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axial (ax)

X= NO₂, CN, CHO, Br, CI, NHCOCH₃, H, C₆H₅, CH₃, OCH₃, NH₂

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ISSN 0040-4020



Available online at www.sciencedirect.com



Tetrahedron

Tetrahedron 60 (2004) 10809-10815

The 5-exo-trig radical cyclization reaction under reductive and oxidative conditions in the synthesis of optically pure GABA derivatives

Verónica Rodríguez, Mario Sánchez, Leticia Quintero* and Fernando Sartillo-Piscil*

Centro de Investigación de la Facultad de Ciencias Químicas, Benemérita Universidad Autónoma de Puebla, 14 Sur Esq. San Claudio, San Manuel, 72570, Puebla México

Received 17 July 2004; revised 16 September 2004; accepted 16 September 2004

Available online 2 October 2004

Abstract—Free radical precursors 4a and 4b were synthesized and treated under reductive or oxidative conditions to obtain the corresponding optically pure pyrrolidinones 5–8, which were subsequently transformed into corresponding optically pure GABA derivatives 9–11. When reductive radical conditions ($4a \rightarrow 7$ and 8) were used, a Ph₁₋₅ migration product 14 was obtained; presumably depending upon the specific conformation of the rotamer precursor and also 14 was found to be a kinetic product.

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1. Introduction

 $\gamma\text{-}Aminobutyric$ acid (GABA) is the most important inhibitory neurotransmitter in the mammalian central nervous system.¹ The GABA derivatives have been subject to further biological studies and they have showed an extensive spectrum of biological activity.² Some of the most important GABA derivatives are Gabapentin (1),³ Vigabatrin (2),⁴ and Baclofen $(3)^5$ which not only have been introduced as anticonvulsant agents but recently have shown to be a potential treatment for neurogenic pain⁶ (see Fig. 1).

Additionally some derivatives of Gabapentin 1 (with alkyl groups or sulfur atom into the cyclohexane ring framework) have been synthesized and tested, and same biological activity compared to Gabepentin itself has been observed.⁷ Thus, developing of novel synthetic routes for the synthesis of the compounds above represents an active synthetic challenge.

In this regard, this work describes a novel and accessible route for the synthesis of optically pure GABA derivatives β-substituted. The key of this work is the use of the (S)-αmethylbenzylamine as chiral auxiliary,8 and the wellstudied 5-exo-trig radical cyclization⁹ as strategy for the

construction of the corresponding 4-substituted pyrrolidinones (5-8). Then, by removing of the chiral auxiliary followed by an aqueous hydrolysis, the corresponding optically pure GABA derivatives β-substituted will be obtained (9–12, Scheme 1).

2. Results and discussion

The synthesis of radical precursors 4a and 4b commences with an allylation reaction of (S)- α -methylbenzylamine followed by N-acylation reaction with bromoacetylbromide to afford 4a. The exposure of 4a to potassium ethylxanthate leads the formation of the radical precursor 4b (Scheme 2).

The 5-exo radical cyclization was achieved by dropwise addition of tributyltin hydride and catalytic amounts of AIBN to 4a in benzene at reflux when the separable pyrrolidinones 7 (43%), 8 (38%) were obtained, and a $Ph_{1.5}$ -migration product 14 (11%) was also obtained, and very small amount of the reduction product ($\sim 3\%$, not showed) was detected (see Scheme 3).



Figure 1. Some GABA derivatives with pharmacological activity.

Keywords: 5-exo Radical cyclization; Reductive and oxidative conditions; GABA derivatives; Ar1,5-migracion.

^{*} Corresponding authors. Tel.: +52 2222 295500x7387; fax: +52 2222 454293 (F.S.-P.); e-mail: fsarpis@siu.buap.mx

^{0040-4020/\$ -} see front matter © 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.053



Scheme 1. Synthetic strategy for the synthesis of optically pure GABA derivatives.

Although the Ar_{1-5} -migration is a well recorded free radical process (neophyl rearrangement),¹⁰ we believe that in this particular case the slow rotation of the amide bond may contribute to the rearrangement. By a simple inspection of the ¹H NMR spectra of **4a** at room temperature, a pair of set sharp peaks were observed, indicating the existence of a mixture of *E/Z* rotamers in a ratio of 3:1, favoring to *Z* rotamer^{11–13} (see Scheme 4).

Geometrically speaking, the formation of pyrrolidinones 7 and 8 is achieved when the *Z*-4a rotamer is present and the Ph_{1-5} -migration product 14 when the *E*-4a rotamer is highly populated or the rotation of the amide bond is quite slow. In order to support this hypothesis, we synthesized the radical precursor 15, expecting with the increment of the molecular polarity (NH bond) a rapid rotation of the amide bond and reducing the half-time of the *E* rotamer which presumably is the responsible for the formation of the Ph_{1-5} migration product 16 (see Scheme 5).

The ¹H NMR spectra of the compound **15** at room temperature did show a single set of sharp peaks, indicating a single rotamer or a very rapid interconversion between both rotamers *E*-**15** and *Z*-**15**. Thus, treating to the radical precursor **15** under the same conditions as **4a**, the reduction product **15** was only observed, and no even traces of the Ph_{1-5} migration product **16** were detected.



Scheme 2. Synthesis of the radical precursors 4a and 4b.

In this sense, we found interesting to study the dynamic nature of the Ph₁₋₅-migration product. Thus, it was observed by adding catalytic amounts of selenol (0.2 equiv as diphenyl diselenide)¹⁴ that the yield of the Ph_{1-5} migration product 14 was increased (from 11% with Bu₃SnH to 18%) with PhSeH). On the other hand, by using a much poorer hydrogen donor like tris(trimethyl)silyl silane (TTMSS), the formation of 14 was inhibited. These results suggest a thermodynamic and kinetic process in the $C \leftrightarrow D \leftrightarrow E$ equilibriums, in which 14 seems to be a kinetic product: only a very fast hydrogen donor like selenol can be capable to capture the radical $\mathbf{\tilde{E}}$.¹⁵ On the other hand, in the presence of the slow hydrogen donor there is not chance to capture the short half-life time radical E, and the reaction course goes back to the formation of the pyrrolidinones 7 and 8 (see Scheme 6).¹⁶ It is important to mention that the ratio of the pyrrolidinones was not changed by using different hydrogen donor.

On the other hand, the 5-*exo*-radical cyclization for xanthate **4b** under pseudo-catalytic oxidizing conditions afforded the pyrrolidinones **5** and **6**, (Scheme 7). This very well-known chain radical process¹⁷ allows forming a carbon–sulfur bond that eventually will afford to optically pure 3-mercaptomethyl- γ -aminoacids **9** and **10**. As it was mentioned before, the incorporation of the sulfur atom in the structure of the GABA derivatives increase significantly the pharmacological activity.⁷

Although **4b** showed the same conformational behavior as **4a**, no evidence for the formation of the Ph_{1-5} -migration



Scheme 3. Synthesis of pyrrolidinones 7 and 8 by 5-*exo trig* radical cyclization of 4a under reductive conditions.



Scheme 4. Dependence of the free radical reaction course upon the conformation of the rotamers.



Scheme 5. Rapid equilibrium between E-15 and Z-15 rotamers.



Scheme 6. Dynamic nature of the Ph_{1,5}-migration product.

product **G** was found. The reasons for the failure might be due to the direct oxidation of the radical **E** by DLP into cationic specie,¹⁸ which is transformed into byproducts like enamines or its corresponding hydrolysis products; but unfortunately none of them were observed or isolated. It might be also due to the slow addition of the radical **E** to the thiocarbonyl of the starting material **4b** (propagation step), so the radical **E** can not be trapped by xanthate group leading the equilibriums toward the formation of the pyrrolidinones **5** and **6** only, this supports the theory of only a very fast hydrogen donor like selenol can be capable to capture the radical **E**. Having the optically pure pyrrolidinones (5–8) in our hands, the assignment of the absolute configuration at C4 was carried out by simple chemical correlation. Birch debenzylation of pyrrolidinone 8 afforded pyrrolidinone 18, which was identical with the sample reported by Meyers and Snyder,¹⁹ except the magnitude of the $[\alpha]_D$ (observed $[\alpha]_D = -20.3$ (*c* 1, CHCl₃), reported by Meyers and Snyder $[\alpha]_D = -6.5$ (*c* 0.26, CH₂Cl₂), see Scheme 8). The optical purity of 18 was determined by HPLC analysis in a chiral stationary phase of Chiralcel OD, hexane/2-propanol (93:7) as eluent.



Scheme 7. Synthesis of pyrrolidinones 5 and 6 by 5-exo radical cyclization oxidative of 4a under oxidative conditions.



Scheme 8. Determination of the absolute stereochemistry at C4 for 8 and 6 by chemical correlation.

Confirmation of the absolute stereochemistry for xanthates **5** and **6** was achieved by reductive removal of the xanthate group from **6** with tributyltin hydride and AIBN, (see Scheme 6).

Finally, pyrrolidinones 7 and 8 were separately debenzylated followed by an alkaline hydrolysis with NaOH

yielding the corresponding (*R*) and (*S*)-3-methyl-4-aminobutyric acids *R*-11 and *S*-12.²⁰ On the other hand,

pyrrolidinones 5 and 6 were separately hydrolyzed under

controlled conditions by adding aqueous NaOH to obtain

the corresponding (R) and (S)-4-(mercaptomethyl)-1-((S)-1-

phenylethyl)pyrrolidin-2-ones **20** and **21**. Then, a Birch debenzylation-acid hydrolysis in one pot produced to the optically pure (R) and (S)-4-amino-3-(mercaptomethyl) butanoic acids R-9 and S-10, (see Scheme 9).

3. Conclusion

A practical procedure for the synthesis of optically pure GABA derivatives is reported. As the result of the use of reductive free radical conditions in the 5-*exo-trig* cyclization reaction for the construction of the pyrrolidinones



framework, a Ph_{1-5} -migration product was observed, which presumably depends upon the conformation of the amide rotamer precursor, and it also showed to be a kinetic product. Besides, another use of the beneficial catalytic effect of the selenol in the free radical reactions is reported, in this case in the Ph_{1-5} -migration reaction.

4. Experimental

4.1. General

Instrumental NMR studies were carried out with Varian mercury 400 and 300 instruments. Internal reference (TMS) for ¹H and ¹³C. Chemical shifts are stated in parts per million. COSY, HSQC, and NOESY experiments have been carried out in order to assign the ¹H and ¹³C spectra completely. High resolution Mass spectra (FAB⁺ ion mode) were realized on a Jeol JMS-SX102A 10 KV and Elemental Analysis at Service de Microanalyse I.C.S.N.-C.N.R.S.-France. IR were carried out with a SHIMADZU FTIR-8400 instrument.

4.1.1. (S)-N-(1-Phenylethyl)prop-2-en-1-amine 13. To a solution of (S)-(-)- α -methylbenzylamine (3 g, 24.7 mmol) and sodium carbonate (3.14 g, 29.7 mmol) in 40 mL of dry acetonitrile at 0 °C was allowed to react for 15 min before to add allyl iodine (4.9 g, 29.7 mmol) dissolved in 5 mL of acetonitrile. The reaction mixture was stirred for 4 h at room temperature before add 30 mL of water, extracted with ethyl acetate, dried with Na₂SO₄ and evaporated under reduced pressure. The residue was purified by column chromatography (2:1, hexane/ethyl acetate) to give 2.85 g of 13 (72%) as a white solid. Mp=103-104 °C; $[\alpha]_D = -62.3$ (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ: 1.38 (d, 3H, J=6.4 Hz), 1.75 (b, 1H), 3.12 (m, 2H), 3.82 (q, 1H, J=6.4 Hz), 5.09 (d, 1H, J = 10 Hz), 5.15 (d, 1H, J = 17.2 Hz), 5.91 (m, 1H), 7.22–7.37 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ : 24.1, 49.9, 57.2, 115.2, 126.1, 126.4, 127.9, 136.5, 144.9. EI-MS *m*/*z* 161 (M+).

4.1.2. (S)-N-Allyl-2-bromo-N-(phenylethyl)acetamide 4a. To a solution of 13 (1 g, 6.2 mmol) and DMAP (1.13 g, 9.24 mmol) in 30 mL of dry THF at 0 °C was allowed to react for 15 min before to add bromoacetyl bromide (1.5 g, 7.44 mmol) dissolved in 5 mL of dry THF. The reaction mixture was warmed up to room temperature and allowed to stir for 2 h before to add 20 mL of water, extracted with ethyl acetate, dried with Na₂SO₄ and evaporated under reduced pressure. The residue was purified by column chromatography through silica gel (5:1, hexane/ethyl acetate) to give 1.53 g (88%) of 13 as colorless oil; $[\alpha]_D = -157.5$ (c 1.7, CHCl₃); NMR data is given separately for each rotamer: Z-4a ¹H NMR (400 MHz, CDCl₃) δ : 1.50 (d, 3H, J=7.2 Hz), 3.64 (dd, 1H, J=18.0, 4.8 Hz), 3.82 (dd, 1H, J=18.0, 4.8 Hz), 3.84 (d, 1H, J=10.8 Hz), 3.89 (d, 1H, J = 10.8 Hz), 5.01 (m, 2H), 5.60 (m, 1H), 6.00 (q, 1H, J=7.2 Hz), 7.25 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ: 16.3, 27.2, 46.2, 51.6, 116.4, 127.2, 127.6, 128.3, 134.6, 139.8, 167.2; *E*-**4a** ¹H NMR (400 MHz, CDCl₃) δ : 1.67 (d, 3H, J=6.8 Hz), 3.44 (dd, 1H, J=16.0, 6.4 Hz), 3.89 (d, 1H, J = 10.0 Hz), 3.99 (d, 1H, J = 10.0 Hz), 4.05 (dd, 1H, J = 16.0, 5.2 Hz), 5.00 (m, 2H), 5.17 (q, 1H)

J=6.8 Hz), 5.71 (m, 1H), 7.23 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ : 18.6, 26.6, 45.5, 56.5, 116.2, 126.4, 127.6, 128.5, 133.5, 139.7, 166.8; IR (CCl₄) 3029 (CH=CH₂), 1646 (C=O) cm⁻¹; Anal. Calcd for C₁₃H₁₆BrNO: C, 55.33; H, 5.72. Found: C, 55.12; H, 5.81.

4.1.3. (S)-N-Allyl-2-ethylxanthate-N-(phenylethyl)acetamide 4b. Radical precursor 4a (1.11 g, 3.93 mmol) was treated with potassium ethylxanthate (0.95 g, 5.9 mmol) in 30 mL of acetone at 0 °C for 2 h. Evaporated under reduced pressure, and the residue was purified by column chromatography through silica gel (5:1, hexane/ethyl acetate) to give 1.2 g (89%) of **4b** as colorless oil; $[\alpha]_D = -165.3$ (*c* 1, CHCl₃); NMR data is reported as a mixture of E/Z rotamers: ¹H NMR (400 MHz, CDCl₃) δ : 1.39 (t, 6H, J = 7.2 Hz), 1.51 (d, 3H, J=7.2 Hz), 1.67 (d, 3H, J=6.8 Hz), 3.49 (dd, 1H, J = 15.2, 6.0 Hz), 3.71 (dd, 1H, J = 18.0, 5.2 Hz), 3.87 (dd, 1H, J = 18.0, 5.2 Hz), 4.02 (d, 1H, J = 16.0 Hz), 4.10 (dd, 1H, J = 15.2, 6.0 Hz), 4.12 (d, 1H, J = 16.0 Hz), 4.23 (q, 1H, J=15.6 Hz), 4.62 (q, 4H, J=7.2 Hz), 4.98 (dd, 2H, J=11.2, 4.0 Hz), 5.13 (dd, 2H, J=14.0, 2.4 Hz), 5.25 (q, 1H, J=7.2 Hz), 5.60 (m, 1H), 5.72 (m, 1H), 6.06 (q, 1H, J=7.2 Hz), 7.18–7.32 (m, 10H); ¹³C NMR (100 MHz, CDCl₃) δ : 13.8, 16.5, 18.2, 40.0, 40.2, 46.0, 46.3, 51.9, 56.1, 70.3, 70.4, 116.3, 117.0, 126.5, 127.3, 127.3, 127.6, 128.2, 128.5, 134.0, 134.3, 139.9, 167.0, 213.7; IR (CCl₄) 1667 (C=O), 1235 (C=S) cm⁻¹; Anal. Calcd for $C_{16}H_{21}NO_2S_2$: C, 59.41; H, 6.54. Found: C, 59.53; H, 6.72.

4.2. The 5-*exo*-radical cyclization under reductive condition (Tin method)

(*R*) and (*S*)-4-Methyl-1-((*S*)-1-phenylethyl)pyrrolidin-2ones **7** and **8**, and *N*-allyl-*N*-ethyl-2-phenylacetamide **14**. To a solution of **4a** (0.2 g, 0.7 mmol) in 30 mL of benzene at 80 °C was added dropwise (in a period of 1 h) Bu₃SnH (0.21 g, 0.71 mmol) and AIBN (20 mg) dissolved in 30 mL of benzene. The reaction mixture was allowed to stir for 2 h before evaporating under reduced pressure. The residue was purified by column chromatography through silica gel (only hexanes until tin residues come out, then a system of 4:1, hexane ethyl acetate is used) to give 60 mg of **7** (43%), 53 mg of **8** (38%), and 16 mg of **14** (12%) all as colorless oils.

4.2.1. (*R*)-4-Methyl-1-((*S*)-1-phenylethyl)pyrrolidin-2one 7. $[\alpha]_D = -93.3$ (*c* 1, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ : 0.96 (d, 3H, *J*=7.2 Hz), 1.51 (d, 3H, *J*=7.2 Hz), 2.02 (dd, 1H, *J*=16.4, 6.4 Hz), 2.37 (m, 1H), 2.50 (dd, 1H, *J*=9.6, 5.6 Hz), 2.59 (dd, 1H, *J*=16.8, 8.4 Hz), 3.41 (dd, 1H, *J*=9.6, 7.6 Hz), 5.50 (q, 1H, *J*=7.2 Hz), 7.29 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ : 16.1, 19.6, 26.3, 39.7, 48.6, 49.3, 126.8, 127.1, 128.2, 139.8, 173.6; IR (CCl₄) 1677 (C=O) cm⁻¹; FABS *m*/*z* 204.1381 (M+H)⁺ (Calcd for C₁₃H₁₇NO₂O₄: 204.1389).

4.2.2. (*S*)-4-Methyl-1-((*S*)-1-phenylethyl)pyrrolidin-2one 8. $[\alpha]_D = -120$ (*c* 1, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ : 1.08 (d, 3H. J = 6.8 Hz), 1.50 (d, 3H, J = 7.6 Hz), 2.10 (dd, 1H, J = 16.4, 7.2 Hz), 2.30 (m, 1H), 2.57 (dd, 1H, J = 16.8, 8.8 Hz), 2.87 (dd, 1H, J = 9.2, 6.0 Hz), 3.01 (t, 1H, J = 8.2 Hz), 5.48 (q, 1H, J = 7.2 Hz), 7.29 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ : 16.1, 19.5, 26.5, 39.7, 48.6, 49.5, 126.7, 127.1, 128.2, 139.9, 173.6; IR (CCl₄) 1681 (C=O) cm⁻¹; FABS m/z 204.1383 (M+H)⁺ (Calcd for C₁₃H₁₇NO: 204.1389).

4.2.3. *N*-Allyl-*N*-ethyl-2-phenylacetamide 14. NMR data is reported as a mixture of *E/Z* rotamers: ¹H NMR (400 MHz, CDCl₃) δ : 1.08 (t, 3H, *J*=7.2 Hz), 1.12 (t, 3H, *J*=7.2 Hz), 3.31 (q, 2H, *J*=7.2 Hz), 3.40 (q, 2H, *J*=7.2 Hz), 3.67 (s, 3H), 3.73 (s, 3H), 3.86 (dt, 2H, *J*=4.8, 2.0 Hz), (d, 2H, *J*=6.0 Hz), 5.06–5.22 (m, 4H), 5.72 (m, 2H), 7.27 (m, 10H); ¹³C NMR (100 MHz, CDCl₃) δ : 12.6, 13.8, 40.7, 40.8, 42.0, 47.4, 50.0, 116.6, 116.8, 126.6, 126.7, 128.2, 128.6, 128.7, 133.2, 133.5, 135.2, 170.3, 170.7; Anal. Calcd for C₁₁H₁₅N: C, 81.94; H, 9.38. Found: C, 82.12; H, 9.24.

4.3. The 5-*exo*-radical cyclization under oxidative conditions (Xanthate method)

(*R*) and (*S*)-4-(Methyl-ethoxythiocarbonil)1-((*S*)-1-phenylethyl)pyrrolidin-2-ones **5** and **6**. A solution of **4b** (0.2 g, 0.61 mmol) and DLP (0.08 g, 0.2 mmol) in 50 mL of benzene was refluxed for 30 min, then, the reaction mixture was evaporated under reduced pressure and the residue was purified by preparative TLC on silica gel (4:1, hexane/ethyl acetate) yielding 0.82 mg of **5** (41%), and 78 mg of **6** (39%).

4.3.1. (*R*)-4-(methyl-ethoxythiocarbonil)1-((*S*)-1-phenylethyl)pyrrolidin-2-one 5. $[\alpha]_D = -72.6 \ (c \ 1, CHCl_3); {}^{1}H$ NMR (400 MHz, CDCl_3) δ : 1.39 (t, 3H, $J = 6.8 \ Hz$), 1.51 (d, 3H, $J = 7.2 \ Hz$), 2.22 (dd, 1H, $J = 16.4, 5.2 \ Hz$), 2.70 (m, 3H), 2.99 (dd, 1H, $J = 13.6, 6.8 \ Hz$), 3.14 (dd, 1H, $J = 13.8, 6.4 \ Hz$), 3.46 (dd, 1H, $J = 12.0, 9.6 \ Hz$), 4.62 (q, 2H, $J = 6.8 \ Hz$), 5.49 (q, 1H, $J = 7.2 \ Hz$), 7.26 (m, 5H); ${}^{13}C$ NMR (100 MHz, CDCl₃) δ : 13.7, 16.0, 30.8, 37.4, 39.6, 47.1, 48.9, 70.2, 127.0, 127.6, 128.6, 139.8, 172.7, 213.8; IR (CCl₄) 1687, 1680 (C=O), 1217 (C=S), 1050 (C-S) cm⁻¹; FABS m/z 324.1082 (M+H)⁺ (Calcd for C₁₆H₂₁NO₂S₂: 324.1089).

4.3.2. (*S*)-4-(Methyl-ethoxythiocarbonil)1-((*S*)-1-phenylethyl)pyrrolidin-2-one 6. $[\alpha]_D = -85.1 (c \ 1.3, CHCl_3)$; ¹H NMR (400 MHz, CDCl_3) δ : 1.40 (t, 3H, J = 7.2 Hz), 1.51 (d, 3H, J = 6.8 Hz), 2.27 (dd, 1H, J = 19.6, 9.6 Hz), 2.61 (m, 2H), 3.0–3.35 (m, 4H), 4.62 (q, 2H, J = 7.2 Hz), 5.48 (q, 1H, J = 6.8 Hz), 7.24 (m, 5H); ¹³C NMR (100 MHz, CDCl_3) δ : 13.7, 16.1, 30.7, 37.3, 39.7, 47.0, 48.8, 70.1, 126.7, 127.2, 128.2, 139.5, 172.2, 213.2; IR (CCl_4) 1683 (C=O), 1216 (C=S), 1050 (C–S) cm⁻¹; FABS *m/z* 324.1082 (M+H)⁺ (Calcd for C₁₆H₂₁NO₂S₂: 324.1089).

4.3.3. (*S*) and (*R*)-4-Methylpyrrolidin-2-ones 18 and 19.¹⁹ Birch debenzylation: A solution of 7 (0.140 g, 0.68 mmol) in 15 mL of dry THF was added to a deep blue solution of Li (0.032 g, 4.5 mmol) in condensed NH₃ (ca. 20 mL) at -70 °C within 10 min. The reaction mixture was allowed to stir for 1 h at -70 °C before to add NH₄Cl (ca. 40 mg), then neutralized with a diluted solution of HCl, extracted with ethyl acetate, dried with Na₂SO₄ and evaporated under reduced pressure. The residue was purified by column chromatography through silica gel (hexane/ethyl acetate, 1:1) afforded the *S*-18 (45 mg, 67%). (*S*)-4-Methylpyrrolidin-2-one 18; $[\alpha]_{\rm D} = -20.3$ (*c* 1.2, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ: 1.13 (d, 3H, J=6.4 Hz), 1.95 (dd, 1H, J=16.8, 7.2 Hz), 2.45 (dd, 1H, J=16.4, 8.8 Hz), 2.53 (m, 1H), 2.96 (dd, 1H, J= 9.6, 6.0 Hz), 3.51 (ddd, 1H, J=8.4, 8.0, 1.2 Hz), 4.25 (b, 1H); ¹³C NMR (100 MHz, CDCl₃) δ: 19.6, 29.4, 38.5, 49.6, 178.5. (*R*)-4-Methylpyrrolidin-2-one **19** was obtained analogously to **18** (65%); [α]_D= +18.1 (*c* 1 CHCl₃).

4.3.4. (*R*) and (*S*)-4-(Mercaptomethyl)-1-((*S*)-1-phenylethyl)pyrrolidin-2-ones 20 and 21. Controlled alkaline hydrolysis: pyrrolidinone 5 (200 mg) were dissolved in 1 mL of ethanol and added 5 mL of an aqueous solution of NaOH (8 N). The reaction mixture was allowed to stir for 4 h at room temperature, extracted with ethyl acetate, dried with Na₂SO₄ and evaporated under reduced pressure; the residue was purified by column chromatography through silica gel (2:1, hexane/ethyl acetate) to give 20 (123 mg, 85%). (R)-4-(Mercaptomethyl)-1-((S)-1-phenylethyl)pyrrolidin-2-one **20**; $[\alpha]_{\rm D} = -97.8$ (*c* 1, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ : 1.48 (d, 3H, J=7.2 Hz), 2.17 (dd, 1H, J = 16.8, 6.0 Hz), 2.28–2.53 (m, 3H), 2.59 (dd, 1H, J =17.2, 8.4 Hz), 2.68 (dd, 1H, J=10.4, 5.2 Hz), 3.43 (dd, 1H, J = 10.0, 7.6 Hz), 5.45 (q, 1H, J = 7.2 Hz), 7.25 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ: 15.9, 28.4, 34.7, 37.0, 46.5, 48.7, 126.7, 127.2, 128.2, 139.4, 172.6; Anal. Calcd for C₁₃H₁₇NOS: C, 60.34; H, 7.28. Found: C, 60.30; H, 7.30. (S)-4-(Mercaptomethyl)-1-((S)-1-phenylethyl)pyrrolidin-2one 21 was obtained analogously to 20 (109 mg, 76%); $[\alpha]_{\rm D} = -172 \ (c \ 1.1, \ CHCl_3); \ ^1H \ NMR \ (400 \ MHz, \ CDCl_3)$ δ: 1.52 (dd, 3H, J=7.2 Hz), 2.25 (dd, 1H, J=16.4, 7.2 Hz), 2.39 (m, 1H), 2.60 (m, 3H), 3.12 (m, 2H), 5.51 (c, 1H, J= 7.2 Hz), 7.30 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ: 16.2, 28.8, 34.7, 37.2, 46.9, 48.9, 126.8, 127.3, 128.4, 173.0; Anal. Calcd for C₁₃H₁₇NOS: C, 60.34; H, 7.28. Found: C, 59.98; H, 7.45.

4.3.5. (R) and (S)-4-Amino-3-(mercaptomethyl)butanoic acids R-9 and S-10. Birch debenzylation-aqueous hydrolysis in one pot, general protocol: Pyrrolidinone 20 (0.230 g, 0.98 mmol) was debenzylated by using the classic Birch method followed by acidic quenching with 10 mL of an aqueous solution at10%, and allowed to stir for 30 min at room temperature, reaction mixture is evaporated under reduced pressure; residue is crystallized in ethanol to afford (R)-4-Amino-3-(mercaptomethyl)butanoic acids R-9 as a with powder (0.125 mg, 86%). Mp = 142 °C; $[\alpha]_D = +32.9$ (c 1, CH₃OH); ¹H NMR (400 MHz, CD₃OD) δ : 2.22 (dd, 1H, J = 16.8, 5.2 Hz), 2.51 (dd, 1H, J = 17.2, 7.6 Hz), 2.85 (m, 3H), 3.18 (dd, 1H, J=10.0, 4.4 Hz), 3.55 (dd, 1H, J = 10.06.8 Hz); ¹³C NMR (100 MHz, CDCl₃) δ : 34.2, 36.0, 42.2, 47.2, 178.5; IR (KBr) 3226 (OH), 2810 (SH), 1679 $(C=0) \text{ cm}^{-1}$; Anal. Calcd for C₅H₁₁NO₂S: C, 40.25; H, 7.43. Found: C, 39.98; H, 7.56. (S)-4-Amino-3-(mercaptomethyl)butanoic acid S-10 was obtained analogously to *R*-9 (82%). Mp=142 °C; $[\alpha]_D = -33.4$ (c 1.02, CH₃OH); IR (KBr) 3220 (OH), 2778 (SH), 1673 $(C=0) \text{ cm}^{-1}.$

4.3.6. (*S*) and (*R*)-3-Methyl-4-aminobutyric acids 11 and 12.²⁰ Alkaline hydrolysis: pyrrolidinone *S*-18 (80 mg, 0.8 mmol) is dissolved in 1 mL of methanol and added 5 mL of an aqueous solution of KOH (8 N). The reaction mixture is refluxed for 5 h before to add an aqueous solution of HCl (until get a pH 7), and evaporated under reduced

pressure, the residue was purified by preparative TLC on silica gel (ethyl acetate/MeOH 4:1) yielding *S*-**11** (79 mg, 85%); $[\alpha]_D = +8.1$ (*c* 1, CH₃OH); ¹H NMR (400 MHz, CD₃OD) δ : 1.04 (d, 3H, *J*=6.8 Hz), 2.18 (m, 1H), 2.30 (d, 2H, *J*=7.0 Hz), 2.88 (m, 2H); ¹³C NMR (100 MHz, CD₃OD) δ : 17.7, 29.6, 43.3, 45.5, 178.6; IR (KBr) 3431 (OH), 2840 (NH₃⁺), 1557 (COO⁻) cm⁻¹. (*R*)-3-Methyl-4-aminobutyric acid **12** was obtained analogously to *S*-**11** (83%); $[\alpha]_D = -8.3$ (*c* 1, CH₃OH).

Acknowledgements

Financial support from CONACyT (grant number: 35102-E) is gratefully acknowledged. V.R. Thanks CONACyT for a graduate scholarship (grant number: 168916). F.S.P. also thanks Benemérita Universidad Autónoma de Puebla (BUAP) for a PROMEP professorship.

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11. The geometry of the E/Z isomers for **4a** was easily determined by applying a computational calculation (Geometry optimizations for both rotamers were performed with the HF/6-31G(d,p)12 method using GAUSSIAN 98. The frequencies were calculated at the same level of theory to confirm that each stationary point corresponds to a minimum. The optimized structures at ab initio were used for obtaining energies at a higher level of theory using the B3LYP/6-311G(d,p)13 method). Above calculation are in agreement with the ¹H NMR spectra, indicating an important intramolecular hydrogen bonding between the carbonyl group and H α (this internal hydrogen bonding is confirmed by theoretic calculation) between H α and carbonyl group in the Z conformation, causing a downfield shift of almost 1 ppm in compare to the *E* rotamer.



Hα, δ: 5.17 ppm

Hα, δ: 6.0 ppm

Relative energie = 0.76 Kcal/mol Relative energie = 0.0 Kcal/mol

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Tetrahedron

Tetrahedron 60 (2004) 10817-10824

Parviflorenes B–F, novel cytotoxic unsymmetrical sesquiterpene-dimers with three backbone skeletons from *Curcuma parviflora*

Kazufumi Toume,^a Masae Takahashi,^a Kentaro Yamaguchi,^b Takashi Koyano,^c Thaworn Kowithayakorn,^d Masahiko Hayashi,^e Kanki Komiyama^e and Masami Ishibashi^{a,*}

^aGraduate School of Pharmaceutical Sciences, Chiba University, 1-33 Yayoi-cho, Inage-ku, Chiba 263-8522, Japan

^bChemical Analysis Center, Chiba University, 1-33 Yayoi-cho, Inage-ku, Chiba 263-8522, Japan

^cTemko Corporation, 4-27-4 Honcho, Nakano, Tokyo 164-0012, Japan

^dDepartment of Horticulture, Faculty of Agriculture, Khon Kaen University, Khon Kaen 40002, Thailand ^eThe Kitasato Institute, 5-9-1 Shirokane, Minato-ku, Tokyo 108-8642, Japan

Received 2 September 2004; revised 15 September 2004; accepted 16 September 2004

Available online 2 October 2004

Abstract—Five novel natural products classified as dimeric sesquiterpenes, named parviflorenes B–F (2–6), possessing three types of novel backbone frameworks, have been isolated from *Curcuma parviflora* (Zingiberaceae). The structures of 2–6 were elucidated by means of spectroscopic studies, and the structure of 2 was further unambiguously established by X-ray crystallographic analysis. Compounds 2, 4, and 6 have an unsymmetrical bis-cadinane skeleton, while compound 3 is a dimer of cadinane and iso-cadinane, and compound 5 possesses another novel carbon framework consisting of two cadinanes with different bond-connection. These new compounds with novel carbon skeletons showed cytotoxicity against tumor cell lines.

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1. Introduction

Curcuma parviflora Wall. (Zingiberaceae) is widely distributed over a forest area of the northern part of Thailand,¹ and is used as an ornamental plant, and is edible, and also it has been said to be used for detoxification of scorpion bites in certain areas. During our search for bioactive natural products from tropical plants,² we investigated the chemical constituents of C. parviflora collected in Thailand, and recently isolated the cytotoxic sesquiterpene-dimer, parviflorene A (1) from this plant.³ We further investigated this plant and here we describe the isolation and structure elucidation of five new sesquiterpene-dimers, parviflorenes B-F (2-6) from the underground part of C. parviflora. These sesquiterpene-dimers may be classified into three groups with different carbon frameworks: (i) biscadinanes (compounds 1, 2, 4, and 6); (ii) a cadinane-isocadinane adduct (compound 3), and (iii) a biscadinane with an alternate bond connection (compound

* Corresponding author. Tel./fax: +81 43 290 2913;

5). These new compounds showed cytotoxicity against KB (human adenocarcinoma) and other tumor cell lines. As described previously,³ this plant also contained cadinane sesquiterpenes, cadalenequinone $(7)^4$ and 8-hydroxycadalene (**8**),⁵ corresponding to monomers for dimers such as **1**.

2. Results and discussion

It was revealed by TLC examination that parviflorene A (1) and its related compounds were contained in the underground part of *C. parviflora*. The EtOAc-soluble fraction of the MeOH extracts, combined with previously obtained EtOAc and *n*-BuOH-soluble fractions of this plant, were subjected to repeated chromatography on silica gel and Sephadex LH-20, followed by purification with HPLC on ODS to give five new compounds, parviflorenes B–F (2–6), in addition to parviflorene A (1).

Parviflorene B (2) was obtained as yellow prisms, $[\alpha]_D^{24} = -16$ (*c* 1.4, MeOH), and the molecular formula was established to be $C_{30}H_{32}O_4$ by its HRFABMS data (*m/z* 456.2260, M⁺, Δ -4.1 mmu). The ¹H NMR spectrum of **2** (Table 1) showed signals due to two singlet methyls and

Keywords: Zingiberaceae; *Curcuma parviflora*; Dimeric sesquiterpene; X-ray analysis; Cytotoxicity.

e-mail: mish@p.chiba-u.ac.jp

^{0040–4020/\$ -} see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.043



four doublet methyls; the chemical shifts of the singlet methyls [$\delta_{\rm H}$ 2.25 (3H, s) and 2.23 (3H, s)] implied that these two methyl groups were attached on sp^2 carbons. The ${}^{13}C$ NMR spectrum of 2 (Table 2) showed 18 aromatic carbons, one carbonyl, 11 sp³ carbons, one of which bore oxygen. Although the ¹H and ¹³C NMR data of 2 were similar to those of 1, one *O*-bearing sp³ carbon ($\delta_{\rm C}$ 81.8), and one carbonyl carbon ($\delta_{\rm C}$ 204.7) were not possessed by compound 1, and compound 2 had two fewer aromatic carbons than 1. Since 10 out of 15 unsaturation degrees were thus accounted for, 2 was deduced to have five rings. The 1 H NMR spectrum of 2 showed six aromatic ring protons [$\delta_{\rm H}$] 6.65 (1H, s), 6.64 (1H, s), 6.70 (1H, s), 6.94 (1H, s), 8.46 (1H, s), and 8.68 (1H, s)]. The ${}^{1}H-{}^{1}H$ COSY spectrum suggested the presence of two isopropyl groups (H₃-12/ H-11/H₃-13 and H₃-27/H-26/H₃-28). The HMBC spectrum of **2** afforded long-range ${}^{1}H^{-13}C$ correlations as shown in Figure 1, which suggested that 2 consisted of three aromatic benzene rings (rings A, C, and E), one cyclohexadienone ring (ring B), and one cyclohexadiene (ring D). The positions of two methyls, two phenols, one carbonyl, one tertiary hydroxyl, and two isopropyl groups were clearly assignable by the HMBC correlations (Fig. 1). In particular, the HMBC correlations for H-4/C-6, H-29/C-7, H-14/C-22, and H-19/C-21 provided evidence for the connections of the A/B, B/C, C/D, and D/E rings, respectively. The methine proton on C-21 was deduced to be equatorially oriented from the two small coupling constants with the vicinal

methylene protons on C-22 ($J_{21,22\alpha} = 3.5 \text{ Hz}$ and $J_{21,22\beta} = 4.5 \text{ Hz}$).

Compound 2 was crystallized from methanol solution to give yellow prisms, and its structure was unambiguously established by X-ray crystallographic analysis (Fig. 2). On the basis of X-ray analysis data, two isopropyl groups on C-6 and C-21 were shown to be axial with opposite orientations on the plane of the molecule. The pentacyclic skeleton of 2 corresponded to an unsymmetric dimer of two cadinane sesquiterpenes with C–C bonds constructed between C-8 and C-14 of the two monomers. This carbon framework was the same as that proposed for parviflorene A (1),³ and the basic skeleton of 1 elucidated from its spectral data³ was, therefore, reasonably corroborated here by the X-ray analysis of 2.⁶

Parviflorene C (**3**) was obtained as a yellow amorphous solid, and was suggested to have the molecular formula $C_{30}H_{32}O_4$ by its HRFABMS data (*m*/*z* 456.2288, M⁺, Δ – 1.3 mmu), which was identical with that of **2**. The ¹H and ¹³C NMR data of **3** closely resembled those of **2**. However, **3** had a different substitution pattern on the B ring; isopropyl and tertiary hydroxyl groups were attached to C-7 and a carbonyl group was present on C-6, as evidenced by the HMBC correlations from the aromatic proton at H-4 ($\delta_{\rm H}$ 7.25) to the carbonyl carbon at C-6 ($\delta_{\rm C}$ 205.8), and from the aromatic proton at H-29 ($\delta_{\rm H}$ 8.38) to the quaternary

Table 1. ¹H NMR data of parviflorene B–F (2–6)

Position	2 ^a	3 ^a	4 ^a (High conc.)	4 ^a (Low conc.)	5 ^a	6 ^b
1						
2	6.56 s	6.90 s	6.56 s	6.57 s	6.63 s	6.63 s ^c
3	7 10 -	7.25 -	(00 -	7.07 -	(72 -	
4	7.12 S	7.25 S	0.99 s	7.07 s	0.73 \$	0.40 S
6					3.28 m	2.54 t (5.0)
7					2.11 dd (14.0, 7.6) α 1.90 ddd (14.0, 11.0, 1.5) β	4.88 t (5.0)
8					1150 uuu (1110, 1110, 110) p	
9						
10						
11	1.99 quint. (6.7)	2.03 m	1.86 quint. (6.7)	1.90 quint. (6.7)	2.54 m	1.87 sext. (5.8)
12	0.74 d (0.7) 0.82 d (6.7)	0.72 (0.9)	0.03 d (0.7) 0.78 d (6.7)	0.00 d (0.7) 0.81 d (6.7)	0.95 d (7.0)	0.19 br s 0.88 d (7.0)
13	8.37 s	8.27 s	8.16 s	8.22 s	$3.57 d (18.5) \alpha$	7.97 s
					3.03 d (18.5) β	
15	2.32 s	2.34 s	2.23 s	2.27 s	2.28 s	2.22 s
16						
17	6.55 s	6.60 s	6.63 s	6.60 s		6.64 s ^c
18	6.60 a	6.61 a	650 .	657 .	7.55 a	6 16 0
20	0.00 \$	0.01 8	0.50 8	0.57 8	1.55 \$	0.40 \$
20	2.37 m	2.35 m	2.30 m	2.37 m		2.25 m
22	3.02 dd (15.0, 3.5) α	2.91 dd (15.0, 3.5)	2.96 dd (15.9, 5.0)	3.01 d (3.5)	7.23 d (7.5)	2.82 dd (14.5, 4.3)
	2.98 dd (15.0, 4.5) β	2.89 dd (15.0, 5.0)	2.95 dd (15.9, 2.7)			2.80 dd (14.5, 2.7)
23					7.13 d (7.5)	
24						
25	1.50 m	1.54 m	1.20 m	1 28 m	2.61 quint (6.7)	1.28 m
20	0.78 d (6.7)	1.34 m 0.82 d (6.6)	0.69 d (6.7)	0.75 d (6.7)	1.35 d (6.7)	1.20 m 0.68 d (6.7)
28	0.70 d (0.7) 0.91 d (6.7)	0.93 d (6.6)	0.76 d (6.4)	0.75 d (0.7)	1.34 d (6.7)	0.00 d (0.7)
29	8.70 s	8.38 s	8.53 s	8.56 s	2.86 s	8.45 s
30	2.29 s	2.30 s	2.26 s	2.26 s	2.33 s	2.22 s
	5.13 s [1-OH]	5.50 s [1-OH]		5.51 s [1-OH]	8.43 s [1-OH]	9.46 s [1-OH] ^d
	5.12 s [16-OH]	5.34 s [16-OH]		5.65 s [16-OH]	3.83 d (1.5) [8-OH]	9.41 s [16-OH] ^d
	4.04 s [6-OH]	3.95 s [7-OH]		4.06 s [6-OH]	3.46 s [9-OH]	5.03 s [7-OH]

^a In CDCl₃. ^b In DMSO- d_6 .

^{c,d} Signals may be reversed.

oxygenated carbon at C-7 ($\delta_{\rm C}$ 82.1). This fact was further supported by the NOE correlation observed between H-29 ($\delta_{\rm H}$ 8.38) and H₃-13 ($\delta_{\rm H}$ 0.81). Compound **3**, therefore, had a different carbon skeleton from that of **1** and **2**, and it was constructed from one cadinane and one isocadinane with C– C bonds between the C-8 carbon of each unit and the C-14 carbon of the other unit. An isocadinane sesquiterpene (**9**) with a similar A–B ring partial structure to **3** was isolated from *Heterotheca inuloides* (Asteraceae), which also simultaneously contained cadinane sesquiterpenes.⁷

Parviflorene D (4) was obtained as a yellow amorphous solid, and its molecular formula was suggested as $C_{30}H_{32}O_4$ by its HRFABMS data (*m*/*z* 456.2339, M⁺, Δ +3.8 mmu), which was also identical with that of **2**. The UV spectrum and ¹H and ¹³C NMR data of **4** closely resembled those of **2**, and interpretation of the 2D NMR data of **4** led to the conclusion that **4** had the same planar structure as that of **2**. However, the CD spectrum of **4** displayed an opposite Cotton effect [λ_{ext} 329 nm ($\Delta \varepsilon$ -21.5), 271 (31.1), and 253 (43.2)], compared with that of **2** [λ_{ext} 333 nm ($\Delta \varepsilon$ 15.2), 271 (-28.0), and 253 (5.9)], suggesting that compound **4** was epimeric with **2** at the C-6 position.^{6,8}

Parviflorene E (5) was obtained as a yellow amorphous solid, and its HRFABMS of 5 showed a molecular ion at m/z

460.2595 ([M]⁺, Δ -1.8 mmu) consistent with the molecular formula $C_{30}H_{36}O_4$. The UV spectrum of 5 showed absorption maxima at λ_{max} (MeOH) 304 and 237 nm, indicating the presence of conjugated systems, but the absorption patterns were different from those of parviflorene A (1) or B (2). The ¹H NMR spectrum of 5 (Table 1) showed signals due to three singlet methyls and four doublet methyls; the chemical shifts of the singlet methyls [$\delta_{\rm H}$ 2.96 (3H, s), 2.33 (3H, s), and 2.28 (3H, s)] implied that these three methyl groups were attached on sp² carbons. The ¹³C NMR spectrum aided by HMQC experiments revealed the presence of 16 sp² carbons as well as 14 sp³ carbons. Since, eight out of 13 degrees of unsaturation were thus accounted for, 5 was deduced to have five rings. The ¹H NMR spectrum of **5** (Table 1) showed the signals of five aromatic hydrogens, which were assignable to two ortho-coupled doublets [$\delta_{\rm H}$ 7.13 (d, J=7.5 Hz) and 7.23 (d, J=7.5 Hz)], two *meta*-oriented broad singlets [$\delta_{\rm H}$ 6.63 (s) and 6.73 (s)], and one isolated singlet [$\delta_{\rm H}$ 7.55 (s)] from the ¹H-¹H COSY spectrum. The ¹H-¹H COSY spectrum of 5 indicated the presence of two isopropyl groups (H₃-12/H-11/H₃-13 and H₃-27/H-26/H₃-28), and revealed that the sp³ methine [H-6: $\delta_{\rm H}$ 3.28 (1H, m)] was adjacent to an sp³ methylene [H₂-7: $\delta_{\rm H}$ 2.11 (1H, dd, J= 14.0, 7.6 Hz) and 1.90 (1H, ddd, J = 14.0, 11.0, 1.5 Hz)]. The HMBC spectrum of **5** afforded long-range ¹H–¹³C

Position	2^{a}	3 ^a	4 ^a	5 ^a	6 ^b
1	153.2 ^c	153.0 ^f	153.0 ^g	155.8	154.1
2	117.0	123.7	117.8	115.9	115.3
3	138.8	139.0	138.1	139.3	135.8
4	120.8	120.5	120.5	119.8	121.8
5	143.4 ^d	131.2	140.8	138.8	139.8
6	82.1	205.8	82.1	41.2	52.3
7	204.0	82.1	206.1	28.6^{i}	70.0
8	127.8	137.3	127.4	98.6	136.9
9	134.4	127.0	134.4	72.9	130.0
10	114.6	121.5	115.1	121.5	118.2
11	38.6	38.0	38.9	30.8	25.4
12	17.2	17.0	17.1	20.3 ^j	19.7
13	16.5	16.7	16.7	15.6	23.4
14	128.2	127.4	128.0	37.0	126.8
15	21.1 ^e	20.9	21.0 ^h	21.3	20.8 ^k
16	153.2 ^c	152.3 ^f	153.3 ^g	150.1	154.5
17	115.8	115.8	116.2	110.8	115.6
18	138.1	137.9	138.9	134.1	136.0
19	122.8	122.9	122.8	117.3	121.2
20	142.1 ^d	143.6	142.8	132.6	142.5
21	46.4	46.7	46.5	141.6	46.2
22	33.7	32.5	33.7	122.1	32.7
23	143.4	135.8	144.1	128.3	132.3
24	131.9	131.6	131.9	132.5	130.7
25	117.0	117.8	117.2	123.4	118.1
26	28.4	28.3	28.8	28.5^{i}	28.1
27	21.7	20.8	21.7	23.6	21.5
28	20.5	21.9	20.8	23.5	20.3
29	125.4	124.4	124.8	23.5	122.9
30	21.3 ^e	21.2	21.1 ^h	20.2^{j}	20.9^{k}

Table 2. ¹³C NMR data of parviflorene B-F (2-6)

In DMSO- d_6 .

c-kSignals may be reversed.

correlations as shown in Figure 3, which suggested that **5** consisted of three aromatic benzene rings (rings A, D, and E), one cyclohexene (ring B), and one dihydropyran (ring C), explained as follows.

