisma orientalis* Juzep. *Acta Pharm. Sin.* 37, 950–954.
- Peng, G.P., Zhu, G.Y., Lou, F.C., 2002a. Terpenoids from *Alisma orientalis* Juzep. *Nat. Prod. Res. Dev.* 14 (6), 7–10.
- Peng, G.P., Zhu, G.Y., Lou, F.C., 2002b. Two novel terpenoids from *Alisma orientalis* Juzep. *Nat. Prod. Res. Dev.* 14 (4), 5–8.

- Peng, G.P., Lou, F.C., Huang, X.F., Tian, G., 2002c. Structure of orientanone from *Alisma orientalis*, a novel sesquiterpene originating from guaiane-type carbon skeleton by isopropyl shift. *Tetrahedron* 58, 9045–9048.
- Peng, G.P., Tian, G., Huang, X.F., Lou, F.C., 2003. Guaiane-type sesquiterpenoids from *Alisma orientalis*. *Phytochemistry* 63, 877–881.
- Sun, X.Z., 1992. In: *Flora Reipublicae Popularis Sinicae*, vol. 8. China Science Press, Ltd., Beijing, pp. 127–145.
- Yamaguchi, K., Ida, Y., Satoh, Y., Nakajima, Y., Shoji, J., 1994. A kaurane derivative isolated from *Alisma orientale*. *Acta Crystallogr., Sect. C: Cryst. Struct. Commun.* C50, 738–740.
- Yoshikawa, M., Hatakeyama, S., Tanaka, N., Matsuoka, T., Yamahara, J., Murakami, N., 1993a. Crude drugs from aquatic plants. II. On the constituents of the rhizome of *Alisma orientale* Juzep. originating from Japan, Taiwan, and China. Absolute stereostructures of 11-deoxyalisols B and B 23-acetate. *Chem. Pharm. Bull.* 41, 2109–2112.
- Yoshikawa, M., Hatakeyama, S., Tanaka, N., Fukuda, Y., Yamahara, J., Murakami, N., 1993b. Crude drugs from aquatic plants. I. On the constituents of *Alismatis rhizoma*. (1). Absolute stereostructures of alisols E 23-acetate, F, and G, three new protostane-type triterpenes from Chinese *Alismatis rhizoma*. *Chem. Pharm. Bull.* 41, 1948–1954.
- Yoshikawa, M., Yamaguchi, S., Matsuda, H., Kohda, Y., Ishikawa, H., Tanaka, N., Yamahara, J., Murakami, N., 1994a. Crude drugs from aquatic plants. IV. On the constituents of *Alismatis rhizoma*. (2). Stereostructures of bioactive sesquiterpenes, alismol, alismoxide, orientalols A, B, and C, from Chinese *Alismatis rhizoma*. *Chem. Pharm. Bull.* 42, 1813–1816.
- Yoshikawa, M., Yamaguchi, S., Matsuda, H., Tanaka, N., Yamahara, J., Murakami, N., 1994b. Crude drugs from aquatic plants. V. On the constituents of *Alismatis rhizoma*. (3). Stereostructures of water-soluble bioactive sesquiterpenes, sulfoorientalols a, b, c, and d, from Chinese *Alismatis rhizoma*. *Chem. Pharm. Bull.* 42, 2430–2435.
- Yoshikawa, M., Murakami, T., Ikebata, A., Ishikado, A., Murakami, N., Yamahara, J., Matsuda, H., 1997. Absolute stereostructures of alismalactone 23-acetate and alismaketone-a 23-acetate, new seco-protostane and protostane-type triterpenes with vasorelaxant effects from Chinese *Alismatis rhizoma*. *Chem. Pharm. Bull.* 45, 756–758.
- Yoshikawa, M., Tomohiro, N., Murakami, T., Ikebata, A., Matsuda, H., Matsuda, H., Kubo, M., 1999. Studies on *alismatis rhizoma*. III. Stereostructures of new protostane-type triterpenes, alisols H, I, J-23-acetate, K-23-acetate, L-23-acetate, M-23-acetate, and N-23-acetate, from the dried rhizome of *Alisma orientale*. *Chem. Pharm. Bull.* 47, 524–528.
- Zeng, L., Pen, X., Zhang, R.Y., 1995. Alizexol A, a novel protostane type of triterpene from *Alisma orientalis* (SAM) Juzep. *Chin. Chem. Lett.* 6, 675–678.