Three singlet methyl groups on sp² carbons were located on C-3, C-18, and C-24 positions based on the HMBC correlations observed from H₃-15 to C-2, C-3, and C-4, from H₃-30 to C-17, C-18, and C-19, and from H₃-29 to C-23, C-24, and C-25, respectively. Five aromatic protons described above were placed on the C-2, C-4, C-19, C-22,



Figure 2. A computer-generated perspective drawing of parviflorene B (2).

and C-23 positions by the HMBC data shown in Figure 3. One isopropyl group was shown to be attached on the C-21 sp² carbon (HMBC correlations: H-26/C-21, H₃-27/C-21, H₃-28/C-21, and H-22/C-26), while another isopropyl group was located on the C-6 sp^3 carbon (HMBC correlations: H₃-12/C-6 and H₃-13/C-6). A phenol hydroxyl group ($\delta_{\rm H}$ 8.43, s) was placed on C-1 ($\delta_{\rm C}$ 155.8) on the basis of the HMBC cross-peaks from OH-1 to C-1, C-2, and C-10. The C-6 sp³ methine carbon was attached on the C-5 aromatic carbon by the HMBC correlation from H-4 ($\delta_{\rm H}$ 6.73) to C-6 ($\delta_{\rm C}$ 41.2), whereas the C-7 methylene was connected to hemiacetal carbon (C-8, $\delta_{\rm C}$ 98.6) from the HMBC correlation from H2-7 to C-8 as well as the ¹H-¹H COSY correlation between H-7 β ($\delta_{\rm H}$ 1.90) and OH-8 ($\delta_{\rm H}$ 3.83, d) presumably through long-range W-type coupling (J =1.5 Hz). Then, C-8 was shown to be adjacent to an sp^3 quaternary carbon (C-9, $\delta_{\rm C}$ 72.9) bearing a tertiary hydroxyl group ($\delta_{\rm H}$ 3.46, s; OH-9) from the HMBC correlation for H_2 -7/C-9, and C-9 was connected to an sp³ methylene (C-14, $\delta_{\rm C}$ 37.0; HMBC correlations: H₂-14/C-9 and H₂-14/ C-8), which in turn was connected to the C-17 aromatic



Figure 1. Key COSY and HMBC correlations for parviflorene B (2).



Figure 3. Key COSY and HMBC correlations for parviflorene E (5).

a In CDCl₃.



Figure 4. Selected coupling constants and NOE correlations for parviflorene E (5).

carbon ($\delta_{\rm C}$ 110.8; HMBC correlations: H₂-14/C-16, H₂-14/ C-17, and H₂-14/C-18). The ¹³C NMR chemical shift of C-16 ($\delta_{\rm C}$ 150.1) implied that this aromatic carbon was oxygenated and, therefore, was connected with the hemiacetal carbon (C-8) through an ether bond to construct a dihydropyran ring (ring C), and the remaining two quaternary carbons, C-9 and C-10, had to be connected to form a cyclohexene ring (ring B). From these results, the planar structure of **5** was constructed as shown in Figure 3. The relative stereochemistry of **5** was elucidated by NOE as well as ¹H–¹H coupling constants data as shown in Figure 4.

NOE correlations were observed for H-14 β /H-7 β , H-7 β /H₃-13, and H₃-13/H-14 β ; these three hydrogen groups were suggested to be oriented β -side, while NOE were observed for H₃-12/H-6 and H₃-12/H-7 α , implying that these hydrogens are located in α -side. These assignments were consistent with the coupling constants data ($J_{6,7\beta}$ =11.0 Hz and $J_{6,7\alpha}$ =7.6 Hz). Observation of NOE between H-14 β and H-7 β and the W-type coupling (J=1.5 Hz) between H-7 β and the hydroxyl proton on C-8 (OH-8), aided by model considerations, strongly suggested that two hydroxyl groups on C-8 and C-9 (OH-8 and OH-9) had α -cis configuration. Thus, the relative configurations of C-6, C-8, and C-9 positions were revealed as shown in Figure 4.⁶ The pentacyclic skeleton of compound **5** is unprecedented and

corresponded to an unsymmetrical cadinane sesquiterpene dimer, which had a different manner of bond connection (type **B** in Figure 5: C-14 and C-2; C-8 and oxygen on C-1) from that of parviflorene A (1) or B (2) (type **A** in Figure 5: C-14 and C-8; C-8 and C-14). A related cadinane-dimer, named dicadalenol (10), has been isolated from *H. inuloides* (Asteraceae),⁷ having a different bond-connection manner (type **C** in Figure 5: C-14 and C-1; C-8 and oxygen on C-2).



Parviflorene F (**6**) was obtained as an amorphous solid, and the molecular formula was established to be $C_{30}H_{34}O_3$ by its HRFABMS data (*m*/*z* 442.2521, M⁺, Δ +1.3 mmu), possessing one less oxygen atom and two more hydrogen atoms than that of **2**. The ¹H and ¹³C NMR spectrum of **6** (Tables 1 and 2) closely resembled those of **2**. However, the ¹H and ¹³C NMR spectra of **6** showed no signals due to a ketone group, but showed signals due to an sp³ oxymethine (δ_H 4.88 and δ_C 70.0; C-7), which was different from those of **2**. The ¹H–¹H COSY spectrum of **6** showed the presence of a proton connectivity network from one isopropyl group [δ_H 1.87 (1H, m; H-11), δ_H 0.19 (3H, br s; H₃-12), and δ_H 0.88 (3H, d, *J*=7.0 Hz; H₃-13)], to sp³ methine [δ_H 4.88 (1H, t, *J*=5.0 Hz; H-7)], and a hydroxyl proton [δ_H 5.03 (1H, br s;



Figure 5. Three manners of dimerization of cadinane sesquiterpenes.



Figure 6. $\Delta\delta$ values of (*R*)- and (*S*)-MTPA esters (**11a** and **11b**).

OH-7)]. This proton-network moiety was revealed to be present at ring B from the HMBC correlation data observed for H-4/C-6, H-11/C-5, H-7/C-8, and H-29/C-7. The rest of the molecule of 6 (rings A, C, D, and E) was revealed to have the same structure as compound 2 on the basis of the ¹H–¹H COSY and HMBC correlation data as well as the comparison of ¹H and ¹³C chemical shifts (Tables 1 and 2). From the NOE clearly observed between H-6 and H-7 as well as the coupling constant $(J_{6,7}=5.0 \text{ Hz})$ and model considerations, these two hydrogens (H-6 and H-7) were assigned as cis. The isopropyl group on C-21 was deduced to be axially oriented from the two small coupling constants with the vicinal methylene protons on C-22 $(J_{21,22\alpha} =$ 4.3 Hz, $J_{21,228} = 2.7$ Hz), indicating the methine proton on C-21 is equatorially oriented.⁹ The absolute configurations of C-6 and C-7 positions of 6 were elucidated as 6S and 7S on the basis of the modified Mosher's method¹⁰ applied on the (R)- and (S)-MTPA esters (11a and 11b, respectively) prepared from 6 (Fig. 6), whose phenol groups were protected by methyl ethers in advance.

The cytotoxic activity of these sesquiterpene-dimers against human adenocarcinoma cells was examined, and the IC_{50} values (µg/mL) against vincristine (VCR)-resistant KB cells are presented in Table 3. All compounds were cytotoxic to KB cell lines, and parviflorene C (**3**) showed a threefold reversal effect of VCR resistance against VCR-resistant KB cell lines.¹¹ Compounds **3** and **6** also exhibited cytotoxicity against LNCaP (human prostate cancer) and TNF-related apoptosis inducing ligand (TRAIL)-resistant KOB (human adult T cell leukemia) cell lines. From these results, these novel dimeric-sesquiterpene skeletons may provide a unique scaffold as new cytotoxic compounds.

Table 3. Cytotoxicity of parviflorenes A-F (1-6) (IC50 values, µg/mL)

	KB/VJ-300		LNCaP	KOB	
	VCR (+)	VCR (-)		TRAIL (+)	TRAIL (-)
1	2.6	3.1	NT	NT	NT
2	2.2	3.0	NT	NT	NT
3	1.1	3.2	3.8	4.8	6.4
4	3.1	2.8	NT	NT	NT
5	5.9	6.8	7.1	>6.3	>6.3
6	2.3	3.0	4.6	5.9	8.1

Tests toward each cell line were carried out in the absence (-) and presence (+) of 100 ng/mL of VCR and 500 ng/mL of TRAIL, respectively, which did not affect the growth of the cells. NT, not tested.

3. Experimental

3.1. Extraction and isolation

The plant C. parviflora was collected at Khon Kaen, Thailand. A voucher specimen is maintained at the Department of Horticulture, Faculty of Agriculture, Khon Kaen University. The air-dried underground part (280 g) was extracted with MeOH and acetone. The combined extract (12.6 g) suspended in water (200 mL) was partitioned against EtOAc ($400 \text{ mL} \times 2$ and 200 mL) and *n*-BuOH (200 mL \times 2). The EtOAc-soluble fraction (8.0 g) and previously obtained EtOAc and n-BuOH-soluble fractions (2.9 g) from the whole plant were combined, and then were subjected to silica gel column chromatography (column A; 4.5×57 cm) eluted with 0–100% EtOAc in hexane. The fraction (1.5 g) eluted with 20-33% EtOAc in hexane was again subjected to silica gel column chromatography (column B; 4.0×33 cm) isocratically eluted with 20% EtOAc in hexane. A portion (277 mg) of the fraction (331 mg) mainly containing parviflorene B (2) was purified by gel filtration with Sephadex LH-20 (column C; 1.5×55 cm) eluted with MeOH, to give parviflorene B (2, 98 mg). A portion (72 mg) of another fraction (82 mg) mainly containing parviflorene D (4) was separated by Sephadex LH-20 (column D; 1.5×55 cm) eluted with MeOH. Finally, the fraction (5.4 mg) in the 95–105 mL elution was purified by HPLC on ODS (Develosil ODS-HG-5, 20×250 mm; eluent, 80% MeOH, flow rate, 8.0 mL/min; detection UV at 310 nm and RI) to give parviflorene C (3, 1.4 mg, $t_R = 25$ min) and parviflorene D (4, 4.4 mg, $t_{\rm R}$ = 42 min). The fraction (34 mg) in the 60–95 mL elution of column D was also purified by HPLC on ODS (Develosil ODS-HG-5, 20×250 mL; eluent, 80% MeOH, flow rate, 8.0 mL/min; detection UV at 310 nm and RI) to give parviflorene D (4, 25.3 mg, $t_{\rm R}$ =42 min). The fraction (1.57 g) of column A eluted with 10-20% EtOAc in hexane was subjected to silica gel column chromatography (column E; 5.0×20 cm) eluted with 15% EtOAc in hexane, followed by separation with Sephadex LH-20 (column F; 1.5×55 cm) eluted with MeOH. The fraction (9 mg) of column F in the 93–102 mL elution was purified by HPLC on ODS (Develosil ODS-MG-5, 10×250 mm; eluent, 95% MeOH, flow rate, 2.0 mL/min; detection UV at 300 nm and RI) to give parviflorene E (5, 3.0 mg, $t_{\rm R}$ = 26 min). The fraction of column A (0.96 g) eluted with 33-50% EtOAc in hexane was partially (0.9 g) subjected to silica gel column chromatography (column G; 3.5×21 cm) eluted with 33–50% EtOAc in hexane, followed by separation with Sephadex LH-20 (column H; 1.5×63 cm) eluted with MeOH. The fraction of column H (237 mg) in the 50-95 mL elution was finally purified by MPLC on ODS (Merck LiChroprep RP-18, 25×310 mm; eluent, 85% MeOH, flow rate, 10.0 mL/min; detection UV at 280 nm and RI) to give parviflorene F (6, 195 mg).

3.1.1. Parviflorene B (2). Yellow crystal from MeOH; mp $> 300 \,^{\circ}\text{C}$; $[\alpha]_{D}^{26} = -16 (c \, 1.4, \text{MeOH})$; IR (KBr) $\nu_{\text{max}} 3420$, 2950, 1690, 1620, 1460, 1300, and 1160 cm⁻¹; ¹H and ¹³C NMR in CDCl₃ (Tables 1 and 2); UV (MeOH) λ_{max} nm (log ε) 378 (3.9), 326 (4.4), 313 (4.4), 275 (4.6), 237 (4.4), and 218 (4.5); CD (0.043 mM, MeOH, 22 °C) $\Delta \varepsilon$ (λ_{ext} nm) -6.4 (380), 0 (358), 3.0 (310), 3.5 (305), 0 (291), -28.1

(275), -1.1 (259), 5.9 (254), 0 (245), -1.8 (242), 0 (239), 16.0 (228), and 9.0 (219); EIMS m/z (%) 456 (M⁺, 15) and 413 (M-(CH₃)₂CH, 100); HRFABMS calcd for C₃₀H₃₂O₄ (M⁺) 456.2301, found m/z 456.2260.

3.1.2. Parviflorene C (3). Yellow amorphous; $[\alpha]_D^{24} = +44$ (*c* 0.7, MeOH); IR (KBr) ν_{max} 3380, 2960, 2925, 2866, 1698, 1682, 1610, 1298, 1161, and 1014 cm⁻¹; ¹H and ¹³C NMR in CDCl₃ (Tables 1 and 2); UV (MeOH) λ_{max} nm (log ε) 387 (3.8), 323 (4.2), 308 (4.2), 285 (4.3), and 220 (4.4); CD (0.014 mM, MeOH, 24 °C) $\Delta \varepsilon$ (λ_{ext} nm) -5.7 (387), 0 (356), 7.0 (335), 0 (310), -20.5 (278), 0 (262), 24.5 (245), 5.8 (230), 14.4 (221), 0 (221), and -6.0 (207); FABMS (NBA) *m/z* 456 (M⁺); HRFABMS calcd for C₃₀H₃₂O₄ (M⁺) 456.2301, found *m/z* 456.2288.

3.1.3. Parviflorene D (4). Yellow amorphous; $[\alpha]_D^{24} = +242$ (*c* 1.15, MeOH); IR (KBr) ν_{max} 3380, 2960, 2920, 2870, 1700, 1670, 1650, 1620, 1300, and 1160 cm⁻¹; ¹H and ¹³C NMR in CDCl₃ (Tables 1 and 2); UV (MeOH) λ_{max} nm (log ε) 378 (3.9), 326 (4.4), 312 (4.3), 275 (4.6), 237 (4.4), and 218 (4.5); CD (0.025 mM, MeOH, 24 °C) $\Delta \varepsilon$ (λ_{ext} nm) 0 (371), -24.2 (329), 0 (306), 6.8 (287), 380 (271), 52.5 (253), 4.5 (232), 0 (228), -25.0 (219), and -9.0 (209); FABMS (NBA) *m/z* 457 [M+H]⁺; HRFABMS calcd for C₃₀H₃₂O₄, (M⁺) 456.2301, found *m/z* 456.2339.

3.1.4. Parviflorene E (5). Yellow amorphous; $[\alpha]_D^{24} = -105$ (*c* 1.17 MeOH); IR (film) ν_{max} 3340, 2960, 2930, 2870, 1710, 1700, and 1630 cm⁻¹; UV (MeOH) λ_{max} nm (log ε) 304 (3.9), 237 (4.7), and 206 (4.9); CD (0.017 mM, MeOH, 24 °C) $\Delta \varepsilon$ (λ_{ext} nm) -2.1 (354), 0 (321), 1.5 (288), 0 (270), -1.2 (254), 0 (250), 28.1 (234), 0 (215), and -10.6 (205); ¹H and ¹³C NMR in CDCl₃ (Tables 1 and 2); EIMS *m/z* (%) 460 (M⁺, 48), 442 (M-H₂O, 26), 424 (M-2H₂O, 18), 381 (18), and 234 (100); FABMS *m/z* 460 (M⁺); HRFABMS calcd for C₃₀H₃₄O₄ (M⁺) 460.2614, found *m/z* 460.2595.

3.1.5. Parviflorene F (6). Yellow amorphous; $[\alpha]_{D}^{2D} = +195$ (*c* 1.0, MeOH); IR (film) ν_{max} 3280, 2950, 2870, 1620, and 1310 cm⁻¹; UV (MeOH) λ_{max} nm (log ε) 340 (4.5), 326 (4.5), 289 (4.1), 258 (4.0), 250 (4.1), and 226 (4.6); CD (0.023 mM, MeOH, 24 °C) $\Delta \varepsilon$ (λ_{ext} nm) -21.9 (339), -19.1 (333), -23.3 (325), -2.2 (301), -16.0 (287), 0 (271), 150 (242), 0 (231), and -37.0 (224); ¹H and ¹³C NMR in DMSO-*d*₆ (Tables 1 and 2); EIMS *m/z* (%) 442 (M⁺, 9), 424 (M-H₂O, 100), and 381 (20); FABMS (NBA) *m/z* 442 [M⁺]; HRFABMS calcd for C₃₀H₃₄O₃, (M⁺) 442.2508, found *m/z* 442.2521.

3.1.6. Preparation of the (*R*)- and (*S*)-MTPA esters (11a and 11b). A solution of 6 (17 mg) in MeOH (1 mL) was treated with 10% TMS-CHN₂ in hexane (0.5 mL) at room temperature for 14 h. The reaction mixture was evaporated and purified over a silica gel column chromatography eluted with hexane/EtOAc to afford dimethyl ether (14 mg), which was then dissolved in dry pyridine (20 μ L), and treated with (*S*)-(+)- α -methoxy- α -(trifluoromethyl)phenylacetyl chloride [(*S*)-MTPA-Cl] (5 μ L) at rt for 14 h. After addition of 3-[(dimethylamino)propyl]amine (3 μ L), the reaction mixture was evaporated and purified over a silica gel

column chromatography eluted with hexane/EtOAc to give the (R)-MTPA ester (11a): ¹H NMR (CDCl₃, at 55 °C) $\delta_{\rm H}$ 6.660 (1H, s; H-2), 6.674 (1H, s; H-4), 2.862 (1H, t, J=5.0 Hz; H-6), 6.364 (1H, br d, J=4.0 Hz; H-7), 1.726 (1H, br s; H-11), 0.414 (3H, br s; H₃-12), 0.735 (3H, d, $J = 5.2 \text{ Hz}; H_3-13$, 8.019 (1H, s; H-14), 6.732 (1H, s; H-17), 6.629 (1H, s; H-19), 2.964 (1H, dd, J=14.7, 4.8 Hz; H-22), 2.913 (1H, dd, J=14.7, 2.8 Hz; H-22'), 0.771 (3H, d, J=6.7 Hz; H₃-27), 0.799 (3H, d, J=6.7 Hz; H₃-28), and 8.304 (1H, s; H-29); EIMS m/z (%) 686 (M⁺, 41), 452 (100), 409 (12), 367 (42), and 352 (9). The (S)-MTPA ester (11b) was also prepared from 6 by the same procedures using (R)-MTPA chloride. Compound 11b: ¹H NMR (CDCl₃, at 55 °C) $\delta_{\rm H}$ 6.667 (1H, s; H-2), 6.694 (1H, s; H-4), 2.883 (1H, t, J=5.0 Hz; H-6), 6.422 (1H, br d, J=4.0 Hz; H-7), 1.880 (1H, br s; H-11), 0.453 (3H, br s; H_{3} -12), 0.855 (3H, d, J=5.2 Hz; H_{3} -13), 8.003 (1H, s; H-14), 6.723 (1H, s; H-17), 6.628 (1H, s; H-19), 2.954 (1H, dd, J=14.7, 4.8 Hz; H-22), 2.904 (1H, dd, J=14.7, 2.8 Hz; H-22'), 0.776 (3H, d, J=6.7 Hz; H₃-27), 0.797 (3H, d, J = 6.7 Hz; H₃-28), and 8.234 (1H, s; H-29); EIMS m/z (%) 686 (M⁺, 9), 452 (100), 409 (11), 367 (37), and 352 (8).

3.2. X-ray crystallographic analysis

All measurements were made on a Rigaku/MSC Mercury CCD diffractometer with graphite monochromated Mo Ka radiation ($\lambda = 0.71069$ Å). The data were collected at a temperature of -173 ± 1 °C to a maximum 2θ value of 57.0°. A total of 0 oscillation images were collected. Crystal data: triclinic, C_{33.5}H₄₆O_{7.5} (M_r 568.73), space group P1 (#1) with a=11.389 (3) Å, b=11.715 (3) Å, c=12.858(3) Å, $\alpha = 83.200$ (3)°, $\beta = 76.259$ (3)°, $\gamma = 69.671$ (3)°, V=1561.5 (6) Å³, Z=2, and $D_{calcd}=1.210$ g/cm³. The structure was solved by direct methods (SIR97) and expanded using Fourier techniques (DIRDIF94). The nonhydrogen atoms were refined anisotropically. Hydrogen atoms were included but not refined. The final cycle of fullmatrix least-squares refinement was based on 5695 observed reflections $(I > 1.50\sigma(I), 2\theta < 56.98^{\circ})$ and 740 variable parameters and converged with unweighted and weighted agreement factors of R = 0.053, $R_w = 0.066$.

Crystal data for the structural analysis have been deposited with the Cambridge Crystallographic Data Centre, CDCC deposition number 233828. The supplementary crystallographic data for this paper can be obtained free of charge from CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (fax: +44 1233 336033; e-mail: deposit@ccdc.cam.ac.uk or http://www.ccdc.cam.ac.uk/).

3.3. Cultured cell lines for cytotoxicity tests and measurement of cell viability

VCR selected multidrug resistant variants of the human epidermoid carcinoma KB cells $(KB/VJ-300)^{12}$ were a gift from Prof. M. Kuwano (Kyushu University School of Medicine). The human prostate carcinoma LNCaP (human prostate cancer) cells were a gift from Prof. S. Egawa (Kitasato University School of Medicine). All these cells were maintained in culture flasks in MEM medium supplemented with 10% fetal calf serum (FCS) and 60 µg/mL of kanamycin. KOB cells (human adult T-cell leukemia)

were a gift from Prof. T. Yamada (Graduate School, Nagasaki University), and were maintained in RPMI-1640 medium supplemented with 10% FCS and 60 μ g/mL of kanamycin.

For the in vitro drug treatment experiments, tumor cells $(1.1 \times 10^4 \text{ cells/mL}$ for KB cells, and $2 \times 10^4 \text{ cells/mL}$ for LNCaP cells) were seeded in 195 µL of culture medium/ well in 96-well plates (Corning Glass Works), and incubated for 24 h at 37 °C in a 5% CO₂–95% air atmosphere. The cells were treated in triplicate with 5 µL of graded concentrations of samples, and were then incubated in a CO₂ incubator at 37 °C for 72 h. Cell viability was determined by the colorimetric assay using MTT.¹³ For KOB cells, 3.5×10^5 cells/mL of the cells were seeded in 95 µL of culture medium/well in 96-well plates, and were treated with 5 µL of graded concentrations of samples in the absence or presence of 0.5 µg/mL of TRAIL, and were then incubated for 42 h at 37 °C in a 5% CO₂–95% air atmosphere. Cell viability was determined by the colorimetric assay using the alamer blue.¹⁴

Acknowledgements

This work was partly supported by a Grant-in-Aid from the Ministry of Education, Culture, Sports, Science, and Technology of Japan.

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- 8. Interestingly, the ¹H NMR spectrum of **4** varied depending on its concentrations. Particularly, the aromatic protons (H-17 and H-19), methylene protons on C-22 (H₂-22), and the isopropyl methyl protons (H₃-27 and H₃-28) resonated at significantly different chemical shifts ($\Delta\delta$ 0.6–0.8 ppm) on the different concentrations (2.0–31 mg/mL in CDCl₃). We assumed this phenomenon was due to intermolecular π – π interaction of the aromatic rings of the compound: Hunter, C. A.; Sanders, J. K. M. J. Am. Chem. Soc. **1990**, *112*, 5525–5534.
- 9. Interestingly, the ¹H NMR spectrum of **6** became sharp depending on its temperature. Particularly, one of the isopropyl methyl groups resonating at high-field ($\delta_{\rm H}$ 0.19, 3H, br s; H₃-12) as a broad singlet at 28 °C appeared sharply as a doublet (J=7.0 Hz) at 50 °C, and the same phenomenon was observed for OH-7 [$\delta_{\rm H}$ 5.30 (br s)], H-7 methine [$\delta_{\rm H}$ 4.88 (t, 5.0)], and H₂-22 methylene [$\delta_{\rm H}$ 2.82 (dd) and 2.80 (dd)] signals.
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Tetrahedron

Tetrahedron 60 (2004) 10825-10832

Synthesis of 5-alkyl(or aryl)pyrrolo[1,2-*a*]quinoxalin-4(5*H*)-ones by denitrocyclisation of *N*-alkyl(or aryl)-1-(2-nitrophenyl)-1*H*pyrrole-2-carboxamides. Evidence of a Smiles rearrangement

Georgios Rotas, Athanasios Kimbaris and George Varvounis*

Department of Chemistry, University of Ioannina, GR-451 10 Ioannina, Greece

Received 16 July 2004; revised 25 August 2004; accepted 16 September 2004

Available online 2 October 2004

Abstract—An efficient method for the synthesis of hitherto unknown alkyl(or aryl)pyrrolo[1,2-*a*]quinoxalin-4(5*H*)-ones **8a–g**, **16** and **17** has been established. The method is based on the synthesis of the corresponding *N*-alkyl(or aryl)-1-(2-nitrophenyl)-1*H*-pyrrole-2-carboxamides **3a–c** and **7a–c**, **e** which undergo denitrocyclisation with NaH in DMF in 4.5 or 2 h. When **3a** was treated with NaH in DMF for 30 min the product of a Smiles rearrangement, **9**, was isolated. Under similar conditions but for 4.5 h **9** was converted into **8a**. This confirms the involvement of a Smiles rearrangement during the denitrocyclisation process. Conversion of **3b** into isomeric pyrroloquinoxalinones **12** and **13** confirms a process involving two pathways, direct denitrocylisation of **3b** and Smiles rearrangement of **3b** followed by denitrocylisation, respectively. Furthermore, denitrocylisation of **7d** into pyrroloquinoxalinones **16** and **17** suggests that similar cyclisation pathways are followed by *N*-arylcarboxamides.

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1. Introduction

Pyrrolo[1,2-*a*]quinoxalines are best known as high-affinity and selective agonists of the 5-HT₃ receptors.^{1–3} More recently, several bispyrrolo[1,2-*a*]quinoxaline derivatives were found to have significant antimalarial activity⁴ whereas certain pyrrolo[1,2-*a*]quinoxalin-4(5*H*)-ones have shown promising antiviral⁵ and antiallergic⁶ properties.

5-Alkylpyrrolo[1,2-*a*]quinoxalin-4(5*H*)-ones may be formed by first synthesising the lactam derivatives^{1–3,7} followed by alkylation.⁵ Other routes include the use of benzimidazolium *N*-ylides in 1,3-dipolar cycloadditions with alkenes^{6,8} and alkynes⁹ or reaction with 2,2-dihydropoly(per)fluoroalkanoates,¹⁰ and by reductive ring-opening/ ring-closure of pyridazinoquinoxalinones.¹¹

2. Results and discussion

Following our previous interest in the synthesis of pyrrolo[1,2-a]quinoxalines¹² we now wish to report our findings that lead to the synthesis of 5-alkyl(or aryl)-pyrrolo[1,2-a]quinoxalin-4(5*H*)-ones by denitrocyclisation

of *N*-alkyl(or aryl)-1-(2-nitrophenyl)-1*H*-pyrrole-2-carboxamides.

Carboxamides **3a–c** were prepared by reacting appropriate 2-trichloroacetylpyrroles **2a–b**, derived by treating 1-arylpyrroles **1a–b** with trichloroacetyl chloride, with diethylamine or cyclohexylamine in 1,4-dioxane for 1.5 h (Scheme 1).

In contrast, the preparation of aromatic carboxamides 7a-e required first hydrolysis of trichloroacetyl derivative 2a to the carboxylic acid 4, conversion of the latter with thionyl chloride into acid chloride 5 and then, without isolation, treatment of 5 with aromatic amines 6a-e for 2 days in a 1:1 pyridine–toluene mixture (Scheme 2). Attempts to react 4-chloro-2-nitroaniline or 2-trifluoromethylaniline with acid chloride failed. This is probably due to a combination of steric and electronic reasons which is reflected in the low yield (45%) of 7b. Railey and Johnson have previously prepared a large number of *N*-alkyl(or aryl)-4,5-dihalo-1*H*-pyrrole-2-carboxamides, as potential antibacterial agents, in a similar manner.¹³

Upon treating carboxamides 3a,c or 7a-c,e with NaH in DMF for 4.5 or 2 h, pyrroloquinoxalinones 8a-f were obtained in excellent yields (77–95%). The work-up involved adding to water and neutralising with 2 N HCl. This led to hydrolysis of the initially formed ester 8e to the carboxylic acid 8d. The ester 8e was isolated when the

Keywords: Pyrroles; Pyrroloquinoxalinones; Denitrocyclisation; Defluorocyclisation; Smiles rearrangement.

^{*} Corresponding author. Tel.: +30 26510 98382; fax: +30 26510 98799; e-mail: gvarvoun@cc.uoi.gr

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Scheme 1. Reagents (a) Cl₃CCOCl, 1,4-dioxane; (b) 70% EtNH₂ in H₂O, CH₂Cl₂ or cyclohexylamine, Et₃N, 1,4-dioxane, 75 °C.



Scheme 2. Reagents (a) 10% aq. NaOH, reflux; (b) $SOCl_2$, reflux; (c) 6a PhNH₂, 6b 2-MeO₂C₆H₄NH₂, 6c 4-ClC₆H₄NH₂, 6d 2,4-F₂C₆H₃NH₂ or 6e 4-MeOC₆H₄NH₂, pyridine.

reaction mixture was poured into ice-water containing sodium bicarbonate. To the best of our knowledge, this intramolecular displacement of an aromatic nitro group by *N*-substituted carboxamides, under mild conditions, to form 5-substituted pyrroloquinoxalinones, is unprecedented in the literature. The closest analogy was published by Nacci et al.^{7c} who cyclised 1-(2-nitrophenyl)-1*H*-pyrrole-2carboxamide to the parent pyrrolo[1,2-*a*]quinoxalin-4(5*H*)one, by heating in DMF containing potassium carbonate (Scheme 3).

It was interesting to note that TLC examination of reactions **3a** or **c** with NaH in DMF after 0.5 h, revealed a new spot together with spots corresponding to starting material and product. We therefore investigated further the reaction of **3a** with NaH in DMF by allowing it to proceed for 0.5 h. After work-up and column chromatography three compounds were isolated, starting material **3a**, pyrroloquinoxalinone **8a** and carboxamide **9** in 13, 27 and 21% yield, respectively (Scheme 4). This result provides conclusive evidence that a Smiles rearrangement is taking place. Furthermore, reaction



Scheme 3. Reagents (a) NaH (60% in oil), DMF.

of **9** with NaH in DMF for 4.5 h gave **8a** as sole product. TLC examination of this reaction after 0.5 h revealed three spots that corresponded to **3a**, **8a** and starting material **9**.

From the above observations we can confidently propose that carboxamide 3a undergoes a Smiles rearrangement to give carboxamide 9. However, although there is ample indication that the transformation of 3a to 9 is reversible, there is no direct evidence as to whether 3a or 9 or both lead to 8a.

Further insight into the mechanism of 4-alkylpyrrologuinoxalinone formation was obtained by reacting carboxamide 3b with NaH in DMF for 1.5 h. This gave a mixture containing pyrroloquinoxalinones 12 and 13 (Scheme 5). Since the separation of these two compounds by column chromatography was not possible, 12 was synthesised unambiguously by N-ethylating 7-methylpyrrolo[1,2-a]quinoxalin-4(5H)-one³ with ethyl iodide in DMF containing NaH. The ¹H NMR spectra of compound **12** and that of the mixture 12/13 were then compared and the peaks corresponding to 12 and 13 assigned. It turned out that 12 and 13 are in a ratio of about 44:36. Compounds 12 and 13 could only have been obtained from the corresponding anions of carboxamides 10 and 11 by denitrocyclisation through paths b and d. Compound 11 can only be obtained from 10 by a Smiles rearrangement that follows path a. Furthermore, based on previous observations (vide infra), it is quite reasonable to assume that the transformation of 10 to 11 is reversible through path c.

In order to verify whether a Smiles rearrangement occurred during the denitrocyclisation of aromatic carboxamides 7a-c,e to pyrroloquinoxalinones 8c,e-g, N-(2,4-difluorophenyl)carboxamide 7d was employed as a potentially useful precursor. When 7d was treated with NaH in DMF for 1.5 h, two products 16 and 17 were obtained in 75 and 15% yield, respectively. A proposed mechanism for this reaction is shown in Scheme 6. The initially formed carboxamide anion 14 could follow two paths a or b. Path a would lead to pyrroloquinoxalinone 16 by direct denitrocylisation whereas path b would lead to intermediate carboxamide 15 via a Smiles rearrangement. Taking into account previous observations (vide infra) it is reasonable to assume that 15 can reverse to 14 through path e and denitrocyclise to 16 through path c. On the other hand, pyrroloquinoxalinone 17 can only be formed through path d, that is, intramolecular nucleophilic displacement of fluoride anion by pyrrolyl anion of



Scheme 4. Reagents (a) NaH (60% in oil), DMF, 30 min; (b) NaH (60% in oil), DMF, 4.5 h.



Scheme 5. Reagents (a) NaH (60% in oil), DMF, 1.5 h.



16 (75%)

17 (15%)

intermediate 15. The formation of 17 strongly suggests that intermediate 15 is its precursor. However, there is no substantial evidence to support that 16 is obtained directly from 14 or from intermediate 15 or from both.

3. Conclusion

In conclusion, we have shown that denitrocyclisation of N-alkyl(or aryl)-1-(2-nitrophenyl)-1H-pyrrole-2-carboxamides is a mild method that gives 5-alkyl(or aryl)pyrrolo [1,2-a]quinoxalin-4(5H)-ones in high yields. For N-alkyl-1-(2-nitrophenyl)-1H-pyrrole-2-carboxamides there is direct evidence that reaction occurs through two pathways: (a) intramolecular nitro group displacement by the anion of N-alkyl-1-(2-nitrophenyl)-1H-pyrrole-2-carboxamides and (b) Smiles rearrangement of N-alkyl-1-(2-nitrophenyl)-1H-pyrrole-2-carboxamide anions into N-(2-nitrophenyl)-Nalkyl(or aryl)-1H-pyrrole-2-carboxamide anions, followed by intramolecular pyrrolyl anion displacement of the nitro group. It is highly probable that N-aryl-1-(2-nitrophenyl)-1Hpyrrole-2-carboxamides give 5-arylpyrrolo[1,2-a]quinoxalin-4(5H)-ones by following similar pathways.

4. Experimental

4.1. General methods

Melting points were taken on a Büchi 510 apparatus and are uncorrected. Infrared spectra were recorded on a Perkin-Elmer 257 spectrometer, as Nujol mulls and liquids between sodium chloride discs. Elemental analyses were performed on a Perkin-Elmer 2400 or a Carlo Erba 1106 elemental analysers. Nuclear magnetic resonance spectra were measured at 360 MHz on a Brüker AM 360 spectrometer or at 400 MHz on a Brüker AMX 400 spectrometer using tetramethylsilane as internal standard. Mass spectra were obtained using a JEOL JMS-AX 505W or a Bruker Apex III high-resolution instruments. Analytical TLC was carried out on Fluka silica gel 60 F₂₅₄. Preparative flash chromatography was carried out for all separations using Merck 9385 silica gel. Solvents and reagents were used as received from the manufacturers, except for dichloromethane, ethanol, ethyl acetate, hexane and methanol that were purified and dried according to recommended procedures.14

4.1.1. Preparation of 2-trichloroacetylpyrroles 2a,b: general procedure A. To a recently distilled solution of trichloroacetyl chloride (41.8 mL, 372 mmol) in dry 1,4dioxane (140 mL), a solution of pyrrole **1a** or **1b** in (0.124 mol) in dry 1,4-dioxane (140 mL) was added dropwise. The reaction mixture was left to stir at room temperature for 6 days and then a saturated solution of potassium carbonate was slowly added until the pH was 7. Water (120 mL) was added and extracted with dichloromethane (3×80 mL). The combined organic extracts were dried (Na₂SO₄) and the solvent removed under reduced pressure to afford a residue, which on crystallisation from ethanol gave 2,2,2-trichloro-1-(1-(2-nitrophenyl)-1*H*pyrrol-2-yl)ethanone **2a** or 2,2,2-trichloro-1-(1-(4-methyl-2-nitrophenyl)-1*H*-pyrrol-2-yl)-ethanone **2b**. **4.1.1.1.** 2,2,2-Trichloro-1-[1-(2-nitrophenyl)-1*H*-pyrrol-2-yl)]ethanone 2a. 4.20 g, 89%, as pale yellow needles (ethanol), mp 115–117 °C. [Found: C, 43.19; H, 2.15; N, 8.41. $C_{12}H_7Cl_3N_2O_3$ requires, 43.21; H, 2.12; N, 8.40%]; ν_{max} (Nujol) 1660, 1520, 1340 cm⁻¹; δ_H (400 MHz; CDCl₃) 6.41 (t, *J*=3.8 Hz, 1H, H-4), 6.96 (s, 1H, H-3), 7.34 (d, *J*=7.8 Hz, 1H, H-6'), 7.54–7.60 (2H, m, H-5, H-4'), 7.65 (t, *J*=7.7 Hz, 1H, H-5'), 8.08 (d, *J*= 8.1 Hz, 1H, H-3'); δ_C (100 MHz; CDCl₃) 111.60, 122.60, 124.76, 125.19, 130.13, 130.34, 133.65, 134.75, 135.03, 145.41, 171.73; *m*/z (EI) 332 (M⁺+1, 32), 215 (92), 187 (82), 171 (100%).

4.1.1.2. 2,2,2-Trichloro-1-[1-(4-methyl-2-nitrophenyl)-1*H*-pyrrol-2-yl)]ethanone 2b. 37.9 g, 88%, as pale yellow needles (ethanol), mp 122–124 °C. [Found: C, 45.03; H, 2.67; N, 8.01. C₁₃H₉Cl₃N₂O₃ requires C, 44.92; H, 2.61; N, 8.06%]; ν_{max} (Nujol) 3124, 1670, 1528, 1347 cm⁻¹; $\delta_{\rm H}$ (400 MHz; CDCl₃) 2.52 (s, 3H, Me), 6.47 (dd, *J*=4.3, 2.7 Hz, 1H, H-4), 7.02 (dd, *J*=2.6, 1.6 Hz, 1H, H-5), 7.29 (d, *J*=7.9 Hz, 1H, H-6'), 7.51 (dd, *J*=7.9, 1.3 Hz, 1H, H-5'), 7.66 (dd, *J*=4.4, 1.5 Hz, 1H, H-3), 7.96 (d, *J*=1.3 Hz, 1H, H-3'); $\delta_{\rm C}$ (100 MHz; CDCl₃) 20.88, 95.44, 111.00, 123.64, 124.31, 125.61, 129.34, 131.71, 132.77, 134.44, 140.59, 145.09, 172.28; *m/z* (EI) 348 (M⁺+2, 6), 346 (M⁺, 7), 302 (5) 300 (5), 283 (7), 265 (5), 229 (50), 201 (40), 183 (100), 154 (21), 84 (30%).

4.1.2. Preparation of 1*H*-pyrrole-2-carboxamides 3a,b: general procedure B. To a solution of 2,2,2-trichloro-1-(1H-pyrrol-2-yl)ethanone 2a or 2b (9.04 mmol) in dichloromethane (40 mL) was added a solution of ethylamine 70% in water (3.5 mL, 54.225 mmol). The reaction mixture was stirred at room temperature for 18 h. Water was added (90 mL), the organic phase separated and the aqueous phase extracted with dichloromethane (3×30 mL). The combined organic phases were dried (Na₂SO₄) and the solvent removed under reduced pressure to afford a residue which was triturated with toluene and filtered. The residue was purified by dissolving in ethyl acetate and precipitating by addition of hexane to afford *N*-ethyl-1-(2-nitrophenyl)-1*H*-pyrrole-2-carboxamide **3b**.

4.1.2.1. *N*-Ethyl-1-(2-nitrophenyl)-1*H*-pyrrole-2-carboxamide 3a. 2.16 g, 92%, as a pale-yellow solid (ethyl acetate/hexane); mp 93–95 °C; ν_{max} (CCl₄) 3450, 2980, 1665, 1350 cm⁻¹; $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.05 (t, *J*= 7.4 Hz, 3H, Me), 3.20 (q, *J*=6.9 Hz, 2H, CH₂), 5.79 (br, s, 1H, NH), 6.26 (t, *J*=3.3 Hz, 1H, H-4), 6.60 (d, *J*=3.9 Hz, 1H, H-3), 6.82 (s, 1H, H-5), 7.37 (d, *J*=7.9 Hz, 1H, H-6'), 7.47 (dd, *J*=8.0, 7.7 Hz, 1H, H-4'), 7.59 (dd, *J*=7.9, 7.7 Hz, 1H, H-5'), 7.98 (d, *J*=8.0 Hz, 1H, H-3'); $\delta_{\rm C}$ (100 MHz; CDCl₃) 14.80, 34.03, 109.54, 112.24, 124.67, 127.01, 127.49, 128.67, 129.58, 133.31, 134.95, 145.02, 160.48; *m*/z (EI) 259 (M⁺, 20), 213 (100), 188 (24), 171 (71) 169 (80), 155 (22), 143 (30), 118 (22%). HRMS (EI): (M⁺), found 259.0961. C₁₃H₁₃N₃O₃ requires 259.0957.

4.1.2.2. *N*-Ethyl-1-(4-methyl-2-nitrophenyl)-1*H*pyrrole-2-carboxamide 3b. 2.22 g, 90%, as a pale-yellow solid (ethyl acetate/hexane); mp 115–117 °C. [Found: C, 61.48; H, 5.47; N, 15.34; C₁₄H₁₅N₃O₃ requires C, 61.53; H,

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5.53; N, 15.38%]; ν_{max} (Nujol) 3290, 1631, 1530, 1353 cm⁻¹; $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.10 (t, J=7.2 Hz, 3H, Me), 2.47 (s, 3H, Me), 3.25 (q, J=7.2 Hz, 2H, CH₂), 5.92 (br, s, 1H, NH), 6.30 (dd, J=3.9, 2.8 Hz, 1H, H-4), 6.65 (dd, J=3.9, 1.6 Hz, 1H, H-3), 6.80 (dd, J=2.8, 1.6 Hz, 1H, H-5), 7.29 (d, J=8.1 Hz, 1H, H-6'), 7.45 (dd, J=8.1, 1.3 Hz, 1H, H-5'), 7.86 (d, J=1.3 Hz, 1H, H-3'); $\delta_{\rm C}$ (100 MHz; CDCl₃) 14.79, 20.80, 34.00, 109.26, 111.91, 125.03, 127.04, 127.49, 128.31, 132.31, 133.83, 139.38, 145.60, 160.49; m/z (EI) 273 (M⁺, 10), 227 (100), 199 (16), 185 (46), 183 (27), 158 (19%).

4.1.3. N-Cyclohexyl-1-(2-nitrophenyl)-1H-pyrrole-2carboxamide 3c. To a solution of 2,2,2-trichloro-1-(1-(2nitrophenyl)-1*H*-pyrrol-2-yl)ethanone **2a** (3 g, 9.04 mmol) in dry 1,4-dioxane (150 mL) under argon was added dropwise freshly distilled cyclohexylamine (5.17 mL, 45.18 mmol) and triethylamine (1.56 mL, 11.296 mmol) in dry 1,4-dioxane. The reaction mixture was stirred at 75 °C for 2 h and then the solvent removed under reduced pressure. Water was added (60 mL), the solution acidified with 2 N hydrochloric acid until the pH was 5 and then extracted with dichloromethane $(3 \times 20 \text{ mL})$. The combined organic extracts were dried (Na_2SO_4) and the solvent removed under reduced pressure to afford a residue which was photosensitive. The residue was suspended in toluene, filtered, and then purified from ethyl acetate/hexane to give the title compound **3c** as a pale-yellow solid (2.75 g, 97%); mp 98–100 °C; ν_{max} (Nujol) 3294, 1627, 1609, 1531, 1357 cm^{-1} ; δ_{H} (400 MHz; CDCl₃) 1.09–1.90 [m, 10H, (CH₂)₅], 3.67-3.74 (m, 1H, CH), 5.73 (s, 1H, NH), 6.32 (dd, J=3.8, 2.8 Hz, 1H, H-4), 6.66 (dd, J=3.8, 1.7 Hz, 1H, H-3), 6.82 (dd, J=2.8, 1.7 Hz, 1H, H-5), 7.44 (d, J=7.8 Hz, 1H, H-6'), 7.53 (dd, J = 8.0, 7.6 Hz, 1H, H-4'), 7.66 (dd, J =7.8, 7.6 Hz, 1H, H-5'), 8.04 (d, J=8.0 Hz, 1H, H-3'); $\delta_{\rm C}$ (100 MHz; CDCl₃) 24.84, 25.44, 33.07, 47.91, 109.48, 111.93, 124.67, 126.98, 127.80, 128.64, 129.55, 133.26, 134.92, 146.01, 159.69; *m*/*z* (EI) 313 (M⁺, 20), 267 (88), 215 (26), 185 (80), 171 (88) 169 (100), 155 (52), 143 (32), 118 (22%). HRMS (EI): (M⁺), found 313.1422, $C_{17}H_{19}N_3O_3$ requires 313.1426.

4.1.4. 1-(2-Nitrophenyl)-1H-pyrrole-2-carboxylic acid 4. A suspension of 2,2,2-trichloro-1-[1-(2-nitrophenyl)-1Hpyrrol-2-yl)]ethanone 2a (11.5 g, 35 mmol) in 10% aqueous sodium hydroxide (70 mL) was heated under reflux for 1.5 h. The resulting solution was allowed to cool and then acidified with 2 N hydrochloric acid to pH 4. The precipitate was filtered, washed with water, dried and recrystallised from propan-2-ol to give the title compound 4 as deep yellow microcrystals (6.81 g, 84%); mp 195.5-197 °C. [Found: C, 57.22; H, 3.52; N, 12.08; C₁₁H₈N₂O₄ requires C, 56.90; H, 3.47; N, 12.06%]; ν_{max} (Nujol) 3257, 3136, 1690, 1669, 1607, 1528, 1350 cm⁻¹; δ_{H} (400 MHz; CDCl₃) 6.39 (t, J=3.3 Hz, 1H, H-4), 6.94 (s, 1H, H-3), 7.20 (d, J= $3.9 \text{ Hz}, 1\text{H}, \text{H-5}, 7.41 \text{ (d}, J = 7.8 \text{ Hz}, 1\text{H}, \text{H-6}^{\prime}), 7.59 \text{ (t}, J =$ 7.7 Hz, 1H, H-4'), 7.68 (t, J = 7.7 Hz, 1H, H-5'), 8.09 (d, J =8.0 Hz, 1H, H-3'), 12.32 (s, 1H, COOH); $\delta_{\rm C}$ (100 MHz; CDCl₃) 110.05, 118.67, 124.19, 124.18, 129.55, 129.93, 130.22, 134.07, 134.28, 145.93, 161.21; m/z (EI) 232 (M⁺, 47), 187 (100), 171 (65), 159 (28) 143 (38), 131 (35), 102 (23), 83 (23), 77 (24%).

4.1.5. Preparation of 1*H*-pyrrole-2-carboxamides 7a–e: general procedure C. A suspension of 1-(2-nitrophenyl)-1H-pyrrole-2-carboxylic acid 4 (1.5 mmol) in freshly distilled thionyl chloride (8 mL) was heated to reflux under argon for 1 h. Removal of excess thionyl chloride, addition of dry benzene (7 mL) and then evaporation under vacuo gave a brownish oily residue. The residue was dissolved in dry benzene (7 mL) and then added dropwise under argon to a solution of the appropriate amine (1.5 mmol) in dry pyridine (7 mL). The reaction mixture was stirred at room temperature for 2 days. The solvents were evaporated in vacuo, a saturated solution of sodium bicarbonate (50 mL) was added and extracted with dichloromethane $(3 \times 20 \text{ mL})$. The combined organic extracts were dried (Na₂SO₄) and the solvent removed under reduced pressure to afford a residue that was purified by flash chromatography (25% ethyl acetate/hexane) to give 1-(2nitrophenyl)-N-phenyl-1H-pyrrole-2-carboxamide 7a, methyl 2-(1-(2-nitrophenyl)-1*H*-pyrrole-5-carboxamido)benzoate 7b, N-(4-chlorophenyl)-1-(2-nitrophenyl)-1H-pyrrole-2carboxamide 7c, N-(2,4-difluorophenyl)-1-(2-nitrophenyl)-1H-pyrrole-2-carboxamide 7d, or N-(4-methoxyphenyl)-1-(2-nitrophenyl)-1*H*-pyrrole-2-carboxamide **7e**.

4.1.5.1. 1-(2-Nitrophenyl)-*N*-phenyl-1*H*-pyrrole-2carboxamide 7a. 387 mg, 84%, as a pale-yellow solid (ethyl acetate/hexane); mp 144–145 °C; ν_{max} (Nujol) 3320, 1637, 1540, 1355 cm⁻¹; $\delta_{\rm H}$ (400 MHz; CDCl₃) 6.17–6.20 (m, 1H, H-4), 6.66–6.68 (m, 2H, H-3, H-5), 6.84 (d, *J* = 7.1 Hz, 1H, H-4"), 7.02–7.11 (m, 2H, H-3", H-5"), 7.20– 7.53 (m, 6H, H-2", H-6", H-6', H-5', H-4', NH), 7.87 (d, *J* = 7.6 Hz, 1H, H-3'); $\delta_{\rm C}$ (100 MHz; DMSO-*d*₆) 109.60, 114.88, 120.36, 123.46, 124.75, 125.98, 128.70, 129.01, 129.19, 129.99, 134.15, 134.72, 139.11, 145.99, 158.79; *m/z* (EI) 307 (M⁺, 33), 261 (22), 215 (32), 169 (100), 140 (17), 114 (12%); HRMS (EI): (M⁺), found 307.0965, C₁₇H₁₃N₃O₃ requires 307.0957.

4.1.5.2. Methyl 2-(1-(2-nitrophenyl)-1*H*-pyrrole-5carboxamido)benzoate 7b. 247 mg, 45%, as a pale-yellow solid (ethyl acetate/hexane); mp 166–168 °C. [Found: C, 62.31; H, 4.35; N, 11.45; $C_{19}H_{15}N_{3}O_5$ requires C, 62.46; H, 4.14; N, 11.50%]; ν_{max} (Nujol) 3257, 3136, 1690, 1669, 1607, 1528, 1350 cm⁻¹; δ_{H} (400 MHz; DMSO-*d*₆) 6.45– 6.48 (m, 1H, H-4), 7.08–7.16 (m, 2H, H-3, H-4"), 7.26–7.27 (m, 1H, H-5), 7.51–7.61 (m, 2H, H-6', H-5"), 7.71 (t, *J*= 7.7 Hz, 1H, H-4'), 7.84 (t, *J*=7.6 Hz, 1H, H-5'), 7.97 (d, *J*= 7.9 Hz, 1H, H-6"), 8.16 (d, *J*=8.0 Hz, 1H, H-3"), 8.26 (d, *J*=8.4 Hz, 1H, H-3'), 11.41 (br, s, 1H, NH); δ_{C} (100 MHz; DMSO-*d*₆) 52.77, 110.15, 114.24, 116.00, 120.21, 122.85, 124.82, 126.73, 129.55, 129.89, 130.16, 130.86, 134.17, 134.26, 134.52, 140.56, 145.84, 158.21, 168.31; *m/z* (EI) 365 (M⁺, 29), 319 (22), 287 (21), 215 (28), 188 (26), 169 (100), 145 (34%).

4.1.5.3. *N*-(**4**-Chlorophenyl)-1-(2-nitrophenyl)-1*H*pyrrole-2-carboxamide 7c. 379 mg, 74%, as a pale-yellow solid (ethyl acetate/hexane); mp 136–137 °C; ν_{max} (Nujol) 3320, 1635, 1520, 1350 cm⁻¹; $\delta_{\rm H}$ (400 MHz; CDCl₃) 6.39 (dd, *J*=3.8, 2.8 Hz, 1H, H-4), 6.86–6.90 (m, 2H, H-3, H-5), 7.18–7.26 (m, 2H, H-3", H-5"), 7.35–7.38 (d, m, 2H, H-2", H-6"), 7.46 (dd, *J*=7.7, 1.5 Hz, 1H, H-6'), 7.57 (t, *J*= 7.6 Hz 1H, H-4'), 7.65–7.70 (m, 2H, H-5', NH), 8.07 (d, *J*= 7.9 Hz, 1H, H-3'); $\delta_{\rm C}$ (100 MHz; DMSO- d_6) 107.51, 109.6, 115.09, 121.73, 124.74, 126.64, 126.98, 128.56, 129.24, 130.01, 134.16, 134.56, 138.07, 145.93, 158.73; *m*/*z* (EI) 343 (M⁺ + 2, 8), 341 (M⁺, 24), 297 (4), 295 (10), 215 (43), 171 (23), 169 (100), 140 (14), 114 (10%); HRMS (EI): (M⁺), found 341.0557, C₁₇H₁₂ClN₃O₃ requires 341.0567.

4.1.5.4. N-(2,4-Difluorophenyl)-1-(2-nitrophenyl)-1Hpyrrole-2-carboxamide 7d. 443 mg, 86%, as a pale-yellow solid (ethyl acetate/hexane); mp 81-83 °C. [Found: C, 59.33; H, 3.29; N, 12.16;. C₁₇H₁₁F₂N₃O₃ requires C, 59.48; H, 3.23; N, 12.24%]; v_{max} (Nujol) 3237, 1639, 1524, 1359 cm⁻¹; $\delta_{\rm H}$ (400 MHz; CDCl₃) 6.42 (t, J=3.3 Hz, 1H, H-4), 6.77 (dddd, J=9.2, 8.3, 2.8, 1.7 Hz, 1H, H-5"), 6.84 (ddd, J = 11.3, 8.3, 2.8 Hz, 1H, H-3''), 6.91-6.92 (m, 2H, H-3)3, H-5), 7.47 (dd, J=7.8, 1.3 Hz, 1H, H-6[']), 7.58 (td, J=8.1, 1.3 Hz, 1H, H-4'), 7.67–7.71 (m, 2H, H-5', NH), 8.03 (td, J=9.2, 6.0 Hz, 1H, H-6''), 8.10 (dd, J=8.1, 1.4 Hz, 1H, H-6'')H-3'); $\delta_{\rm C}$ (100 MHz; CDCl₃) 103.47 (dd, J = 26.6, 23.7 Hz), 110.13, 111.09 (dd, J=21.5, 3.7 Hz), 113.75, 122.40 (dd, J=10.4, 3.8 Hz), 122.95 (dd, J=9.0, 2.1 Hz), 124.99, 126.84, 128.63, 129.21, 129.81, 133.52, 134.47, 146.00, 153.77 (dd, J = 246.2, 11.8 Hz), 158.29, 158.94 (dd, J =246.0, 11.4 Hz); $\delta_{\rm F}$ (376 MHz; CDCl₃) -115.37 (tdd, J= 8.3, 6.0, 4.6 Hz, 1F, F-4"), -126.27 (ddddd, J=11.0, 9.2, 4.6, 2.9, 1.7 Hz, 1F, F-2"); *m*/*z* (EI) 343 (M⁺, 23), 215 (36), 169 (100), 119 (51), 105 (85%).

4.1.5.5. *N*-(**4**-Methoxyphenyl)-1-(2-nitrophenyl)-1*H*pyrrole-2-carboxamide 7e. 390 mg, 77%, as a pale-yellow solid (ethyl acetate/hexane); mp 107–109 °C;. ν_{max} (Nujol) 3315, 1630, 1510, 1345 cm⁻¹; $\delta_{\rm H}$ (400 MHz; CDCl₃) 3.75 (s, 3H, OMe), 6.37–6.39 (m, 1H, H-4), 6.79 (d, *J*=7.3 Hz, 2H, H-3", H-5"), 6.83–6.87 (m, 2H, H-3, H-5), 7.30 (d, *J*= 7.3 Hz, 2H, H-2", H-6"), 7.46 (d, *J*=7.8 Hz, 1H, H-6'), 7.52–7.56 (m, 2H, H-4', NH), 7.66 (t, *J*=7.8 Hz, 1H, H-5'), 8.05 (d, *J*=8.1 Hz, 1H, H-3'); $\delta_{\rm C}$ (100 MHz; CDCl₃) 55.35, 109.53, 113.85, 114.42, 122.00, 124.71, 127.07, 128.71, 129.13, 129.95, 132.07, 134.13, 134.77, 145.99, 155.51, 158.51; *m*/z (EI) 337 (M⁺, 85), 291 (14), 215 (40), 169 (100), 149 (30), 134 (15); HRMS (EI): (M⁺), found 337.1060, C₁₈H₁₅N₃O₄ requires 337.1063.

4.1.6. Preparation of pyrrolo[1,2-*a*]quinoxalin-4(5*H*)ones 8a,b: general procedure D. To a stirred solution of carboxamide 3a or c (0.65 mmol) in dry DMF (10 mL) under argon, was added NaH (60% in oil, 1.10 mmol) and left to stir at room temperature for 1.5 h. The reaction mixture was poured into water (50 mL), neutralised with 2 N HCl and extracted with ethyl acetate (3×10 mL). The combined organic phases were dried (Na₂SO₄) and evaporated in vacuo. The oily residue was triturated with hexane, the solid collected and purified by flash chromatography (25% ethyl acetate/hexane) to afford 5-ethylpyrrolo[1,2-*a*]quinoxalin-4(5*H*)-one 8a or 5-cyclo-hexylpyrrolo[1,2-*a*]quinoxalin-4(5*H*)-one 8b.

4.1.6.1. 5-Ethylpyrrolo[1,2-*a*]quinoxalin-4(5*H*)-one **8a.** 119 mg, 85%, as a pale-yellow solid (ethyl acetate/ hexane); mp 78–80 °C. [Found: C, 73.45; H, 5.63; N, 13.14;. C₁₃H₁₂N₂O requires C, 73.56; H, 5.70; N, 13.20%]; ν_{max} (Nujol) 3118, 1638 cm⁻¹; $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.29 (t, *J*=7.0 Hz, 3H, CH₃), 4.25, (q, *J*=7.2 Hz, 2H, CH₂), 6.58 (dd, J=3.6, 2.8 Hz, 1H, H-2), 7.12–7.19 (m, 2H, H-3, H-8), 7.24–7.27 (m, 2H, H-6, H-7), 7.57 (dd, J=2.8, 1.6 Hz, 1H, H-1), 7.61 (d, J=7.6 Hz, 1H, H-9); $\delta_{\rm C}$ (100 MHz; CDCl₃) 13.19, 36.58, 112.85, 113.61, 115.19, 115.96, 116.21, 123.07, 123.75, 124.63, 125.98, 129.56, 155.59; m/z (EI) 212 (M⁺, 79), 197 (39), 184 (100) 167 (25), 155 (17%).

4.1.6.2. 5-Cyclohexylpyrrolo[**1**,**2**-*a*]**quinoxalin-4**(*5H*)**one 8b.** 149 mg, 86%, as a pale-yellow solid (ethyl acetate/ hexane); mp 109–111 °C. [Found: C, 76.38; H, 6.88; N, 10.46;. C₁₇H₁₈N₂O requires C, 76.66; H, 6.81; N, 10.52%]; ν_{max} (Nujol) 3115, 1640, 1609 cm⁻¹; δ_{H} (400 MHz; CDCl₃) 1.25–2.63 [m, 10H, (CH₂)₅], 4.75 (br, s, 1H, CH), 6.63–6.64 (m, 1H, H-2), 7.22–7.16 (m, 2H, H-3, H-8), 7.28 (t, *J*= 7.3 Hz, 1H, H-7), 7.60–7.56 (m, 2H, H-1, H-6), 7.66 (d, *J*= 7.8 Hz, 1H, H-9); δ_{C} (100 MHz; CDCl₃) 25.32, 26.51, 29.15, 55.98, 112.08, 113.03, 114.84, 115.30, 116.43, 122.43, 123.79, 124.31, 124.82, 129.76, 155.97; *m*/*z* (EI) 266 (M⁺, 18), 184 (100), 155 (12), 129 (8%).

4.1.7. Preparation of pyrrolo[1,2-*a*]quinoxalin-4(5*H*)ones 8c,d,f,g: general procedure E. To a stirred solution of the corresponding carboxamide 7a-c,e (0.65 mmol) in dry DMF (10 mL) under argon, was added NaH (60% in oil, 1.10 mmol) and left to stir at room temperature for 4.5 h. The reaction mixture was poured into water (50 mL), neutralised with 2 N HCl and the precipitate filtered, washed with water and dried to afford 5-phenylpyrrolo[1,2-*a*]quinoxalin-4(5*H*)-one 8c, 2-(4-oxo-pyrrolo[1,2-*a*]quinoxalin-5(4*H*)-yl)benzoic acid 8d.

4.1.7.1. 5-Phenylpyrrolo[**1**,2-*a*]**quinoxalin-4**(*5H*)-**one 8c.** 178 mg, 89%, as a pale-yellow solid (ethyl acetate/ hexane); mp 168–170 °C. [Found: C, 78.10; H, 4.68; N, 10.74;. $C_{17}H_{12}N_2O$ requires C, 78.44; H, 4.65; N, 10.76%]; ν_{max} (Nujol) 3110, 1652 cm⁻¹; $\delta_{\rm H}$ (400 MHz; CDCl₃) 6.50 (dd, J=8.4, 1.1 Hz, 1H, H-6), 6.75 (dd, J=3.9, 2.6 Hz, 1H, H-2), 7.11 (dd, J=3.9, 1.4 Hz, 1H, H-3), 7.18 (td, J=7.8, 1.1 Hz, 1H, H-8), 7.26 (td, J=7.7, 1.2 Hz, 1H, H-7), 7.40 (d, J=7.2 Hz, 2H, H-2', H-6'), 7.56 (t, J=7.3 Hz, 1H, H-4'), 7.63 (t, J=7.4 Hz, 2H, H-3', H-5'), 8.16 (dd, J=8.1, 1.2 Hz, 1H, H-9), 8.29 (dd, J=2.6, 1.4 Hz, 1H, H-1); $\delta_{\rm C}$ (100 MHz; CDCl₃) 112.40, 113.16, 115.26, 116.73, 118.22, 122.68, 122.99, 123.03, 125.46, 128.76, 129.57, 129.93, 130.98, 136.70, 154.32; *m*/z (EI) 260 (M⁺, 100), 259 (100), 231 (6) 205 (7), 178 (6), 166 (7), 115 (8), 102 (8%).

4.1.7.2. 2-(4-Oxopyrrolo[1,2-*a***]quinoxalin-5(4***H***)-yl)benzoic acid 8d.** 162 mg, 82%, as a pale-yellow solid (2 N NaOH/2 N HCl); mp 260–262 °C. [Found: C, 70.73; H, 4.03; N, 9.17; C₁₈H₁₂N₂O₃ requires C, 71.05; H, 3.97; N, 9.21%]; ν_{max} (Nujol) 2900, 3135, 1709, 1651 cm⁻¹; δ_{H} (250 MHz; DMSO-*d*₆) 6.43 (d, *J*=8.1 Hz, 1H, H-6), 6.73– 6.76 (m, 1H, H-2), 7.11 (m, 1H, H-3), 7.17 (t, *J*=7.5 Hz, 1H, H-8), 7.25 (t, *J*=7.5 Hz, 1H, H-7), 7.49 (d, *J*=7.7 Hz, 1H, H-6¹), 7.69 (t, *J*=7.6 Hz, 1H, H-4¹), 7.86 (t, *J*=7.6 Hz, 1H, H-5¹), 8.17 (d, *J*=7.7 Hz, 2H, H-9, H-3¹), 8.29 (m, 1H, H-1), 12.80 (bs, 1H, CO₂H); δ_{C} (63 MHz; DMSO-*d*₆) 112.63, 113.32, 115.45, 116.63, 118.42, 123.07 (2C), 123.34, 125.76, 129.57, 130.56, 131.37, 131.68, 132.13, 134.13, 136.74, 154.70, 165.79; *m*/*z* (EI) 304 (M⁺, 60), 259 (100%).

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4.1.7.3. Methyl 2-(4-oxopyrrolo[1,2-*a*]quinoxalin-5(4*H*)-yl)benzoate 8e. 159 mg, 77%, as a pale-yellow solid (ethyl acetate /hexane); mp 159–161 °C. [Found: C, 71.40; H, 4.48; N, 8.77; C₁₉H₁₄N₂O₃ requires C, 71.69; H, 4.43; N, 8.80%]; v_{max} (Nujol) 3148, 1724, 1660 cm⁻¹; δ_{H} (250 MHz; DMSO- d_{6}) 3.55 (s, 1H, Me), 6.41 (d, J=8.1 Hz, 1H, H-6), 6.74–6.77 (m, 1H, H-2), 7.09–7.11 (m, 1H, H-3), 7.16 (t, J=7.8 Hz, 1H, H-8), 7.26 (t, J=7.7 Hz, 1H, H-7), 7.54 (d, J=7.7 Hz, 1H, H-6'), 7.72 (t, J=7.6 Hz, 1H, H-4'), 7.88 (t, J=7.6 Hz, 1H, H-5'), 8.18 (d, J=7.7 Hz, 2H, H-9, H-3'), 8.30–8.31 (m, 1H, H-1); δ_{C} (63 MHz; DMSO- d_{6}) 52.26, 112.68, 113.12, 113.27, 115.45, 116.45, 118.52, 122.92, 123.13, 123.31, 125.73, 128.94, 129.68, 131.16, 131.84, 134.59, 136.92, 154.61, 164.37; *m*/*z* (EI) 318 (M⁺, 56), 259 (100%).

4.1.7.4. 5-(4-Chlorophenyl)pyrrolo[**1**,2-*a*]**quinoxalin-4-(5***H***)-one 8f.** 182 mg, 82%, as a pale-yellow solid (ethyl acetate/hexane); mp 236–238 °C. [Found: C, 68.93; H, 3.82; N, 9.47; C₁₇H₁₁ClN₂O requires C, 69.28; H, 3.76; N, 9.50%]; ν_{max} (Nujol) 3088, 1660 cm⁻¹; $\delta_{\rm H}$ (400 MHz; DMSO- d_6) 6.55 (dd, J=8.3, 1.1 Hz, 1H, H-6), 6.77 (dd, J= 3.8, 2.8 Hz, 1H, H-2), 7.13 (dd, J=3.8, 1.4 Hz, 1H, H-3), 7.21 (td, J=7.8, 1.1 Hz, 1H, H-8), 7.29 (td, J=7.7, 1.2 Hz, 1H, H-7), 7.48 (d, J=8.6 Hz, 2H, H-2', H-6'), 7.70 (d, J= 8.6 Hz, 2H, H-3', H-5'), 8.19 (dd, J=8.1, 1.2 Hz, 1H, H-9), 8.32 (dd, J=2.8, 1.4 Hz, 1H, H-1); $\delta_{\rm C}$ (100 MHz; DMSO- d_6) 113.46, 114.11, 116.21, 117.62, 119.27, 123.45, 124.03, 126.47, 130.92, 131.66, 132.54, 134.32, 136.51, 155.20; *m/z* (EI) 294 (M⁺, 100), 258 (18), 230 (11) 204 (6), 167 (7), 130 (12), 115 (9).

4.1.7.5. 5-(4-Methoxyphenyl)pyrrolo[**1,2**-*a*]**quinoxalin-4-(5***H***)-one 8g.** 208 mg, 95%, as a pale-yellow solid (ethyl acetate/hexane); mp 207–209 °C. [Found: C, 74.16; H, 4.90; N, 9.58; $C_{18}H_{14}N_2O_2$ requires C, 74.47; H, 4.86; N, 9.65%]; ν_{max} (Nujol) 3148, 1724, 1660 cm⁻¹; δ_{H} (400 MHz; DMSO-*d*₆) 3.85 (s, 3H, Me), 6.55 (d, *J*= 8.1 Hz, 1H, H-6), 6.74 (dd, *J*=3.8 Hz, 2.7 Hz, 1H, H-2), 7.09 (dd, *J*=3.8, 1.5 Hz, 1H, H-3), 7.20–7.14 (m, 3H, H-8, H-2', H-6'), 7.25 (td, *J*=7.5, 1.1 Hz, 1H, H-7), 7.29 (d, *J*= 8.8 Hz, 2H, H-3', H-5'), 8.12 (d, *J*=8.1 Hz, 1H, H-9), 8.27 (dd, *J*=2.7, 1.5 Hz, 1H, H-1); δ_{C} (100 MHz; DMSO-*d*₆) 55.60, 112.55, 113.35, 115.33, 115.42, 117.09, 118.36, 123.01, 123.15, 123.27, 125.67, 128.25, 130.80, 131.52, 154.78, 159.39; *m*/z (EI) 290 (M⁺, 100), 275 (11), 167 (18), 145 (8%).

4.1.8. Reaction of *N***-ethyl-1-(2-nitrophenyl)-1***H***-pyrrole-2-carboxamide 3a with NaH in DMF.** To a stirred solution of carboxamide **3a** (170 mg, 0.65 mmol) in dry DMF (10 mL) under argon, was added NaH (60% in oil, 44 mg, 1.10 mmol) and left to stir at room temperature for 30 min. The reaction mixture was added to a cold saturated aqueous sodium bicarbonate solution (50 mL), neutralised with 2 N HCl and extracted with ethyl acetate (3×10 mL). The combined organic phases were dried (Na₂SO₄) and evaporated in vacuo. The oily residue was triturated with hexane, the solid collected and purified by flash chromatography (25, 50% ethyl acetate/hexane) to afford *N*-ethyl-1-(2-nitrophenyl)-1*H*-pyrrole-2-carboxamide **3a**, 5-ethyl-pyrrolo[1,2-*a*]quinoxalin-4(5*H*)-one **8a** and *N*-ethyl-*N*-(2-nitrophenyl)-1*H*-pyrrole-2-carboxamide **9**.

4.1.8.1. *N*-Ethyl-*N*-(2-nitrophenyl)-1*H*-pyrrole-2-carboxamide 3a. 29 mg, 17%, as a pale-yellow solid (ethyl acetate/hexane), mp 93–95 °C, identical in all respects to an authentic sample (vide infra).

4.1.8.2. 5-Ethylpyrrolo[1,2-*a*]**quinoxalin-4**(5*H*)-**one 8a.** 51 mg, 37%, as a pale-yellow solid (ethyl acetate/ hexane), mp 78–80 °C, identical in all respects to an authentic sample (vide infra).

4.1.8.3. *N*-Ethyl-*N*-(2-nitrophenyl)-1*H*-pyrrole-2-carboxamide **9.** 36 mg, 21%, as a pale-yellow solid (ethyl acetate/hexane); mp 165.5–167.5 °C. [Found: C, 60.30; H, 5.26; N, 16.25; C₁₃H₁₃N₃O₃ requires C, 60.22; H, 5.05; N, 16.21%]; ν_{max} (Nujol) 3268, 1615 cm⁻¹; $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.29 (t, *J*=7.2 Hz, 3H, Me), 3.60 (br, s, 1H, CH₂), 4.27–4.32 (m, 1H, CH₂) 4.94 (br, s, 1H, H-4), 5.96 (s, 1H, H-3), 6.87 (s, 1H, H-5), 7.50 (dd, *J*=7.8, 1.4 Hz, 1H, H-6'), 7.66 (td, *J*=7.6, 1.3 Hz, 1H, H-4'), 7.77 (td, *J*=7.7, 1.5 Hz, 1H, H-5'), 8.12 (dd, *J*=8.1, 1.5 Hz, 1H, H-3'), 9.69 (br, s, 1H, NH); $\delta_{\rm C}$ (100 MHz; CDCl₃) 13.47, 46.00, 110.35, 112.99, 121.87, 125.05, 126.35, 129.83, 132.93, 134.55, 136.78, 147.67, 161.18; *m/z* (EI) 259 (M⁺, 27), 213 (65), 184 (6) 166 (54), 151 (25), 131 (7), 106 (42), 94 (100), 66 (26%).

4.1.9. Preparation of 5-ethyl-7-methylpyrrolo[1,2-a]quinoxalin-4(5H)-one 12. To a stirred solution of 7-methylpyrrolo[1,2-a]quinoxalin-4(5H)-one¹ (300 mg, 1.51 mmol) in dry DMF (10 mL) under argon, was added NaH (60% in oil, 65 mg, 1.61 mmol) and left to stir at room temperature for 1 h. Ethyl iodide (242 mg, 1.55 mmol) was added and the reaction mixture stirred for 15 min. Water (50 mL) was added and extracted with ethyl acetate $(3 \times 20 \text{ mL})$. The combined organic extracts were dried (Na₂SO₄) and evaporated in vacuo. The oily residue was purified by flash chromatography (25% ethyl acetate/ hexane) to afford 5-ethyl-7-methylpyrrolo[1,2-a]quinoxalin-4(5H)-one 12 (270 mg, 79%) as a colourless solid (propan-2-ol/hexane); mp 104-105 °C. [Found: C, 73.98; H, 6.29; N, 12.36; C₁₄H₁₄N₂O requires C, 74.31; H, 6.24; N, 12.38%]; ν_{max} (Nujol) 3130, 1651 cm⁻¹; δ_{H} (400 MHz; $CDCl_3$) 1.34 (t, J=7.2 Hz, 3H, CH_2CH_3), 2.42 (s, 3H, Me), 4.27 (g, J=7.2 Hz, 2H, CH₂CH₃), 6.60 (dd, J=3.9, 2.8 Hz, 1H, H-2), 6.97 (dd, J = 8.2, 0.8 Hz, 1H, H-8), 7.06 (s, 1H, H-6), 7.17 (dd, J = 3.9, 1.6 Hz, 1H, H-3), 7.50 (d, J = 8.2 Hz, 1H, H-9), 7.55 (dd, J = 2.8, 1.6 Hz, 1H, H-1); $\delta_{\rm C}$ (100 MHz; CDCl₃) 12.65, 35.94, 68.54, 111.94, 112.75, 114.39, 115.50, 115.66, 121.85, 122.94, 123.27, 128.76, 135.38, 155.15; *m/z* (EI) 226 (M⁺, 100), 211 (43), 198 (96), 181 (25), 169 (14%).

4.1.10. Reaction of *N***-ethyl-1-(4-methyl-2-nitrophenyl)-**1*H***-pyrrole-2-carboxamide 3b with NaH in DMF.** Following general procedure D carboxamide **3b** (0.65 mmol) in dry DMF (10 mL) and NaH (60% in oil, 1.10 mmol) at room temperature for 1.5 h gave a mixture (118 mg, 80%) of 5-ethyl-7-methylpyrrolo[1,2-*a*]quinoxalin-4(5*H*)-one **12** (44% in mixture from ¹H NMR) and 5-ethyl-8-methylpyrrolo[1,2-*a*]quinoxalin-4(5*H*)-one **13** (44% in mixture from ¹H NMR); $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.29–1.34 (m, 4H, 2×CH₂CH₃ of **12** and **13**), 2.41 (s, 3H, Me of **12**), 2.42 (s, 3H, Me of **13**), 4.24–4.30 (m, 4H, 2×CH₂CH₃ of **12** and **13**), 6.59–6.62 (m, 2H, 2×H-2 of **12** and **13**), 6.97 (d, J=8.2 Hz, 1H, H-8 of **12**), 7.06 (s, 1H, H-6 of **12**), 7.07 (d, J=7.8 Hz, H-7 of **13**), 7.14–7.18 (m, 3H, H-3 of **12** and **13**, H-6 of **13**), 7.45 (s, 1H, H-9 of **13**), 7.50 (d, J=8.2 Hz, 1H, H-9 of **12**), 7.55 (dd, J=2.8, 1.6 Hz, 1H, H-1 of **12**).

4.1.11. Reaction of N-(2,4-difluorophenyl)-1-(2-nitrophenyl)-1*H*-pyrrole-2-carboxamide 7d with NaH in DMF. Following general procedure E carboxamide 7d (0.65 mmol) in dry DMF (10 mL) and NaH (60% in oil, 1.10 mmol) at room temperature for 4.5 h gave, after purification by flash chromatography (33% ethyl acetate/hexane), 5-(2,4-difluorophenyl)pyrrolo[1,2-*a*]quinoxalin-4(5*H*)-one 16 and 8-fluoro-5-(2-nitrophenyl)pyrrolo[1,2-*a*] quinoxalin-4(5*H*)-one 17.

4.1.11.1. 5-(2,4-Difluorophenyl)pyrrolo[1,2-a]quinoxalin-4(5H)-one 16. 144 mg, 75%, as a pale-yellow solid (ethyl acetate/hexane); mp 200–202 °C. [Found: C, 69.07; H, 3.65; N, 9.35; C₁₇H₁₀F₂N₂O requires C, 68.92; H, 3.40; N, 9.46%]; ν_{max} (Nujol) 3122, 1659 cm⁻¹; $\delta_{\rm H}$ (400 MHz; DMSO- d_6) 6.63 (d, J = 8.2 Hz, 1H, H-6), 6.77 (dd, J = 3.9, 2.8 Hz, 1H, H-2), 7.17 (dd, J=3.9, 1.5 Hz, 1H, H-3), 7.23 (td, J=7.8, 1.3 Hz, 1H, H-7), 7.31 (td, J=7.8, 1.3 Hz, 1H, H-8), 7.36 (ddd, J = 8.8, 8.7, 2.8, Hz, 1H, H-5'), 7.61 (ddd, J=10.1, 9.2, 2.8 Hz, 1H, H-3'), 7.67 (td, J=8.8, 6.2 Hz, 1H, H-6') 8.19 (dd, J = 8.1, 1.3 Hz, 1H, H-9), 8.32 (dd, J =2.8, 1.5 Hz, 1H, H-1); $\delta_{\rm C}$ (100 MHz; DMSO- d_6) 105.94 (dd, J=27.1, 24.2 Hz), 113.37 (dd, J=22.5, 3.5 Hz), 113.56, 113.78, 115.93, 116.52, 119.27, 120.62 (dd, J=13.5, 3.9 Hz), 122.49, 123.55, 124.01, 126.32, 130.34, 133.51 (d, J = 10.4 Hz), 154.38, 158.90 (dd, J = 251.1, 13.3 Hz), 162.80 (dd, J = 248.6, 11.8 Hz); $\delta_{\rm F}$ (376 MHz; DMSO- d_6) -107.70 (dddd, J=9.2, 8.7, 8.6, 6.2 Hz, 1F, F-4'), -117.05 (ddd, J=10.1, 8.8, 8.6 Hz, 1F, F-2'); m/z (EI) 296 (M⁺, 100), 277 (57), 247 (6%).

4.1.11.2. 8-Fluoro-5-(2-nitrophenyl)pyrrolo[1,2*a*]quinoxalin-4(5*H*)-one 17. 32 mg, 15%, as a pale-yellow solid (ethyl acetate/hexane); mp 264–265 °C. [Found: C, 63.32; H, 3.33; N, 12.86; C₁₇H₁₀FN₃O₃ requires C, 63.16; H, 3.12; N, 13.00%]; ν_{max} (Nujol) 1661, 1514, 1346 cm⁻¹; $\delta_{\rm H}$ (400 MHz; DMSO-*d*₆) 6.64 (dd, J=9.0, 5.1 Hz, 1H, H-6), 6.81 (t, J = 3.6 Hz, 1H, H-2), 7.10 (td, J = 9.0, 2.7 Hz, 1H, H-7), 7.16 (dd, J=3.8, 1.2 Hz, 1H, H-3), 7.77 (d, J=7.8 Hz, 1H, H-3'), 7.87 (td, J = 7.8, 1.3 Hz, 1H, H-5'), 8.01 (td, J=7.7, 1.3 Hz, 1H, H-4'), 8.26 (dd, J=9.8, 2.7 Hz, 1H,H-9), 8.32–8.36 (m, 2H, H-1, H-6'); $\delta_{\rm C}$ (100 MHz; DMSO d_6) 103.31 (d, J = 28.4 Hz), 112.84 (d, J = 22.8 Hz), 113.91, 114.02, 118.35 (d, J=9.6 Hz), 119.88, 122.25, 124.25 (d, J=11.4 Hz), 126.16, 126.95, 129.86, 131.13, 132.65, 135.80, 147.26, 153.85, 158.36 (d, J=240.3 Hz); $\delta_{\rm F}$ $(376 \text{ MHz}; \text{DMSO-}d_6) - 118.25 \text{ (ddd, } J = 9.8, 9.0, 5.1 \text{ Hz},$ 1F, F-8); *m*/*z* (EI) 323 (M⁺, 39), 277 (100), 247 (15), 222 (8%).

Acknowledgements

This research was funded by the programme "Heraklitos" of the Operational Programme for Education and Initial Vocational Training of the Hellenic Ministry of Education under the 3rd Community Support Framework and the European Social Fund (to G. R.) and partly by a PENED grant (91ED102) (to A. K.) from the General Secretariat of Research and Technology, Athens. We thank Professor Dr. B. Stanovnik of the University of Ljubljana and Professor N. Hadjiliadis of the University of Ioannina for elemental analyses. We are particularly grateful to A. Cakebread and R. Tye for mass spectra, and, J. Cobb for NMR spectra of fluorine containing compounds, obtained on machines funded by the University of London Intercollegiate Research Services Scheme.

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Tetrahedron

Tetrahedron 60 (2004) 10833-10841

Cycloaddition approach to benzo-annulated indane-based α-amino acid derivatives

Sambasivarao Kotha* and Arun Kumar Ghosh

Department of Chemistry, Indian Institute of Technology-Bombay, Powai, Mumbai 400076, India

Received 13 April 2004; revised 25 August 2004; accepted 16 September 2004

Available online 7 October 2004

Abstract—Synthesis of various benzo-annulated indane-based α -amino acid (AAA) derivatives are reported via a [4+2] and [2+2+2] cycloaddition reactions as key steps. Here, *o*-xylylene based AAA moiety has been used as a reactive intermediate and by adopting this strategy various indane-based constrained AAA derivatives are prepared. © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

During the past several years, research in our laboratory has focused on the synthesis of new building blocks which are suitable for the design of various constrained α -amino acid (AAA) derivatives.¹ These investigations are directed towards the preparation of α, α -dialkylated amino acid derivatives that are not accessible by conventional modes such as Bucherer-Berg (BB) method. Our efforts have led to the development of several building blocks, which are useful for the generation of various unusual AAA derivatives. Within this context, we proposed o-xylylene (or *o*-quinodimethane) intermediate 1 containing an AAA moiety as a versatile reactive intermediate for the generation of various benzo-annulated indane-based AAA derivatives (e.g., 5) via a Diels-Alder (DA) strategy. It is hoped that introduction of new type of building blocks (e.g., 1) as reaction partners in DA chemistry would significantly extend the impact of these methodologies in peptide-based drug design. Indane-based AAA derivatives such as 4, are a special class of constrained phenylalanine (Phe) derivative **3**, used in several instances to modify various biologically active peptides.²⁻⁵ Also, indane-based AAA are useful building blocks for the design of 'ladder-like' parallel tapes in crystal engineering studies.⁶ Indanylglycine (Ind) **4** in which phenyl ring is covalently connected to the α -carbon through a C-methylene group and thereby providing the rigid aromatic ring lacking rotational freedom of the Pheside chain. Compound 4, a component of a potent ligand of

angiotensin-II receptor with agonistic or antagonistic activity was extensively used in the synthesis of various biologically active peptides.³ In this regard, Chasing and coworkers synthesized several 2-indanyl-glycine derivatives for analyzing the binding pockets of Phe⁽⁷⁾ (S7) and Phe⁽⁸⁾ (S8), two important aromatic residues for pharmacological properties of substance P (SP).³ Cyclic analogues of phenylglycine, Ind, α -methyl-40-phosphonophenylglycine (MPPG) have been synthesized and exhibited interesting pharmacological properties towards glutamate receptors.⁷

Topographical consideration is the major approach to the rational design of peptide ligands to explore the side-chain interaction for binding to their receptors and for signal transduction. This aspect has been realized by incorporation of constrained AAA such as **5** (R=H) into the backbone of polypeptide and also in non-peptide templates.⁸ When the C-terminal Phe residue in chemotactic agent HCO-Met-Leu-Phe-OH (FMLP) was replaced by Ind, the resulting modified analog has shown to be highly active in the superoxide production (Fig. 1).^{4–5}





Keywords: Amino acids and derivatives; Fullerenes; Sulfur heterocycles; Cycloadditions; Diels–Alder reactions.

^{*} Corresponding author. Tel.: +91 22 25767160; fax: +91 22 2572 3480; e-mail: srk@chem.iitb.ac.in

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In addition, the indane moiety is an important structural element, which can fix the relative position of functional groups to impart a greater specificity for biological activity. Also, a number of indane-based derivatives have been synthesized and were shown to be pharmacologically active compounds.^{9–16}

Several approaches towards the parent o-xylylene^{17–18} or oquinodimethane (o-QDM) **2** intermediate have been explored and these are summarized in Scheme 1. Thermal generation of **2** by various roots was realized at higher temperature except in case of sultine **7**. In 1991, Dittmer and co-workers had reported the syntheses of the sultine **7** from o-xylenedibromide **9** in high yield using PTC, tetrabutylammonium bromide (TBAB) conditions.¹⁹ In this respect, sultine **7** was found to be a suitable precursor to generate the parent o-xylylene **2** at around 80 °C.



Scheme 1.

In a related study, Martin and co-workers explored several routes to generate the *o*-xylylene intermediate and they have reported various C_{60} fullerene adducts using sultine as a latent diene component (Scheme 2).²⁰ Later on, Chung and co-worker have reported various heterocyclic sultine derivatives capable of participating in DA chemistry.²¹

Realization of the proposition shown in Scheme 3 depends on finding suitable conditions to generate the key precursor related to 1. Although, several methods are available for the generation of the parent o-xylylene 2, these methods may not be applicable for the intermediates related to 1 due to the presence of two reactive functional groups, i.e., amino and carboxyl. Our initial experience to prepare o-xylylene



intermediates containing AAA moiety from benzocyclobutene intermediates was not favorable.²² In the past, we had demonstrated that sultine intermediate can be trapped with a dienophile containing AAA moiety.²³ Therefore, the attention was focussed towards the generation of **1** via sultine intermediate. Compounds such as **5** may be prepared by BB²⁴ method starting from benzofused 2-indanone derivative, that involves drastic conditions for the hydrolysis of hydantoin intermediate. Due to this reason many sensitive substrates do not survive and only simple derivatives can be prepared by BB method.

2. Strategy

In a program directed towards the synthesis of highly functionalized benzo-annulated indane-based (AAA) derivatives such as **5**, we identified [4+2]-cycloaddition (DA) strategy as a possible route (Scheme 3). Herein, we report the realization of this strategy to prepare various benzo-annulated indane-based AAA derivatives by trapping the reactive intermediate **1** with different dienophiles.

Approaches based on the DA strategy have been well established in the literature where the sultine derivatives were used as a latent diene equivalents.^{20–21} However, there are no example known where the sultine derivative containing AAA moiety, and, in this regard we attempted to prepare various indane-based AAA derivatives by utilizing sultine methodology. To expand upon 'Building Block Approach' for highly functionalized benzo-annulated indane-based AAA derivatives, generation of **1** via sultine derivative **20** is an attractive proposition.

3. Results and discussion

Towards the synthesis of compounds such as **5**, the required indane derivative **17** was synthesized according to the reported procedure via a four-step sequence starting from ethyl isocyanoacetate **15** (Scheme 4).²⁵ The dihydroxy indane derivative **17** was then converted to the corresponding dibromide **18** under PBr₃ conditions (Scheme 5).²⁶ The IR spectrum of compound **18** showed characteristic strong





Scheme 4. (i) Propargyl bromide, K₂CO₃, TBAHS, CH₃CN, 80 °C; (ii) HCl, EtOH, rt; (iii) Ac₂O, DCM, rt; (iv) Rh(PPh₃)₃Cl, 2-butyne-1,4-diol, EtOH, 80 °C, 65%.



Scheme 5. (i) PBr₃, C_6H_6 , rt, 81%; (ii) DMAD, Zn, ultrasound or NaI, DMF, or Bu₄NI or KI, 18-crown-6.

absorption bands at 3225, 1737 and 1654 cm^{-1} , indicating the presence of amino, ester and carbonyl functional groups, respectively. The M^+ peak at m/z 433 in its mass spectrum supported its structural formulation. The ¹H NMR spectrum of 18, showed the absence of two hydroxyl protons. The two singlets at δ 4.65 and 6.03 are due to the methylene protons neighbouring the bromine atom and amide proton, respectively. It also exhibited a clear doublet at δ 3.25 (J = 16.8 Hz) and another doublet at δ 3.61 (J=16.8 Hz) due to diastereotopic protons (H_a and H_b) present in the indane ring and a singlet at δ 7.21 accounts for aromatic protons. The 11-line ¹³C NMR spectrum resonating indicated the symmetry present in the molecule 18. Towards our objective, initially, various attempts to generate o-xylylene intermediate 19 from the corresponding dibromo derivative **18** were unsuccessful (Scheme 5).²⁷

Next, the dibromo compound 18 was reacted with sodium hydroxy methanesulfinate (rongalite)²⁸ in presence of tetrabutylammonium bromide (TBAB) in DMF at 0 °C, the two isomeric sultine-based AAA derivatives 20 and 20a were obtained in 72% combined yield (1:1) (Scheme 6). IR spectrum of the first isomer 20 (mp: 180-181 °C) showed absorption bands at 1453, 1169 cm^{-1} due to the presence of S=O and S-O functionalities suggesting the conversion of dibromide 18 into sultine derivative 20. The absorption bands at 1668 and 1732 cm^{-1} are due to amide and ester carbonyl groups, respectively. The ¹H NMR spectrum of **20** showed doublets at δ 3.53 (*J*=15.4 Hz), δ 4.4 (*J*=15.3 Hz), δ 4.92 (J=13.5 Hz) and δ 5.27 (J=13.2 Hz) due to the presence of four diastereotopic protons in oxathiin-3-oxide (sultine) moiety. It further displayed an ABq at δ 3.30 (d, J=16.8 Hz) and δ 3.62 (d, J=16.8 Hz), due to diastereotopic -CH₂ protons (H_a and H_b) which are present in the indane ring system and singlets at δ 7.12 and 7.14 for the aromatic protons. The structure of compound 20 was further confirmed by its characteristic 16-line ¹³C NMR spectrum and also supported the unsymmetrical nature of 20. The 13 C NMR signals are assigned by reference to related literature



20 and **20a Scheme 6.** (i) Rongalite, TBAB, DMF, 0 °C, 72%.

examples.²⁹ The ¹³C NMR spectrum of **20** showed two signals at δ 57.0 (*C*H₂S=O) and 63.9 (*C*H₂O-S) due to the presence of two characteristic carbons of oxathiin-3-oxide (sultine) moiety. The second isomer **20a** (mp: 198–199 °C) also displayed similar spectral parameters. Since these two isomers have very similar IR, ¹H NMR and ¹³C spectra, it was not possible to assign the exact stereochemistry with the available data. Isomers **20** and **20a** are presumably diastereoisomers (each as a racemic mixture).

Having the sultines 20 and 20a in hand, their DA chemistry with various dienophiles, was then explored. Subsequent oxidation of the DA adducts with DDQ³⁰ gave benzoannulated derivatives (21-25). In a separate experiment both sultine derivatives 20 and 20a were treated with dimethyl acetylenedicarboxylate (DMAD) in xylene at 120 °C to give the DA adduct which was contaminated with the aromatized product (Scheme 7). Therefore, isolation of the DA products was not attempted and they were treated with DDQ to give aromatized products 21-25. In the ¹H NMR spectrum the presence of singlet at δ 7.67 and 8.11 are due to the presence of two different types of aromatic protons. The disappearance of four peaks at δ 3.53 (J=15.4 Hz), 4.4 (J=15.3 Hz), 4.92 (J=13.5 Hz) and 5.27(J=13.2 Hz) indicated the absence of four diastereotopic protons in oxathiin-3-oxide (sultine) moiety and a strong absorption band at 1747 (C=O) cm^{-1} in the IR spectrum confirm the structure of **21**. The 13-line ¹³C NMR spectrum resonating indicated the symmetry present in the molecule 21.



Scheme 7. (i) DMAD, toluene, 120 °C; (ii) DDQ, C₆H₆, 80 °C, 78%.

We were pleased to discover that the sultine **20** (or **20a**) on treatment with various dienophiles delivered highly functionalized indane derivatives, which on subsequent oxidation gave aromatized products. These products are characterized with the complementary spectral data (¹H and ¹³C) and the results are summarized in Table 1.

During the DA sequence, we found that both the sultine derivatives **20** and **20a** gave the same sulphone **26** in refluxing toluene in the absence of a dienophile (Scheme 8).



Scheme 8. (i) Toluene, 120 °C, 88%.





^a Yields refer to combined isolated yields for both the DA reaction and DDQ oxidation sequence.

In the ¹H NMR spectrum of **26**, the methylene proton attached to the sulphone group appeared as a singlet at δ 4.32 and the M⁺ peak at m/z 337 in its mass spectrum revealed its structural formulation. The 11-line ¹³C NMR spectrum resonating indicated the symmetry present in the molecule **26**. In view of several functionalization reactions of sulfones,³¹ availability of intermediate **26** can further expand the scope of the 'Building Block Approach'.¹

A possible mechanism based on the earlier observation³² for the formation of sulphone **26** from sultine derivatives **20** and **20a** under thermal condition is shown in Scheme 9. Under thermal conditions, retro Diels–Alder occurs to generate SO_2 and *o*-xylylene intermediate. Later on the in situ generated intermediate **19** react with SO_2 by a [4+1] cheletropic addition to form more stable sulphone **26**. Here, the formation of **19** is also supported by the formation of various DA products (**21–25**), which are shown in Table 1.



Scheme 9. Possible mechanism for the sulphone derivative 26.

Next, the sultine **20** (or **20a**) was reacted with a dienophile such as methyl 2-acetamido acrylate at 120 °C in toluene to give the highly constrained bis AAA derivative **27**, in 33% isolated yield (Scheme 10).



Scheme 10. (i) Methyl 2-acetamidoacrylate, 120 °C, 33%.

Along similar lines, the sultine **20** (or **20a**) was reacted with *N*-phenyl maleimide to give a pair of DA adducts **28** and **29** in 96% combined yield (Scheme 11). Both these compounds **28** (mp 162–164 °C) and **29** (mp 142–144 °C) gave almost identical ¹H, ¹³C NMR and mass spectral data. Based on this data we concluded that they are diastereoisomeric compounds and stereochemistry of the individual isomers could not be determined.

In view of various applications of fullerene-based AAA derivatives in bioorganic chemistry,³³ we turned our



Scheme 11. (i) N-phenyl maleimide, toluene, 120 °C, 96% (combined).



Scheme 12. (i) C₆₀, toluene, 120 °C.

attention to incorporate the AAA moiety in fullerene system. For example the hydrophobic character and their ability to act as an electron sink make fullerene-based AAA derivatives an attractive building block for biological application. In this context sultine (or **20a**) was reacted with Buckminister fullerene (C_{60}) in toluene at reflux temperature to give the DA product **30** in 49% yield. The FAB mass spectral data (M+H, 994) of the DA product **30** is in agreement with the structure of the DA adduct **30** (Scheme 12).

4. Conclusions

General methods for cyclic AAA synthesis is an important stepping stone for the design of unnatural peptides and complex AAAs. In this respect, we have shown that *o*xylylene derivative containing AAA moiety derived from sultine **20** (or **20a**) can be trapped with various dienophiles. It is worth mentioning that compounds, of the type **23–25**, are not accessible by the currently available BB method, due to the presence of the keto functionality. Moreover, the synthesis of the starting keto precursor required for BB method is not a trivial exercise. We believe that the methodology reported here can be extended to the synthesis of a new class of benzo-annulated AAA derivatives that may play an important role in combinatorial chemistry.

5. Experimental

5.1. General

Analytical thin layer chromatography (TLC) were performed on $(10 \times 5 \text{ cm})$ glass plate coated with Acme's silica gel G or GF 254 (containing 13% calcium sulfate as a binder). Visualization of the spots on TLC plate was achieved either by exposure to iodine vapor, or UV light. Flash chromatography was performed using Acme's silica gel (100–200 mesh) and the column was usually eluted by a ethyl acetate and petroleum ether mixture. Melting points are uncorrected. Infrared spectra (FT-IR) were recorded as KBr pellets unless otherwise mentioned. UV spectra were taken in dry chloroform as the solvent. Acetonitrile and carbon tetrachloride were distilled over phosphorous pentoxide. Dry THF and diethyl ether were obtained by distillation over sodium-benzophenone ketyl. For all the reactions, anhydrous magnesium sulfate was used as drying agent after worked up. Glycine ethyl ester hydrochloride, trimethylorthoformate, dimethyl acetylenedicarboxylate (DMAD), methyl propiolate, Wilkinson's catalyst, were purchased from Aldrich Chemical Co. (Milwaukee, WI, USA). Methyl 2-acetamidoacrylate was obtained from Lancaster Synthesis, Lancashire LA3 3BN, England (UK). ¹H and ¹³C NMR samples were made in chloroform-d

solvent and chemical shifts are reported in δ scale using tetramethylsilane as the internal standard. The standard abbreviation s, d, t, q and m refer to singlet, doublet, triplet, quartet and multiplets, respectively. 60 MHz ¹H NMR spectra were recorded on EM-300 spectrometer. 300, 400 MHz ¹H and 75.4, 100 MHz ¹³C NMR spectra were recorded on Bruker spectrometer. Coupling constants (*J*) are in Hertz.

5.1.1. 2-Acetylamino-5,6-bis-bromomethyl-indane-2carboxylic acid ethyl ester (18). To a stirred solution of the diol 17^{25} (800 mg, 2.6 mmol) in dry benzene (30 ml) was added phosphorous tribromide (1.6 g, 6.3 mmol) at 0 °C and the mixture was stirred at rt for 6 h. Then, the reaction mixture was poured into ice-cold water (40 ml) and then extracted with chloroform $(3 \times 100 \text{ ml})$. The combined organic extract was washed with water, brine, and then dried over magnesium sulfate. Evaporation of the solvent and purification of the crude product by a silica gel column using 30% ethyl acetate-hexane mixture as an eluent gave the dibromide 18 as colourless pellets (914 mg, 81%). Mp 172–174 °C. IR (KBr): ν_{max} 3225 (NH), 1737 (COOEt), 1654 (NCO) cm⁻¹. UV (CHCl₃): λ_{max} (ε M⁻¹ cm⁻¹) 274 (1272), 243 (2764) nm. ¹H NMR (300 MHz, CDCl₃): δ 1.25 (t, J=7.1 Hz, 3H), 1.96 (s, 3H), 3.25 (d, J=16.8 Hz, 2H),3.61 (d, J = 16.8 Hz, 2H), 4.22 (q, J = 7.1 Hz, 2H), 4.65 (s, 4H), 6.03 (s, 1H), 7.21 (s, 2H). ¹³C NMR (75.4 MHz, CDCl₃): δ 14.1, 23.2, 30.3, 43.2, 61.9, 65.9, 127.2, 135.6, 141.8, 170.2, 172.6. Anal.: for C₁₆H₁₉NO₃Br₂ Calcd: 44.36 (C), 4.42 (H), 3.23 (N)%; found: 44.57 (C), 4.43 (H), 3.01 (N)%.

5.1.2. 2-Acetylamino-7-oxo-1,2,3,5,7,8-hexahydro-6-oxa- $7\lambda^{\circ}$ -thia-cyclopenta[b]naphthalene-2-carboxylic acid ethyl ester (20 and 20a). To a suspension of rongalite (710 mg, 0.43 mmol) in DMF (15 ml) was added dibromo compound 18 (200 mg, 0.46 mmol) and tetrabutylammonium bromide (TBAB) (148 mg, 0.43 mmol) at 0 °C, and the resulting solution was stirred at 0 °C for 3 h, and then at rt for 4 h. The reaction mixture was quenched with water (25 ml) and extracted with chloroform $(3 \times 150 \text{ ml})$, dried over magnesium sulfate and concentrated. The crude product was purified on a silica gel column. Elution with 60% ethyl acetate-hexane mixture gave the fast moving compound **20** as a white solid (56 mg, 36%). Mp: 180– 181 °C. UV (CHCl₃): λ_{max} ($\varepsilon \text{ M}^{-1} \text{ cm}^{-1}$) 240 (780), 276 (1092) nm. IR (KBr): ν_{max} 3324 (NH), 1732 (COOEt), 1668 (NCO), 1453, 1169 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.24 (t, J = 7.3 Hz, 3H), 1.95 (s, 3H), 3.30 (d, J = 16.8 Hz, 2H), 3.53 (d, J = 15.4 Hz, 1H), 3.62 (d, J = 16.8 Hz, 2H), 4.22 (q, J=7.3 Hz, 2H), 4.40 (d, J=15.3 Hz, 1H), 4.92 (d, J=13.5 Hz, 1H), 5.27 (d, J=13.2 Hz, 1H), 6.11 (s, 1H), 7.12 (s, 1H), 7.14 (s, 1H). ¹³C NMR (75.4 MHz, CDCl₃): δ 14.2, 23.1, 43.2, 57.8, 61.7, 63.4, 65.7, 122.1, 125.5, 125.7, 133.1, 140.2, 141.1, 169.8, 172.6. HRMS (EI): m/z for C₁₆H₁₉O₅S Calcd: (M+H): 338.1062; found: 338.1057.

Further elution of the column with the same solvent system gave **20a** as a white solid (56 mg, 36%). Mp: 198–199 °C. UV (CHCl₃): λ_{max} (ε M⁻¹ cm⁻¹) 240 (901), 276 (1224) nm. IR (KBr): ν_{max} 3329 (NH), 1735 (COOEt), 1658 (NCO), 1459, 1163 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.24 (t, *J*=7.2 Hz, 3H), 1.95 (s, 3H), 3.27 (d, *J*=16.8 Hz,
2H), 3.54 (d, J=15.3 Hz, 1H), 3.63 (d, J=16.8 Hz, 2H), 4.21 (q, J=7.2 Hz, 2H), 4.31 (d, J=15.2 Hz, 1H), 4.92 (d, J=13.5 Hz, 1H), 5.23 (d, J=13.5 Hz, 1H), 6.15 (s, 1H), 7.07 (s, 1H), 7.10 (s, 1H). ¹³C NMR (75.4 MHz, CDCl₃): δ 14.1, 23.0, 43.1, 43.2, 57.0, 61.8, 63.9, 65.8, 122.0, 124.8, 126.0, 132.5, 140.2, 140.8, 170.2, 172.6. HRMS (EI): m/zfor C₁₆H₁₉O₅S Calcd: (M+H): 338.1062; found: 338.1063.

5.2. General procedure for the DA reaction with sultine derivative with various dienophiles and subsequent DDQ oxidation of the DA adduct

A solution of the sultine (1 equiv) and dienophile (2-3 equiv) in toluene was refluxed until the starting materials have been disappeared. At the conclusion of the reaction (TLC monitoring), the solvent was removed at reduced pressure and the crude product was purified by silica gel column chromatography. Elution of the column with ethyl acetate/petroleum ether mixture gave the required DA adduct always mixed with small amount aromatized product. Subsequently, that DA adduct (1 equiv) and DDQ (1.5-2 equiv) in dry benzene was refluxed (22-48 h). The reaction mixture was quenched with water and extracted with ethyl acetate. The combined organic layer was washed with 2% KOH, water, brine, and dried with MgSO₄. The solvent was evaporated and the crude product was charged on a silica gel column. Elution of the column with ethyl acetate/petroleum ether mixture gave the desired product.

5.2.1. 2-Acetylamino-2,3-dihydro-1H-cyclopenta[b]naphthalene-2,6,7-tricarboxylic acid ethyl ester dimethyl ester (21). A solution of the sultine 20 (15 mg, 0.04 mmol) and dimethyl acetylenedicarboxylate (DMAD) (20 mg, 0.14 mmol) in dry toluene (2 ml) was stirred at 120 °C in a sealed tube for 48 h. At the conclusion of reaction (TLC monitoring), the solvent was removed at reduced pressure. The crude product was purified by silica gel column chromatography. Elution of the column with 50% ethyl acetate/petroleum ether gave the required DA adduct (15 mg) as a semi-solid. Subsequently, the DA adduct and DDQ (22 mg, 0.08 mmol) in dry benzene (2 ml) was refluxed for 48 h. At the conclusion of reaction (TLC monitoring), the reaction mixture was worked up according to the general procedure. The crude product was purified by a silica gel column chromatography. Elution of the column with 50% ethyl acetate/petroleum ether gave the desired product 21 (14.4 mg, 78%) as a semi-solid. UV (CHCl₃): $\lambda_{\text{max}} (\varepsilon \text{ M}^{-1} \text{ cm}^{-1})$ 254 (12,906) nm. IR (KBr): ν_{max} 3321 (NH), 1737 (COOR), 1631 (NCO) cm^{-1} . ¹H NMR (300 MHz, CDCl₃): δ 1.23 (t, J=6.9 Hz, 3H), 1.95 (s, 3H), 3.49 (d, J=16.8 Hz, 2H), 3.71 (d, J=16.8 Hz, 2H), 3.94 (s, 6H), 4.21 (q, J=6.9 Hz, 2H), 6.11 (s, 1H), 7.67 (s, 2H), 8.11 (s, 2H). ¹³C NMR (75.4 MHz, CDCl₃): δ 14.2, 23.2, 42.8, 52.6, 61.8, 66.1, 123.6, 128.2, 129.7, 133.2, 142.3, 168.0, 172.6. HRMS (EI): *m/z* for C₂₂H₂₃NO₇ Calcd: (M+H): 414.1552; found: 414.1548.

5.2.2. 2-Acetylamino-2,3-dihydro-1H-cyclopenta[b]naphthalene-2,6,-dicarboxylic acid 2-ethyl ester 6methyl ester (22). A solution of the sultine 20 (14 mg, 0.04 mmol) and methyl propiolate (17.5 mg, 0.20 mmol) in dry toluene (2 ml) was stirred at 120 °C in a sealed tube for 24 h. At the conclusion of reaction (TLC monitoring), the

solvent was removed at reduced pressure. The crude product was purified by silica gel column chromatography. Elution of the column with 40% ethyl acetate/petroleum ether mixture gave the required DA adduct (6.5 mg) as a semisolid. Subsequently, the DA adduct and DDQ (6 mg, 0.05 mmol) in dry benzene (2 ml) was refluxed for 48 h. At the conclusion of the reaction (TLC monitoring), the reaction mixture was worked up according to the general procedure. The crude product was purified by a silica gel column chromatography. Elution of the column with 40% ethyl acetate/petroleum ether gave the desired product 22 (6.4 mg, 43%) as a white solid. Mp: 102–104 °C. UV (CHCl₃): λ_{max} ($\epsilon M^{-1} cm^{-1}$) 250 (10,533), 288 (6021) nm. IR (KBr): ν_{max} 3428 (NH), 1729 (COOR), 1650 (NCO) cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.25 (t, J=7.3 Hz, 3H), 1.95 (s, 3H), 3.49 (dd, J = 16.8, 5.1 Hz, 2H), 3.71 (dd, J = 17.2, 7.1 Hz, 2H), 3.97 (s, 3H), 4.23 (q, J = 7.3 Hz, 2H), 6.0 (s, 1H), 7.69 (s, 1H), 7.75 (s, 1H), 7.8 (d, J=8.4 Hz, 1H), 8.0 (dd, J=8.4, 1.5 Hz, 1H), 8.53 (s, 1H). HRMS (EI): m/z for C₂₀H₂₁NO₅ Calcd: (M+H): 356.1497; found: 356.1495.

5.2.3. 2-Acetylamino-6,9-dioxo-2,3,6,9-tetrahydro-1Hcvclopenta[b]anthracene-2-carboxylic acid ethyl ester (23). A solution of the sultine 20 (20 mg, 0.059 mmol) and 1,4-benzoquinone (12.8 mg, 0.12 mmol) in dry toluene (2 ml) was stirred at 120 °C in a sealed tube for 14 h. At the conclusion of reaction (TLC monitoring), the solvent was removed at reduced pressure. The crude product was purified by a silica gel column chromatography. Elution of the column with 50% ethyl acetate/petroleum ether gave the required DA adduct (20.5 mg) as a yellow solid. Subsequently, the DA adduct and DDQ (15.5 mg, 0.07 mmol) in dry benzene (2 ml) was refluxed for 30 h. At the conclusion of the reaction (TLC monitoring), the reaction mixture was worked up according to the general procedure. The crude product was purified by a silica gel column chromatography. Elution of the column with 50% ethyl acetate/petroleum ether gave the desired product 23 (20 mg, 90%) as a yellow solid. Mp: >290 °C. UV (CHCl₃): λ_{max} (ε $M^{-1} cm^{-1}$) 250 (20,416), 300 (18,053), 420 (5771) nm. IR (KBr): ν_{max} 3321 (NH), 1737 (COOR), 1668 (CO), 1629 (NCO) cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.23 (t, *J*= 7.1 Hz, 3H, ester CH₃), 1.98 (s, 3H, NCOCH₃), 3.55 (d, J =17.4 Hz, 2H), 3.77 (d, J = 17.4 Hz, 2H), 4.23 (q, J = 7.1 Hz, 2H), 6.16 (s, 1H), 7.04 (s, 2H), 7.85 (s, 2H), 8.52 (s, 2H). ¹³C NMR (75.4 MHz, CDCl₃): δ 14.0, 23.2, 42.0, 62.0, 65.9, 125.1, 127.9, 128.3, 134.6, 143.7, 139.9, 170.3, 172.5, 184.7. HRMS (EI): m/z for $C_{22}H_{19}O_5N$ Calcd: (M+H): 378.1341; found: 378.1334.

5.2.4. 2-Acetylamino-6,11-dioxo-2,3,6,11-tetrahydro-1Hcyclopenta[b]naphthacene-2-carboxylic acid ethyl ester (24). A solution of the sultine 20 (20 mg, 0.059 mmol) and 1,4-naphthoquinone (13.0 mg, 0.12 mmol) in dry toluene (2 ml) was stirred at 120 °C in a sealed tube for 14 h. At the conclusion of reaction (TLC monitoring), the solvent was removed at reduced pressure. The crude product was purified by a silica gel column chromatography. Elution of column with 50% ethyl acetate/petroleum ether gave the required DA adduct (17.5 mg) as a yellow solid. Subsequently, the DA adduct and DDQ (26.0 mg, 0.114 mmol) in dry benzene (2 ml) was refluxed for 48 h. At the conclusion of reaction (TLC monitoring), the reaction mixture was worked up according to the general procedure. The crude product was purified by a silica gel column chromatography. Elution of column with 50% ethyl acetate/petroleum ether gave the desired product **24** (17 mg, 89%) as a yellow solid. Mp: 300–302 °C. UV (CHCl₃): λ_{max} (ϵ M⁻¹ cm⁻¹) 248 (38,982), 302 (32,233), 400 (4116) nm. IR (KBr): ν_{max} 3319 (NH), 1726 (COOR), 1665 (CO), 1621 (NCO) cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.25 (t, *J*= 7.3 Hz, 3H), 1.99 (s, 3H), 3.56 (d, *J*=17.2 Hz, 2H), 3.78 (d, *J*=17.2 Hz, 2H), 4.24 (q, *J*=7.2 Hz, 2H), 6.16 (s, 1H), 7.81–7.85 (m, 2H), 7.88 (s, 2H), 8.37–8.41 (m, 2H), 8.75 (s, 2H). ¹³C NMR (75.4 MHz, CDCl₃): δ 14.0, 23.2, 42.8, 62.0, 66.0, 125.0, 127.4, 129.0, 129.4, 134.1, 134.4, 135.0, 143.5, 170.3, 172.5, 184.7. HRMS:(EI) *m/z* for C₂₆H₂₁O₅N Calcd: (M+H): 428.1497; found: 428.1499.

5.2.5. 2-Acetylamino-6,13-dioxo-2,3,6,13-tetrahydro-1Hcyclopenta[b]pentacene-2-carboxylic acid ethyl ester (25). A solution of the sultine 20 (15 mg, 0.044 mmol) and 1,4-anthraquinone (18.5 mg, 0.089 mmol) in dry toluene (2 ml) was stirred at 120 °C in a sealed tube for 24 h. At the conclusion of reaction (TLC monitoring), the solvent was removed at reduced pressure. The crude product was purified by a silica gel column chromatography. Elution of the column with 50% ethyl acetate/petroleum ether gave the required DA adduct (22 mg) as a yellow solid. Subsequently, the DA adduct and DDQ (16.8 mg, 0.75 mmol) in dry benzene (2 ml) was refluxed for 48 h. At the conclusion of reaction (TLC monitoring), the reaction mixture was worked up according to the general procedure. The crude product was purified by a silica gel column chromatography. Elution of the column with 50% ethyl acetate/petroleum ether gave the desired product 25 (19.5 mg, 92%) as a yellow solid. Mp: 216–218 °C. UV (CHCl₃): λ_{max} ($\varepsilon M^{-1} cm^{-1}$) 248 (14,653), 306 (17,722), 410 (2443) nm. IR (KBr): ν_{max} 3328 (NH), 1736 (COOR), 1670 (CO), 1626 (NCO) cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.25 (t, J=7.3 Hz, 3H), 1.99 (s, 3H), 3.57 (d, J=17.2 Hz, 2H), 3.79 (d, J=17.2 Hz, 2H), 4.24 (q, J=7.2 Hz, 2H), 6.11 (s, 1H), 7.70-7.73 (m, 2H), 7.92 (s, 2H), 8.12-8.15 (m, 2H), 8.86 (s, 2H), 8.94 (s, 2H). FAB-MS (EI): *m*/*z* 419 (M-NHAc), 441 (M – NHAc + Na + H).

5.2.6. 6-Acetylamino-2,2-dioxo-1,2,3,5,6,7-hexahydro- $2\lambda^{\circ}$ -thia-s-indacene-6-carboxylic acid ethyl ester (26). A solution of the sultine 20 (20 mg, 0.059 mmol) in dry toluene (2 ml) was stirred at 120 °C in a sealed tube for 24 h. At the conclusion of reaction (TLC monitoring), the solvent was removed at reduced pressure. The crude product was purified by a silica gel column chromatography. Elution of the column with 50% ethyl acetate/petroleum ether gave the required compound 26 (14 mg, 88%) as a white solid. Mp: 210–211 °C. UV (CHCl₃): λ_{max} (ε M⁻¹ cm⁻¹) 278 (1905) nm. IR (KBr): ν_{max} 3440 (NH), 1689 (COOEt), 1629 (NCO), 1428, 1137 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.23 (t, J = 7.3 Hz, 3H), 1.97 (s, 3H), 3.31 (d, J = 16.8 Hz, 2H), 3.60 (d, J = 16.8 Hz, 2H), 4.23 (q, J = 7.1 Hz, 2H), 4.32 (s, 4H), 6.03 (s, 1H), 7.14 (s, 2H). ¹³C NMR (75.4 MHz, CDCl₃): δ 14.0, 23.1, 43.1, 56.8, 61.9, 65.6, 122.1, 129.8, 141.1, 170.1, 172.6. MS (EI): *m/z* 437 (M⁺). Anal.: for C₁₆H₁₉SNO₅ Calcd: 56.95 (C), 5.68 (H), 4.16 (N)%; found 57.20 (C), 5.79 (H), 3.69 (N)%.

5.2.7. 2,6-Bis-acetylamino-2,3,5,6,7,8-hexahydro-1Hcyclopenta[b]naphthalene-2,6,-dicarboxylic acid 2-ethyl ester 6-methyl ester (27). A solution of the sultine 20 (20 mg, 0.059 mmol) and methyl 2-acetamidoacrylate (17 mg, 0.2 mmol) in dry toluene (3 ml) was stirred at 120 °C in a sealed tube for 14 h. At the conclusion of reaction (TLC monitoring), the solvent was removed at reduced pressure. The crude product was purified by a silica gel column chromatography. Elution of the column with 60% ethyl acetate/petroleum ether gave the required DA adduct 27 (8 mg, 33%) as a white solid. Mp: 208-210 °C. UV (CHCl₃): λ_{max} (ϵ M⁻¹ cm⁻¹) 246 (32,956) nm. IR (KBr): ν_{max} 3303 (NH), 1750 (COOR), 1626 (NCO) cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.24 (t, J=7.3 Hz, 3H), 1.93 (s, 3H), 1.95 (s, 3H), 1.99-2.16 (m, 1H), 2.51-2.56 (m, 1H), 2.74–2.94 (m, 3H), 3.20–3.28 (m, 3H), 3.56 (dd, J = 16.8, 5.54 Hz, 2H), 3.76 (s, 3H), 4.21 (q, J=7.1 Hz, 2H), 5.69 (s, 1H), 6.13 (s, 1H), 6.9 (s, 1H), 6.95 (s, 1H). HRMS (EI): m/z for C₂₂H₂₈NO₆ Calcd: (M+H): 417.2025; found: 417.2022.

5.2.8. 7-Acetylamino-1,3-dioxo-2-pheny1,2,3,3a,4,6,7,8, 10,10a-decahydro-2-aza-dicyclopenta[b,g]naphthalene-7-carboxylic acid ethyl ester (28 and 29). A solution of the sultine 20 (20 mg, 0.059 mmol) and N-phenyl maleimide (20 mg, 0.115 mmol) in dry toluene (2 ml) was stirred at 120 °C in a sealed tube for 12 h. At the conclusion of reaction (TLC monitoring), the solvent was removed at reduced pressure. The crude product was purified by a silica gel column chromatography. Elution of the column with 50% ethyl acetate/petroleum ether gave a fast moving DA adduct **28** (13 mg, 46%) as a white solid. Mp: 162–164 °C. UV (CHCl₃): λ_{max} (ε M⁻¹ cm⁻¹) 244 (1727), 274 (1826) nm. IR (KBr): 3317 (NH), 1734 (COOEt), 1668 (NCO) cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.24 (t, J=7.2 Hz, 3H), 1.93 (s, 3H), 2.93–2.98 (m, 2H), 3.17 (d, J = 16.8 Hz, 2H), 3.19-3.24 (m, 2H), 3.44-3.45 (m, 2H), 3.59 (d, J=16.8 Hz, 2H), 4.20 (q, J=7.2 Hz, 2H), 5.91 (s, 1H), 6.93 (dd, J=7.9, 1.5 Hz, 2H), 7.03 (s, 2H), 7.28–7.38 (m, 3H). ¹³C NMR (75.4 MHz, CDCl₃): δ 14.1, 23.1, 30.0, 40.2, 43.2, 61.6, 65.9, 124.1, 126.1, 127.2, 128.5, 128.9, 133.8, 139.0, 170.0, 172.6, 178.5. HRMS (EI): m/z for $C_{26}H_{26}O_5N_2$ Calcd: (M+H): 447.1919; found: 447.1908.

Further elution of the column with the same solvent system gave another DA adduct **29** as a white solid (13 mg, 36%). Mp: 142–144 °C UV (CHCl₃): λ_{max} ($\varepsilon M^{-1} cm^{-1}$) 246 (1657), 274 (1808) nm. IR (KBr): ν_{max} 3321 (NH), 1732 (COOEt), 1667 (NCO) cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.23 (t, J=7.3 Hz, 3H), 1.94 (s, 3H), 2.92–2.96 (m, 2H), 3.21 (d, J=16.8 Hz, 2H) 3.20–3.25 (m, 2H), 3.44–3.46 (m, 2H), 3.57 (d, J=16.8 Hz, 2H), 4.19 (q, J=7.3 Hz, 2H), 6.01 (s, 1H), 6.89 (dd, J=8.0, 1.8 Hz, 2H), 7.04 (s, 2H), 7.30–7.38 (m, 3H). ¹³C NMR (75.4 MHz, CDCl₃): δ 14.0, 23.2, 30.0, 40.2, 43.2, 61.6, 65.9, 124.1, 126.3, 128.5, 129.0, 131.6, 133.8, 139.1, 170.0, 172.8, 178.5. HRMS (EI): m/z for C₂₆H₂₆O₅N₂ Calcd: (M+H): 447.1919; found: 447.1915.

5.2.9. DA adduct of 2-acetylamino-2,2-dioxo-1,2,3,5,6,7hexahydro- $2\lambda^6$ -thia-s-indacene-6-carboxylic acid ethyl ester (20 and 20a) with C₆₀ (30). A solution of the sultine 20 (14 mg, 0.041 mmol) and C₆₀ (30 mg, 0.041 mmol) in dry toluene (2 ml) was stirred at 120 °C in a sealed tube for 10 h. At the conclusion of reaction (TLC monitoring), the solvent was removed at reduced pressure. The crude product was purified by a silica gel column chromatography. Elution of column with 25% ethyl acetate/petroleum ether gave the DA adduct **30** (20.4 mg, 49%) as a black solid. Mp: 246 °C. UV (CHCl₃): λ_{max} (ε M⁻¹ cm⁻¹) 260 (44,760) nm. IR (KBr): ν_{max} 3423 (NH), 1723 (COOEt), 1648 (NCO) cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.25 (brs, 3H), 2.03 (s, 3H), 3.47 (d, *J*=16.0 Hz, 2H), 3.79 (d, *J*=16.0 Hz, 2H), 4.26 (q, *J*=7.3 Hz, 2H), 4.40 (d, *J*=14.1 Hz, 2H), 4.78 (d, *J*= 14.2 Hz, 2H), 6.14 (s, 1H), 7.51 (s, 2H). FAB-MS (EI): *m/z* 994 (M+H), 1017 (M+Na+H).

Acknowledgements

We gratefully acknowledge the CSIR, New Delhi for financial support and the RSIC Mumbai for recording the spectral data, AG thanks the IIT-Bombay for the award of research fellowship.

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Tetrahedron

Tetrahedron 60 (2004) 10843-10850

Aluminum dodecatungstophosphate (AlPW₁₂O₄₀) as a non-hygroscopic Lewis acid catalyst for the efficient Friedel–Crafts acylation of aromatic compounds under solvent-less conditions

Habib Firouzabadi,* Nasser Iranpoor* and Farhad Nowrouzi

Department of Chemistry, Shiraz University, Shiraz 71454, Iran

Received 1 August 2004; revised 24 August 2004; accepted 16 September 2004

Available online 1 October 2004

Abstract—Stable and non-hygroscopic aluminum dodecatungstophosphate (AlPW₁₂O₄₀), which is prepared easily from cheap and commercially available compounds was found to be an effective catalyst for Friedel–Crafts acylation reactions using carboxylic acids, acetic anhydride and benzoyl chloride in the absence of solvent under mild reaction conditions. © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

Heteropolyacids (HPAs) show very high catalytic activity for some acid-catalyzed reactions. They have been used for hydration of alkenes^{1–3} and polymerization of tetrahydrofuran.^{4–6} Strong Brønsted acidity^{7,8} and the softness of the heteropoly anions are responsible for their high catalytic activities in these reactions.⁹ HPAs are usually solids that are insoluble in non-polar solvents but highly soluble in polar ones. The use of HPAs in non-polar solvents improves product selectivity and also provides easy separation of them.⁴ Heterogeneous catalysis has become attractive in view of the increasingly strict environmental legislation, in view of their isolation and separation from the reaction media. Solid acid catalysts are harmless to the environment with respect to corrosiveness, safety, quantity of waste, and separability with certainly some exceptions.

Cesium salts of HPAs are good examples of active and useful solid acid catalysts. They exhibit an excellent performance in several organic reactions owing to their high surface acidity and also probably to their unique basicity.^{10,11} The catalytic activities of solid acids are usually suppressed significantly in the presence of water. Hence, although H-ZSM-5 has a high Si:Al ratio is known to

0040–4020/\$ - see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.049

have fair tolerance in aqueous solutions.¹² It has been demonstrated that the acidic cesium salts of HPAs are very water-tolerant catalysts for hydration of olefins¹³ and hydrolysis of esters.¹⁴ This nature was assigned to moderate hydrophobicity of the catalysts.¹⁵

We have recently started to study new catalytic applications of heteropoly acids in organic reactions.^{16a-h}

Friedel–Crafts acylation of aromatic compounds is the most important and practical route for the synthesis of aromatic ketones that are used in manufacturing of fine chemicals as well as pharmaceuticals.¹⁷ The acylating agents for the synthesis of aromatic ketones by Friedel–Crafts reactions are mostly acid anhydrides or acyl chlorides. For this purpose, varieties of catalysts have been reported.^{18,19} A literature survey indicates that the use of carboxylic acids, as acylating agents, are scarcely reported.²⁰ Using carboxylic acids as acylating agents is a superior method with respect to the procedures utilizing acyl chlorides and anhydrides for the preparation of aryl ketones. Carboxylic acids are stable and more available compounds and their handlings are much easier than their corresponding acyl chlorides and anhydrides.

Now, we introduce aluminum dodecatungstophosphate $(AlPW_{12}O_{40})$ as a stable, non-hygroscopic, easily available, cheap and highly effective catalyst for Friedel–Crafts acylation of aromatic compounds using carboxylic acids, acetic anhydride and benzoyl chloride as acylating agents.

Keywords: Freidel–Crafts acylation; Poly oxometalate; Aluminum dodecatungstophosphate; Solvent-free.

^{*} Corresponding author. Tel.: +98 711 2284822; fax: +98 711 2280926 (H.F.); fax: +98 711 2280926 (N.I.); e-mail addresses: firouzabadi@chem.susc.ac.ir; iranpoor@chem.susc.ac.ir

2. Results and discussion

2.1. Acylation of aromatic compounds with acetic anhydride catalyzed by AlPW₁₂O₄₀

The reaction of acetic anhydride and aromatic compounds by Friedel–Crafts acylation reactions is the most practical route for the synthesis of methyl aryl ketones. Investigation of the application of $AlPW_{12}O_{40}$ as an easily available, cheap and non-hygroscopic Lewis acid for this purpose is of practical importance. In order to optimize the reaction conditions, acylation of anisole with acetic anhydride in the presence of different molar ratios of $AlPW_{12}O_{40}$ and various solvents were studied (Scheme 1).

MeOPh +
$$Ac_2O(2 eq) \xrightarrow{AlPW_{12}O_{40} (3 Mol\%)} MeCOAr$$

Scheme 1.

The results of this investigation are shown in Table 1. The results indicate that the most suitable solvent for this purpose is *n*-hexane in which the catalyst reacts under completely heterogeneous conditions and the best result was obtained when no solvent was used.

The optimum ratio of the catalyst was found to be $3 \mod \%$ with respect to anisole for the reaction conducted in *n*-hexane or solvent-less conditions. The optimized molar ratio of acetic anhydride was found to be 2 mol equivalents with respect to 1 mol equivalent of anisole under similar reaction conditions (Scheme 2).

ArH +
$$Ac_2O(2 \text{ eq}) \xrightarrow{AlPW_{12}O_{40} (3 \text{ Mol}\%)}{\text{neat,60-70 °C}}$$
 MeCOAr

Scheme 2.

This reaction condition was applied for the preparation of different aryl ketones from electron rich aromatic compounds with success. The results of this study are tabulated in Table 2.

In order to show the efficiency and the catalytic activity of $AlPW_{12}O_{40}$ for the acylation of anisole with acetic anhydride, we have tabulated our results in Table 3 with some other recently used Lewis acids such as Sc(OTf)₃, Bi(OTf)₃ and In(OTf)₃.

The results show that aluminum dodecatungstophosphate is a more efficient and more effective catalyst than the triflates used for similar reaction and also does not require aqueous workup which makes the process of the isolation of the product much easier and not a time-consuming process.

2.2. Benzoylation of aromatic compounds with benzoyl chloride in the presence of $AIPW_{12}O_{40}$

We have studied optimization of the reaction conditions for the benzoylation of anisole under solvent-free conditions (Scheme 3).

ArH + PhCOCI
$$\xrightarrow{AIPW_{12}O_{40} (4 \text{ Mol}\%)}_{\text{Neat, 60-70°C}} PhCO-Ar$$

Scheme 3.

It was found that the reaction was complete after 2 h by using of 4 mol% of the catalyst at 60–70 °C to produce ortho and *para* isomers (1:4 ratio). We applied similar reaction conditions for the preparation of other aryl ketones using different aromatic compounds. The reactions proceeded well and the desired products were isolated in excellent yields (Table 4). We have observed that the rate of acylation and benzoylation of 1,3-dimethoxy benzene was slower than anisole. Para acylation or benzoylation of anisole do not encounter that much steric resistance against the approach of bulky Lewis acid complex and acylium ions from the para position. However, in 1,3-dimethoxy benzene the situation is quiet different and the bulky complex should only approach from the ortho position of OMe groups which encounters ortho steric effects. This effect causes retardation in the rates of the reactions. Bezovlation of anthracene under solvent-free conditions was sluggish in the presence of this catalyst and gave a low yield. However, 9benzoylanthracene was isolated in 88% yield when the reaction in *n*-hexane was heated under reflux conditions. We also applied similar reaction conditions for benzovlation of naphthalene. This reaction was not successful and a mixture of 1 and 2 substituted benzoyl naphthalenes in low yields plus unreacted starting material was isolated. Benzoylation of benzene under similar reaction conditions failed. However, this reaction was successful in a sealed tube at 120 °C overnight with 10 mol% of AlPW12O40 (97% GC yield).

2.3. Direct acylation of anisole with carboxylic acids in the presence of $AIPW_{12}O_{40}$ as a reusable catalyst

We have also studied acylation of anisole with carboxylic acids in the presence of 6 mol% of $AlPW_{12}O_{40}$ at 120 °C (Scheme 4).

Table 1. Effect of solvents upon Friedel–Crafts acylation of anisole catalyzed by $AIPW_{12}O_{40}$

Entry	Solvent	Temp. (°C)	<i>T</i> (h)	Conv. (%) ^a
1	CH ₂ Cl ₂	40	5	63
2	CHCl ₃	62	3	76
3	CH ₃ CN	82	3	38
4	CH ₃ NO ₂	100	4	63
5	<i>n</i> -Hexane	69	2.5	93
6	None	60	1	100

^a Conversion was determined by GC analysis.

$$MeO \longrightarrow + RCO_2H \xrightarrow{AIPW_{12}O_{40}(6 \text{ Mol}\%)}{120 \text{ °C}, R=C_1-C_{11}, Ph} MeO \longrightarrow R$$
(50 mmol) (1 mmol)

Scheme 4.

This acylation was carried out with different carboxylic acids (C_2-C_{12}) in good yields (Table 5). In this study, it was observed that by increasing the chain length of the carboxylic acids from C_2 to C_{12} the regioselectivity of the reactions increased and *p*-isomers were mostly produced. We have not also observed any ester formation in any of the reactions we conducted in the presence of AlPW₁₂O₄₀. We

have found that this catalyst is reusable for these reactions and can be easily recovered from the reaction mixture by simple filtration followed by washing the mixture by *n*hexane/EtOAc (1:1). Drying of the catalyst was performed at 120 °C under vacuum overnight. We have used the recovered catalyst for the reaction of anisole with hexanoic acid with success five times without noticeable change in its catalytic activity. In Table 6, the results are compiled in order to show the effectiveness of AlPW₁₂O₄₀ with respect to the recently reported catalysts $Cs_{2.5}H_{0.5}PW_{12}O_{40}$ and HZSM-5 for acylation of anisole with different carboxylic acids. The results show that AlPW₁₂O₄₀ promotes the reactions more effectively than the reported catalysts.

Table 2. Acylation of aromatic compounds with acetic anhydride catalyzed by $AIPW_{12}O_{40}$

Entry	ArH	Time (h)	Product	Yield ^a (%)	Ref.
1	OMe	0.75	MeCO-OMe	94 (<i>Ortho:Para</i> =12:88)	18p,t
2	OMe	0.25	MeCO-OMe	95	18k,y
3	MeO	2	MeO MeCO	88 ^b	18k
4	OEt OEt	0.6	MeCO OEt	93	19a
5		6	СОМе	91	18n
6		2.5	MeCO	92°	18f
7		5.5	MeCO	90	181
8		4	COMe	85	18j
9	OMe	1.6	COMe OMe	91	18i
10		2.6	COMe	90 ^d	18i

^a Isolated yields and the purity of the products were determined by ¹H NMR spectroscopy and the ratio of the isomers was determined by GC.

^b 2,6-Dimethoxyacetophenone was also detected in 12% yield.

^c Other regioisomers were detected in 8%.

^d Ac₂O (4 equiv/mol) was used.

Table 3. Comparison of the results used for the acylation of anisole with acetic anhydride catalyzed by $AlPW_{12}O_{40}$ and some other catalysts

Entry	Catalyst	Mol%	Temp. (°C)	<i>T</i> (h)	Yield (%)	Ref
1	Sc(OTf) ₃	20	50	18	99	181
2	Sc(OTf) ₃	20	50	2 ^a	90	18n
3	Bi(OTf) ₃	5	50	2	80	18p
4	$In(OTf)_3$	1	50	1 ^b	96	18w
5	$AlPW_{12}O_{40}$	3	60	0.8	94	c

^a 2 equiv of LiClO₄ was used as an additive. ^b 1 equiv of LiClO₄ was used as an additive.

^c Results are shown in Table 2.

Table 4. Benzoylation of aromatic compounds with benzoyl chloride catalyzed by $AlPW_{12}O_{40}$

Entry	ArH	Time (h)	Product	Yield ^a	Ref.
1	OMe	2	PhCO	94 (<i>Ortho:Para</i> =1:4)	18p,t
2	ОМе	1	PhCO-OMe	97	18k
3	MeO	3.5	MeO PhCO—OMe	90 ^b	18d
4	OEt	2	PhCO-OEt	90	19Ь
5		4.5	COPh	91	18u
6		3.5	PhCO	92°	18q,v
7		2.5	PhCO	91	18q
8		3	PhCO	92 ^d	18f
9	OMe	1.7	COPh OMe	94	18a
10		4	COPh	88°	18x

^a Isolated yields and the purity of the products were determined by ¹H NMR spectroscopy and the ratio of the isomers was determined by GC. ^b Formation of 2,6-dimethoxybenzophenone was detected in 10%.

^c Formation of 2,6-dimethylbenzophenone was detected in 8%.

^d Other regioisomers were detected in 7%.

^e Reaction was carried out in 2 mL of *n*-hexane under reflux conditions.

Table 5. Acylation of anisole with carboxylic acids catalyzed by reusable AlPW_{12}O_{40} at 120 $^{\circ}\text{C}^{a}$

Entry	Acid	Time (h)	Yield (%) ^b Ref. ^{20b}
1	Acetic	3	85 ^c
2	Propionic	3.5	88^{d}
3	Butyric	5	87
4	Hexanoic	3	0^{e}
5	Hexanoic	1.5	91
6	Octanoic	4	85
7	Dodecanoic	8	90
8	Benzoic	10	92

^a The catalyst was filtered, washed with *n*-hexane: ethyl acetate, dried at 120 °C/(-20) Torr/overnight and reused.

^b Isolated yield based on carboxylic acid conversion determined by ¹H NMR spectroscopy and GC analysis.

^c 12% regioisomer was determined.

^d 8% regioisomer was detected.

^e Reaction was carried out in the presence of 1 g silica gel.

Table 6. Comparison of some of the results obtained using $AlPW_{12}O_{40}$ with some other catalysts used for acylation of anisole with carboxylic acids^a

Acid	Cat.	Yield (%) (para-product)
Acetic	HZSM-5	0.025
	40%CsPW/SiO ₂	62
	$AlPW_{12}O_{40}$	73
Hexanoic	HZSM-5	19.92
	40%CsPW/SiO ₂	63.2
	$AlPW_{12}O_{40}$	91
Octanoic	HZSM-5	4
	40%CsPW/SiO ₂	32.9
	$AlPW_{12}O_{40}$	85
Dodecanoic	HZSM-5	0
	40%CsPW/SiO ₂	24.8
	AlPW ₁₂ O ₄₀	91

^a The results were extracted from Ref. 20b,c.

2.4. Acylation of aromatic compounds with carboxylic acids in the presence of trifluoroacetic anhydride catalyzed with AlPW₁₂O₄₀ at room temperature

In this part of our studies, we have found that with 3 mol% of AlPW₁₂O₄₀ in the presence of trifluoroacetic anhydride (1.4 equiv) acylation of anisole proceeded easily in 0.25 h in an excellent yield (Scheme 5).





In order to show the effect of the catalyst in this reaction, we performed acylation of anisole with acetic acid in the presence of trifluoroacetic anhydride (1.4 equiv) at room temperature in the absence of the catalyst. We have observed that the corresponding ketone was produced in 88% yield (GC) after 4 h (Table 7, entry 1) whereas the desired ketone was isolated in 94% yield after 0.25 h in the presence of the catalyst (Table 7, entry 2). We also tried the reaction of anisole with TFAA in the presence of the

catalyst. We observed that the reaction did not proceed at all even after 24 h. We have found that this is a general method and can be applied easily to a wide range of liquid and crystalline carboxylic acids. Therefore, acylation of anisole was conducted with acetic acid, octanoic acid, phenyl acetic acid and benzoic acid in the presence of AlPW₁₂O₄₀ (3 mol%) and trifluoroacetic anhydride in excellent yields (Table 7, entries 1–5). 2-Methylanisole was also acylated with these acids in the presence of this catalyst and the anhydride in excellent yields (90-96%) with high regioselectivity (Table 7, entries 9-12). 1,2-Diethoxybenzene reacted with the above acids under similar reaction conditions to give the desired compounds as the sole products of these reactions in 91-94% yields (Table 7, entries 13-16). We also tried acylation of anthracene with acetic acid, phenyl acetic acid and benzoic acid. The reactions were slow and needed higher temperatures therefore, the reactions were performed at 50 °C and the desired ketones were isolated in 69-78% yields (Table 7 entries 17-19). The reaction of 2-methylnaphthalene with acetic acid was also conducted at 50 °C in the presence of the catalyst and trifluoroacetic anhydride. This reaction was sluggish and after prolonged reaction times the desired ketone was isolated in only 53% yield (Table 7 entry 20). The reaction of 2-methoxynaphthalene with acetic acid, octanoic acid, phenyl acetic acid and benzoic acid went smoothly at room temperature in excellent yields (Table 7, entries 21-24). Acylation of furan and pyrrole is not a straightforward reaction and usually in the presence of acid catalysts undergo polymerization reactions. We found that AlPW₁₂O₄₀ is a suitable catalyst for such important acylation reactions.

Acylation of furan with acetic acid, phenyl acetic acid and benzoic acid was conducted in the presence of TFAA and AlPW₁₂O₄₀ at 0 °C successfully in 94–98% isolated yield (Table 7, entries 25–27).

Acylation of pyrrole also conducted smoothly in the presence of TFAA, $AlPW_{12}O_{40}$ with acetic and benzoic acids at -23 °C. The desired pyrrole ketones were isolated in 92–98% yield (Table 7, entry 28–29). The formation of any polymeric materials was not observed in these reactions.

3. Conclusion

AlPW₁₂O₄₀ is a cheap, easily available, non-hygroscopic, heterogeneous, non-corrosive and environmentally benign compound. In this work it has been used as an effective catalyst for the efficient preparation of ketones via Friedel– Crafts acylation reactions with acetic anhydride, benzoyl chloride and with a range of easily available caboxylic acids in the presence of triflouroacetic anhydride. This general method can be applied easily to a wide range of liquid and crystalline carboxylic acids. The method has many advantages such as cheapness and availability of reactants and also non-aqueous and easy work-up of the reaction due to the heterogeneous conditions. The isolation of products was easily accomplished by a simple extraction and evaporation of the organic solvent.

Table 7. AIPW ₁₂ O ₄₀ catalyzed a	cylation of aromatic com	pounds with compounds with	a carboxylic acids in the	presence of trifluoroacetic anhydride
12 10 2	2		2	

Entry	ArH	RCO ₂ H	Product	Time (h)	Yield (%) ^a	Ref.
1 2 3 4 5 6 7 8	OMe OMe OMe	R = Me R = Me $R = PhCH_2$ R = Ph R = Me $R = PhCH_2$ R = Ph	R O OMe OMe R OMe R OMe R OMe OM	4 0.25 1.5 2 2.5 2 2.5 3.75	88 ^{b.c} 94 ^c 91 94 96 97 90 91	18p,t 18p,t 20b,d 18w 18p,t 18y 18c 18k
9 10 11 12	OMe	$R = Me$ $R = PhCH_2$ $R = C_7H_{15}$ $R = Ph$		0.12 0.4 0.34 0.75	96 92 93 90	18y 18c 18k
13 14 15 16	OEt	$R = Me$ $R = C_7H_{15}$ $R = PhCH_2$ $R = Ph$	R OEt	0.65 1.5 2.5 3.5	93 94 91 91	19a 19b
17 18 19		$R = Me$ $R = PhCH_2$ $R = Ph$		2.5 10 8.5	78 ^d 71 ^d 69 ^d	18i 18b 18x
20		R=Me		2	53°	18j
21 22 23 24	OMe	$R = Me$ $R = C_7H_{15}$ $R = PhCH_2$ $R = Ph$	O O O O Me	0.25 0.34 2 8	98 95 90 88	18i 18m 18m 18a
25 26 27 28 29		R = Me $R = PhCH_2$ R = Ph R = Me R = Ph		1.25 1.5 2.5 0.25 0.4	$94^{\rm f}_{95^{\rm f}}$ $98^{\rm f}_{92^{{ m g.h}}}$ $98^{{ m g.g}}_{98^{ m g}}$	18n 19c 18g 18h 18h

^a Isolated yields and the purity of the products were determined by ¹H NMR spectroscopy, ¹³C NMR spectroscopy and GC.

^b Reaction was performed without using AlPW₁₂O₄₀.

^c 8% of regioisomer was detected.

^d Reaction was carried out in 2 mL *n*-hexane at 50 °C.

^e Reaction was carried out at 50 °C.

^f Reaction was carried out at 0 °C with 2 equiv of furan.

^g Reaction was carried out in 2 mL *n*-hexane at -23 °C.

^h 8% 3-acetyl pyrrole was detected by GC.

4. Experimental

4.1. General remarks

Chemicals were purchased from Fluka or Merck and $AlPW_{12}O_{40}$ was prepared from $H_3PW_{12}O_{40}$ and aluminium nitrate according to the literature method.²¹ Aromatic

compounds, carboxylic acids, acetic anhydride and acyl chloride were purified by distillation or recrystallization before use. The purity determination of the products was accomplished by GC on a Shimadzu model GC-14A instrument or by TLC on silica gel polygram SIL G/UV 254 plates. The FTIR spectra were recorded on a Shimadzu model 8300 instrument. The NMR spectra were recorded on a Bruker Advance DPX 250 MHz spectrometer and in all cases $CDCl_3$ was used as a solvent and TMS as an internal standard.

4.2. Typical procedure for acylation of anisole with acetic anhydride; a

Anisole (2 mmol, 0.216 g), acetic anhydride (4 mmol, 0.408 g) and AlPW₁₂O₄₀ (0.06 mmol, 0.232 g) were combined, and the mixture was stirred at 60–70 °C for 45 min (Table 3). After completion of the reaction (monitored by TLC and GC), Et₂O (10 mL) and powder of sodium hydrogen carbonate (0.8 g) was added to the mixture in order to destroy the unreacted acetic anhydride. The solid phase was isolated and washed with Et₂O (2× 5 mL). Evaporation of the combined ethereal solutions afforded the almost pure product in 94% yield (0.504 g).

4.3. Typical procedure for benzoylation of anisole with benzoyl chloride

Anisole (2 mmol, 0.216 g), benzoyl chloride (3 mmol, 0.421 g) and AlPW₁₂O₄₀ (0.08 mmol, 0.232 g) were combined and the mixture was stirred at 60–70 °C for 2 h (Table 4, entry 1). After completion of the reaction (monitored by TLC and GC), Et₂O (10 mL) and powder of sodium hydrogen carbonate (0.8 g) was added to the reaction mixture in order to destroy the unreacted benzoyl chloride. The solid phase was recovered by filtration and was washed with Et₂O (2×5 mL). Evaporation of the combined ethereal solutions afforded the almost pure product in 94% yield (0.796 g).

4.4. General procedure for acylation of anisole with carboxylic acids

To a mixture of carboxylic acid (2 mmol) and anisole (100 mmol), $AlPW_{12}O_{40}$ (12 mol%, 0.362 g) was added and the reaction mixture was stirred at 120 °C for the appropriate reaction times (Table 5). After completion of the reaction (monitored by TLC and GC), the catalyst was filtered. The unreacted anisole was separated by vacuum distillation and the desired ketone was isolated with high purity (GC) in good yield.

4.5. General procedure for acylation of aromatic compounds using carboxylic acids and triflouroacetic anhydride (TFAA) catalyzed with AlPW₁₂O₄₀

Aromatic compound (2 mmol), carboxylic acid (2 mmol), trifluoroacetic anhydride (2.8 mmol, 0.39 mL) and AlPW₁₂O₄₀ (0.03 mmol, 0.174 g) were mixed together and stirred by a magnetic stirrer at room temperature for the appropriate reaction times (Table 7). After completion of the reaction (monitored by TLC and GC), Et₂O (10 mL) and powder of sodium hydrogen carbonate (1.2–1.5 g) was added to destroy the unreacted TFAA or carboxylic acid. The solid phase was recovered by filtration and was washed with Et₂O (2×5 mL). Evaporation of the combined ethereal solutions afforded the highly pure product.

4.6. Spectral data of the unknown ketones prepared by the presented protocol

4.6.1. Table 7 entry 11: $\mathbf{R} = \mathbf{C}_7 \mathbf{H}_{15}$. Colourless viscous oil, [Found: C, 77.38; H, 9.65. $C_{16}H_{24}O_2$ requires C, 77.41; H, 9.65%]. Bp (*P*/32 mmHg)=218 °C IR (KBr) (ν C=O)= 1610, 1691 cm⁻¹ $\delta_{\rm H}$ (250 MHz, CDCl₃) 7.83–7.77 (2H, m, Ph), 6.83 (1H, d, *J*=8.4 Hz), 2.92 (t, 2H, ²*J*_{*HH*}=12 Hz, – COC*H*₂CH₂), 2.25 (s, 3H), 1.67 (m, 2H), 1.56 (m, 2H), 1.42 (m, 2H), 1.36 (m, 2H), 1.21 (m, 2H), 0.88 (t, 3H, ²*J*_{*HH*}=7.5 Hz, C*H*₃CH₂). ¹³C NMR: δ (ppm)=199.53, 161.59, 130.7, 129.72, 128.1, 126.6, 109.15, 55.4, 38.2, 31.7, 29.4, 24.7, 22.6, 16.2, 14.1. MS (20 eV): *m/z* (%)=248 (M⁺, 3.1), 177 (6.9), 164 (71.5), 150 (10.1), 149 (100), 91 (23.5), 41 (16.2).

4.6.2. Table 7 entry 14: $\mathbf{R} = \mathbf{C}_7 \mathbf{H}_{15}$. Light yellow solid, mp=43 °C, [Found: C, 73.94; H, 9.57. $\mathbf{C}_{18}\mathbf{H}_{28}\mathbf{O}_3$ requires C, 73.97; H, 9.58%] **IR** (KBr) (ν C=O)=1677 cm⁻¹, δ_{H} (250 MHz, CDCl₃) 6.5–7.4 (m, 3H, Ph), 4.08 (q, 4H), 2.9 (t, 2H, $^2J_{HH}$ =12 Hz, -COCH₂CH₂), 1.5 (t, 6H, $^2J_{HH}$ =15 Hz, OCH₂CH₃), 1.71 (m, 2H), 1.41 (m, 2H), 1.3 (m, 2H), 1.18 (m, 2H), 0.98 (m, 2H), 0.81 (t, 3H, $^2J_{HH}$ =7.5 Hz, CH₃CH₂). δ_{C} (63 MHz, CDCl₃)=196.6, 153.2, 148.8, 144, 130.4, 122.9, 120, 115, 112.4, 64, 42, 32, 30, 23, 15, 14, MS (20 eV): m/z (%)=292 (M⁺, 10.8), 221 (8.6), 208 (100), 193 (73.2), 180 (14.8), 165 (38.4), 137 (36.3), 109 (17.5), 81 (14), 57 (16.1).

4.6.3. Table 7 entry 15: $\mathbf{R} = \mathbf{PhCH}_2$. Yellow solid, mp= 82 °C, [Found: C, 76.1; H, 7.02. $C_{18}H_{20}O_3$ requires C, 76.05; H, 7.04%] IR (KBr) (ν C=O)=1700, 1596 cm⁻¹ $\delta_{\rm H}$ (250 MHz, CDCl₃) 7.34–7.2 (5H, m, Ph), 7.62 (1H, d, J= 8.37 Hz), 7.46 (1H, s),6.86 (1H, d, J= 8.4 Hz), 4.22 (2H, s), 4.08–4.19 (4H, m, $-OCH_2CH_3$), 1.5–1.41 (6H, m, OCH_2CH_3). $\delta_{\rm C}$ (63 MHz, CDCl₃)=196.3, 148, 144, 135.1, 131, 130, 129.3, 128, 126, 120, 114, 64.4, 45.13, 14.6. MS (20 eV): m/z (%)=284 (M⁺, 3.3), 194 (12.5), 193 (100), 165 (32.1), 137 (27.3), 109 (10.4), 91 (19.5).

Acknowledgements

The authors are thankful to Iran TWAS Chapter Based at ISMO and the Shiraz University Research Council for the support of this work.

Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tet.2004.09. 049

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Tetrahedron

Tetrahedron 60 (2004) 10851-10857

Efficient synthesis of CCR5 antagonist, 2,3-dihydro-1-benzothiepine derivatives by improved intramolecular Claisen type reaction using dialkylcarbonate

Tomomi Ikemoto,* Tatsuya Ito, Atsuko Nishiguchi and Kiminori Tomimatsu

Chemical Development Laboratories, Takeda Pharmaceutical Company Limited, 2-17-85 Jusohonmachi, Yodogawa-ku, Osaka 532-8686, Japan

Received 3 September 2004; revised 14 September 2004; accepted 15 September 2004

Available online 2 October 2004

Abstract—The efficient synthesis of 2,3-dihydro-1-benzothiepine derivatives **4** has been developed. The intramolecular Claisen type reaction of the new products, 4-(*o*-formylphenylthio)butyrate **9**, with alcoholate in dialkylcarbonate as a solvent afforded **4** in good yields. According to this new procedure, we have accomplished the practical preparation of CCR5 antagonist **1** as a candidate for oral HIV-1 therapy.

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1. Introduction

Recently, various small-molecular nonpeptide compounds have been reported as CC chemokine receptor 5 (CCR5) antagonist, because CCR5 was found to be a coreceptor for the entry of macrophage-tropic human immunodeficiency virus type 1 (HIV-1) into host cells.^{1,2} Shiraishi et al. previously reported that TAK-779 as a CCR5 antagonist appeared to be a candidate for the therapy of HIV-1 infected individuals.³ Furthermore, N-[4-[N-methyl-N-(tetrahydropyran-4-yl)aminomethyl]phenyl]-7-(4-propoxyphenyl)-1,1-dioxo-2,3-dihydro-1-benzothiepin-4-carboxamide 1 showed CCR5 antagonistic activity when orally administered.⁴ Hence, an efficient preparation of **1** on a large scale was required to support pharmacological and toxicological evaluations. In the early report,⁴ the oxidation of 2,3dihydro-1-benzothiepine 4f with H₂O₂ followed by arylation and hydrolysis gave the acid 2, which was amidated with the aniline **3** to provide the desired **1** (Scheme 1). For the preparation of 2,3-dihydro-1-benzothiepine 4, there have been some syntheses based on the same generation (Scheme 2).^{4,5} Compound 4 was led from β -oxo-ester 6 prepared from the ketone 5. These methods via 6, however, had some drawbacks, for example, the production of overreduced compound 8 in the reduction of 7, and the requirement of several processes to 4 from 5. There were

similar problems in the preparation of **4f** via β -oxo-ester. There has been no report of synthesis of **4** based on an other generation to the best of our knowledge. On the other hand, 2,3-dihydro-1-benzoxepine and 2,3-dihydro-1-benzazepine could be synthesized from the analogues of **9** with alcoholate according to the Claisen type reaction.^{4,5a,6} The



Scheme 1.

Keywords: 2,3-Dihydro-1-benzothiepine; Claisen type condensation; Dialkylcarbonate; CCR5 antagonist.

^{*} Corresponding author. Tel.: +81 6 6300 6378; fax: +81 6 6300 6251; e-mail: ikemoto_tomomi@takeda.co.jp

^{0040–4020/\$ -} see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.039



Scheme 2.

preparation of 4 from 9 attracted our interest regarding large-scale preparations because of its simple and wasteless protocol, even though the yields based on this generation were not sufficiently high. In this paper, we announce the facile and efficient synthesis of 4, and the application for 1.

2. Results and discussion

2.1. Development of new preparation of 2,3-dihydro-1benzothiepines

We synthesized 9, having various substituted groups, to develop a new synthetic procedure for 4. The synthesis of new products 9a-e was accomplished by thioalkylation of *o*-halogenobenzaldehyde 10 with 4-mercaptobutyrate 11^7 and K_2CO_3 in DMF overnight at room temperature

Table 1. Thioalkylation of o-halogenobenzaldehydes^a

(Table 1), although there has been no report about the synthesis of 9. Most thioalkylations of 10 gave 9 in good to excellent yields, while 10d having an electron-donating group at the p-position of the halogen gave lower yields (entry 5).

To optimize the reaction conditions, the intramolecular cyclization of 9a for producing 4a was examined (Table 2). First, the conditions for synthesizing benzoxepines⁵ were applied to the cyclization of 9a. The treatment of 9a was conducted with NaOEt in EtOH to give many products. After the workup, the chromatographic purification provided the desired 4a in 27% yield and the hydrolyzed 12 in 18% yield (entry 1). The combinations of NaOEt and 1,2-dimethoxyethane (DME) or NaH and THF decreased the hydrolysis to increase the yields of **4a** (entries 2 and 3). However, these reactions could neither depress the production of impurities of which the structures were unknown, nor increase the total yields of cyclization (4a+12). It was thought that the value of pK_a at the α carbon of ester was insufficient for cyclization of 9a, resulting in the production of impurities.⁸ To decrease the value of pK_a , the reaction of **9a** was performed with the combination of NaOEt and diethylcarbonate as a solvent, which was converted to the malonate derivatives.⁹ Surprisingly, this treatment made a breakthrough in the yield of cyclization, giving 4a in 71% yield without detection of the malonate derivative and 12 (entry 4). For this reaction, 1.2 equiv of diethylcarbonate affected the cyclization to increase the yield of 4a (entry 6 vs 1). Moreover, the same reaction of 9a with NaOEt using ethyl formate as a solvent, expected to produce a similar effect, gave a slightly higher yield of 4a than that using DME as a solvent (entry 2 vs 5).¹⁰

For research into the scope and limitation of the intramolecular Claisen type reaction, a variety of **9** reactions were next performed under the conditions of alcoholate/ dialkylcarbonate (Table 3). The use of dimethylcarbonate



Entry	Su	Yield (%) ^b	
	10	11	
1	10a ($R^1 = R^2 = R^3 = H, X = F$)	11a ($R^4 = Et$)	9a : 67
2	10a	11b ($R^4 = Me$)	9b : 95
3	10b ($R^1 = R^2 = H, R^3 = Br, X = F$)	11a	9c : 78
4	10c $(R^1 = Cl, R^2 = R^3 = H, X = Cl)$	11a	9d : 70
5	10d $(R^1 = R^2 = H, R^3 = OMe, X = Br)$	11a	9e : 27

^a General procedure. To a suspension of 10 (1.0 g, 1.0 equiv), K_2CO_3 (2.0 equiv) and DMF (3 v/w) was added 11 (2 equiv), and the whole suspension was stirred overnight at room temperature.

^b Isolated yield.

Table 2. Optimization of reaction conditions^a



Entry	Solvent	Base	Conditions	Isolated yield (%)		
				9a	4a	12
1	EtOH	20% NaOEt in EtOH	rt, 3 h, then refluxed, 1 h	ND	27	18
2	DME	20% NaOEt in EtOH	rt, 45 min	ND	37	6
3	THF	NaH	rt, 1 h, then refluxed, 0.5 h	ND	44	6
4	(EtO) ₂ CO	20% NaOEt in EtOH	rt, 2 h	ND	71	ND
5	HCO ₂ Et	20% NaOEt in EtOH	rt, 4 h	26	58	ND
6	EtOH, (EtO) ₂ CO (1.2 eqiv)	20% NaOEt in EtOH	rt, 3 h, then refluxed, 1 h	ND	53	4

^a The mixture of **9a** (1.0 g, 1.0 equiv), base (1.2 equiv) and solvent (20 v/w) was reacted.

Table 3. The intramolecular Claisen type reaction^a



Entry	Substrate	Time (h)	Isolated yield (%)
1 ^b	9b	15	4b ($R^1 = R^2 = R^3 = H$, $R^4 = Me$): 71
2	9c	2	4c ($R^1 = R^2 = H$, $R^3 = Br$, $R^4 = Et$): 32
3	9d	7	4d ($R^1 = Cl$, $R^2 = R^3 = H$, $R^4 = Et$): 46
4	9e	6	4e ($R^1 = H$, $R^2 = R^3 = OMe$, $R^4 = Et$): 70

^a To a solution of **9** (1.0 g) and diethyl carbonate (20 v/w) was added 20% NaOEt in EtOH (1.2 equiv), and the whole mixture was stirred at room temperature.

^b 28% NaOMe in MeOH (1.2 equiv) and (MeO)₂CO (20 mL) were used.

instead of diethylcarbonate as a solvent also affected the intramolecular condensation of **9b** to give **4b** in 71% yield (entry 1). The yields of **4** were decreased by the electron-withdrawing groups at the *o*- or *p*-position of thiobutyrate, while the treatment of **9e** having an electron donor group at the *p*-position of the sulfur atom afforded **4e** in 70% yield (entries 2, 3 and 4).

The reaction mechanism to 4 from 9 was not clear, although the addition of dialkylcarbonate was required for improvement in these reactions. We deduced that the treatment of 9with alcoholate and dialkylcarbonate converted it to the malonate derivative **13**, which smoothly afforded the cyclization to give **4**. However, the role of dialkylcarbonate in the intramolecular cyclization also might be thought to the acceleration of dehydration of **16** (Scheme 3).

2.2. Large-scale preparation of 1 as a candidate for orally administered HIV-1 therapy

According to the new procedure for 2,3-dihydro-1-benzothiepines, we tried to develop a large-scale preparation of the desired 1 (Scheme 4). The biarylaldehyde 10f was selected as an intermediate, because the intramolecular condensation of 9c as a substrate afforded a lower yield, as mentioned. Although there were many syntheses of biaryl compounds, the preparation of **10f** was attempted using the convenient procedure of the Suzuki-Miyaura reaction in one-pot for large-scale preparation.^{11,12} The treatment of **17** was conducted with magnesium in THF under the refluxing condition, followed by cooling to -10 °C and boronation with trimethoxyborane, which was reacted with 10b, aqueous K₂CO₃, and a catalytic amount of Pd(PPh₃)₄ under the refluxing condition to give **10f** in excellent yield. The coupling reaction also proceeded smoothly when 0.05 mol% of Pd(OAc)₂ and 0.2 mol% of PPh₃ instead of Pd(PPh₃)₄ were employed. As a result, all reactions from the Grignard reaction of 17 to the cross-coupling reaction were performed in one-pot to give 10f almost quantitatively. After workup followed by extraction and concentration, 10f was used without further purification. The thioalkylation of crude 10f with 11a using K₂CO₃ in DMF as a base, followed





Scheme 4.

by cyclization with the combination of NaOEt in EtOH solution and diethyl carbonate at room temperature provided 1-benzothiepine 4g in one-pot. However, it was difficult to adopt these conditions for a large-scale preparation, because the reaction time was unstable in the thioalkylation when the reaction was scaled up to multi-decade grams. It was thought that the thioalkylation of 10f with 11a proceeded heterogenously. The use of DBU instead of K₂CO₃ afforded a stable reaction time, because the salt was dissolved in DMF. The reactions to 4g in two steps afforded 48% yield using only the crystallization procedure.

The preparation of **2** from **4g** was conducted using two methods. The hydrolysis of **4g** with 2 N NaOH gave carboxylic acid, which was oxidated with H_2O_2 in AcOH. However, the oxidation was not completed, giving **2** containing the sulfoxide. The oxidation from the sulfide **4g** to the sulfone **18** was accomplished in 94% yield by the reaction of 30% hydrogen peroxide in acetic acid. The sulfone **18** was hydrolyzed with aqueous K_2CO_3 solution in a mixture of THF and MeOH under the refluxing condition to give **2** in 95% yield. The hydrolysis using NaOH as a base provided some by-products, and the hydrolysis of **18** also was not completed under the acidic conditions.

Finally, the acid-chloride generated from **2** was amidated with the amine $3 \cdot 2$ HCl (**3** dihydrochloride)¹³ to give **1**. The reaction in *N*,*N*-dimethylacetamide (DMAc) proceeded smoothly, although the acid **2** was left in **1** when DMF or 1-methyl-2-pyrrolidone was used. After the reaction was completed, the addition of water to the reaction mixture caused crystallization to give the high-quality target product **1** in 82% yield.

3. Conclusion

We have developed a facile and novel synthetic preparation of 2,3-dihydro-1-benzothiepine derivatives **4**. The new compound, alkyl 4-(*o*-formylphenylthio)butyrate **9**, was prepared from *o*-halogenobenzaldehyde **10** and mercaptobutylate **11**. The improved intramolecular Claisen type reaction of **9** with alcoholate in dialkylcarbonate as a solvent provided **4** in good yields. Furthermore, the practical preparation of **1** as a candidate for orally administered HIV-1 therapy was accomplished by employing the new method, and did not require any chromatographic purification. This synthetic route was consisted of five steps in 35% yield, as compared with the previous nine steps.

4. Experimental

4.1. General

Melting points were recorded on a Büchi B-540 micro melting apparatus and were uncorrected. IR spectra were recorded on a Horiba FT-210 spectrophotometer. ¹H and ¹³C NMR spectra were recorded on a Bruker DPX-300 spectrometer. ¹H NMR spectra are reported as follows: chemical shifts in ppm (δ) downfield from tetramethylsilane as an internal standard, multiplicity (s, singlet; d, doublet; t, triplet and m, mutiplet), coupling constants spectra (Hz) and integration. ¹³C NMR spectra recorded in ppm (δ) relative to the central line for CDCl₃ at 77 ppm and DMSO-d₆ at 39.7 ppm. The column chromatography was performed on BW820 (Fuji Silysia Chemical Ltd) Elemental and HRMS analyses were performed at Takeda Analytical Research Laboratories, Ltd.

4.1.1. General procedure for the preparation of 9. Compound **11** (2 equiv) was added to a suspension of **10** (1.0 g) and K_2CO_3 (2 equiv) in DMF (3 v/w), and stirred overnight at room temperature. The reaction mixture was diluted in AcOEt, and washed successively with water, 0.5 N HCl, and brine. The organic layer was dried by Na_2SO_4 and concentrated in vacuo. The residue was purified by column chromatography on silica-gel with *n*-hexane–AcOEt.

4.1.2. Ethyl 4-(2-formylphenylthio)butyrate (9a), from 10a and 11a. A pale yellow oil; yield: 67%; ¹H NMR (300 MHz, CDCl₃) δ 1.26 (t, J=7.1 Hz, 3H), 1.95–2.07 (m, 2H), 2.49 (t, J=7.2 Hz, 2H), 3.02 (t, J=7.1 Hz, 2H), 4.13 (q, J=7.1 Hz, 2H), 7.27–7.34 (m, 1H), 7.45–7.53 (m, 2H), 7.83–7.86 (m, 1H), 10.38 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 14.2, 23.9, 32.4, 33.0, 60.5, 125.5, 128.3, 132.1, 134.0, 134.2, 141.3, 172.7, 191.4; IR (neat) 1731, 1695, 1196 cm⁻¹; HRMS (EI) calcd for C₁₃H₁₆O₃S ([M]⁺) 252.0820; Found 252.0820.

4.1.3. Methyl 4-(2-formylphenylthio)butyrate (9b), from 10a and 11b. A pale yellow oil; yield: 95%; ¹H NMR (300 MHz, CDCl₃) δ 1.96–2.06 (m, 2H), 2.50 (t, J=7.2 Hz, 2H), 3.01 (t, J=7.1 Hz, 2H), 3.68 (s, 3H), 7.27–7.34 (m, 1H), 7.46–7.55 (m, 2H), 7.81–7.84 (m, 1H), 10.36 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 23.8, 32.3, 32.7, 51.6, 125.5, 128.2, 132.1, 134.0, 134.1, 141.3, 173.1, 191.3; IR (neat) 1735, 1693, 1197 cm⁻¹; HRMS (EI) calcd for C₁₂H₁₄O₃S ([M]⁺) 238.0664; Found 238.0664.

4.1.4. Ethyl 4-(4-bromo-2-formylphenylthio)butyrate (9c), from 10b and 11a. A pale yellow solid; yield: 78%; ¹H NMR (300 MHz, CDCl₃): δ 1.26 (t, *J*=7.1 Hz, 3H), 1.95–2.05 (m, 2H), 2.48 (t, *J*=7.1 Hz, 2H), 3.00 (t, *J*=7.2 Hz, 2H), 4.13 (q, *J*=7.1 Hz, 2H), 7.35 (d, *J*=8.5 Hz, 1H), 7.60–7.64 (m, 1H), 7.94 (d, *J*=2.3 Hz, 1H), 10.32 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 14.2, 23.9, 32.7, 32.9, 60.6, 119.6, 130.4, 134.1, 135.5, 136.7, 140.3, 172.6, 189.8; IR (KBr) 1732, 1678, 1458, 1178 cm⁻¹; mp 48–49 °C; HRMS (FAB) calcd for C₁₃H₁₆O₃BrS ([MH]⁺) 331.0004; Found 331.0004. Anal. Calcd for C₁₃H₁₅O₃SBr: C, 47.14; H, 4.56; S, 9.68; Br, 24.12. Found: C, 47.19; H, 4.48; S, 9.58; Br, 24.23.

4.1.5. Ethyl 4-(2-chloro-6-formylphenylthio)butyrate (9d), from 10c and 11a. A pale yellow oil; yield: 70%; ¹H NMR (300 MHz, CDCl₃) δ 1.26 (t, *J*=7.2 Hz, 3H), 1.84–1.94 (m, 2H), 2.43 (t, *J*=7.2 Hz, 2H), 2.95 (t, *J*=7.2 Hz, 2H), 4.11 (q, *J*=7.2 Hz, 2H), 7.42 (dd, *J*=7.9, 7.2 Hz, 1H), 7.71 (d, *J*=7.9 Hz, 1H), 7.84 (d, *J*=7.7 Hz, 1H), 10.77 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 14.1, 24.7, 32.8, 36.0, 60.5, 126.9, 129.8, 135.0, 136.8, 140.2, 141.2, 172.5, 192.0; IR (neat) 1739, 1685, 1037 cm⁻¹; HRMS (FAB) calcd for C₁₃H₁₆O₃CIS ([MH]⁺) 287.0509; Found 287.0509.

4.1.6. Ethyl 4-(3,4-dimethoxy-6-formylphenylthio)butyrate (9e), from 10d and 11a. A pale yellow solid; yield: 27%; ¹H NMR (300 MHz, CDCl₃) δ 1.24 (t, J=7.1 Hz, 3H), 1.80–2.07 (m, 2H), 2.43 (t, J=7.1 Hz, 2H), 2.92 (t, J=7.4 Hz, 2H), 3.93 (s, 3H), 3.99 (s, 3H), 4.12 (q, J=7.1 Hz, 2H), 7.02 (s, 1H),7.41 (s, 1H), 10.48 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 13.6, 23.9, 32.2, 35.1, 55.5, 55.8, 60.0, 110.0, 114.4, 129.2, 133.3, 148.3, 153.3, 172.1, 189.7; IR (nujol) 1718, 1671, 1272 cm⁻¹; mp 87–88 °C; HRMS (FAB) calcd for C₁₅H₂₀O₅S ([M]⁺) 312.1031; Found 312.1031. Anal. Calcd for C₁₅H₂₀O₅S: C, 57.67; H, 6.45; S, 10.26; O, 25.61. Found: C, 57.50; H, 6.47; S, 10.37.

General procedure for the preparation of **4**. 20% NaOEt (1.2 equiv) in EtOH (or 28% NaOMe in MeOH) was added to a solution of **9** (1.0 g) and diethyl carbonate (20 v/w) (or dimethyl carbonate), and stirred at room temperature. After cooling to 0 °C, the reaction mixture was neutralized with 1 N HCl. The resulting solution was extracted with AcOEt, and the organic layer was washed with water, dried by Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography on silica-gel with *n*-hexane–AcOEt.

4.1.7. Ethyl 2,3-dihydro-1-benzothiepin-4-carboxylate (**4a**). A yellow oil; yield: 71%; ¹H NMR (300 MHz, CDCl₃) δ 1.35 (t, J=7.1 Hz, 3H), 2.96–3.00 (m, 2H), 3.16–3.21 (m, 2H), 4.28 (q, J=7.1 Hz, 2H), 7.18–7.27 (m, 2H), 7.36–7.48 (m, 2H), 7.81 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 14.2, 32.8, 35.3, 61.1, 127.0, 128.5, 132.2, 133.4, 134.4, 136.8, 138.8, 139.4, 168.1; IR (neat) 1703, 1267, 1240 cm⁻¹; HRMS (EI) calcd for C₁₃H₁₄O₂S ([M]⁺) 234.0715; Found 234.0715.

4.1.8. Methyl 2,3-dihydro-1-benzothiepin-4-carboxylate (4b). A white solid; yield: 71%; ¹H NMR (300 MHz, CDCl₃) δ 2.96–3.00 (m, 2H), 3.16–3.21 (m, 2H), 3.83 (s, 3H), 7.18–7.27 (m, 2H), 7.36–7.48 (m, 2H), 7.81 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 32.9, 35.2, 52.2, 127.0, 128.6, 132.2, 133.0, 134.5, 136.7, 138.8, 139.7, 168.5; IR (KBr) 1706, 1631, 1428 cm⁻¹; mp 52–54.3 °C; HRMS (FAB) calcd for C₁₂H₁₂O₂S ([M]⁺) 220.0558; Found 220.0558. Anal. Calcd for C₁₂H₁₂O₂S: C, 65.43; H, 5.49; S, 14.56. Found: C, 65.57; H, 5.43; F, 14.32.

4.1.9. Ethyl 7-bromo-2,3-dihydro-1-benzothiepin-4-carboxylate (4c). A yellow solid; yield: 32%; ¹H NMR (300 MHz, CDCl₃) δ 1.35 (t, J=7.1 Hz, 3H), 2.95–2.99 (m, 2H), 3.15–3.20 (m, 2H), 4.28 (q, J=7.1 Hz, 2H), 7.26– 7.34 (m, 2H), 7.51–7.52 (m, 1H), 7.69 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 14.7, 33.1, 35.8, 61.7, 121.1, 131.6, 134.1, 135.3, 136.9, 138.2, 138.3, 139.2, 168.0; IR (KBr) 1704, 1629, 1465 cm⁻¹; mp 82–83 °C; HRMS (FAB) calcd for C₁₃H₁₃O₂SBr ([M]⁺) 311.9820; Found 311.9820. Anal. Calcd for C₁₃H₁₃O₂SBr: C, 49.85; H, 4.18; S, 10.24; Br, 25.51. Found: C, 50.13; H, 4.12; S, 10.11; Br, 25.55.

4.1.10. Ethyl 9-chloro-2,3-dihydro-1-benzothiepin-4-carboxylate (4d). A yellow oil; yield: 46%; ¹H NMR (300 MHz, CDCl₃) δ 1.35 (t, *J*=7.1 Hz, 3H), 2.99–3.03 (m, 2H), 3.15–3.20 (m, 2H), 4.28 (q, *J*=7.1 Hz, 2H), 7.12– 7.27 (m, 1H), 7.26–7.36 (m, 2H), 7.77 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 14.3, 33.2, 34.3, 61.2, 126.9, 129.3, 133.5, 134.1, 135.6, 137.9, 138.6, 138.9, 167.8; IR (neat) 1712, 1631 cm⁻¹. HRMS (FAB) calcd for C₁₃H₁₃O₂ClS ([M]⁺) 268.0325; Found 268.0325.

4.1.11. Ethyl 7,8-dimethoxy-2,3-dihydro-1-benzothiepin-4-carboxylate (4e). A yellow oil; yield: 70%; ¹H NMR (300 MHz, CDCl₃) δ 1.35 (t, *J*=7.1 Hz, 3H), 2.95–2.98 (m, 2H), 3.16–3.21 (m, 2H), 3.88 (s, 3H), 3.89 (s, 3H), 4.28 (q, *J*=7.1 Hz, 2H), 6.86 (s, 1H), 6.96 (s, 1H), 7.74 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 14.3, 32.8, 35.7, 55.9, 60.9, 114.7, 116.6, 129.4, 130.9, 131.8, 139.2, 147.8, 148.8, 168.1; IR (neat) 1700, 1594, 1502 cm⁻¹; HRMS (FAB) calcd for C₁₅H₁₈O₄S ([M]⁺) 294.0926; Found 294.0926.

4.1.12. Ethyl 7-(4-propoxyphenyl)-2,3-dihydro-1-benzothiepin-4-carboxylate (4g). Under an argon atmosphere, a solution of 17 (37.1 g, 172.4 mmol) and THF (90 mL) was added dropwise to a suspension of magnesium (4.3 g, 177.3 mmol) and THF (270 mL) under a refluxing condition, and the whole was stirred for 1.5 h under the same conditions. After cooling to -11 °C, a solution of trimethoxyborane (17.9 g, 172.4 mmol) and THF (90 mL) was added dropwise to the reaction mixture and stirred for 1 h at -10 °C. After warming to room temperature, palladium (II) acetate (11 mg, 0.049 mmol) and triphenylphosphine (52 mg, 0.197 mmol) were added to the resulting mixture, and stirred for 30 min at room temperature. Compound **10b** (20.0 g, 98.5 mmol), K₂CO₃ (71.5 g, 517.2 mmol) and distilled water (85 mL) were added to the resulting mixture, and the whole was refluxed for 4 h. After cooling to room temperature, 2 N HCl (450 mL) was added dropwise to the reaction mixture at 20-30 °C and separated. The aqueous solution was extracted with toluene (450 mL), and the combined organic solution was washed successively with 2 N HCl (300 mL), 2 N NaOH (300 mL×2), 2 N HCl (300 mL) and 20% NaCl solution $(300 \text{ mL} \times 2)$. Activated charcoal (1.0 g) was added to the organic solution, and the mixture was stirred for 20 min at room temperature. The charcoal was filtered off and washed with toluene (50 mL). The filtrate and washing were concentrated in vacuo to give crude 2-fluoro-5-(4-propoxyphenyl)benzaldehyde (10f, 30.5 g) as a brown oil. An analytically pure sample of 10f was obtained by chromatography on silica-gel as a white solid; ¹H NMR (300 MHz, CDCl₃) δ 1.06 (t, J=7.4 Hz, 3H), 1.77–1.90 (m, 2H), 3.96 (t, J=6.6 Hz, 2H), 6.95–6.98 (m, 2H), 7.18–7.25 (m, 1H), 7.46-7.50 (m, 2H), 7.74-7.78 (m, 1H), 8.00-8.04 (m, 1H), 10.41 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 10.5, 22.6, 70.0, 115.0, 116.7, 117.0, 124.1, 126.3, 128, 131.1, 134.3, 137.8, 159.2, 162.1, 165.6, 187.2; IR (KBr) 2875, 1691, 1608, 1484, 1216 cm⁻¹; mp 42–43 °C; HRMS (FAB) calcd for $C_{16}H_{15}O_2F$ ([M]⁺) 258.1056; Found 258.1056. Anal. Calcd for C₁₆H₁₅O₂F: C, 74.40; H, 5.85; F, 7.36. Found: C, 74.66; H, 5.67; F, 7.28.

DBU (34.4 mL, 229.9 mmol) was added dropwise to a solution of the above crude 10f and 11a (32.6 mL, 229.9 mmol) in DMF (59 mL) at 0-10 °C under an argon atmosphere, and the whole was stirred for 1 h at 20-30 °C. After diethyl carbonate (590 mL) was added the reaction mixture, 20% NaOEt in EtOH (156.0 g, 459.8 mmol) was added dropwise to the resulting mixture, and the whole was stirred for 3 h at 20-30 °C. After cooling to 5 °C, 2 N HCl (338 mL) was added dropwise to the reaction mixture, and separated. The aqueous solution was extracted with AcOEt (290 mL), and the combined organic layer was washed successively with water (300 mL), 5% NaHCO₃ solution (300 mL) and 5% NaCl solution (300 mL). Activated charcoal (3.5 g) and tri-n-butylphosphine (4 mL) were added to the organic solution, and the mixture was stirred for 20 min. The charcoal was filtered off, and washed with AcOEt (60 mL). After the filtrate and washing was concentrated in vacuo, the resulting residue was dissolved in diisopropyl ether (60 mL) under a refluxing condition. After cooling to room temperature, stirring for 1.5 h at room temperature and stirring for 2 h at 0 °C, the resulting crystals were collected by filtration, washed with cold diisopropyl ether (60 mL) and dried in vacuo to give 4g (20.3 g, yield, 48% based on **10b**) as a pale yellow solid. ¹H NMR (CDCl₃) δ 1.06 (t, J=7.4 Hz, 3H), 1.36 (t, J=7.1 Hz, 3H), 1.77–1.90 (m, 2H), 3.00 (t, J=5.3 Hz, 2H), 3.22 (t, J=5.6 Hz, 2H), 3.96 (t, J = 6.6 Hz, 2H), 4.29 (q, J = 7.1 Hz, 2H), 6.94-7.00(m, 2H), 7.36–7.57 (m, 5H), 7.87 (s, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 10.5, 14.4, 22.6, 32.7, 35.6, 61.1, 69.6, 114.9, 126.7, 127.9, 132.0, 132.5, 132.7, 133.6, 136.7, 137.2, 139.6, 139.7, 159.1, 168.1; IR (KBr) 2927, 1704, 1240, 819 cm⁻¹; mp 87–88 °C; HRMS (FAB) calcd for $C_{22}H_{24}O_3S\left([M]^+\right)$ 368.1446; Found 368.1446. Anal. Calcd for $C_{22}H_{24}O_3S$: C, 71.71; H, 6.56; S, 8.70. Found: C, 71.72; H, 6.77; S, 8.64.

4.1.13. Ethyl 7-(4-propoxyphenyl)-1,1-dioxo-2,3-dihydro-1-benzothiepin-4-carboxylate (18). After 4g (15.0 g, 40.7 mmol) was dissolved in acetic acid (135 mL) at 56 °C, 30% H₂O₂ (9.5 g, 83.5 mmol) in acetic acid (15 mL) was added dropwise to a solution at the same temperature and stirred for 3 h at 65-70 °C. Water (15 mL) and 5% Na₂SO₃ solution (60 mL) were added dropwise to the reaction mixture, and then the whole was cooled to room temperature and stirred for 2 h at the same temperature. The resulting crystals were collected by filtration, washed successively with acetic acid/water (3/2, 15 mL) and water (150 mL), and dried in vacuo to give 18 (15.4 g, yield, 94%) as a white solid. ¹H NMR (300 MHz, CDCl₃) δ 1.06 (t, J=7.4 Hz, 3H), 1.38 (t, J = 7.2 Hz, 3H), 1.78–1.90 (m, 2H), 3.14 (t, J =6.3 Hz, 2H), 3.64 (t, J=7.1 Hz, 2H), 3.98 (t, J=6.6 Hz, 2H), 4.32 (q, J=7.2 Hz, 2H), 6.99–7.02 (m, 2H), 7.53–7.57 (m, 2H), 7.65–7.69 (m, 2H), 7.89 (s, 1H), 8.18 (d, J =8.0 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 10.5, 14.3, 22.6, 25.0, 55.8, 61.7, 69.7, 115.2, 126.7, 127.9, 128.4, 130.5, 131.8, 132.6, 132.9, 137.9, 138.3, 146.1, 160.0, 166.6; IR (KBr) 2933, 1706, 1606, 1517, 1243 cm⁻¹; mp 139– 140 °C; HRMS (FAB) calcd for $C_{22}H_{24}O_5S$ ([M]⁺) 400.1344; Found 400.1344. Anal. Calcd for C₂₂H₂₄O₅S: C, 65.89; H, 6.04; S, 8.01. Found: C, 65.72; H, 5.87; S, 7.96.

4.1.14. 7-(4-Propoxyphenyl)-1,1-dioxo-2,3-dihydro-1benzothiepin-4-carboxylic acid (2). A solution of K₂CO₃ (342 g, 2.47 mol) and water (4.2 L) was added dropwise to a solution of 18 (495 g, 1.24 mol), THF (4.95 L) and MeOH (2.48 L), and the whole was refluxed for 6.5 h. Under the same condition, 3 N HCl (1.85 L) was added dropwise to the reaction mixture and the whole was cooled to room temperature. Under the same temperature, 6 N HCl (84 mL) was added dropwise to the resulting mixture and stirred for 1 h with ice-bathing. The resulting crystals were collected by filtration, washed with THF/MeOH/water (1/1/3, 2.97 L), and dried in vacuo to give 2 (443 g, yield, 96%) as a light white-yellow solid. ¹H NMR (300 MHz, DMSO- d_6) δ 0.98 (t, J=7.4 Hz, 3H), 1.70–1.78 (m, 2H), 2.95 (t, J = 6.2 Hz, 2H), 3.73 (t, J = 6.4 Hz, 2H), 3.98 (t, J =6.5 Hz, 2H), 7.05 (d, J=8.7 Hz, 2H), 7.75 (d, J=8.7 Hz, 2H), 7.86–7.88 (m, 2H), 8.02–8.05 (m, 2H); ¹³C NMR (75 MHz, DMSO-*d*₆) δ 10.6, 22.2, 25.4, 53.9, 69.3, 115.3, 126.4, 127.1, 128.6, 129.9, 132.1, 132.4, 134.0, 136.8, 138.8, 144.9, 160.0, 168.5; IR (KBr) 3500, 1673, 1608, 1519, 1294, 1251 cm⁻¹; mp 271–272 °C; HRMS (FAB) calcd for C₂₀H₂₀O₅S ([M]⁺) 372.1031; Found 372.1031. Anal. Calcd for C₂₀H₂₀O₅S: C, 64.50; H, 5.41; S, 8.61. Found: C, 64.40; H, 5.47; S, 8.55.

4.1.15. *N*-[**4**-[*N*-**Methyl**-*N*-(**tetrahydropyran-4-yl**)**aminomethyl**]**phenyl**]-**7**-(**4**-**propoxyphenyl**)-**1**,**1**-**dioxo-2**,**3dihydro-1-benzothiepin-4-carboxamide** (1). Thionyl chloride (0.70 g, 5.91 mmol) was added dropwise to a suspension of **2** (2.0 g, 5.37 mmol) in *N*,*N*-dimethylacetoamide (10 mL) at room temperature, and the whole was stirred for 2 h at room temperature. The reaction mixture was added dropwise to a suspension of **3**·2HCl (1.9 g, 6.44 mmol) and NEt₃ (5.8 mL, 41.87 mmol) in N,N-dimethylacetamide (10 mL) at 0-10 °C, and stirred for 1 h at room temperature. Water (20 mL) was added to the resulting mixture at room temperature and stirred for about 1 h at the same temperature. The resulting crystals were collected by filtration, washed with water (5 mL) and MeOH (5 mL), and dried in vacuo to give 1 (2.5 g, yield, 82%) as a white solid. ¹H NMR (300 MHz, CDCl₃) δ 1.06 (t, J=7.4 Hz, 3H), 1.64–1.88 (m, 6H), 2.20 (s, 3H), 2.60– 2.67 (m, 1H), 3.13 (t, J=6.6 Hz, 2H), 3.33–3.41 (m, 2H), 3.56 (s, 2H), 3.69 (t, J = 6.3 Hz, 2H), 3.95-4.07 (m, 4H), 6.98 (d, J = 8.7 Hz, 2H), 7.29–7.34 (m, 3H, 7.48–7.61 (m, 6H), 8.08–8.16 (m, 2H); 13 C NMR (75 MHz, CDCl₃) δ 10.5, 22.6, 25.0, 29.2, 37.5, 57.3, 57.5, 59.6, 67.7, 69.7, 115.2, 120.3, 126.3, 128.1, 128.4, 129.4, 130.4, 130.6, 132.0, 133.2, 136.4, 136.5, 137.3, 139.3, 146.1, 160.1, 166.6; IR (KBr) 3485, 2948, 1654, 1635, 1606, 1519, 1315, 1292, 1130 cm⁻¹; mp 239.6–240.8 °C; HRMS (FAB) calcd for $C_{33}H_{39}N_2O_5S$ ([MH]⁺) 575.2580; Found 575.2580. Anal. Calcd for C33H38N2O5S; C, 68.96; H, 6.66; N, 4.87; S, 5.58. Found: C, 68.79; H, 6.56; N, 4.95; S, 5.66.

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Tetrahedron 60 (2004) 10859-10868

Synthesis of polyhydroxy cavitands and intramolecular inclusion of their octaester derivatives

Eric Efrain Dueno and Kirpal S. Bisht*

Department of Chemistry, University of South Florida, 4202 East Fowler Avenue, Tampa, FL 33620, USA

Received 21 May 2004; revised 14 September 2004; accepted 15 September 2004

Available online 8 October 2004

Abstract—A facile and efficient synthesis of novel cavitands containing eight hydroxyl groups was accomplished in eight steps beginning from commercially available resorcinol and the corresponding aldehydes. The synthesis combines two classical approaches to cavitand chemistry and yields the target octol cavitand molecules in gram quantities with no chromatographic separations. A variety of octaester cavitand derivatives were then prepared from the parent octols and their spectral properties are reported. © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

The acid-catalyzed condensation of resorcinol and aldehydes produces resorcinarenes such as compound 1 (Fig. 1). Resorcinarenes are known to posses hydrophilic (upper rim) and hydrophobic (lower rim) regions and a cavity, which can accommodate small organic molecules.¹ Cavitands of the general structure 2 (Fig. 1) are synthesized from resorcinarenes via bridging reactions of the hydroxyl groups.² The bridges serve to impart conformational rigidity to the existing aryl skeleton and form the concave cavity of the cavitand.³ As a result of these structural motifs resorcinarenes and cavitands have enjoyed use as surfactants and liquid crystals, in the complexation of metals, ammonium compounds, alcohols, diols, sugars, amino acids and carboxylic acids.^{2b} The introduction of functional groups onto cavitand scaffolds has been the focus of intensive research, to elucidate various structural features that provide valuable insight into factors governing host-guest chemistry.^{2d} Functionalized cavitand substrates offer interesting molecular platforms for metal ligand exchange complexes,^{2,4} polymeric materials,⁵ self-assembled systems,^{2d} and as prospective reaction sites, and unique catalytic platforms.^{2b,d} The availability of such material will be useful in synthesis of a wide array of novel materials and applications such as metal complexing agents,⁶ sensors,⁷ water soluble cavitands,⁸ in phase transfer extraction of heavy metals,⁹ for complexation of fullerenes¹⁰ and related molecules,^{11,12} for new bioorganic and biomimetic chemistries,¹³ for self assembled systems,¹⁴ for cavitand based crown ethers,¹⁵ as stationary phases,¹⁶ and as novel cavitand polymers.^{5,17}

However, functionalized cavitands have largely been unexplored because of their unavailability stemming from the difficult synthesis. The main difficulties associated with the functionalization of cavitands are solubility related, which are manifested in two ways. First, the poor solubility of cavitand compounds leads to problems in chromatographic separations as tedious column separations, render lengthy synthetic sequences cumbersome. Second, the solubility of cavitands in reaction media is limited and thus reaction yields are not uniformly high.^{2c} One method for the introduction of functional groups onto cavitands is a stepwise manner^{2c} beginning from tetrabromocavitands such as compound 2 in (Fig. 1). The bromo atoms are substituted via lithium exchange and subsequent quenching with an appropriate electrophilic reagent introduces hydroxyl, thiol, or carboxylic ester functionalities respectively.^{2d} An alternative synthetic method for the introduction of functional groups onto cavitands is known as the modular approach, which consists of coupling reactions of resorcinarenes with a suitable bridging component and then further synthetic elaboration on the bridging component to introduce the functional moieties.^{2c}

In our laboratories, we are interested in the design and synthesis of polyhydroxy cavitands with well-defined structures.^{18a,b} We sought to blend the stepwise and modular approaches to create polyhydroxy cavitands **3** and **4**, that are reminiscent of their parent compounds the resorcinarenes, and to which various functional groups can be appended through phosphorylation, sulfonylation, acylation, alkylation, halogenation, and oxidation reactions on the eight hydroxyl groups. The combination of the synthetic

Keywords: Cavitands; Functionalization; Polyhydroxy cavitands; Octaester cavitands.

^{*} Corresponding author. Tel.: +1 8139740350; fax: +1 8139743203; e-mail: kbisht@cas.usf.edu

^{0040–4020/\$ -} see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.042



Figure 1. Resorcinarenes, cavitands, polyhydroxy cavitands.

approaches towards synthesis of functionalized cavitands proved highly effective in our hands. We, thus, secured the target molecules in gram quantities while performing no chromatographic separations throughout the entire synthetic sequence. We subsequently synthesized several octaester derivatives of the octols **3** and **4**, including acetate, mesylate, isopropionate, *p*-anisoate, and *p*-phenylbenzoate in excellent yields. In this article we report on a simple and efficient synthesis of well defined polyhydroxy cavitands, their derivatives, and spectral properties.

2. Results and discussion

Our studies began by utilizing heptaldehyde or acetaldehyde, as the two alkyl aldehydic components for the condensation reactions with resorcinol to produce the resorcinarenes.¹ Aromatic bromination reactions resulted in the formation of the tetrabromides which served to introduce a suitable functionality for the stepwise approach.³ Cavitand formation using Kaifer's sealed tube method on the brominated resorcinarenes yielded the desired tetrabromide tetramethylene bridged cavitands.¹⁹ The aryl bromide positions were further functionalized to the esters and subsequently reduced to give the tetrol cavitands.²⁰ Further functionization of the tetrol was conceived through a modular coupling approach using 2,2-bis(phenyldioxymethyl)propionic acid,²¹ a well estab-lished synthetic building block in the synthesis of macromolecules.²² However, this proved difficult, as we screened several DCC and DMAP mediated coupling, procedures. Additionally, the reactions mixtures were complicated by the formation of urea, which presented difficulty in the purification, and identification of the product. However, employing a Mitsunobu protocol, using a triphenylphosphine and DIAD mediated coupling in THF, we were able to isolate the product in quantitative yield by precipitation of the reaction mixture into cold methanol. Moreover, we envisioned that catalytic hydrogenolysis to remove the protected alcohols would secure the target molecules (Scheme 1). Thus, we coupled the tetrol

cavitands with a benzyl ketal protected acid, and subsequently removed the benzyl-protecting group to give the polyhydroxy cavitands 3 and 4 in excellent yields with no need of column chromatography.

2.1. Synthesis of octol cavitands 3 and 4

Synthesis of octol cavitand **3** (Scheme 1) began from acid catalyzed condensation of resorcinol **5** and heptaldehyde **6** in equal molar quantities in a refluxing solution of 95% (EtOH/H₂O), which afforded resorcinarene 8 in 96% yield. Subsequent aromatic bromination using N-bromosuccinamide in a solution of 2-butanone gave tetrabromide 10 in 75% yield. Cs₂CO₃ catalyzed cavitand formation with bromochloromethane as the bridging reagent in 1:1 DMF/ DMSO using a sealed tube reaction vessel afforded after recrystallization from CH₂Cl₂/hexanes bromo-substituted cavitand 12 in 89% yield. (CAUTION: the reaction vessel must be able to tolerate an internal pressure of several atmospheres). Metal-halogen exchange between bromo cavitand 12 and *n*-butyllithum and subsequent nucleophilic attack on ethylchloroformate afforded tetraester 14 in 80% yield, after recrystallization from 80% EtOAc/EtOH. Reduction of the tetraester 14 using lithium aluminum hydride afforded tetrol 16 in 85% yield. Mitsunobu esterification protocol of tetrol 16 with 2,2-bis(phenyldioxymethyl)propionic acid 18 afforded tetrabenzylketal 19 in 96% yield. Palladium catalyzed hydrogenation of tetrabenzylketal **19** afforded octol cavitand **3** in 94% yield. Using the same protocol we synthesized octol cavitand 4 from resorcinol 5 and acetaldehyde 7 (Scheme 1). We confirmed the structure of 3 and 4 by 1 H NMR, 13 C NMR, MALDI analysis, and ¹H–¹H COSY.

2.2. Synthesis of octaester cavitands 21 and 22

In order to test the viability of the octols 3 and 4 to further modification utilizing esterification protocols, we synthesized octaacetate cavitands 21 and 22 (Scheme 2) upon reaction of cavitands 3 and 4, respectively, with acetic anhydride in dry THF at rt using triethylamine and a



Scheme 1. Synthesis of polyhydroxy cavitands 3 and 4. Reagents and conditions: (a) 95% EtOH/H₂O, HCl, reflux, 8=96%, 9=82%. (b) NBS, 2-butanone, rt, 10=75%, 11=71%. (c) BrCH₂Cl, Cs₂CO₃, 1:1 DMF/DMSO, 88 °C, sealed tube, 12=89%, 13=94%. (d) *n*-BuLi, ethylchloroformate, THF, -78 °C, 14=80%, 15=89%. (e) LiAlH₄, THF, rt, 16=85%, 17=87%. (f) 18, triphenylphosphine, DIAD, THF, rt, 19=96%, 20=98%. (g) Pd/C, H₂, EtOAc, THF, 3=94%, 4=95%. Yields correspond to pure compounds.

catalytic amount of DMAP. Upon completion of the reaction, removal of the solvent under reduced pressure gave a residue, which was partitioned between methylene chloride and water. Evaporation of the methylene chloride yielded **21** in 98% and **22** in 97% as white solids.

2.3. ¹H NMR spectra of 21 and 22

Before recording spectra we ensured that the samples were free of solvents used during the synthesis and crystallization such as DMSO, MeOH, CH_2Cl_2 , H_2O , etc. This was accomplished by repeatedly dissolving the compound in solutions of dry THF and dry toluene and then distillation of the solvent through a vigreux column followed by drying under heated vacuum overnight. The ¹H NMR spectrum of **21** recorded in CDCl₃ at 20 °C (Fig. 2) showed a singlet corresponding to the four aromatic protons (H₁) of the resorcinarene rings at 7.2 ppm. A double doublet characteristic of methylene-bridged protons³ is observed at 5.7 ppm (four outer protons, H₂), and at 4.1 ppm (four inner protons, H₃). A 8H singlet corresponding to the methylene protons (H₄) adjacent to the resorcinarene rings at 5.0 ppm, a triplet at 4.7 ppm corresponding to four benzylic protons (H_7) , and a singlet corresponding to the methylene groups (H_6) adjacent to the prochiral center at 4.0 ppm were also observed. Further upfield, were three separate resonances for the alkyl chains; a quartet for the methylene groups (H₈) at 2.2 ppm, a broad multiplet at 1.5–1.2 ppm for the methylene groups (H_9-H_{12}) , and a triplet at 0.88 ppm for the terminal methyl groups (H_{13}) . Also observed was a 12H singlet corresponding to the methyl groups (H₅) adjacent to the prochiral center at 1.1 ppm. Interestingly, acetate group resonances that would have been consistent with literature at around 2.1 ppm were not present in the ¹H NMR spectrum of **21**.²³ Similarly, the ¹H NMR spectrum of **22** recorded in CDCl₃ at 20 °C (Fig. 2) showed proton resonances at expected chemical shift values but did not exhibit acetate signals at chemical shift values consistent with literature.²³ Instead, a signal upfield at 1.5 ppm that integrated to twenty-four protons was observed for the acetate groups. In ¹H NMR spectra of **21** and **22**, the upfield signal shifts in the acetate methyl resonances are characteristic of aromatic shielding and we suspected that intramolecular inclusion of the acetate groups within the



Scheme 2. Synthesis of octaacetate cavitands 21 and 22. Reagents and conditions: (a) acetic anhydride, TEA, DMAP, THF, rt, 21=98%, 22=97%. Yields correspond to pure compounds.



Figure 2. ¹H NMR spectra of octaacetates 21 and 22 in $CDCl_3$ at 20 °C at 360 MHz.

cavitand cavity was responsible. The inclusion is believed to be intramolecular as evidenced by the fact that ¹H NMR spectra recorded at 5, 10, 15, 20 and 25 mM concentrations resulted in no change of the spectra. Additionally, ¹H NMR spectral data of **21** and **22** recorded in deuterated solvents of different polarity, viz., benzene- d_6 , 1,2-dichlorobenzene- d_4 , dimethylsulfoxide- d_6 , and methanol- d_4 , resulted in no change of the spectrum.

2.4. Variable temperature ¹H NMR spectra of 21 and 22

The ¹H NMR spectra of **21**, recorded at different probe temperatures in $CDCl_3$ at 360 MHz (Fig. 3), showed a

downfield shift of 0.7 ppm (from 1.5 to 2.2 ppm) in acetate methyl resonance upon cooling from 20 to -50 °C for octaacetate **21**. Similarly, the spectra of **22** also show a downfield field shift for the acetate resonances upon cooling to -50 °C (Fig. 3). This suggests that at 20 °C there is a higher percentage of the 'bound' species present, however exchange is fast on the NMR timescale and an averaged signal is observed at 1.5 ppm. At intermediate temperature, i.e. -20 °C broad signals are observed, which correspond to both the free and bound species being present and in intermediate-rate exchange. At -50 °C, the acetyl groups are mostly uncomplexed and a higher percentage of the 'free' species is favored giving rise to a single signal at 2.2 ppm.

Importantly, when octaacetate **21** was heated in DMSO- d_6 , inside the 5 mm probe of a 360 MHz spectrometer, a sharpening of the acetate signals at 1.5 ppm was observed in the temperature range of 300–380 K, which arises from a higher rate of exchange between the bound and the free species at higher temperatures (Fig. 4).

2.5. Synthesis of octaester cavitands 23-26

In order to examine the intramolecular inclusion phenomenon further, we treated the octol cavitand **3** with a series of electrophiles that afforded the octaester cavitands **23–26** (Scheme 3). We selected octol **3** as the starting material due to its higher solubility in organic solvents than octol **4**, which we attribute to increased chain length, i.e. hexyl chains versus methyl chains. Acyl halides were of different size and steric bulk, including mesyl chloride (as the sulfur analogue of the acetates) and isobutyryl chloride. Additionally we examined aromatic esters of *p*-anisoyl chloride and *p*-biphenylcarbonyl chloride. The reactions were performed in dry THF at rt using triethylamine and catalytic amount of DMAP. Each of the electrophiles reacted quantitatively and afforded the desired products with no need of column chromatography.



Figure 3. Variable temperature ¹H NMR spectra of octaacetates 21 and 22 in CDCl₃ at 360 MHz.



Figure 4. Parts of the variable temperature ¹H NMR spectra of octaacetate 21 in DMSO at 360 MHz.

2.6. ¹H NMR spectra of 23 and 24

Similar to octaacetates **21** and **22**, the ¹H NMR spectrum of octamesylate **23** recorded in CDCl₃ at 360 MHz showed

most resonances at expected chemical shift values²³ however, the mesylate methyl signal was shifted upfield at 1.5 ppm. Variable temperature ¹H NMR experiments on 23 revealed a downfield shift in the mesylate resonance upon cooling from 20 to -50 °C. In contrast, the octaisopropionate 24 (Fig. 5) showed most resonances at expected chemical shift values²³ and only a small upfield resonance shift ($\Delta \delta = 0.07$) to 1.06 ppm for the methyl protons in the isopropyl esters was observed, compared to literature value of 1.13 ppm. Interestingly, the isopropyl methyls were observed in a double doublet-splitting pattern (Fig. 6), suggesting chemical non-equivalence of the two-methyl groups. Although the upfield ¹H NMR shift is fairly small excluding total encapsulation of the isopropyl group, however, influence of the aromatic cavity such that the methyl groups are oriented towards and away from it cannot be discounted for the non-equivalence of the isopropyl groups.

2.7. Variable temperature ¹H NMR of 24

During acquisition of the ¹H NMR spectra of **24**, cooling from 20 to -20 °C (Fig. 6), the isopropyl group signals began to merge while shifting downfield. The merging and



Scheme 3. Synthesis of octaester cavitands 23–26. Reagents and conditions: (a) mesyl chloride, TEA, DMAP, THF, rt, 23=98%. (b) Isobutyryl chloride, TEA, DMAP, THF, rt, 24,=98%. (c) *p*-Anisoyl chloride, TEA, DMAP, THF, rt, 25=97%. (d) *p*-Biphenylcarbonyl chloride, TEA, DMAP, rt, 26=95%. Yields correspond to pure compounds.



Figure 5. ¹H NMR spectra of octaisopropionate 24 in CDCl₃ at 500 MHz.

shifting of the signals at lower temperatures was consistent with our previous findings for the octaesters **21**, **22**, and **23** although not as pronounced, which we attribute to the larger size of the isopropyl moiety.

2.8. ¹H NMR spectra of 25 and 26

Figure 7 shows the spectrum of octaanisoate 25 here we observed all resonances at expected values²³ with no intramolecular inclusion of the methoxy signal. Finally, in octaphenylbenzoate compound 26 interactions of the cavitand cavity and the phenylbenzoate group was not observed. From these results one can glean certain information about the intramolecular inclusion phenomenon in these novel cavitand esters. It is apparent from the spectral data obtained in this study that the intramolecular inclusion is directly related to size of the ester moiety or 'guest'. When the ester moiety is 'small' such as the acetate and mesylate, the cavitand is able to accommodate the guest. The cavity contains aromatic rings that effectively shield the acetate signals causing upfield shifts in the ¹H NMR spectra. When one substitutes a larger ester group such as the isopropyl, the intramolecular inclusion is only



Figure 6. Parts of the variable temperature ¹H NMR spectra of octaisopropionate **24** in CDCl₃ at 500 MHz.



Figure 7. ¹H NMR spectra of octaanisoate 25 in CDCl₃ at 360 MHz.

partial as a direct result the larger size of the ester moiety. When the ester size is increased to the aromatic system no inclusion was observed within our detection limits.

3. Conclusion and outlook

We have demonstrated a highly efficient and facile methodology for the synthesis of polyhydroxy functionalized cavitands and their octaester derivatives. The synthetic strategy relied upon the combination of classical methods of stepwise and modular approaches for the synthesis of the target molecules, which proceeded in excellent yield without the need of column chromatography. ¹H NMR experiments showed that intramolecular inclusion of the ester moieties is directly related to their size and steric bulk. Increasing the steric bulk or size of the ester moiety resulted in partial or no intramolecular inclusion. Such cavitands offer unique molecular platforms for host–guest chemistries, sensor development, metal complexation, as well as new polymers and self-assembled systems, and as potential reaction sites, and novel catalytic platforms.

4. Experimental

4.1. General

Synthesis of the requisite starting tetrol cavitand **17** has been reported elsewhere.^{3,19,20} All reactions were carried out under a nitrogen atmosphere in flame-dried glassware.

Commercially available reagents were purchased from Acros Chemical Co. or Aldrich Chemical Co. and were used without further purification. Cs₂CO₃ was dried by heating to 250 °C for 24 h and then cooled to ambient temperature in a vacuum desiccator. *N*-bromosuccinimide (NBS) was recrystallized from boiling water 1 g to 100 mL. Triphenylphosphine was recrystallized from boiling MeOH. 2,2-Bis(phenyldioxymethyl)propionic acid **18** was prepared from 2,2-bis(hydroxymethyl)propionic acid and benzaldehyde dimethyl acetal according to literature procedures.²⁰ Triethylamine was distilled from potassium hydroxide under a nitrogen atmosphere and stored over potassium hydroxide. DMF and DMSO were stored over molecular sieves (3 \AA) for 24 h and degassed prior to use. Tetrahydrofuran was distilled from sodium benzophenone ketyl under a nitrogen atmosphere.

Thin layer chromatography was performed using Whatman silica gel 60 Å with fluorescent indicator on 250-micrometer thickness glass backed with hexanes/ethyl acetate as the mobile phase. NMR spectra were recorded on a Bruker 250 and 360 MHz, and an Inova 500 MHz spectrometers in CDCl₃ or DMSO containing 0.03% TMS. Chemical shifts are listed downfield in ppm relative to tetramethylsilane. Coupling constants are given in Hz. Mass spectrometric analysis was performed on a Bruker Autoflex MALDI-TOF MS with DHB as the matrix. HRESIMS data were obtained using either PEG (polyethylene glycol) or the PPG (polypropylene glycol) as internal standard. Data were obtained on JEOL Model JMS-T100LC (The AccuTOF) by spraying in MeOH.

4.1.1. Resorcinarene (8).^{2b} Resorcinol **5** (20 g, 0.181 mol) was dissolved in 120 mL of 95% ethanol/water, and 36 mL of 37% aqueous HCl. The solution was cooled in an ice bath and 25.3 mL (0.181 mol) of heptaldehyde 6 was added drop wise over a period of 1 h with an addition funnel. The mixture was allowed to slowly warm to rt. The reaction mixture was then maintained at 80 °C for 12 h, and the yellow needles that separated were collected by filtration and washed with cold 1:1 ethanol-water until the material was pale yellow, and neutral to pH paper. Drying under vacuum at 100 °C for 16 h afforded 35.9 g (96%) of 8: 1 H NMR (250 MHz, DMSO) δ 0.829 (t, 12H, CH₃), 1.21 (m, 32H, $CH_2CH_2CH_2CH_2$), 2.07 (m, 8H, $CH_2 \alpha$ to methine), 4.23 (t, 4H, methine, J=7.3 Hz), 6.13 (s, 4H, ArH), 7.12 (s, 4H, ArH), 8.85 (s, 8H, Ar-OH); MALDI-TOF m/z 847 (M+Na⁺).

4.1.2. Tetrabromide resorcinarene (10).^{2b} To a stirred orange solution of resorcinarene **8** (30 g, 0.036 mol) in 181 mL of 2-butanone was added 38.5 g (0.216 mol) of *N*-bromosuccinimide in portions. After 5 min, the product began to precipitate. The mixture was stirred for an additional 3 h, and the product was collected by filtration and washed with cold 2-butanone, drying under vacuum at 100 °C for 12 h afforded 30.9 g (75%) of **10** as a white solid: ¹H NMR (250 MHz, DMSO) δ 0.818 (t, 12H, CH₃), 1.28 (m, 32H, CH₂CH₂CH₂CH₂), 2.13 (m, 8H, CH₂ α to methine), 4.34 (t, 4H, methine, *J*=7.1 Hz), 7.31 (s, 4H, ArH), 9.06 (s, 8H, Ar-OH); MALDI-TOF *m*/*z* 1159 (M+Na⁺).

4.1.3. Tetrabromide cavitand (12).^{2b} To a sealed tube (Ace Glass Co., pressure tube, 200 ml volume) was added tetrabromide resorcinarene **10** (3 g, 0.0026 mol) and 26 mL of degassed dry DMF and 26 mL of degassed dry DMSO and (5.74 g, 0.0415 mol) of dry Cs_2CO_3 . After stirring at rt for 1 h the solution developed a deep purple color. Then was added 2.7 mL (0.415 mol) of CH₂BrCl in one portion. The reaction was then heated to 88 °C for 3 h. (CAUTION: modifications of the reaction conditions may result in much larger internal pressures.) The sealed tube was then cooled in an ice bath for 2 h and poured into (600 ml of 2% HCl), the solid was filtered and slowly recrystallized from

CH₂Cl₂/hexanes, drying under vacuum at 100 °C for 12 h afforded 2.78 g (89%) of **12** as off white needles: ¹H NMR (250 MHz, CDCl₃) δ 0.92 (t, 12H, CH₃), 1.39 (m, 32H, CH₂CH₂CH₂CH₂), 2.24 (m, 8H, CH₂ α to methine), 4.40 (d, 4H inner OCH₂, *J*=7.31 Hz), 4.88 (t, 4H, methine, *J*=7.8 Hz), 5.97 (d, 4H outer OCH₂, *J*=7.31 Hz), 7.03 (s, 4H, ArH); MALDI-TOF *m*/*z* 1207 (M+Na⁺).

4.1.4. Tetraester cavitand (14).^{2b} Before metal-halo exchange it was important to drive off reactive solvents such as CH₂Cl₂ trapped within the crystal lattice, this was most easily accomplished by repeatedly dissolving the material in solutions of dry THF and dry toluene and then distillation of the solvent through a vigreux column. Tetrabromide cavitand 12 (10 g, 0.0084 mol) was dissolved in 844 mL of dry THF and cooled to -78 °C in a dry ice-acetone bath, then was added 52.5 mL of *n*-butyllithium, 1.6 M solution in hexanes. The mixture was stirred for 2 h at -78 °C and then was added 8.02 mL (0.065 mol) of ethyl chloroformate all in one portion. The mixture was allowed to warm to rt over a 12 h period after which time was added 22 mL of water. The THF was removed in vacuo and the residue was partitioned between methylene chloride (250 mL) and water (100 mL). The aqueous layer was extracted twice with methylene chloride (250 ml) and the combined organic layers were washed with brine (200 ml) dried over sodium sulfate and concentrated in vacuo. Overnight recrystallization from 80% EtOAc/EtOH and drying under vacuum at 100 °C for 12 h afforded 8.17 g (80%) of 14 as white needles: ¹H NMR (250 MHz, CDCl₃) δ 0.91 (t, 12H, CH₃), 1.09–1.49 (m, 44H, CH₂CH₂CH₂CH₂, and OCH₂CH₃), 2.20 (m, 8H, CH₂ a to methine), 4.29 (q, 8H, OCH₂CH₃), 4.59 (d, 4H inner OCH₂, J=7.31 Hz), 4.88 (t, 4H, methine, J = 7.8 Hz), 5.97 (d, 4H outer OCH₂, J = 7.31 Hz), 7.03 (s, 4H, ArH); MALDI-TOF m/z 1183 $(M + Na^+)$.

4.1.5. Tetrol cavitand (16). Tetraester cavitand 14 (5 g, 0.0041 mol) was dissolved in 130 mL of dry THF and transferred via canula to a flask containing 1.55 g (0.041 mol) of LiAlH₄ in 16 mL of dry THF. The mixture was stirred at rt for 12 h and then was carefully added 2 mL of water followed by 2 mL of 10% NaOH. The mixture was filtered through celite, concentrated in vacuo, and the residue was partitioned between methylene chloride (150 mL) and water (50 mL). The aqueous layer was extracted twice with methylene chloride (100 mL) and the combined organic layers were washed with brine (100 mL) dried over sodium sulfate and concentrated in vacuo. Drying under vacuum at 100 °C for 12 h afforded 3.46 g (85%) of 16 as a white solid: ¹H NMR (500 MHz, CDCl₃) δ 0.91 (t, 12H, CH₃), 1.17 (s, 12H, CH₃ α to prochiral center), 1.31 (m, 32H, CH₂CH₂CH₂CH₂), 1.43–1.54 (m, 24H, OCOCH₃), 2.23 (m, 8H, CH₂ α to methine), 4.04 (s, 16H, CH₂ α to prochiral center), 4.14 (d, 4H inner OCH₂, J=7.32 Hz), 4.78 (t, 4H, methine, J = 7.8 Hz), 5.04 (s, 8H, CH₂OCO), 5.78 (d, 4H outer OCH₂, J=7.32 Hz), 7.16 (s, 4H, ArH); ¹³C NMR (90 MHz, CDCl₃) δ 14.3, 17.9, 20.1, 22.84, 27.9, 29.6, 32.0, 37.0, 46.3, 56.9, 65.6, 100.2, 121.4, 122.7, 138.4, 154.4, 170.6, 172.6; MALDI-TOF m/z 1015 (M+Na⁺). HRESIMS m/z: 1015.5547 $[M+Na]^+$ (Calcd for $C_{60}H_{80}O_{12}Na$, 1015.5558).

4.1.6. Tetrabenzyl ketal cavitand (19). Tetrol cavitand 16 (1 g, 0.0010 mol) was dissolved in 5 mL of dry THF, then was added 1.37 g (0.006 mol) of 2,2-bis(phenyldioxymethyl)propionic acid 18 and 2.12 g (0.008 mol) of triphenylphosphine. Then was added, drop wise at rt, 1.6 mL of diisopropyl diazodicarboxylate. The reaction was stirred for 24 h and precipitated into cold methanol, the product was collected and washed with cold methanol. Drying under vacuum at 100 °C for 12 h afforded 1.73 g (96%) of **19** as a white solid: ¹H NMR (360 MHz, CDCl₃) δ 0.91 (t, 12H, CH₃), 1.17 (s, 12H, CH₃ α to prochiral center), 1.31 (m, 32H, CH₂CH₂CH₂CH₂), 1.43–1.54 (m, 24H, OCOCH₃), 2.23 (m, 8H, CH₂ α to methine), 4.04 (s, 16H, $CH_2 \alpha$ to prochiral center), 4.14 (d, 4H inner OCH_2 , J = 7.32 Hz), 4.78 (t, 4H, methine, J = 7.8 Hz), 5.04 (s, 8H, CH₂OCO), 5.78 (d, 4H outer OCH₂, J=7.32 Hz), 7.16 (s, 4H, ArH); ¹³C NMR (90 MHz, CDCl₃) δ 14.3, 17.9, 20.1, 22.8, 27.9, 29.6, 32.0, 36.9, 46.3, 56.9, 65.6, 100.2, 121.4, 122.7, 138.4, 154.4, 170.6, 172.6; MALDI-TOF m/z 1831 $(M+Na^{+})$. HRESIMS *m/z*: 1831.8692 $[M+Na]^{+}$ (Calcd for C₁₀₈H₁₂₈O₂₄Na, 1831.8693).

4.1.7. Octol cavitand (3). Tetrabenzyl ketal cavitand 19 (1 g, 0.0005 mol) was dissolved in 7.8 mL of dry THF, and 71 mL of EtOAc then was added $(0.100 \text{ g}, \text{Pd}(\text{OH})_2 \ 10\%)$. The apparatus for catalytic hydrogenolysis was evacuated and filled with $H_2(g)$ to 40 psi. The reaction mixture was then stirred for 24 h, and the Pd(OH)₂ was removed by filtration through celite. The filtrate was concentrated in vacuo to a volume of about 5 mL and was precipitated into cold methanol, the product was collected and washed with cold methanol, drying under vacuum at 100 °C for 12 h afforded (0.728 g, 94%) of 3 as a white solid: ¹H NMR (360 MHz, CDCl₃) δ 0.91 (t, 12H, CH₃), 1.17 (s, 12H, CH₃) α to prochiral center), 1.31 (m, 32H, CH₂CH₂CH₂CH₂), 1.43-1.54 (m, 24H, OCOCH₃), 2.23 (m, 8H, CH₂ α to methine), 4.04 (s, 16H, $CH_2 \alpha$ to prochiral center), 4.14 (d, 4H inner OCH₂, J=7.32 Hz), 4.78 (t, 4H, methine, J=7.8 Hz), 5.04 (s, 8H, CH₂OCO), 5.78 (d, 4H outer OCH₂, J=7.32 Hz), 7.16 (s, 4H, ArH); ¹³C NMR (90 MHz, CDCl₃) δ 14.3, 17.9, 20.07, 22.8, 27.9, 29.6, 32.0, 37.0, 46.3, 56.9, 65.6, 100.2, 121.4, 122.7, 138.4, 154.4, 170.6, 172.6; MALDI-TOF *m*/*z* 1479 (M+Na⁺). HRESIMS *m*/*z*: 1479.7513 $[M+Na]^+$ (Calcd for $C_{80}H_{112}O_{24}Na$, 1479.7441).

4.1.8. Tetrabenzyl ketal cavitand (20). Tetrol cavitand 17 $(0.450 \text{ g}, 6.23 \times 10^{-4} \text{ mol})$ was dissolved in 3.16 mL of dry THF, then was added 0.842 g $(3.7 \times 10^{-3} \text{ mol})$ of 2,2bis(phenyldioxymethyl)propionic Acid 18 and 1.24 g $(5.08 \times 10^{-3} \text{ mol})$ of triphenylphosphine. Then was added, drop wise at rt, 1.0 mL of diisopropyl diazodicarboxylate, the reaction was then stirred for 24 h and precipitated into cold methanol, the product was collected and washed with cold methanol. Drying under vacuum at 100 °C for 12 h afforded 1.73 g (98%) of **20** as a white solid: ¹H NMR (360 MHz, CDCl₃) δ 1.10 (s, 12H, CH₃ α to prochiral center), 1.78 (d, 12H, CH₃ chain), 3.62 (d, 8H, CH₂ α to prochiral center), 4.25 (d, 4H inner OCH₂, J=7.32 Hz), 4.62 (d, 8H, CH₂ α to prochiral center), 5.04 (s, 8H, CH₂OCO), 5.12 (t, 4H, methine, J=7.8 Hz), 5.49 (s, 4H CH benzyl), 5.78 (d, 4H outer OCH₂, J=7.32 Hz), 7.25-7.69 (m, 24H, ArH); ¹³C NMR (400 MHz, CDCl₃) δ 13.3, 15.6,

30.9, 39.1, 39.5, 39.5, 39.7, 39.9, 40.1, 53.8, 57.3, 99.7, 119.1, 127.5, 138.9, 152.8, 156.8; MALDI-TOF m/z 1551 (M+Na⁺). HRESIMS m/z: 1551.5488 [M+Na]⁺ (Calcd for C₈₈H₈₈O₂₄Na, 1551.5563).

4.1.9. Octol cavitand (4). Tetrabenzyl ketal cavitand 20 1 g (6.1 \times 10⁻⁴ mol) was dissolved in 8.6 mL of dry THF and 86 mL of EtOAc and 0.100 g of 10% Pd(OH)₂ was added. The apparatus for catalytic hydrogenolysis was evacuated and filled with $H_2(g)$ to 40 psi. The reaction mixture was then stirred for 24 h, and the $Pd(OH)_2$ was removed by filtration through celite. The filtrate was concentrated in vacuo to a volume of about 5 mL and was precipitated into cold methanol, the product was collected and washed with cold methanol, drying under vacuum at 100 °C for 12 h afforded 0.728 g (95%) of **4** as a white solid: ¹H NMR (360 MHz, CDCl₃) δ 1.05 (s, 12H, CH₃ α to prochiral center), 1.79 (d, 12H, CH₃ chain), 3.63 (m, 16H, CH₂ α to prochiral center), 4.13 (d, 4H inner OCH₂, J=7.32 Hz), 5.00 (q, 4H, methine, J = 7.8 Hz), 5.08 (s, 8H, CH₂OCO), 5.83 (d, 4H outer OCH₂, J=7.32 Hz), 7.23 (s, 4H, ArH); ¹³C NMR (500 MHz, CDCl₃) δ 55.1, 59.4, 95.4, 96.7, 99.1, 103.3, 111.9 126.6 135.3, 195.7, 196.2, 197.0; MALDI-TOF *m*/*z* 1200 (M+Na⁺). HRESIMS *m*/*z*: 1199.4334 $[M+Na]^+$ (Calcd for C₆₀H₇₂O₂₄Na, 1199.4311).

4.2. Synthesis of the octaester cavitands

4.2.1. Octaacetate cavitand (21). To a solution of octol 3 0.100 g (6.7×10^{-5} mol) in 6.86 mL of dry THF was added 0.075 mL of triethylamine, 0.054 mL of acetic anhydride, and a few crystals of DMAP (15 mg). The reaction mixture was stirred for 1 h, after which time TLC analysis (hex/ EtOAc 1:1) showed complete consumption of the starting material. The THF was removed in vacuo and the residue was partitioned between methylene chloride (30 mL) and water (10 mL). The aqueous layer was extracted with methylene chloride $(2 \times 30 \text{ mL})$ and the combined organic layer was washed with brine (30 mL) dried over sodium sulfate and concentrated in vacuo to give 0.122 g (99%) of **21** as a white solid: ¹H NMR (500 MHz, CDCl₃) δ 0.92 (t, 12H, CH₃), 1.17-1.31 (m, 44H, CH₃ α to prochiral center and CH₂CH₂CH₂CH₂), 1.43–1.54 (m, 24H, OCOCH₃), 2.23 (m, 8H, CH₂ α to methine), 4.04 (s, 16H, CH₂ α to prochiral center), 4.14 (d, 4H inner OCH₂, J=7.32 Hz), 4.78 (t, 4H, methine, J=7.8 Hz), 5.04 (s, 8H, CH₂OCO), 5.78 (d, 4H outer OCH₂, J = 7.32 Hz), 7.16 (s, 4H, ArH); ¹³C NMR (125 MHz, CDCl₃) δ 14.3, 17.9, 20.1, 22.8, 27.9, 29.6, 32.0, 36.9, 46.2, 56.8, 65.6, 100.1, 121.3, 122.7, 138.4, 154.4, 170.5, 172.5; MALDI-TOF m/z 1816 (M+Na⁺). HRESIMS m/z: 1815.8290 $[M+Na]^+$ (Calcd for C₉₆H₁₂₈O₃₂Na, 1815.8286).

4.2.2. Octaacetate cavitand (22). Following a procedure similar to the one described above, a solution of 0.1 g, $(8.3 \times 10^{-5} \text{ mol})$ of octol **4** in 8.31 mL of dry THF, 0.094 mL of triethylamine, 0.067 mL of acetic anhydride, and DMAP (15 mg) gave 0.122 g (97%) of **22** as a white solid: ¹H NMR (500 MHz, CDCl₃) δ 1.20 (s, 12H, CH₃ α to prochiral center) 1.52 (m, 24H, OCOCH₃) 1.78 (d, 12H, CH₃ chain), 4.04 (s, 16H, CH₂ α to prochiral center) 4.14 (d, 4H inner OCH₂, J=7.32 Hz) 4.78 (t, 4H, methine, J=7.8 Hz) 5.04 (s, 8H, CH₂OCO) 5.78 (d, 4H outer OCH₂,

J=7.32 Hz) 7.16 (s, 4H, ArH); ¹³C NMR (400 MHz, CDCl₃) δ 15.9, 17.6, 31.1, 46.1, 56.7, 65.4, 99.7, 99.9, 120.6, 122.4, 136.1, 153.9, 170.4, 172.4; MALDI-TOF m/z 1536 (M+Na⁺). HRESIMS m/z: 1535.5108 [M+Na]⁺ (Calcd for C₇₆H₈₈O₃₂Na, 1535.5156).

4.2.3. Octamesylate cavitand (23). Following a procedure similar to the one described above, a solution of 0.050 g $(3.3 \times 10^{-5} \text{ mol})$ of octol **3** in 3.43 mL of dry THF, 0.037 ml of triethylamine, 0.023 mL of methanesulfonyl chloride, and DMAP (5 mg), gave 0.07 g (98%) of **23** as a white solid: 1 H NMR (360 MHz, CDCl₃) δ 0.92 (t, 12H, CH₃), 1.19–1.31 (m, 44H, CH₃ α to prochiral center and CH₂CH₂CH₂CH₂CH₂), 1.39-1.54 (m, 24H, OSOCH₃) 2.22 (m, 8H, CH₂ α to methine), 4.06 (s, 16H, $CH_2 \alpha$ to prochiral center), 4.13 (d, 4H inner OCH₂, J=7.2 Hz), 4.74 (t, 4H, methine, J=7.2 Hz), 5.06 (s, 8H, CH₂OCO), 5.77 (d, 4H outer OCH₂, J =7.2 Hz), 7.10 (s, 4H, ArH); 13 C NMR (90 MHz, CDCl₃) δ 13.9, 17.3, 22.5, 27.7, 29.3, 29.8, 31.7, 36.5, 36.8, 46.6, 57.1, 69.0, 99.7, 121.0, 122.7, 138.6, 154.2, 170.8; MALDI-TOF m/z 2105 (M+Na⁺). HRESIMS m/z: 2103.392 [M+Na]⁺ (Calcd for C₈₈H₁₂₈O₄₀S₈Na, 2103.564).

4.2.4. Octaisopropanoate cavitand (24). Following a procedure similar to the one described above, a solution of 0.050 g, $(3.3 \times 10^{-5} \text{ mol})$ of octol **3** in 3.43 mL of dry THF, 0.043 ml of triethylamine, 0.032 mL of isobutyryl chloride, and DMAP (5 mg), gave 68 mg (99%) of 24 as a white solid: ¹H NMR (500 MHz, CDCl₃) δ 0.89 (t, 12H, CH₃), 1.03 and 1.01 (2d, 48H, CH(CH₃)₂), 1.22–1.31 (m, 44H, CH₃ α to prochiral center and CH₂CH₂CH₂CH₂), 2.24 (m, 8H, CH₂ α to methine), 2.45 (sep, 8H, CH α to $CH_3)_2$, 4.12–4.20 (m, 16H, $CH_2 \alpha$ to prochiral center, 4H inner OCH₂), 4.68 (t, 4H, methine, J=7.7 Hz), 5.04 (s, 8H, CH₂OCO), 5.79 (d, 4H outer OCH₂, J=7.0 Hz), 7.18 (s, 4H, ArH); ¹³C NMR (125 MHz, CDCl₃) δ 13.5, 17.4, 18.3, 22.2, 27.3, 29.0, 29.6, 29.8, 31.34, 33.4, 36.3, 45.9, 56.6, 64.6, 99.2, 120.9, 120.9, 137.8, 153.8, 172.3, 175.9; MALDI-TOF m/z 2040 (M+Na⁺). HRESIMS m/z: 2040.072 $[M+Na]^+$ (Calcd for $C_{112}H_{160}O_{32}Na$, 2040.0790).

4.2.5. Octaanisoate cavitand (25). Following a procedure similar to the one described above, a solution of 0.050 g, $(3.3 \times 10^{-5} \text{ mol})$ of octol **3** in 3.43 mL of dry THF, 0.037 ml of triethylamine, 0.052 g of p-anisoyl chloride, and DMAP (5 mg) gave 0.085 g (98%) of 25 as a white solid: ¹H NMR (360 MHz, CDCl₃) δ 0.88 (t, 12H, CH₃), 1.25–1.50 (m, 44H, CH₃ α to prochiral center, and CH₂CH₂CH₂CH₂), 2.18 (m, 8H, CH₂ α to methine), 3.82 (s, 24H, ArOCH₃), 4.27 (d, 4H inner OCH₂, J=7.13 Hz), 4.49 (s, 16H, CH₂ α to prochiral center), 4.78 (t, 4H, methine, J = 8.0 Hz), 5.01 (s, 8H, CH₂OCO), 5.62 (d, 4H outer OCH₂, J=7.27 Hz), 6.87 (d, 16H, ArH, J=8.9 Hz), 7.16 (s, 4H, ArH), 7.95 (d, 16H, ArH, J=8.61 Hz); ¹³C NMR (90 MHz, CDCl₃) δ 13.7, 17.6, 22.3, 27.4, 28.9, 29.3, 30.0, 31.4, 36.5, 46.5, 55.1, 57.2, 65.4, 99.3, 113.3, 131.0, 137.8, 146.2, 154.0, 163.3, 165.8, 172.5; MALDI-TOF m/z 2552 (M+Na⁺). HRESIMS m/z: 2552.0413 [M+Na]⁺ (Calcd for C₁₄₄H₁₆₀O₄₀Na, 2552.0383).

4.2.6. Octa *p*-phenyl benzoate cavitand (26). Following a procedure similar to the one described above, a solution of

 $0.050 \text{ g} (3.3 \times 10^{-5} \text{ mol}) \text{ of octol } 3 \text{ in } 3.43 \text{ mL of dry THF},$ 0.037 ml of triethylamine, 0.065 g of *p*-biphenyl carbonyl chloride, and DMAP (5 mg) gave 0.094 g (95%) of 26 as a white solid: ¹H NMR (360 MHz, CDCl₃) δ 0.88 (t, 12H, CH₃), 1.25–1.50 (m, 44H, CH₃ α to prochiral center, and CH₂CH₂CH₂CH₂), 2.18 (m, 8H, CH₂ a to methine), 3.82 (s, 24H, ArOCH₃), 4.27 (d, 4H inner OCH₂, J=7.13 Hz), 4.49 (s, 16H, $CH_2 \alpha$ to prochiral center), 4.78 (t, 4H, methine, J=8.0 Hz), 5.01 (s, 8H, CH₂OCO), 5.62 (d, 4H outer OCH₂, J=7.27 Hz), 6.87 (d, 16H, ArH, J=8.9 Hz), 7.16 (s, 4H, ArH), 7.95 (d, 16H, ArH, J=8.61 Hz); ¹³C NMR (90 MHz, CDCl₃) δ 13.7, 17.6, 22.3, 27.4, 29.0, 29.3, 30.0, 31.4, 36.5, 46.5, 55.1, 57.2, 65.4, 99.3, 113.3, 131.0, 137.8, 146.2, 154.0, 163.3, 165.8, 172.5; MALDI-TOF m/z 2920 (M+Na⁺). HRESIMS m/z: 2920.2148 [M+Na]⁺ (Calcd for C₁₈₄H₁₇₆O₃₂Na, 2920.2042).

Acknowledgements

We are grateful for financial support from the Frasch Foundation and acknowledge efforts of Dr. T. Gauthier (MALDI); Mr. R. Federspiel (VT NMR); Dr. Umesh Goli (FSU, HRMS); Dr. Pratap Chand (¹³C NMR). We also thank Dr. M. McLaughlin for helpful discussions.

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Tetrahedron 60 (2004) 10869-10876

Thermal and microwave assisted reactions of 2,5-disubstituted thienosultines with [60]fullerene: non-Kekulé biradicals and self-sensitized oxygenation of the cycloadduct

Chih-Chin Chi, I-Feng Pai and Wen-Sheng Chung*

Department of Applied Chemistry, National Chiao Tung University, 1001 Ta-Hseuh Rd., Hsinchu 30050, Taiwan, ROC

Received 21 July 2004; revised 13 September 2004; accepted 15 September 2004

Available online 1 October 2004

Abstract—Refluxing an o-dichlorobenzene solution of 2,5-disubstituted thienosultines 10a-f with [60]fullerene for 2-24 h gave both 1:1 and 2:1 cycloadducts in 37-79% isolated yields. The reaction was highly accelerated by microwave irradiation giving comparable yields of cycloadducts. Sultines 10a-f underwent cheletropic extrusion of SO₂ to form the corresponding non-Kekulé biradical intermediates 11a-f, which were subsequently trapped by [60] fullerene to form corresponding cycloadducts. The activation energy barriers (ΔG_c^{\neq}) determined for the boat-to-boat inversion of these 4',5',6',7'-tetrahydrobenzo[c]thieno-[5',6':1,2][60]fullerene adducts **12a–f** were found to be in the range of 13.5–14.8 kcal/mol. Unexpectedly, one of the monoadduct 12a was found to be labile when kept in air under ambient light. Two new products 15 (a sulfine-enone) and 16 (an endione) were isolated from the decomposed 12a and were found to derive from self-sensitized singlet oxygen reaction on the 2,5-dimethylthieno moiety of 12a. © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

Functionalization of [60]fullerene is fascinating and promising for new ferromagnetic materials and biological application because of its unique spherical structure, photochemistry, and radical quenching properties.¹ The Diels-Alder reaction of [60]fullerene with ortho-quinodimethanes $(o-QDMs)^2$ has been developed extensively and represents one of the most powerful methods for synthesizing a large variety of functionalized fullerenes.^{1,2} o-QDM is usually obtained from thermal or photochemical elimination of a small molecule from various precursors shown in Scheme 1.²

The diazene 1 can form o-QDM cleanly, but its fivemembered ring heterocyclic analogues are usually unstable at room temperature.³ Isochromanone 2 will undergo thermal decarboxylation only under harsh conditions (such as flash vacuum pyrolysis at 500 °C) unless it is substituted with electron-donating groups on the aromatic moiety.⁴ Among the precursors for o-QDM, sulfolene⁵ **3** and sultine⁶ 4 are good choices because they undergo pyrolysis at

reasonably low temperatures and yet they are usually stable indefinitely at room temperature.

Despite many reports of using hetero-o-QDM to functionalize [60] fullerene,^{1,7} relatively little is known about whether the non-Kekulé biradicals⁸ such as trimethylenemethane (TMM) or tetramethyleneethane (TME) will add to [60]fullerene efficiently. A pioneering work on the cycloaddition of TMM with C_{60} has been reported by Wudl,^{9a} where the TMM biradical was generated from a 7-alkylidene-2,3-diazabicycloheptene, but the cycloadduct of the TMM- C_{60} was not isolated due to its similar polarity with C_{60} .^{9a} Palladiummediated addition of TMM to C_{60} has also been successfully carried out by Luh.^{9b} Meanwhile, Ohno et al. reported^{9c} the first success in using TME biradical to functionalize C_{60} , where 3,4-fused pyrrolo-3-sulfolenes were used as TME precursors. As our continuous interests in the research of





Keywords: Non-Kekulé biradical; TME biradical; Microwave; Cycloaddition; [60]Fullerene; Singlet oxygen reaction.

^{*} Corresponding author. Tel.: +88 635131517; fax: +88 635723764; e-mail: wschung@cc.nctu.edu.tw

^{0040-4020/\$ -} see front matter © 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.040

heteroaromatic-fused sultines **5** and their use in the generation of TME biradicals^{10a,e} or hetero-*o*-QDMs,^{10b-d} we report here the reaction of C₆₀ with 2,5-disubstituted-thienosultines **10a–f** and an interesting self-sensitized singlet oxygen reaction of one of the fullerene adducts.



2. Results and discussion

The syntheses of heterocyclic sultines **10a–d** through the reaction of corresponding bis(chloromethyl)thiophenes with Rongalite (sodium formaldehyde sulfoxylate) have been reported elsewhere.^{10a,e} Sultines **10e** and **10f** were synthesized from **8** and **9** by a similar method, where the dialkylthio **8** and diarylthio **9** were obtained from lithium exchange of 2,5-dibromothiophene,¹¹ followed by thiolation, and then chloromethylation^{3a,12} (Scheme 2). When refluxed in *o*-dichlorobenzene (ODCB) with C₆₀ as a limiting reagent for different duration (2 h for **10a,c–f** and 24 h for **10b**), sultines **10a–f** underwent extrusion of SO₂, and the resulting non-Kekulé biradicals **11a–f** were intercepted as a mixture of 1:1 cycloadducts **12a–f** and 2:1 bisadducts **13a–f** in ca. 2–3:1 ratio (Scheme 3 and



Scheme 2. Reagents and conditions: (a) 2.1 equiv *n*BuLi, THF, -78 °C, 30 min; 2.1 equiv RSSR, -78 °C, 1 h; (b) 37% HCHO, conc. HCl, rt, 8 h (for 8); (c) ClCH₂OCH₃, ZnCl₂, CHCl₃, rt, 3 h (for 9); (d) Rongalite, TBAB, DMF, rt, 10–24 h.

Table 1). When solutions of sultines **10a–f** with C_{60} were irradiated with microwave (900 W) under 180 °C for only 4 min, comparable yields of cycloadducts were obtained in all cases except **10b**; where sultine **10b** was almost completely converted to sulfolene instead of reacting with the C_{60} , therefore, only trace of adduct **12b** was observed. The isolated yields for these cycloadducts **12** and **13** were in the range of 37–79% (58–96% based on consumed C_{60} , Table 1). It is remarkable that microwave assisted synthesis dramatically shortened the reaction time needed compared to those by conventional heating.^{13d} Furthermore, the ratio of monoadduct **12** vs. bisadduct **13** also increased from 2–3:1 to 3.5–6:1 when microwave was applied.

All fullerene adducts were separated by column chromatography on silica gel using cyclohexane/toluene as eluents. The low product yield for sultine **10b** was mainly due to its poor reactivity because there was no solubility problem. The bisadducts **13a–f** are mixtures of regioisomers and are usually difficult to be separated; therefore, their characterizations were only done by FAB-MS and ¹H NMR spectroscopy. The UV–vis spectra of compounds **12a–f** revealed a typical weak absorption band around 435 nm, which is characteristic of a dihydrofullerene structure for monoadduct.¹ Further support of the monoadducts **12a–f** came from FAB-MS by detecting their molecular ion peaks (M+H⁺) at following *m*/*z* ratios: 859 for **12a**, 899 for **12b**, 921 for **12c**, 983 for **12d**, 921 for **12e**, and 1047 for **12f**.

¹H NMR spectra for **12a** showed that the methylene protons bridging C_{60} and thiophene displayed a singlet (δ 4.43) at 25 °C and the singlet became a well-resolved AB quartet below 5 °C. Variable-temperature NMR experiments revealed that these cycloadducts were in their boat forms and the boat-to-boat inversion rates can be determined. The coalescence temperature T_c of the two methylene doublets and their coupling constants were used to determine the activation energy barriers $\Delta G_c^{\neq,10c,13,14}$ For **12a**, the two doublets coalesced to a broadened singlet at 291 K (T_c) and an activation energy, ΔG_c^{\neq} , of 14.0±0.2 kcal/mol was calculated (Fig. 1a). Although different T_c values are expected for the two diastereotopic methylenes of 12c due to molecular asymmetry, they happen to have same $T_{\rm c}$ values (290 K) and therefore have same activation energy barriers ΔG_c^{\neq} (Fig. 1b). Furthermore, despite the large variation in 2,5-disubstituents of **12a–f** (such as dimethyl, dichloro, diphenyl, bis(methylthio), and bis(phenylthio) groups), their ΔG_c^{\neq} are all within 13.5–14.8 kcal/mol



Sultine	Microwave ^a , yield (%)			Conventional heating ^b , yield (%)		
	Time (min)	Monoadduct 12	Bisadduct 13	Time (h)	Monoadduct 12	Bisadduct 13
10a	4	47 (64) ^c	11 (15) ^c	2	38 (73) ^c	12 (23) ^c
10b	4	d	d ´	24	$28 (44)^{c}$	$9(14)^{c}$
10c	4	$39(64)^{c}$	e	2	$47(62)^{c}$	$16(20)^{c}$
10d	4	53 (81) ^c	$9(14)^{c}$	2	$52(65)^{c}$	$20(25)^{c}$
10e	4	$41(52)^{c}$	$10(13)^{c}$	2	$52(62)^{c}$	$27(32)^{c}$
10f	4	51 (76) ^c	15 (22) ^c	2	51 (63) ^c	27 (33) ^c

Table 1. Results of the cycloaddition reactions of sultines 10a-f with C_{60} under microwave irradiation or conventional heating

^a Power 900 W, ODCB, 180 °C.

^b ODCB, reflux.

^c Based on consumed C₆₀.

 d Sultine 10b was completely converted to sulfolene without reacting with C_{60}

e Trace.

(Table 2 and Figs. S1–S4). Compared to other 4,7dihydrobenzo[*d*]thiophene-[5,6-*f*]-fullerene adducts such as **14a** and **14b**,^{13b} these 4,7-dihydrobenzo[*c*]thiophene-[5,6-*e*]-fullerene adducts **12a–f** have larger ΔG_c^{\neq} by 2.8 kcal/mol on average.

The activation energy barriers of compounds **12a–f** are close to those of pyrazines^{10c} and other carbocyclic fused [60]fullerenes.¹⁴ Factors that may affect the activation energy barriers are dependent on the nature of the heterocyclic systems, such as bond lengths and angles,^{7k,13b} tortional and angular constraints,^{13b} electronic and steric effects.^{10c} Bond order may have played an important role here on the magnitude of ΔG_c^{\neq} , because **12a–f** which contain two *exocyclic* double bonds in the bridged cyclohexene ring have larger values in ΔG_c^{\neq} , whereas compounds **14a,b** with an *endo* double bond character in the cyclohexene rings have smaller ones.^{7f,13b,c} This observation is consistent with that reported by Illescas et al.^{7f} where a correlation was found between the activation energies ΔG_c^{\neq} and the bond lengths of the cyclohexene double bond across the fullerene junction; that is, as the bond length increases the barrier also increases.

The ${}^{13}C$ NMR spectrum of **12a** showed only 20 peaks (with two peaks superimposed) when measured at 70 °C (well

above T_c), where rapid ring inversion is expected and the symmetry of the molecule is simplified from C_s to $C_{2,v}$, therefore, an overall of 21 peaks are expected. The ¹³C NMR spectrum for **12a** showed the characteristic quaternary sp³-carbon atoms of the 6,6-ring junction on the C₆₀ cage at δ 66.9, the α -methyls of thiophene at δ 13.3, and the methylene bridges between thiophene and C₆₀ at δ 41.4. There are 17 other peaks of quaternary sp² carbons between 134 and 158 ppm for compound **12a**. All spectroscopic features of **12a–f** are consistent with the monoadducts of thienobiradicals **11a–f** that add to C₆₀ on its 6,6-ring junction. Complete spectroscopic data of **12a–f** are summarized in Section 3.

Research on the oxygenation^{15–19} of fullerene derivatives has attracted considerable attention because it provides an excellent method for ring-opening of the fullerene cage.^{1,19} Many oxidizing reagents, such as ozone,¹⁵ iodosobenzene,¹⁶ and *m*-CPBA,¹⁸ have been employed to prepare fullerene oxides. Above all, photo-induced ring opening of [60]fullerene cage by reaction with singlet oxygen (¹O₂) seems to have drawn the most attention.¹⁹ We were surprised to find that, short exposure of a non-degassed solution of **12a** in CS₂ to ambient light led to its quick decomposition, where two new products **15** and **16** (in about 1:2 ratio) could be isolated from this solution in 50–73% yields. Compound



Figure 1. Various temperature ¹H NMR spectra (600 MHz, CDCl₃/CS₂=1:2) of cycloadducts: (a) 12a, for which T_c is 291 K; and (b) 12c, for which both the two T_c values are 290 K.

Table 2. Activation free energies	(ΔG_c^{\neq}) of [60]fullerene	adducts 12a-f from dynamic	^I H NMR ^a studies
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Thienoadduct (substituents)	$T_{\rm c}$ (K)	$\Delta \nu (\text{Hz})^{\text{b}}$	$J_{\rm AB}~({\rm Hz})^{\rm b}$	$\Delta G_{\rm c}^{\neq}$ (kcal/mol)	Reference
12a ^c (2,5-Dimethyl)	291	53.8	14.0	14.0 ± 0.2	This work
12b ^d (2,5-Dichloro)	286	95.6	14.1	13.5 ± 0.2	This work
12c ^c (2-Methyl-5-phenyl)	290	48.7	14.1	14.2 ± 0.2	This work
	290	42.6	14.0	14.2 ± 0.2	This work
12d ^c (2,5-Diphenyl)	304	59.1	14.1	14.8 ± 0.2	This work
12e ^c (2.5-Bis(methlthio))	298	120.9	14.5	14.6 ± 0.2	This work
12f ^c (2,5-Bis(phenylthio))	298	152.3	14.4	14.0 ± 0.2	This work
14a SCO ₂ CH ₃	245	26.2	14.7	12.0±0.2	13b
\sim	241	14.8	13.9	11.9 ± 0.2	13b
14b	223	51.0	15.4	10.7±0.2	13b
~	231	86.7	15.4	11.0 ± 0.2	13b

^a Various temperature measurements were taken in a 600 MHz NMR (**12e** in a 500 MHz NMR). The activation free energies were obtained using equation: $k_c = 2.22 \ (\Delta v_{AB}^2 + 6J_{AB}^2)^{1/2}, \ \Delta G_c^{\neq} = 4.58 \ T_c \ (10.32 + \log \ (T_c/k_c)) \times 10^{-3} \ \text{kcal/mol.}$

^b Data are reported at the highest temperature that affords well-separated quartet: -10 °C for **12a**, -15 °C for **12b**, 7 °C for **12c**, 20 °C for **12d**, -5 °C for **12e**, and 0 °C for **12f**.

^c In CDCl₃/CS₂ = 1:2.

^d In d_4 -ortho-dichlorobenzene.

12a was found to be stable if either oxygen or light was kept from the system.

The new product **15** shows a molecular ion peak (M+H⁺) at m/z=891, which is 32 units more than the cycloadduct **12a**. Furthermore, FT-IR spectrum of **15** shows medium signals at 1700 and 1529 cm⁻¹. Compound **16** shows strong vibrations at 1685, 1636 and 1617 cm⁻¹ (Fig. S5, Supporting Information), but its molecular ion peak is the same with **12a** (m/z=859). The ¹³C NMR of compound **15** shows the characteristic absorption of two carbonyl carbons at 197.7 (C=O) and 193.0 ppm (C=S \rightarrow O) as well as two quaternary carbons of C₆₀ moiety at 65.8 and 65.5 ppm. Compound **16** shows the characteristic absorption of only one carbonyl carbon at 200.6 ppm and one quaternary carbon of C₆₀ moiety at 65.3 ppm.

Based on all structural information available for **15** and **16** and literature precedents, ^{19–21} we speculated that singlet oxygen reaction must have occurred on the thiophene moiety first and forming an endoperoxide **17**. Subsequent rearrangement of the endoperoxide **17** should lead to two fullerene products: one, an asymmetrical sulfine-enone **15**, and the other, a symmetrical endione **16** (see Scheme 4). Similar reactions have been reported in the methylene blue sensitized singlet oxygen reactions of cyclohexenone-fused thiophenes^{20b,c} and 2,5-dimethylthiophene.²¹ All spectroscopic data for **15** and **16** are consistent with the proposed structures.

It should be noted that other thienoadducts, such as 2,5dichloro-, diphenyl-, bis(methylthio)-, bis(phenylthio)-, and 2-methyl-5-phenylthienofullerenes **12b–f**, are free from the singlet oxygen induced reaction and can be stored for several months in air with ambient light. There are two possibilities from the experimental results: (a) singlet oxygen was not formed in the solution of **12b–f**; therefore, they are stable in air and ambient light, or (b) singlet oxygen did form, however, the other cycloadducts **12b–f**, with different 2.5-substituents, simply did not react with it. We thus carried out two series of experiments to clarify possible reasons for the big differences in singlet oxygen reactivity towards **12a** and **12b–f**. In the first series of experiments, we found that adding 2,5-dimethylthiophene to the solution of other fullerene derivatives (e.g., **12d**) did result in the photoinduced oxygenation of 2,5-dimethylthiophene (entry



3 in Table 3 of Supporting Information). The results suggest that singlet oxygen is formed in all solutions of fullerene adducts **12a–f** and yet it reacts with them in very different rates.²² In the second series of experiments, we prepared *d*-chloroform solutions of various 2,5-disubstituted thiophenes (**A–F**) with C₆₀ (1.5 mol %) as a singlet oxygen sensitizer and irradiated them simultaneously with tungsten lamp to make sure that they were exposed to the same doses of light. The relative reactivity of singlet oxygen towards these 2,5-disubstituted thiophenes are: 2,5-dimethylthiophene **A** (37%), 2,5-bis(methylthio)thiophene **E** (26%), 2-methyl-5-phenylthiophene **C** (<6%) (Table 3, Supporting Information).^{21c} The rest of the other 2,5-disubstituted-thiophenes (**B**, **D**, and **F**) did not show any observable reactivity toward singlet oxygen (namely <1% conversion) even after 42 h irradiation by tungsten lamp.

In summary, we report here the syntheses of a series of thieno-fused fullerenes which represents one of the rare examples in the derivatization of [60]fullerene through TME non-Kekulé biradicals.9 When ODCB solutions of sultines 10a-f and C₆₀ were refluxed for 2-24 h or under microwave irradiation (900 W, <180 °C) for only 4 min, moderate to good yields (37-79%) of cycloadducts 12a-f were obtained in all cases except 10b. To our surprise, of the six thieno-[60]fullerene adducts 12a-f, only 12a was unstable in the presence of oxygen and ambient light. The decomposed products from 12a were characterized and found to derive from singlet oxygen reaction on the 2,5dimethylthiophene moiety of 12a. Why some of the fullerene derivatives attract oxygen on standing, while others resist oxidation, deserves further photophysical and theoretical calculation study.

3. Experimental

3.1. General

3.1.1. 2,5-Bis(methylthio)thiophene 6. The preparation of **6** followed a literature¹¹ procedure. To a solution of 2,5dibromothiophene (5.00 g, 20.7 mmol) in diethyl ether (100 mL) at -78 °C was added *n*-butyllithium (2.5 M in hexane, 43.4 mmol) via syringe under nitrogen. After the solution was stirred for 30 min, dimethyl disulfide (4.28 g, 45.5 mmol) in ether (20 mL) was added dropwise with vigorous stirring. The mixture was stirred at -78 °C for 1 h and then slowly warmed to room temperature. An ice-cold saturated ammonium chloride solution (50 mL) was added. The two layers were separated, and the aqueous layer was extracted with ether $(3 \times 30 \text{ mL})$. The organic layers were combined, dried over MgSO₄, filtered, and concentrated. The residue was subjected to silica gel chromatography using hexane as the eluent to yield 2.89 g (16.4 mmol, 79%) of **6** as a colorless oil: $R_f = 0.68$ (hexane); ¹H NMR (300 MHz, CDCl₃) δ 6.90 (2H, s), 2.48 (6H, s); ¹³C NMR (75.4 MHz, CDCl₃) δ 139.1 (Cq), 131.0 (CH), 21.9 (CH₃); MS (EI) m/z 178/177/176 (M⁺, 9/5/95), 161 (M⁺ - CH₃, 100), 114 (M⁺-CH₃-SCH₃, 42), 69 (42); HRMS *m/z* calcd for C₆H₈³²S₃ 175.9789, found 175.9788.

3.1.2. 2,5-Bis(phenylthio)thiophene 7. Follow the same procedures as in the preparation of **6**. 2,5-Dibromothiophene

(5.00 g, 20.7 mmol) was allowed to react with *n*-butyllithium (2.5 M in hexane, 43.4 mmol), and then with diphenyl disulfide (9.48 g, 43.4 mmol). The crude product was purified by column chromatography (hexane) to give 5.79 g (19.3 mmol, 93%) of **7** as a colorless solid: mp 42– 43 °C (hexane); R_f =0.33 (hexane); ¹H NMR (300 MHz, CDCl₃) δ 7.30–7.18 (10H, m), 7.16 (2H, s); ¹³C NMR (75.4 MHz, CDCl₃) δ 137.4 (Cq), 136.9 (Cq), 135.5 (CH), 129.1 (CH), 128.1 (CH), 126.7 (CH); MS (EI) *m/z* 303/302/ 301/300 (M⁺, 3/16/24/100), 299 (M⁺ – 1, 23), 190 (78); HRMS *m/z* calcd for C₁₆H₁₂S₃: C, 63.96; H, 4.03. Found: C, 63.73; H, 4.17.

3.1.3. 3,4-Bis(chloromethyl)-2,5-bis(methyl)thiophene 8.¹² Concentrate hydrochloric acid (12 M, 20 mL) was added to the mixture of 6 (2.00 g, 11.3 mmol) and 37% formaldehyde aqueous solution (1.70 g, 56.7 mmol). The mixture was stirred at room temperature for 8 h, then poured into water (30 mL), and extracted with CH_2Cl_2 (3×20 mL). The organic layers were combined, washed with water $(2 \times 30 \text{ mL})$, brine, dried over MgSO₄, filtered, and concentrated in vacuum to yield 2.71 g (9.92 mmol, 87%) of 8 as a pale green solid. Product 8 decomposed after chromatography on silica gel; therefore, crude product was used without further purification. Mp 75.5-77 °C (a colorless solid after recrystallization from hexane). (lit.23 78–78.5 °C); $R_f = 0.75$ (hexane/ethyl acetate = 4:1); ¹H NMR (300 MHz, CDCl₃) δ 4.77 (4H, s), 2.47 (6H, s); ¹³C NMR (75.4 MHz, CDCl₃) δ 139.2 (Cq), 138.9 (Cq), 37.2 (CH₂), 21.7 (CH₃); MS (EI) *m*/*z* 276/275/274/273 (M⁺, 7/6/ 30/15), 272 (M⁺-1, 61), 239 (30), 237 (M⁺-Cl, 100), 186 (38), 69 (57); HRMS m/z calcd for $C_8H_{10}^{35}Cl_2S_3$ 271.9324, found 271.9318.

5,7-Bis(methylthio)-1,4-dihydro-1H-3 λ^4 -3.1.4. thieno[3,4-d][2,3]oxathiin-3-oxide, sultine 10e. A solution of 8 (2.86 g, 10.5 mmol), Rongalite (3.33 g, 20.9 mmol), and TBAB (1.71 g, 5.24 mmol) in DMF (50 mL) was stirred at room temperature for 24 h. The mixture was diluted with H_2O (40 mL) and extracted with CH_2Cl_2 (3×20 mL). The organic layer was dried over MgSO₄, concentrated, and purified by column chromatography (6:1 hexane/ethyl acetate) to give 0.68 g (2.55 mmol, 24%) of 10e, as a white solid after recrystallization from a solvent of CH₂Cl₂ and hexane: mp 71–73 °C (CH₂Cl₂/hexane); $R_f = 0.45$ (hexane/ethyl acetate = 4:1); ¹H NMR (300 MHz, CDCl₃) δ 5.19, 5.10 (2H, ABq, J=14.7 Hz), 3.91, 3.81 (2H, A'B'q, J = 15.9 Hz), 2.44 (3H, s), 2.42 (3H, s); ¹³C NMR (75.4 MHz, CDCl₃) δ 137.1 (Cq), 132.8 (Cq), 132.0 (Cq), 126.8 (Cq), 58.8 (CH₂), 51.1 (CH₂), 21.3 (CH₃), 21.3 (CH₃); MS (EI) m/z 268/267/266 (M⁺, 3/5/13), 265 (M⁺ - 1, 22), 202 (M⁺ – SO₂, 32), 201 (40), 187 (100); HRMS *m/z* calcd for $C_8H_{10}O_2S_4$ 265.9565, found 265.9577; Anal. Calcd for C₈H₁₀O₂S₄: C, 36.07; H, 3.78. Found: C, 36.42; H, 4.20.

3.1.5. 3,4-Bis(chloromethyl)-2,5-bis(phenylthio)thiophene 9 and 5,7-bis(phenylthio)-1,4-dihydro-1*H***-3** λ ⁴**- thieno[3,4-***d***][2,3]oxathiin-3-oxide, sultine 10f.** Adapted from a literature method.^{4b} To a stirred solution of 7 (2.00 g, 6.66 mmol) and zinc chloride (1.4 g, 10 mmol) in dry chloroform (20 mL) was added dropwise chloromethylmethyl ether (1.6 g, 20 mmol) via syringe. The mixture

turned dark green upon addition. The reaction mixture was stirred at room temperature under nitrogen for 3 h, and then poured into ice-water (20 mL), and CH₂Cl₂ (40 mL) was added. After stirring for 10 min, the layers were separated and the organic layer was washed with water (20 mL) and dried over MgSO₄. The solvent distilled off in vacuum to yield 2.67 g of crude product as viscous orange oil. Without purification, the crude product was dissolved in DMF (50 mL), and added Rongalite (5.16 g, 33.5 mmol) and TBAB (1.08 g, 3.45 mmol). The reaction mixture was stirred at room temperature for 10 h. The mixture diluted with H₂O (40 mL) and extracted with CH₂Cl₂ (3×20 mL). The organic layer was dried over MgSO₄, concentrated, and purified by column chromatography (10:1 hexane/ethyl acetate) to give 0.29 g (0.47 mmol, 11% overall yield from 7 in two steps) of 10f.

3.1.6. Data of compound 9. a colorless liquid; R_f =0.3 (hexane); ¹H NMR (300 MHz, CDCl₃) δ 7.32–7.20 (10H, m), 4.85 (4H, s); ¹³C NMR (75.4 MHz, CDCl₃) δ 141.5 (Cq), 136.6 (Cq), 135.7 (Cq), 129.3 (CH), 128.9 (CH), 127.3 (CH), 37.1 (CH₂); MS (EI) *m*/*z* 398/399/400 (M⁺, 87/76/13), 396 (M⁺ – 1, 100), 348 (37), 231 (51), 216 (69), 215 (81), 203 (61), 184 (43), 171 (40), 51 (50), 50 (74), 38 (55); HRMS *m*/*z* calcd for C₁₈H₁₄³⁵Cl₂S₃ 395.9637, found 395.9625.

3.1.7. Data of compound 10f. A white solid after recrystallization from a solvent of CH₂Cl₂ and hexane; mp 71–72 °C (CH₂Cl₂/hexane); R_f =0.65 (hexane/ethyl acetate=4:1); ¹H NMR (300 MHz, CDCl₃) δ 7.30–7.18 (10H, m), 5.17, 5.10 (2H, ABq, *J*=15.0 Hz), 3.87, 3.81 (2H, A'B'q, *J*=16.4 Hz); ¹³C NMR (75.4 MHz, CDCl₃) δ 136.0 (Cq), 135.7 (Cq), 135.5 (Cq), 134.2 (Cq), 130.6 (Cq), 129.5 (CH), 129.4 (CH), 128.0 (CH); MS (EI) *m*/*z* 390/391 (M⁺, 2/14), 326 (M⁺ – SO₂, 80), 325 (M⁺ – 1 – SO₂, 100), 216 (38), 215 (41), 184 (67), 77 (40), 51 (71); HRMS *m*/*z* for C₁₈H₁₄O₂S₄ 389.9878, found 389.9885; Anal. Calcd for C₁₈H₁₄O₂S₄: C, 55.36; H, 3.61. Found: C, 55.19; H, 3.73.

3.1.8. Cycloaddition reactions of thienosultines 10a–f with C₆₀. General procedure. A solution of C₆₀ (50 mg, 0.069 mmol) and thienosultine (1.5 equiv. for 10a–f) in ODCB (20 mL) was refluxed under nitrogen or irradiated with microwave (900 W, ≤ 180 °C). The resulting brown reaction mixture was evaporated to dryness under reduced pressure. The residue was subjected to silica gel chromatography with cyclohexane/toluene (4/1) as the eluent. The reaction conditions and yields after purification are shown in Table 1.

3.1.9. 1',3'-Dimethyl-4',5',6',7'-tetrahydrobenzo[*c*]thieno-[5',6':1,2][60]fullerene (12a). A brown solid; mp>495 °C; R_f =0.75 (cyclohexane); ¹H NMR (25 °C, 600 MHz, *o*-C₆D₄Cl₂) δ 4.43 (4H, s), 2.56 (6H, s); ¹H NMR (5 °C, 600 MHz, *o*-C₆D₄Cl₂) δ 4.43, 4.38 (4H, ABq, *J*= 13.5 Hz), (2H, ABq, *J*=13.3 Hz), 2.55 (6H, s); ¹³C NMR (70 °C, 150 MHz, *o*-C₆D₄Cl₂) δ 159.9, 148.5, 147.3, 147.0, 146.6, 146.3, 146.2, 146.1, 145.5, 143.9, 143.4, 143.1, 142.9, 142.4, 141.1, 136.4, 135.8, 134.7, 66.9 (sp³ C of C₆₀), 41.4 (CH₂), 13.3 (CH₃); FAB-MS (MNB) *m*/*z* 859 (M+H⁺, 4), 858 (M⁺, 4), 721 (90), 461 (100), 460 (95); UV (CHCl₃) λ_{max} , nm (log ε) 434 (3.33), 308 (4.47), 256 (4.97); HRMS (FAB+) calcd for C₆₈H₁₀S 858.0504, found 858.0523.

3.1.10. Adduct 13a. A brown solid; $R_f = 0.45$ (cyclohexane); FAB-MS (MNB) m/z 997/998 (M+H⁺, 5/3), 857/858 (4/3), 721 (100); HRMS (FAB+) calcd for C₇₆H₂₀S₂ 996.1006, found 996.1751.

3.1.11. 1',3'-Dichloro-4',5',6',7'-tetrahydrobenzo[*c*]thieno-[5',6':1,2][60]fullerene (12b). A brown solid; mp>495 °C; R_f =0.75 (cyclohexane); ¹H NMR (25 °C, 600 MHz, o-C₆D₄Cl₂) δ 4.49 (4H, s); ¹H NMR (-15 °C, 600 MHz, o-C₆D₄Cl₂) δ 4.54, 4.38 (4H, ABq, J=14.1 Hz); ¹³C NMR (70 °C, 150 MHz, o-C₆D₄Cl₂) δ 156.5, 148.5, 147.3, 147.1, 146.5, 146.3, 146.2, 145.8, 145.4, 143.8, 143.4, 142.9, 142.4, 141.1, 136.2, 135.8, 122.7, 65.8 (sp³ C of C₆₀), 40.6 (CH₂); FAB-MS (MNB) *m*/*z* 899 (M+H⁺, 3), 898 (M⁺, 3), 720 (8), 460 (100); UV (CHCl₃) λ_{max} , nm (log ε) 434 (3.57), 310 (4.37), 257 (4.89); HRMS (FAB+) calcd for C₆₆H₄Cl₂S 897.9712, found 897.9510.

3.1.12. Adduct 13b. A brown solid; $R_f = 0.63$ (cyclohexane); FAB-MS (MNB) m/z 1074/1076/1077/1078/1079/ 1080 (M+H⁺, 0.7/0.9/0.9/1/1/0.8), 720 (8), 307 (100); HRMS (FAB+) calcd for C₇₂H₈Cl₄S₂ 1075.8821, found 1075.8821.

3.1.13. 1'-Methyl-3'-phenyl-4',5',6',7'-tetrahydrobenzo[c]thieno-[5',6':1,2][60]fullerene (12c). A brown solid; mp>495 °C; $R_f = 0.6$ (cyclohexane); ¹H NMR (25 °C, 600 MHz, 1:2 CDCl₃/CS₂) δ 7.53-7.51 (2H, m), 7.38-7.36 (2H, m), 7.29-7.26 (1H, m), 4.66 (2H, br s), 4.52 (2H, br s), 2.67 (3H, s); ¹H NMR (-20 °C, 600 MHz, 1:2 CDCl₃/ CS₂) & 7.53-7.52 (2H, m), 7.40-7.38 (2H, m), 7.31-7.28 (1H, m), 4.72, 4.63 (2H, ABq, J=14.1 Hz), 4.56, 4.48 (2H, ABq, J = 14.0 Hz), 2.69 (3H, s); ¹³C NMR (25 °C, 150 MHz, 1:2 CDCl₃/CS₂) δ156.4 (br), 147.4, 146.2, 146.2, 146.0, 145.6, 145.2, 144.7, 144.5, 142.9, 142.4, 141.9, 141.8, 141.4 (br), 140.0, 134.6, 133.6, 133.1, 132.6, 128.8 (CH), 128.6 (CH), 127.4 (CH), 65.5 (sp³ C of C₆₀), 65.4 (sp³ C of C₆₀), 40.5 (CH₂), 34.0 (CH₂), 12.8 (CH₃); FAB-MS (MNB) *m*/*z* 921 (M+H⁺, 5), 920 (M⁺, 5), 720 (100); UV (CHCl₃) λ_{max} , nm (log ε) 487 (3.34), 432 (3.64), 257 (5.07); HRMS (FAB+) calcd for $C_{73}H_{12}S$ 920.0661, found 920.0663.

3.1.14. Adduct 13c. A brown solid; $R_f = 0.3$ (cyclohexane); FAB-MS (MNB) m/z 1121 (M+H⁺, 8), 721 (18), 392 (100); HRMS (FAB+) calcd for C₈₆H₂₄S₂ 1120.1321, found 1120.1461.

3.1.15. 1',3'-Diphenyl-4',5',6',7'-tetrahydrobenzo[*c*] thieno-[5',6':1,2][60]fullerene (12d). A brown solid; mp>495 °C; R_f =0.54 (cyclohexane); ¹H NMR (25 °C, 600 MHz, 1:2 CDCl₃/CS₂) δ 7.59–7.57 (4H, m), 7.42–7.39 (4H, m), 7.33–7.31 (2H, m), 4.76 (2H, br s), 4.69 (2H, br s); ¹H NMR (-20 °C, 600 MHz, 1:2 CDCl₃/CS₂) δ 7.59–7.58 (4H, m), 7.44–7.41 (4H, m), 7.34–7.32 (2H, m), 4.79, 4.69 (4H, ABq, *J*=14.2 Hz); ¹³C NMR (-20 °C, 150 MHz, 1:2 CDCl₃/CS₂) δ 156.0, 155.9, 147.1, 145.98, 145.96, 145.8, 145.7, 145.32, 145.27, 145.1, 145.1, 144.95, 144.91, 144.88, 144.4, 144.2 (2C), 142.7, 142.6, 142.1, 142.1, 141.7, 141.6, 141.6, 141.5, 141.3, 141.1, 139.78, 139.76, 137.9, 135.7, 134.7, 133.9, 132.9, 128.8 (CH), 128.5 (CH), 127.7 (CH), 65.0 (sp³ C of C₆₀), 40.1 (CH₂); FAB-MS (MNB) *m/z* 984/ 983 (M+H⁺, 2/3), 982 (M⁺, 3), 721 (24), 461 (67), 392 (100); UV (CHCl₃) λ_{max} , nm (log ε) 432 (3.55), 257 (5.10); HRMS (FAB+) calcd for C₇₈H₁₄S 982.0817, found 982.0905.

3.1.16. Adduct 13d. A brown solid; R_f =0.2 (cyclohexane); FAB-MS (MNB) m/z 1246/1245 (M+H⁺, 4/5), 721 (67), 461 (57), 392 (100); HRMS (FAB+) calcd for C₉₆H₂₈S₂ 1244.1634, found 1244.1669.

3.1.17. 1',3'-Bis(methylthio)-4',5',6',7'-tetrahydrobenzo[*c*]thieno-[5',6':1,2][60]fullerene (12e). A brown solid; mp>495 °C; R_f =0.45 (cyclohexane); ¹H NMR (25 °C, 500 MHz, 1:2 CDCl₃/CS₂) δ 4.62 (4H, br s), 2.46 (6H, s); ¹H NMR (-25 °C, 500 MHz, 1:2 CDCl₃/CS₂) δ 4.74, 4.49 (4H, ABq, *J*=14.5 Hz), 2.46 (6H, s); ¹³C NMR (70 °C, 150 MHz, *o*-C₆D₄Cl₂) δ 157.3, 148.5, 147.3, 147.0, 146.5, 146.2, 146.2, 146.0, 145.5, 143.9, 143.4, 142.9, 142.4, 141.1, 136.2, 135.8, 135.0, 66.5 (sp³ C of C₆₀), 41.8 (CH₂), 23.2 (CH₃); FAB-MS (MNB) *m*/*z*; 923 (M+H⁺, 3), 614 (32), 462 (100), 444 (26); UV (CHCl₃) λ_{max} , nm (log ε) 435 (3.67), 310 (4.80), 256 (5.31); HRMS (FAB+) calcd for C₆₈H₁₀S₃ 921.9946, found 922.0023.

3.1.18. Adduct 13e. A brown solid; R_f =0.41 (cyclohexane: toluene=9:1); FAB-MS (MNB) m/z 1124 (M+H⁺, 3), 1123 (M⁺, 3) 720 (18), 442 (25), 308 (100); HRMS (FAB+) calcd for C₇₆H₂₀S₆ 1123.9892, found 1123.9897.

3.1.19. 1',3'-Bis(phenylthio)-4',5',6',7'-tetrahydrobenzo[c]thieno-[5',6':1,2][60]fullerene (12f). A brown solid; mp> 495 °C; $R_f = 0.35$ (cyclohexane); ¹H NMR (40 °C, 600 MHz, 1:2 CDCl₃/CS₂) δ 7.32-7.30 (4H, m), 7.17-7.15 (4H, m), 7.10–7.09 (2H, m), 4.66 (4H, br s); ¹H NMR $(-30 \degree C, 600 \text{ MHz}, 1:2 \text{ CDCl}_3/\text{CS}_2) \delta 7.28-7.24 (4\text{H}, \text{m}),$ 7.19-7.16 (4H, m), 7.11-7.10 (2H, m), 4.78, 4.52 (4H, ABq, J = 14.4 Hz; ¹³C NMR (40 °C, 150 MHz, 1:2 CDCl₃/CS₂) δ 156.0, 147.5, 146.3, 146.1, 145.6, 145.4, 145.3, 144.9, 144.5, 144.4, 143.0, 142.4, 141.9, 141.9, 141.4, 140.1, 136.8, 135.2, 131.2, 129.0 (CH), 128.3 (CH), 126.7 (CH), 65.0 (sp³ C of C₆₀), 40.7 (CH₂); FAB-MS (MNB) *m/z* 1047 $(M+H^+, 5), 1046 (M^+, 5), 766 (5), 720 (20), 613 (40), 460$ (100); UV (CHCl₃) λ_{max} , nm (log ε) 435 (3.67), 310 (4.80), 256 (5.31); HRMS (FAB +) calcd for C₇₈H₁₄S₃ 1046.0259, found 1046.0404.

3.1.20. Adduct 13f. A brown solid; R_f =0.28 (cyclohexane/toluene=9:1); FAB-MS (MNB) m/z 1373 (M+H⁺, 5), 1372 (M⁺, 5), 721 (32), 442 (37), 308 (100); HRMS (FAB+) calcd for C₉₆H₂₈S₆ 1372.0518, found 1372.0522.

3.1.21. Synthesis of 15 and 16. An air-saturated solution of **12a** (25 mg, 0.029 mmol) in CS₂ (15 mL) was stirred at room temperature under the irradiation of room light and monitored by TLC chromatography until the starting compound disappeared which took about 3 days. After removal of the solvent, the residue was purified by silica gel chromatography (3–1:2 cyclohexane/CHCl₃) to give 7.3 mg of **15** (28%) and 11.6 mg of **16** (45%).

3.1.22. Data of 15. A brown solid, mp>495 °C; $R_f=0.3$

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(cyclohexane: CHCl₃=1:1); FT-IR (KBr, cm⁻¹) 1700 (C=O), 1529 (C=C); ¹H NMR (25 °C, 600 MHz, CDCl₃) δ 4.56 (2H, br s), 4.45 (2H, br s), 2.71 (3H, s), 2.53 (3H, s); ¹³C NMR (25 °C, 150 MHz, CDCl₃) δ 197.7 (C=O), 193.0 (C=S \rightarrow O), 155.5, 155.4, 147.7 (2C), 146.6 (2C), 146.3 (2C), 145.8, 145.6 (3C), 145.5 (2C), 145.2, 144.7 (2C), 143.2, 142.7, 142.6, 142.3, 142.2, 142.1 (2C), 141.7 (2C), 140.2 (2C), 137.9, 135.8, 135.7, 65.8 (sp³ C of C₆₀), 65.5 (sp³ C of C₆₀), 49.1 (CH₂), 42.6 (CH₂), 30.9 (CH₃), 30.3 (CH₃); FAB-MS (MNB) *m*/*z* 891 (M+H⁺, 2), 890 (M⁺, 3), 889 (M⁺ – 1, 2), 721 (100); HRMS (FAB+) calcd for C₆₈H₁₀O₂S 890.0402, found 890.0382.

3.1.23. Data of 16. A brown solid; mp>495 °C; R_f =0.25 (cyclohexane/CHCl₃=2:3); FT-IR (KBr, cm⁻¹) 1685 (C=O); ¹H NMR (25 °C, 600 MHz, CDCl₃) δ 4.31 (4H, s), 2.63 (6H, s); ¹³C NMR (25 °C, 150 MHz, CDCl₃) δ 200.6 (C=O), 155.2, 147.5, 146.6, 146.4, 145.7, 145.7, 145.6, 145.2, 144.9, 144.7, 143.2, 142.7, 142.3, 142.1, 141.7, 140.3, 135.7, 65.3 (sp³ C of C₆₀), 41.7 (CH₂), 29.2 (CH₃); FAB-MS (NMB) 859 (M+H⁺, 2), 858 (M⁺, 2), 720 (100); HRMS (FAB+) calcd for C₆₈H₁₀O₂ 858.0681, found 858.0712.

Acknowledgements

We thank the National Science Council of the Republic of China for financial support (Grant No. NSC92-2113-M-009-013). We also thank the National Chiao Tung University for supporting the Center of Interdisciplinary Molecular Science.

Supplementary data

Various temperature ¹H NMR spectra of compounds **12b**, **d–f**, ¹H NMR spectra of compounds **6**, **8**, **9**, and **10e**, ¹H and/ or ¹³C NMR spectra of compounds **12**, **13**, **15** and **16** and FTIR spectra of adducts **15** and **16**.

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tet.2004.09. 040

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Tetrahedron

Tetrahedron 60 (2004) 10877-10882

Stable structures of 12-crown-O₃N complexes with Li⁺ or Na⁺ in aqueous and acetonitrile solutions

Katsuhiko Okano,^a Hiroshi Tsukube^b and Kenzi Hori^{c,*}

^aTokyo R&D Center, Daiichi Pharmaceutical Co. Ltd, Kitakasai, Edogawa, Tokyo 134-8630, Japan ^bDepartment of Chemistry, Graduate School of Science, Osaka City University, Osaka 558-8585, Japan ^cDepartment of Applied Chemistry and Chemical Engineering, Faculty of Engineering, Yamaguchi University,

Tokiwadai, Ube 755-8611, Japan

Received 29 June 2004; revised 13 September 2004; accepted 15 September 2004

Available online 8 October 2004

Abstract—Molecular geometries of crown ether derivatives play an important role in capturing and transporting alkali metal ions such as Li^+ and Na^+ . As the selectivity of ions is observed in solutions, it is necessary to know their molecular structures in solutions. Recently, we investigated stable conformations of 12-crown-O₃N and its Li^+ complex in aqueous solution by the combination of three programs, the CONFLEX, Gaussian 98, and BOSS programs. In the present study, we applied the same procedure to investigate stable structures of 12-crown-O₃N complexes with an alkali ion in aqueous and acetonitrile solutions. It was confirmed that the stable structures of Li^+ and Na^+ complexes in solutions are highly dependent on the polarity of the solvents.

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1. Introduction

Crown ethers have an interesting function in that they capture and transport alkali metal ions selectively. For example, it is known that 15-crown-5 selectively forms a complex with Na⁺ and 18-crown-6 with $K^{+,1}$ As a reason for the selectivity, the relation between ion size and cavity size of crown ethers has been considered to be the most important since Pedersen first synthesized the cyclic ethers.² However, in polar solvents such as water, alcohol, and so on, 12-crown-4 with the optimal hole for Li⁺ shows little selectivity for the small ion. It was experimentally found that the stability of complexes depends greatly on the solvents used.¹ These results show the importance of not only the ion-hole relation but also the solvent effect for the ion selectivity. Later, it was pointed out that the solvent effect rather than spatial factors largely participates in ion selectivity.³

Theoretical calculation has been a powerful tool to solve the mechanism of selective capture and transportation of alkali ions by the use of crown ether derivatives.⁴ For example, it was observed that an amine side arm is effective for improving the Li⁺ selectivity of crown derivatives.⁵ That is,

0040–4020/\$ - see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.041

the interaction energy between Li^+ and an amine arm is important for the selectivity. The presence of an amine arm increases the interaction energy between Li^+ and the 12-crown-O₃N derivatives so that solvent molecules around Li^+ are removed to form their stable complex.

As crown ethers function in solution, solvent effects should be included in efforts to clarify the mechanism of the ion selectivity. Previous theoretical works, however, estimated the effect partially by including a few solvent molecules in molecular orbital calculations.⁶ Furthermore, the crown ether derivatives have a flexible ring and this property is considered to greatly affect the selectivity of ions. In a previous paper,⁷ we used three programs, the Conflex,⁸ Gaussian 98⁹ and BOSS¹⁰ programs, for investigating geometrical and solvent effects on ion selectivity.

The first program can generate stable conformations of 12-crown-O₃N **1** and its alkali ion complex. The second one calculates energies at the B3LYP/6-31G**//RHF/6-31G* level of theory, and the last one calculates differences in free energy of solvation between two conformers by using results of Monte Carlo (MC) simulations. It was confirmed that this combination of programs makes it possible to study stable conformations of the (12-crown-O₃N)Li⁺ complex in aqueous solution. The results of the calculations agree well with experimental findings for **1** in alcohol (Drawing 1).

Keywords: Stable conformations in solution; Monte Carlo simulation; Ab initio molecular orbital calculations; 12-Crown-O₃N.

^{*} Corresponding author. Tel.: +81 836859238; fax: +81 836859201; e-mail: kenji@sparklx.chem.yamaguchi-u.ac.jp



12-crown-O₃N 1

Drawing 1.

In the present study, we used the same procedure to study the stability of 12-crown- O_3N complexes with alkali ions not only in water but also in acetonitrile. Neither Li⁺ nor Na⁺ complexes were observed in the former solvent. By means of this procedure we were able to compare their stability in acetonitrile with that in water.

2. Method of calculations

2.1. Quantum mechanical calculations and Monte Carlo simulations

The Gaussian 98 program was used for optimizing a complex of Li⁺ or Na⁺ with four solvents, the solvated complex, at the RHF/6-31G* level of theory. The electrical charges for Monte Carlo (MC) simulations were calculated by applying the Merz–Singh–Kollman¹¹ scheme at the same level of theory. MC simulations were performed by using the BOSS Version 3.5 program.¹²

An orthorhombic box for MC calculations contains a solute as well as 500 TIP4P waters or 267 acetonitriles as solvents. These calculations make it possible to calculate the difference in free energy of solvation $\Delta\Delta G_{sol}$. Metropolis sampling and periodic boundary conditions were employed. The range of the movement was chosen to yield an acceptance rate of 30–40% for new configurations. The run for each window involved the equilibration for 10⁶ configurations, followed by averaging over an additional 10^6 configurations.

2.2. Calculation of $\Delta\Delta G_{sol(AB)}$ and potential energy profile in solutions

The potential curves for solvated complexes in a vacuum were calculated by the use of energies at the B3LYP/ 6-31G**//RHF/6-31G*level of theory. The results are listed in Table 1. The statistical perturbation theory (SPT)^{13,14} was

Table 1	Та	ble	1
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applied to calculate the difference in free energy of solvation $\Delta\Delta G_{\rm sol(AB)}$ between two structures, A and B, which have the ion in the different positions. MC simulations in aqueous and acetonitrile solutions were used for calculating $\Delta\Delta G_{\rm sol(AB)}$. Thermodynamic cycle for calculating the energy is shown below (Drawing 2).



Drawing 2.

From the thermodynamic cycle, $\Delta G_{AB(sol)}$ (sol = acetonitrile or water), a free energy difference in solution could be expressed to Eq. 1.

$$\Delta G_{AB(sol)} = \Delta G_{AB(g)} + (\Delta G_{sol(B)} - \Delta G_{sol(A)})$$
$$= \Delta G_{AB(g)} + \Delta \Delta G_{sol(AB)} \cong \Delta E_{AB(g)}$$
$$+ \Delta \Delta G_{sol(AB)}$$
(1)

 $\Delta G_{AB(g)}$, a free energy difference in gas phase is approximated to $\Delta E_{AB(g)}$, an energy difference between two structures, A and B, in the gas phase obtained from ab initio MO or density functional theory (DFT) calculations. $\Delta \Delta G_{sol(AB)}$, the difference in free energy of solvation between two structures, A and B, was from MC simulations. Potential energies in solutions were calculated by adding the $\Delta \Delta G_{sol(AB)}$ to the potential energies for solvated complexes in the a vacuum. As will be shown later, $\Delta E_{AB(g)}$ and $\Delta G_{AB(sol)}$ are plotted along the positions of the alkali ion.

3. Results and discussion

3.1. The most stable geometry of the (12-crown-O₃N)M⁺ complex with solvent molecules in the gas phase

It is necessary to calculate potential energies including bulk

r	LW	NW	LA	NA	
0	0.00 (0.00)	0.00 (0.00)		0.00 (0.00)	
0.3	-0.46(-0.79)	3.73 (0.27)	0.00 (0.00)	3.73 (2.01)	
0.6	-1.02(135)	9.90 (3.58)	1.51 (1.59)	9.27 (9.48)	
0.9	4.71 (6.08)	16.79 (7.76)	9.62 (7.20)	11.05 (12.90)	
1.2	5.08 (10.41)	25.17 (8.31)	12.46 (10.62)	15.23 (15.88)	
1.5	0.50 (15.10)	23.66 (7.24)	16.04 (14.36)	17.40 (18.43)	
1.8	-1.38(13.68)	23.52 (10.08)	24.23 (19.77)	22.38 (20.45)	
2.1	-5.04(11.54)	24.38 (10.55)	25.10 (21.22)	27.56 (21.80)	
2.4	-8.28(9.56)	23.63 (11.35)	25.07 (21.19)	34.09 (22.30)	
2.7	-12.72 (8.69)	24.41 (11.64)			
3.0	-13.52(10.98)	24.97 (14.36)			
3.3	-8.13 (12.12)	30.02 (19.04)			



Figure 1.

solvent effects to ascertain whether or not the 12-crown-O₃N complexes with an ion are stable or not in polar solvents. First of all, the geometry of the 12-crown-O₃N complex with Li⁺ and four water molecules as coordinated solvents was optimized at the RHF/6-31G* level of theory. Similar optimizations for the Na⁺ complex were performed and are shown in Figure 1. These optimized structures represented as MS-R, where M ($L = Li^+$ or $N = Na^+$) and S (W = water or A = acetonitrile) show the metal ion and the solvent, respectively. R shows the position of the ion as will be discussed later. The most stable structures of the solvated complexes are designated as LW-00 and NW-00.In the former, only one H₂O coordinates to Li⁺ and the other three waters make hydrogen bonds with the coordinated water. In contrast, two waters in NW-00 coordinate to Na⁺ since the Na-O2 and Na-O3 lengths are ca. 2.5 Å. The other two make hydrogen bonds with the coordinated waters according to atomic distances (Na-O1: 3.13 Å and Na-O4: 3.77 Å). The interaction between the 12-crown- O_3N ring and the ions prevents two waters from coordinating to the ion.

Similar solvated complexes, in which four acetonitriles were included instead of waters, with the alkali ions were optimized at the same level of theory. They are represented as LA-00 and NA-00. In LA-00, two acetonitriles coordinate to Li^+ and one makes a hydrogen bond with the H–N fragment of the ring. It is necessary to emphasize that one acetonitrile of LA-00 locates under the 12-crown-O₃N ring. The ion size of Li⁺ is too small to accommodate the fourth solvent on the same side where the ion is located. The NA-00 possesses four solvents over the crown ring. In the structure, three acetonitriles coordinate to Na⁺ within 2.7 Å and the other makes a hydrogen bond with the N–H

fragment of the crown ring, that is, the N4–H5 is 2.78 Å in length. The geometry difference between the LW-00 and the NW-00 complexes comes from the difference in the ion size.

3.2. The coordination geometry depends on the ion position in the vacuum

To obtain dependence of coordination geometry of solvents on the alkali ion position, we optimized a solvated complex with an ion, changing its position by 0.3 Å from that in the most stable structure along the axis perpendicular to the O₃N plain. As written above, *R* in the MS-R description is the number showing how long the ion locates away from the most stable position 'M' by $\Delta r = 0.3R$ Å (Drawing 3).



Drawing 3.

LA-01 in Figure 1 indicates the acetonitrile complex with Li^+ at the position $\Delta r = 0.3$ Å, and only the geometries of the solvent molecules were optimized at the RHF/6-31G* level of theory. The geometry of the crown ring was fixed at that of the most stable complex. The optimized structures



Figure 2.

LA-R, NW-R, and NA-R, depending on the ion position, are shown in Figure 2.

The same coordination number of Na⁺ was obtained for the structures from NW-00 to NW-06 (Δr =0.3–1.8 Å). Four waters can interact with Na⁺ in the geometry with more than Δr =2.1 Å. In NW-11 (Δr =3.3 Å), the Na⁺ has a tetrahedral geometry of waters since there is space enough to accommodate the solvent waters between the crown rings and the ion.

As discussed before, LA-00 has an acetonitrile located under the crown ring. On the other hand, all the acetonitriles locate on the same side of the crown ring in LA-01 as shown in Figure 1. It is impossible to continually change the coordination geometry of solvents from LA-00 to LA-01. The continuous change in geometry is strictly required, however, for calculating the difference in free energy of solvation by using SPT. Therefore, LA-01 is used for the initial geometry for calculating $\Delta G_{AB(sol)}$ along the ion position in acetonitrile. The ion size makes it possible to make a complex of Na⁺ coordinated with three acetonitriles in NA-00, whose geometry is similar to that of LA-01. In NA-07 (2.1), four acetonitriles coordinate to Na^+ , although the ion does not adopt a tetrahedral geometry. This is due to the size of the organic solvent as well as the space between the ion and 12-crown-O₃N.

3.3. The relative free energy change depends on the position of Li^+

We previously reported that the coordination number of waters, which changes from one to four, depends on the position of Li⁺ of LW in the vacuum. While the most stable structure is LW-00, the potential energy curve in the vacuum (Fig. 3) has another minimum around LW-09 (Δr = 2.7). While the latter minimum is less stable by ca. 9 kcal mol⁻¹ than LW-00, the latter geometry in aqueous solution (closed circles) is more stable by ca. 15 kcal mol⁻¹ than the former. The potential curve shows that no stable





Figure 4.

complex of 12-crown- O_3N with Li⁺ forms in aqueous solution. This is consistent with the experimental results as mentioned above.

The curves of LA are different from those for LW. The potential curve in the vacuum (open square) goes up from $\Delta r = 0.0-1.8 \text{ Å}$ by ca. 25 kcal mol⁻¹ and is almost the same in the region with more than $\Delta r = 1.8$ Å, as shown in Figure 3. The coordination number of Li⁺ changes from three such as with LA-02 ($\Delta r = 0.6$ Å) to four as for LA-07. While one acetonitrile still interacts with the NH fragment in LA-06, all the solvents coordinate to Li⁺ according to their Li-O distances. This change happens between LA-06 and LA-07. However, the coordination geometry of acetonitriles around Li⁺ is not tetrahedral even in LA-07, since the solvent molecules cannot enter the space between the 12-crown- O_3N and Li⁺. The potential curve for LA in acetonitrile (closed square) almost overlaps with that in the vacuum. This result means that 12-crown-O₃N tends to form a complex with Li⁺ in acetonitrile. This change accompanying the difference of the solvent polarity is very important for understanding the solvent effect on the ion selectivity.

3.4. The relative free energy change depends on the position of Na^+

The potential curves for the Na⁺ complex in aqueous and acetonitrile solutions are shown in Figure 4. The curve for NW in the vacuum (open triangles) becomes unstable by 10 kcal mol⁻¹ with the increase of Δr from 0.0 to 1.2 Å. In the region of more than 1.2 Å, the potential is almost constant. The number of the water molecules coordinating to Na⁺ change from 3 to 4 around $\Delta r = 1.2 - 1.5$ Å, as shown in Figure 2. This change is related to the shape of the potential curve. In aqueous solution (closed triangles), the maximum of ca. 25 kcal mol⁻¹ is seen around $\Delta r = 1.2$ Å. For the longer distances, the $\Delta G_{AB(sol)}$ value of ca. 24 kcal mol⁻¹ keeps almost constant. The most stable structure was also calculated to be NW-00 in water. The snapshot of NW-11 ($\Delta r = 3.3 \text{ \AA}$) is shown in Figure 5a. In this structure, $Na(OH_2)_4^+$ interacts with 12-crown-O₃N and the solvent waters surrounding these two fragments.

Near the point of $\Delta r = 1.2$ Å, the coordination number of Na⁺ changes from three to four between NA-06 and NA-07, as shown in Figure 2. The snap shot of the MC simulation for NA-11 is displayed in Figure 5b. Acetonitriles as solvents surround the (12-crown-O₃N)Na⁺(NCCH₃)₄, in which the Na⁺ ion still interacts with the crown ring. The potential curve for acetonitrile (open squares) goes up monotonically until $\Delta r = 2.4$ Å by ca. 20 kcal mol⁻¹ in the vacuum. That in acetonitrile (closed squares) showed almost the same tendency, until $\Delta r < 1.5$ Å, as that in the vacuum. The most stable structure in the organic solvent was calculated to be NA-00, which is also the most stable in aqueous solution.

The $\Delta G_{AB(sol)}$ between NA-00 and NA-08 ($\Delta r = 2.4$ Å) was calculated to be ca. 34 kcal mol⁻¹ (Table 1). This value is larger by 9 kcal mol⁻¹ than that in water. Therefore, it is







considered that the complex in acetonitrile is more stable than that in water. These results of calculations indicate that Na^+ is likely to interact with the 12-crown-O₃N ring. That is, their interaction energy is larger than that of the ion with solvents. It is, therefore, considered that the azaethers tend to make sandwich complexes with Na⁺ in acetonitrile and, probably, in aqueous solutions.¹⁵ This expectation is consistent with our experimental results.

4. Concluding remarks

In the present study, the stable structures of alkali ion complexes of the cyclic azaether not only in the aqueous but also in acetonitrile solution were investigated by using the method mentioned in the previous paper.⁷ Even in the organic solvent, the present procedure is applicable for examining whether or not the stable complex with an alkali ion and crown ether derivatives structures forms.

For the Li^+ complex, it was confirmed that LW-10, the uncomplexed structure, is the most stable in aqueous solution; that is, 12-crown-O₃N makes no stable complex with Li^+ . On the other hand, LA-01, the complexed structure, is the most stable in acetonitrile. NA-00 is the most stable in acetonitrile and the same tendency was obtained for an aqueous solution. It is considered that Na⁺ can complex with the crown ring in both solutions.

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Tetrahedron

Tetrahedron 60 (2004) 10883-10886

Photoinduced ene-reaction of 9-methylene-9,10-dihydrophenanthrene with alkenes

Akira Sugimoto, Ryoichi Hiraoka, Masahiro Kanayama Yasueda, Hirofumi Mukae and Kazuhiko Mizuno^{*}

Department of Applied Chemistry, Graduate School of Engineering, Osaka Prefecture University, 1-1 Gakuen-cho, Sakai, Osaka 599-8531, Japan

Received 3 August 2004; revised 10 September 2004; accepted 13 September 2004

Available online 1 October 2004

Abstract—Irradiation of 9-methylene-9,10-dihydrophenanthrene (1) in the presence of 1,1-diphenylethene or styrene in benzene afforded ene-reaction adduct in good yield. In the absence of arylalkenes, the dimerized product of 1, 9-[2-(9-phenanthryl)ethyl]-9,10-dihydrophenanthrene, and 9-methylphenanthrene were obtained as major products. © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

Previously, we have reported the chemical properties of 9-methylene-9,10-dihydrophenanthrene (1) which was prepared by the photolysis of 9-(2-anilinoethyl)phenanthrene^{1,2} (Eq. 1). Similarly, 1-methylene-1,2-dihydronaphthalene and 2-methylene-2,3-dihydrobenzo[b]thiophene were also prepared from the corresponding 2-anilinoethyl derivatives and their chemical properties were reported.² In the photolysis of 2-(2-anilinoethyl)naphthalene, however, 2-[2-(2naphthyl)ethyl]-1,2-dihydronaphthalene (2) as an eneadduct of the corresponding *exo*-methylene compound (3) with another 3 (Eq. 2), was exclusively obtained and neither 3 nor 2-methylnaphthalene was obtained.³ Lots of enereactions have been reported so far and their mechanisms have been discussed in relation to the Diels-Alder reaction.^{4–8} However, photoinduced ene-reactions are less known,^{9–12} because [2+2]photocycloaddition proceeds predominantly.¹³ The chemical properties of 5-methylene-1,3-cyclohexadiene and its benzologues^{14–23} have also been attracted considerable attention from the synthetic and mechanistic viewpoints, but photochemistry of them was rarely noted. We now report the photoinduced ene-reaction of 1 to clarify the photochemical reactivity of $1.^3$



2. Results and discussion

A degassed benzene solution of 1 in a sealed Pyrex-tube was irradiated with a 300 W high-pressure mercury lamp. The reaction products obtained were 9-[2-(9-phenanthryl)ethyl]-9,10-dihydrophenanthrene (4) and 9-methylphenanthrene (5) (Eq. 3) and no other product was detected by ¹H NMR

Keywords: Photoreaction; Ene reaction; Isomerization; Aromatization. * Corresponding author. Tel./fax: +81 72 254 9289;

e-mail: mizuno@chem.osakafu-u.ac.jp

^{0040–4020/\$ -} see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.032

Solvent	Yie	ld/%	Recovery/%	
	4	5	1	
C ₆ H ₆	34	21	45	
$C_{6}H_{12}$	37	31	32	
CH ₃ CN	31	69	0	
CH ₃ OH	37	63	Trace	

Table 1. Solvent effect on the photoreaction^a of 1

^a The values were estimated from the intensities of ¹H NMR spectra (CDCl₃) of the irradiated mixture after 8 h irradiation.

spectrum. The formation of **4** and **5** can be readily interpreted by the ene-reaction and 1,3-hydrogen shift reaction, respectively. The former is a photochemically forbidden reaction and the latter is an allowed one if the reactions proceed by a concerted mechanism.²⁴ From the mechanistic viewpoint, irradiation was carried out in several solvents as shown in Table 1. In all cases, the products were the same as those in benzene. The photoreaction proceeded almost completely in polar solvents, and **5** was obtained as major product, although the yield of **4** did not depend on the solvent polarity. It was suggested that the formation of **5** might involve a polar intermediate.



Based on these results, the photoinduced ene-reaction of 1 in the presence of arylalkenes as enophiles were investigated. Irradiation of a degassed benzene solution of 1 containing a large excess (20-fold excess) of 1,1-diphenylethene afforded 9-(3,3-diphenylpropyl)phenanthrene 6 in a 69% isolated yield. Similar irradiation of 1 in the presence of styrene, 1-phenylpropene, and 1,3-pentadiene gave the corresponding ene-adducts 7–9. However, the photoreaction of 1 with 1-hexene and butyl vinyl ether did not afford ene-products, but 4 and 5. In the cases of stilbene and cinnamaldehyde, 1 was almost recovered. The results are summarized in Table

Table 2. Photoreaction of 1 with alkenes in benzene^{a,b}

2. The ene-adducts 6-8 were identified by comparing their spectroscopic data with those of authentic compounds, which were prepared by the other methods without irradiation. The photoproduct 9 was converted into the hydrogenated derivative, 9-hexylphenanthrene, and identified.

The incident light (Pyrex filtered light: >280 nm) is competitively absorbed by 1 (λ_{max} =278 nm) and the added alkenes, although the amounts of alkenes are large excess.¹ 1,1-Diphenylethene, styrene, and 1-phenylpropene reacted with the excited singlet of 1 regioselectively to give substituted propane derivatives 6, 7, and 8, respectively. These ene-adducts bear phenanthrylmethyl group regioselectively to the sterically less-hindered carbon atom of the alkenes. However, irradiation of 1 in the presence of (*E*)stilbene gave only (*Z*)-stilbene and unchanged 1 was recovered, presumably due to most of the light absorption by (*E*)-stilbene. Phenylacetylene reacted with 1 upon irradiation to give a complex mixture.



The photoreaction of 1 in the presence of a large amount of 1,3-pentadiene (120-fold) also afforded an ene-adduct in 15%. However, the less amount of 1,3-pentadiene (20-fold) did not give the ene-adduct, but **5** was obtained in 96% yield. In this case, the excited singlet state of **1** may rearrange to **5** before being trapped by 1,3-pentadiene.

Mono-olefins such as 1-hexene and butyl vinyl ether gave no ene-adduct. These results suggested that the reactive

Alkene ^c	Yield/%			
	Ene-adduct	4	5	1
Ph ₂ C=CH ₂	6 (69)	nd	nd	nd
PhCH=CH ₂	7 (62)	nd	(4)	(4)
PhCH=CHMe	8 (21)	nd	(7)	(19)
(E)-PhCH=CHPh	(0)	(0)	(0)	(100)
CH ₂ =CHCH=CHMe	9 (0)	(0)	96	nd
CH ₂ =CHCH=CHMe ^d	9 (15)	(10)	nd	nd
$CH_2 = CH(CH_2)_3 Me$	(0)	(18)	(16)	(44)
Bu-O-CH=CH ₂	(0)	(20)	(12)	(35)
PhCH=CHCHO	(0)	(0)	(0)	(75)

^a Isolated yields and recoveries were obtained based on 1 used.

^b nd: not determined.

^c 20 equiv of an alkene based on **1** used.

^d 120 equiv of 1,3-pentadiene based on **1** used.

alkene has to be conjugated with a phenyl group or with a C=C double bond. Carbonyl compounds such as 2-propenal, β -ionone, and cinnamaldehyde gave no ene-adduct in detectable amount under similar reaction conditions and unchanged **1** was recovered on irradiation, suggesting that an abstraction of hydrogen atom by carbonyl group did not take place under the reaction conditions.

Compound **1** reacted with typical electron-acceptors such as tetracyanoethene and dimethyl acetylenedicarboxylate in the dark at room temperature to give ene-adducts, respectively. But it did not react with styrene, maleic anhydride, and fumaronitrile.² 1,1-Diphenylethene did not react with **1** even in refluxing benzene. On the basis of these observations, the present photoreaction is regarded as a photoinduced ene-reaction, although it has been stated that photo-initiated ene-reactions do not occur at less polar conditions.¹³

As mentioned before,³ irradiation of 2-(2-anilinoethyl)naphthalene affords the ene-adduct 2 without formation of the exo-methylene compound 3, while that of 9-(2anilinoethyl)phenanthrene affords the *exo*-methylene compound 1 without formation of the ene-adduct 4. These observation may be explained by the difference in their absorption spectra. Thus, the compounds 3 and 1 show an absorption maximum at 297 and 278 nm (log $\varepsilon = 4.14$), respectively.²⁵ On the other hand, the absorption maximum of 2-(2-anilinoethyl)naphthalene was observed at 276 nm $(\log \varepsilon = 3.83)$ and that of 9-(2-anilinoethyl)phenanthrene was at 298 nm (log $\varepsilon = 4.00$).² Namely, in the case of naphthalene derivative, most of the incident light should be absorbed by the photo-product 3 which may change into the ene-adduct 2. In the case of 1, the starting amine should absorb the incident light preferentially and therefore after the consumption of starting amine, the photo-induced enereaction of **1** might take place.

3. Experimental

Mp's were determined with a Yanaco micromelting point apparatus (MP-500) and are uncorrected. Spectra were obtained with a Hitachi infrared spectrophotometer (type-215), an NMR spectrometer (JNM-GX-270) (270 MHz) using tetramethylsilane as internal standard, a gas chromatograph-mass spectrometer (Shimadzu-LKB 9000) (mass), and a Shimadzu UV spectrophotometer (UV-160A).

3.1. Materials

9-Methylene-9,10-dihydrophenanthrene **1** was prepared by the method described previously.^{1,2} As **1** gradually changed into the methyl isomer, photoreactions were performed without separation of the two isomers (the starting material **1** was contaminated with 5–10% of **5**). The yields in Tables 1 and 2 are based on the amount of **1** at the start of the irradiation.

3.2. Photoreactions

3.2.1. Preparative irradiation. A general procedure is shown for the irradiation of 1 with 1,1-diphenylethene: a

solution of **1** [58 mg, purity 91% contained **1** (0.27 mmol)] and 1,1-diphenylethene (973 mg, 5.4 mmol) in benzene (20 mL) in a Pyrex tube was degassed by three freezepump-thaw cycles under argon, and the tube was sealed under reduced pressure. The solution was irradiated externally with 300 W high-pressure mercury lamp (Eikosha PIH-300) for 8 h at room temperature. After irradiation, the solvent was removed under reduced pressure and the residue was chromatographed on silica gel with hexane-benzene (5:1, v/v) to give 3-(9-phenanthryl)-1,1diphenylpropane 6 as a colorless solid (70 mg, 69%), mp 129.5–130.5 °C (from hexane); δ (CDCl₃) 2.53–2.62 (2H, m, CH₂), 3.05-3.10 (2H, t, CH₂), 4.05-4.10 (1H, t, CH), and 7.14-7.92 (17H, m) and 8.63-8.74 (2H, m) (together 19H ArH); m/z 372 (M+, 46%), 205 (22), 192 (100), and 191 (29). Found: C, 93.39; H, 6.19. Calcd for C₂₉H₂₄: C, 93.51; H, 6.49%.

The spectral data of the other photo ene-adducts, **7**, **8**, and **9** isolated from the irradiation mixtures by chromatography were compared with those of the corresponding authentic compounds are identified after hydrogenation.

3.2.2. Preparation of 9-[2-(9-phenanthryl)ethyl]-9,10**dihydrophenanthrene 4.** A solution of **1** [61 mg, purity 90% contained 1 (0.286 mmol)] in acetonitrile (20 mL) was irradiated. Chromatography of the irradiated mixture on silica gel with benzene-hexane (1:3, v/v) gave 4 from the third eluent (12 mg, 22%), mp 153–154 °C (from chloro-form–ethanol); ν_{max} (KBr)/cm⁻¹ 3060, 3030, 3010, 2920, 1500, 1485, 1455, 1440, 880, 740, and 720; λ_{max} (hexane)/ nm 212, 254, and 298 (log ε 4.79, 4.79, and 4.14); δ (CDCl₃) 1.86–1.95 (2H, m, CH₂), 2.94–3.28 (5H, m, two CH₂ and CH), and 7.21-7.82 (15H, m) and 8.59-8.69 (2H, m) (together 17H ArH); δ (CDCl₃) 31.31, 33.90, 34.08, 38.83, 122.40, 123.13, 123.55, 124.14, 124.26, 125.82, 125.88, 126.04, 126.44, 126.54, 127.05, 127.17, 127.47, 127.61, 127.96, 128.32, 129.01, 129.60, 130.67, 131.13, 131.86, 133.66, 134.12, 135.24, 136.41, 140.82; *m*/*z* 384 (M+, 33%), 193 (22), 192 (100), 191 (19), 179 (26), and 178 (27). Found: C, 93.46; H, 6.02. Calcd for C₃₀H₂₄: C, 93.71; H, 6.29%.

3.2.3. Preparation of authentic compounds. 1,1-Diphenyl-3-(9-phenanthryl)propane 6 (ene-adduct with 1,1diphenylethene) was obtained by the hydrogenation of 3-(9-phenanthryl)-1,1-diphenylpropene (mp 141–141.5 °C) which was prepared by the reaction of methyl 3-(9phenanthryl)propanoate with phenylmagnesium bromide followed by the dehydration and 5% Pd-C catalyzed hydrogenation, mp 128.5–129.5 °C; λ_{max} (hexane)/nm 210, 253, 277, 286, and 298 (log ε 4.65, 4.74, 4.12, 4.01, and 4.08); δ (CDCl₃) 2.57 (2H, q, J = 7.7 Hz, CH₂), 3.07 (2H, t, J=7.7 Hz, CH₂), 4.07 (1H, t, J=7.7 Hz, CH), 7.17–7.21 (2H, m, ArH), 7.28-7.36 (10H, m, ArH), 7.51-7.65 (5H, m, ArH), 7.79 (1H, dd, J=1.5, 7.3 Hz, ArH), 7.90 (1H, d, J= 8.0 Hz, ArH), 8.64 (1H, d, J=7.9 Hz, ArH), 8.72 (1H, d, J=8.2 Hz, ArH); δ (CDCl₃) 32.00, 36.37, 51.52, 122.50, 123.27, 124.42, 126.02, 126.18, 126.22, 126.34, 126.55, 126.65, 128.00, 128.06, 128.60, 129.70, 130.75, 131.18, 131.90, 136.34, 144.82.

3.2.4. 3-Phenyl-1-(9-phenanthryl)propane 7. Ene-adduct

with styrene. This compound was prepared by the hydrogenation of 3-(9-phenanthryl)-1-phenylpropene (mp 105–107.5 °C) which was obtained by 3 steps from 9-phenanthrylethenyl phenyl ketone (mp 136.5–138 °C), mp 84–85 °C (from pentane); λ_{max} (hexane)/nm 211, 253, 277, 286, and 298 (log ε 4.57, 4.77, 4.13, 4.02, and 4.07); δ (CDCl₃) 2.15 (2H, quintet, J=7.7 Hz, CH₂), 2.79 (2H, t, J=7.7 Hz, CH₂), 3.13 (2H, t, J=7.7 Hz, CH₂), 7.15–7.36 (2H, m, ArH), 7.50-7.68 (5H, m, ArH), 7.51-7.65 (5H, m, ArH), 7.81 (1H, d, J=7.5 Hz, ArH), 8.01 (1H, d, J=7.9 Hz, ArH), 8.63 (1H, d, J=8.2 Hz, ArH), 8.72 (1H, d, J=8.2 Hz, ArH); δ (CDCl₃) 31.89, 33.10, 36.12, 122.48, 123.25. 124.42, 125.88, 125.96, 126.11, 126.15, 126.52, 126.62, 128.05, 128.40, 128.56, 129.67, 130.73, 131.26, 131.91, 136.40, 142.17; m/z 296 (M+, 44%), 192 (100), and 191 (42). Found: C, 93.15; H, 6.8. Calcd for C₂₃H₂₀: C, 93.2; H, 6.8%.

3.2.5. 1-(9-Phenanthryl)-2-benzylpropane 8. Ene-adduct with 1-phenylpropene. This compound was prepared by the following route: propiophenone to 1-(9-phenanthryl)-2benzoylpropane (mp 140-140.5 °C) and from this to 8, mp 112–115 °C (from pentane); λ_{max} (hexane)/nm 211, 253, 277, 286, and 298 (log ε 4.56, 4.75, 4.12, 4.03 and 4.12); δ $(CDCl_3)$ 0.93 (3H, d, J=7.6 Hz, CH_3), 2.27 (1H, m, CH), 2.61 (1H, dd, J=7.7, 13.7 Hz, CH₂), 2.78 (2H, dd, J=8.8, 13.8 Hz, CH₂), 3.25 (1H, dd, J=5.2, 13.7 Hz, CH₂), 7.17-7.34 (5H, m, ArH), 7.50-7.66 (5H, m, ArH), 7.78-7.84 (2H, m, ArH), 8.64 (1H, d, J=8.2 Hz, ArH), 8.71 (1H, d, J= 8.2 Hz, ArH); δ (CDCl₃) 20.15, 36.11, 41.01, 44.26, 122.49, 123.22, 124.73, 125.95, 125.96, 126.07, 126.42, 126.62, 127.52, 128.07, 128.26, 129.36, 129.75, 130.80, 131.45, 131.75, 135.41, 141.19; m/z 310 (M+, 45%), 192 (100), and 191 (73). Found: C, 92.53; H, 6.94. Calcd for C₂₄H₂₂: C, 92.86; H, 7.14%.

3.2.6. 9-Hexylphenanthrene.²⁶ The hydrogenated product of 9, which corresponds to the ene-adduct with 1,3pentadiene. This was prepared from 9-phenanthrenecarbaldehyde by the reaction with pentylmagnesium bromide followed by dehydration and hydrogenation, mp 78–79 °C (from pentane); λ_{max} (hexane)/nm 253, 277, 285, and 298 (log ε 4.76, 4.13, 4.00 and 4.06); δ (CDCl₃) 0.90 (3H, t, J=7.1 Hz, CH₃), 1.25–1.41 (4H, m, CH₂CH₂), 1.43– 1.54 (2H, m, CH₂), 1.81 (2H, quintet, J=7.7 Hz, CH₂), 3.11 (2H, t, J=7.8 Hz, CH₂), and 7.52–7.71 (5H, m, ArH), 7.78– 7.84 (1H, m, ArH), 8.03-8.14 (1H, m, ArH), 8.65 (1H, d, J = 8.4 Hz, ArH), and 8.61–8.76 (1H, m, ArH); δ (CDCl₃) 14.40, 22.94, 29.81, 30.49, 32.03, 33.73, 122.48, 123.25, 124.54, 125.86, 125.96, 126.09, 126.48, 126.59, 128.04, 129.63, 130.73, 131.39, 132.00, 137.05; *m/z* 262 (M+, 60%), 191 (100), and 179 (28). Found: C, 91.51; H, 8.66. Calcd for C₂₀H₂₂: C, 91.55; H, 8.45%.

Acknowledgements

This work is partially supported by Tokyo Ohka Foundation

for the Promotion of Science and Technology and by a Grant-in-Aid for Scientific Research (Nos. 15350026 and 16655018) and by a Grant-in-Aid for Scientific Research on Priority Areas (417) (No. 15033264) from the Ministry of Education, Science, Sports, and Culture of Japan.

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Tetrahedron

Tetrahedron 60 (2004) 10887-10898

Photoreactions of β-aziridinylacrylonitriles and acrylates with alkenes: formation of head-to-head adducts and application to the preparation of pyrrolizidine alkaloid

Keitaro Ishii,* Takuya Sone, Yukio Shimada, Takahide Shigeyama, Masahiro Noji and Shigeo Sugiyama

Meiji Pharmaceutical University, 2-522-1 Noshio, Kiyose, Tokyo 204-8588, Japan

Received 3 July 2004; revised 10 September 2004; accepted 13 September 2004

Available online 1 October 2004

Abstract—The photochemical C,C-bond cleavage of *N*-benzyl β -aziridinylacrylonitrile **1** and acrylate **2** and the subsequent [3+2] cycloaddition with electron-deficient alkenes afforded head-to-head adducts selectively and efficiently. Irradiation of *N*-phenyl aziridine **3** with acrylonitrile gave adducts, but photoreaction of *N*-benzoyl aziridine **4** and thermal reactions of **3** and **4** with alkenes yielded C(γ),N-cleaved products instead of cycloadducts. *N*-trityl aziridine **5** also reacted with electron-deficient alkenes, affording 2,3-*cis*-pyrrolidine derivatives exclusively. A formal synthesis of a pyrrolizidine alkaloid, isoretronecanol (**27**), starting from **5** was achieved in a convenient manner.

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1. Introduction

The 1,3-dipolar cycloaddition of azomethine ylides with alkenes is an important and useful strategy for the construction of nitrogen-containing five-membered hetero-cycles.¹ One of the method for the generation of azomethine ylide is the heating or irradiation of aziridines, most of which bear an adjacent electron-withdrawing or phenyl group.² However, mild and efficient methods for the C,C-bond cleavage of aziridines have not been widely studied.

We have investigated photochemical reactions of α , β unsaturated γ , δ -epoxy nitriles systematically.³ These studies have revealed that carbonyl ylides photochemically generated from epoxy nitriles undergo 1,3-diporlar cycloaddition with electron-rich alkenes to afford tetrahydrofurans.^{3d} On the basis of these studies, we became interested in extending the photochemistry of epoxy nitriles to that of β -aziridinylacrylonitrile.

As part of these studies, we reported in a previous letter that direct irradiation or heating of β -aziridinylacrylonitrile 1 with electron-deficient alkenes causes the ring-opening of 1

e-mail: ishiikei@my-pharm.ac.jp



Scheme 1.

and subsequent cycloaddition reactions, leading to head-tohead adducts selectively and efficiently (Scheme 1).⁴ In this paper, we describe the details of the reactions of nitrile 1, the photochemical behavior of β -aziridinylacrylate 2, and the effects of N-substituents in the aziridine ring [*N*-phenyl, *N*-benzoyl and *N*-trityl aziridines 3–5 (Fig. 1)] on the cycloaddition with alkenes. Furthermore, we describe that using the cycloadducts 5, the formal preparation of a pyrrolizidine alkaloid, isoretronecanol (27),⁵ was achieved conveniently.



Figure 1.

Keywords: Aziridine; Photolysis; [3+2] Cycloaddition; Pyrrolidine; Pyrrolizidine.

^{*} Corresponding author. Tel./fax: +81 424958783;

^{0040–4020/\$ -} see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.031

2. Results and discussion

The *N*-benzylnitrile **1** and ester **2** were prepared from aldehyde **6**⁶ with diethyl cyanomethylphosphonate and diethyl ethoxycarbonylmethylphosphonate in 75% yield (E/Z=44:31) and 75% yield (E/Z=68:7), respectively. *N*-phenyl ester (*E*)-**3** and *N*-trityl ester (*E*)-**5** were synthesized from the corresponding alcohol **7**⁷ and aldehyde **8**,⁸ respectively, as shown in Scheme 2. *N*-Benzoyl ester



Scheme 2. Reagents and conditions: (i) $(EtO)_2 P(O)CH_2CN$, NaH, THF, 0 °C; (ii) $(EtO)_2 P(O)CH_2CO_2Et$, NaH, THF, 0 °C; (iii) oxalyl chloride, DMSO, CH_2Cl_2 , -78 °C; (iv) $(EtO)_2P(O)CH_2CO_2Et$, NaH, CH_2Cl_2 , 0 °C; (v) TFA, MeOH, $CHCl_3$; (vi) $(PhCO)_2O$, NEt₃, $CHCl_3$.



(Z)-1 $\xrightarrow{\text{ii}}$ (E)-1

Scheme 3. Reagents and conditions: (i) $\lambda = 254$ nm, acetonitrile, rt; (ii) $\lambda > 280$ nm, acetone, rt.

Table 1. Photochemical and thermal reactions of aziridine 1 with alkenes or an alkyne^a

(*E*)-4 was prepared in 58% yield by the detritylation and benzoylation of (*E*)-5 (Scheme 2).

Direct irradiation of a solution of (*Z*)-1 in acetonitrile with a low-pressure mercury lamp in a quartz test tube at rt (conversion 83%) afforded dimers **9A** ($32\%^9$) and **9B** (14%) (Scheme 3). On triplet sensitization, the nitrile (*Z*)-1 in acetone with a high-pressure mercury lamp in a Pyrex test tube at rt (conversion 58%) selectively underwent (*E*/*Z*)-isomerization of the side chain leading to (*E*)-1 ($64\%^9$) (Scheme 3).

Since the photolysis of nitrile **1** had given cycloadducts **9** in moderate yield, the reactions of **1** and electron-deficient alkenes or an alkyne were studied. The results are summarized in Table 1 and Figure 2. No significant differences in reactivity between (*E*)- and (*Z*)-**1** were observed (entries 1-4).



Figure 2.

Entry	(<i>E</i>)/(<i>Z</i>)-1	Alkene or alkyne	Reaction time (h)	Conversion (%)	Products and yields (%) ^{9,b}
1	(<i>E</i>)	Acrylonitrile	6	100	(E)-10a (52) and (E) -10b (26)
2	(Z)	Acrylonitrile	7	100	(Z)-10a (52) and (Z) 10b (15)
3	(E)	Methyl acrylate	2	87	(E)-11a (37)
4	(Z)	Methyl acrylate	4	98	(Z)-11a (38) and (Z)-11b (48)
5	(<i>Z</i>)	tert-Butyl acrylate	2 [3] ^c	90 [86]	12a (23) [21] and 12b (49) [13]
6	(Z)	2-Cyclopentenone	2.5	91	13 (39)
7	(Z)	<i>N</i> -Phenylmaleimide	2 [2]	84 [81]	14 (39) [42]
8	(Z)	Methyl propiolate	0.75	66	15 (49)

^a A 0.060 mol L^{-1} solution of 1 in acetonitrile with 10 equiv of alkene or alkyne was irradiated at rt.

^b Isolated yield.

 $^{\rm c}$ Values in square brackets are yields of thermal reactions of 1 with 10 equiv of alkene in refluxing xylene.

The reactions **1** and mono-substituted alkenes selectively afforded 3-substituted pyrrolidines in moderate yields $(62-86\%^9)$ (entries 1–5). The photoreactions of nitrile **1** and dimethyl fumarate or dimethyl acetylenedicarboxylate gave only dimethyl maleate and a complex mixture, respectively.



Figure 3.



Figure 4.

Table 2. Photocher	nical reactions	of aziridines	2–5	with	alkenes
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On the other hand, the reactions of 1 and non-activated (bicyclo[2.2.1]hept-2-ene and cyclohexene) or electron-rich alkenes (ethyl vinyl ether) gave only dimers 9 instead of the adducts with alkenes.

The thermal 1,3-dipolar cycloaddition of an azomethine ylide derived from an aziridine bearing an ester function and electron-deficient alkenes normally affords products possessing the electron-withdrawing group (EWG) at the C(4) position in the pyrrolidine (Fig. 3).^{2b} However, the position of the EWG in our cycloadducts was at C(3). In order to investigate the mechanism of the cycloaddition step between electron-deficient alkenes and ring-cleaved intermediate A (Fig. 3), thermal reactions of 1 and alkenes were performed. A solution of (Z)-1 with *tert*-butyl acrylate or N-phenylmaleimide was heated in refluxing xylene and gave the same adducts 12 (a 21% and b 13%) and 14 $(42\%^9)$ as yielded by the photoreactions, respectively. The results may suggest that the C,C-bond cleavage of aziridine 1 proceeds photochemically or thermally and the cycloaddition occurs thermally.

We next investigated ethyl β -aziridinylacrylate **2** possessing an ester group, which is easily transformed to other functional groups. Direct irradiation of a solution of (*E*)-**2** in acetonitrile with a low-pressure mercury lamp in a quartz test tube at rt (conversion 98%) afforded dimers **16A** (19%⁹) and **16B** (7%⁹) (Fig. 4).

On triplet sensitization, ester (*E*)-**2** in acetone with a highpressure mercury lamp in a Pyrex test tube at rt (conversion 84%) selectively underwent (*E*/*Z*)-isomerization of the side chain leading to (*Z*)-**2** (25%⁹).

Since the photochemical behavior of ester 2 was similar to that of nitrile 1, the reactions of 2 and acrylonitrile were studied (Table 2, Fig. 4).

The structures of the cycloadducts **9–15** were deduced mainly on the basis of their spectral data and were discussed in the previous communication.⁴ Particularly, in the ¹H NMR spectra of the *N*-benzyl pyrrolidines **9–12**, the signals due to H-(3') for 2,3-*cis*-pyrrolidines appear in a lower field (δ 3.18–3.46) than those of 2,3-*trans*-pyrrolidines (δ 2.74–2.85) (Table 3).

The molecular ion peak in the mass spectrum (MS) of 17 indicates the 1:1 adducts of 2 and acrylonitrile. The regioand stereochemistries of 17a and 17b were determined by the H–H and C–H COSY spectra. In particular, the configurations at the 2',3'-positions of 17a and 17b were deduced from a comparison of the chemical shifts

Entry	Aziridine	Alkene	Reaction time (h)	Conversion (%)	Products and yields (%) ^{9,b}
1	(E)- 2	Acrylonitrile	3	100	17a (42) and 17b (21)
2	(E)- 3	Acrylonitrile	2	52	18a (10) and 18b (6)
3	(E)- 4	Acrylonitrile	5	74	20 (30)
4	(E)- 5	Acrylonitrile	1.3	40	22 (43)
5	(E)- 5	Methyl acrylate	4	57	23 (61)

^a A 0.060 mol L^{-1} solution of 2–5 in acetonitrile with 10 equiv of alkene was irradiated at rt.

^b Isolated yield.

Table 3. The chemical shift of H-C(3') in the ¹H NMR spectra for 9–12, 16 and 17

cis-Adduct	δ	trans-Adduct	δ
(E)-10a (Z)-10a ^a (E)-11a ^a (Z)-11a ^a 12a ^a	3.18–3.25 m 3.29 ddd 3.20–3.28 m 3.29–3.37 m 3.18–3.24 m	(<i>E</i>)-10b (<i>Z</i>)-10b ^a (<i>E</i>)-11b (<i>Z</i>)-11b 12b ^a	2.82 ddd 2.85 ddd 2.81 ddd 2.83 ddd 2.74 ddd
17a 9A ^a 9B ^a	3.46 dd 3.36 dd	17b 16A ^a 16B	2.84 ddd 2.49 dd 2.62 dd

^a The stereochemistry was also determined by phase-sensitive NOESY spectrum.

(δ 3.16–3.21 for **17a** and δ 2.84 for **17b**) with those of the compounds described in Table 3.

The molecular ion peak in the mass spectrum (MS) of **16A** and **16B** shows that they are the dimers of **2**. The regioand stereochemistries of **16A** and **16B** were determined from the H–H and C–H COSY spectra and from a comparison of the spectral data with those of **9A** and **9B**. In particular, in the ¹H NMR spectrum, the 2',3'-trans configuration in the pyrrolidine ring was deduced from a comparison of the chemical shifts at the 3'-position of **16A** (δ 2.49) and **16B** (δ 2.62) with the data described in Table 3. Furthermore, in the NOESY spectrum of **16A**, the crosspeaks showed 2',3'-trans and 3',4'-trans configurations in the pyrrolidine ring (Fig. 5). However, the stereochemistries at C(2") for **16A** and **16B** could not be determined.



Figure 5. Phase-sensitive NOESY for 16A and 23.

As the photochemical reactions of *N*-benzyl aziridines **1** and **2** with electron-deficient alkenes afforded the cycloadducts in moderate yields, the effects of other N-substituents in the aziridine ring on the cycloaddition were studied. Aziridines substituted with phenyl or benzoyl groups, which possess stronger electron-withdrawing characteristics than the benzyl group, were supposed to react with electron-rich or non-activated alkenes.^{2b}

Irradiation of a solution of (E)-3 and acrylonitrile in acetonitrile with a low-pressure mercury lamp in a quartz test tube afforded the adducts **18a** and **18b** (Table 2, Fig. 4). The yields of adducts from 3 were reduced in comparison with those from the *N*-benzyl aziridine 2. Aziridine (E)-3 also did not react with electron-rich alkene (ethyl vinyl ether) photochemically giving a complex mixture. On the other hand, the thermal reaction of (E)-3 and 3,4-dihydro-2*H*-pyran in refluxing xylene yielded no adducts but underwent an electrocyclic reaction leading to benzazepine **19** (Fig. 4). This type of rearrangement is also observed by thermal reaction¹⁰ or treatment with silica gel¹¹ of 1-phenyl-2-vinylaziridines.

The structures of **18a** and **18b** were deduced from the chemical shifts for H–(C3') in the ¹H NMR spectra in comparison with those of the adducts shown in Table 3; the signal (δ 3.28) for **18a** appears in a lower field than that for **18b** (δ 3.06–3.11). Furthermore, in the phase-sensitive NOESY spectrum of **18a** the crosspeak between H-2' and H-3' was observed. The structure of **19** was determined on the basis of its spectral data. In particular, the molecular ion peak in MS indicates that **19** is an isomer of **3**, and the ¹H NMR spectrum shows the signals due to four aromatic protons, two isolated alkenic protons and amino moiety (see Section 4).

An acetonitrile solution of **4** and acrylonitrile was irradiated with a low-pressure mercury lamp in a quartz test tube affording the C(γ),N-bond-cleaved product **20** (Table 2, Fig. 4). The thermal reaction of (*E*)-**4** and *tert*-butyl acrylate in refluxing xylene yielded no adducts but a mixture of C(γ),N-bond-cleaved compounds (mainly isomers of **20**) and pyridine derivative **21** (4%⁹) (Fig. 4).¹²

The structure of **20** was deduced from the spectral data (see Section 4). The structure of **21** was determined by a comparison of the spectral data with those of reference 12. The *N*-benzoyl substituent indicated a tendency to cleave the $C(\gamma)$, N-bond on thermal and photochemical reactions.

In order to improve the stereoselectivity at the 2,3-position of the pyrrolidine ring on the cycloaddition, we chose trityl group, which is more bulky than benzyl group, as the N-substituent of aziridine. Acetonitrile solutions of *N*-trityl aziridine **5** with acrylonitrile and methyl acrylate were irradiated with a low-pressure mercury lamp in quartz test tubes affording the adducts **22** and **23**, respectively (Table 2, Fig. 4).

The regio- and stereochemistries of 22 and 23 were determined by the H–H COSY and the phase-sensitive NOESY spectra. In particular, the crosspeaks between H-2' and H-3', between H-3' and H_a-4' and between H_a-4' and H_a-5' are observed in the NOESY spectra of 23 (Fig. 5).

In the case of the reaction of *N*-trityl aziridine **5**, the relative configuration between C(2') and C(3') in the isolated adducts was absolutely *cis*. In the transition state **B** for the formation of **22** and **23**, both the acrylate moiety of the aziridine-ring-cleaved intermediate and the substituent R of alkenes were presumably orientated on the opposite side of the trityl group because of the steric hindrance (Fig. 4).

Since the photolysis of *N*-trityl aziridine **5** and methyl acrylate gave 2,3-*cis*-pyrrolidine **23** in moderate yield, we were interested in the synthesis of a pyrrolizidine alkaloid, isoretronecanol (**27**), using the stereochemistry of **23**. Hydrogenolysis of the side chain in **23** over Pd/C gave no reduced product. After detritylation of **23** with trifluoroacetic acid, reduction of the double bond in **24** over Pd/C proceeded successfully, affording propionate **25** (64%). Cyclization of **25** in toluene gave pyrrolizidine **26**⁵ in 87% yield, which can be transformed by authentic methods⁵ into **27** (Scheme 4).

To clarify the chemical behavior and the utility of



Scheme 4. Reagents and conditions: (i) TFA, rt; (ii) 10% Pd/C, H₂ (1 bar), AcOEt; (iii) toluene, 110 °C.

 β -aziridinylacrylates, further work with 2,3-disubstituted aziridines and the synthetic application of the cycloadducts is currently in progress.

3. Summary

In conclusion, the photoreactions of *N*-benzyl β -aziridinylacrylonitrile **1** and acrylate **2** with electron-deficient alkenes afforded novel head-to-head adducts selectively and efficiently. Aziridines **3** and **4**, possessing the N-conjugated substituent had a tendency to cleave the C(γ),N-bond. *N*-trityl aziridine **5** also reacted with electron-deficient alkenes, yielding 2,3-*cis*-pyrrolidine derivatives selectively. A formal synthesis of a pyrrolizidine alkaloid, isoretronecanol (**27**), starting from the pyrrolidine **23** was achieved in a convenient manner.

4. Experimental

4.1. General

Melting points and boiling points are uncorrected. Melting points were measured with a Yanaco MP-3 apparatus and boiling points were measured with a Büchi Kugel Rohr GKR-50 apparatus. UV spectra were recorded on a Hitachi 124 spectrometer and IR spectra on a Hitachi 215 spectrometer. NMR spectra were obtained with a JEOL JNM-AL300 (300 MHz; AL3), a JEOL JNM-AL400 (400 MHz; AL4) or JEOL JNM-LA500 (500 MHz; LA) spectrometers in CDCl₃ using tetramethylsilane as an internal standard. Mass spectra (MS) and high-resolution MS (HRMS) were taken on a JEOL JMS-700 spectrometer. Column chromatography was performed with Merck silica gel 60 (230–400 mesh) and Chromatorex NH (Fuji Silysia Chemical LTD.), and preparative TLC with Wakogel B-5F.

An Eikosha 60 W low-pressure mercury lamp and a Riko 400 W high-pressure mercury lamp were used for irradiation. The photolysis solutions were purged with argon both before and during irradiation.

4.2. Preparations of aziridines

4.2.1. (*E*)-**3**-(**1-Benzylaziridin-2-yl)acrylonitrile** (*E*)-**1** and (*Z*)-**3**-(**1-benzylaziridin-2-yl)acrylonitrile** (*Z*)-**1**. To a suspension of NaH [1.48 g, 61.8 mmol; prepared from a NaH dispersion (60%, 2.47 g) by washing it twice with hexane (30 mL)] in dry THF (125 mL) was added dropwise a solution of diethyl cyanomethylphosphonate (10.9 g, 61.8 mmol) in dry THF (125 mL) at 0 °C. After the mixture had been stirred for 10 min at 0 °C, a solution of *N*-benzylaziridinecarbaldehyde **6**⁶ (6.64 g, 41.2 mmol) in dry THF (40 mL) was added dropwise, and stirring was continued for 1.5 h at 0 °C. Ice/water was added to the mixture, and the organic phase was extracted with diethyl ether. The ethereal extract was washed with brine, dried with MgSO₄, and concentrated in vacuo, giving a residue that was subjected to flash column chromatography [hexane–ethyl acetate (9:1)] to afford (*E*)-1 (3.30 g, 44%) and (*Z*)-1 (2.34 g, 31%).

Compound (*E*)-1. Bp 130 °C at 0.35 mm Hg; IR (film): 2300 cm⁻¹ (C \equiv N); ¹H NMR (AL4): δ 1.84 (d, 1H, J= 6.4 Hz, H-3'), 1.92 (d, 1H, J= 3.2 Hz, H-3'), 2.09–2.14 (m, 1H, H-2'), 3.47, 3.58 (each d, 2H, J= 13.6 Hz, CH₂Ph), 5.59 (dd, 1H, J= 16.4, 0.8 Hz, H-2), 6.55 (dd, 1H, J= 16.4, 6.8 Hz, H-3), 7.26–7.42 (m, 5H, Ph); ¹³C NMR (AL4): δ 37.7 (t, C-3'), 39.0 (d, C-2'), 64.0 (t, CH₂Ph), 99.8 (d, C-2), 117.1 (s, C-1), 127.2, 127.6 128.3 (3d, 5 C in Ph), 137.9 (s, C in Ph), 153.9 (d, C-3); EI-MS m/z 184 (M⁺, 19%), 104 (4), 91 (100), 77 (3), 65 (10), 51 (3), 39 (7). Anal. Calcd for C₁₂H₁₂N₂: C, 78.23; H, 6.57; N, 15.20%. Found: C, 78.01; H, 6.72; N, 14.85%.

Compound (*Z*)-1. An oil; IR (CHCl₃): 2220 cm⁻¹ (C≡N); ¹H NMR (AL4): δ 1.94 (d, 1H, *J*=6.4 Hz, H-3'), 2.02 (d, 1H, *J*=3.6 Hz, H-3'), 2.56–2.62 (m, 1H, 2'-H), 3.46, 3.66 (each d, 2H, *J*=13.2 Hz, CH₂Ph), 5.39 (dd, 1H, *J*=10.8, 0.8 Hz, H-2), 6.10 (dd, 1H, *J*=16.4, 9.3 Hz, H-3), 7.26–7.42 (m, 5H, Ph); ¹³C NMR (AL4): δ 36.1 (t, C-3'), 39.0 (d, C-2'), 64.1 (t, CH₂Ph), 99.7 (d, C-2), 115.6 (s, C-1), 127.2, 127.8, 128.3 (3d, 5C in Ph), 137.8 (s, C in Ph), 154.2 (d, C-3); EI-MS *m/z* 184 (M⁺, 15%), 104 (4), 91 (100), 77 (3), 65 (9), 51 (3), 39 (6); HRMS calcd for C₁₂H₁₂N₂: 184.1000. Found: 184.1004.

4.2.2. Ethyl (*E*)-3-(1-benzylaziridin-2-yl)acrylate (*E*)-2 and ethyl (*Z*)-3-(1-benzylaziridin-2-yl)acrylate (*Z*)-2. By analogy with the synthesis of 1, aldehyde 6 (6.79 g, 42.1 mmol) was treated with NaH (1.52 g, 63.2 mmol) and diethyl ethoxycarbonylmethylphosphonate (14.2 g, 63.2 mmol) in dry THF at 0 °C, and the resulting mixture was stirred for 1 h at 0 °C. Flash column chromatography [hexane–ethyl acetate (9:1)] of the reaction mixture afforded esters (*E*)-2 (6.61 g, 68%) and (*Z*)-2 (680 mg, 7%).

Compound (*E*)-**2**. An oil; IR (film): 1710 cm⁻¹ (C=O); ¹H NMR (AL4): δ 1.27 (t, 3H, J=6.9 Hz, CH₃), 1.84 (d, 1H, J=6.6 Hz, H-3'), 1.95 (d, 1H, J=3.0 Hz, H-3'), 2.09–2.17 (m, 1H, H-2'), 3.51, 3.54 (each d, 2H, J=13.5 Hz, CH₂Ph), 4.18 (q, 2H, J=6.9 Hz, OCH₂), 6.05 (d, 1H, J=15.8 Hz, H-2), 6.69 (dd, 1H, J=15.8, 7.9 Hz, H-3), 7.24–7.34 (m, 5H, Ph); ¹³C NMR (AL4): δ 14.3 (q, CH₃), 36.8 (t, C-3'), 39.3 (d, C-2'), 60.3, 64.3 (2t, OCH₂, CH₂Ph), 121.8 (d, C-2), 127.0, 127.6, 128.2 (3d, 5C in Ph), 138.3 (s, C in Ph), 147.8 (d, C-3), 165.8 (s, C-1); EI-MS *m/z* 231 (M⁺, 2%), 186 (9),

158 (98), 140 (36), 112 (47), 96 (19), 91 (100), 83 (27); HRMS calcd for $C_{14}H_{17}NO_2$: 231.1259. Found: 231.1259.

Compound (*Z*)-**2**. An oil; IR (film): 1705 cm⁻¹ (C=O); ¹H NMR (AL4): δ 1.30 (t, 3H, *J*=7.1 Hz, CH₃), 1.87 (d, 1H, *J*=6.6 Hz, H-3'), 1.92 (d, 1H, *J*=3.2 Hz, H-3'), 3.43, 3.65 (each d, 2H, *J*=13.4 Hz, CH₂Ph), 3.43–3.48 (m, 1H, H-2'), 4.20 (q, 2H, *J*=7.1 Hz, OCH₂), 5.80 (dd, 1H, *J*=11.5, 8.5 Hz, H-3), 5.86 (d, 1H, *J*=11.5 Hz, H-2), 7.24–7.34 (m, 5H, Ph); ¹³C NMR (AL4): δ 14.4 (q, CH₃), 36.1 (t, C-3'), 37.4 (d, C-2'), 60.7, 64.2 (2t, OCH₂, CH₂Ph), 120.9 (d, C-2), 126.9, 127.8, 128.2 (3d, 5C in Ph), 138.5 (s, C in Ph), 149.7 (d, C-3), 166.2 (s, C-1); EI-MS *m*/*z* 231 (M⁺, 3%), 186 (6), 158 (90), 140 (31), 112 (38), 96 (18), 91 (100), 83 (28); HRMS calcd for C₁₄H₁₇NO₂: 231.1259. Found: 231.1250.

4.2.3. Ethyl (E)-3-(1-phenylaziridin-2-yl)acrylate 3. To a solution of oxalyl chloride (410 mg, 3.2 mmol) in dry CH₂Cl₂ (7.0 mL) was added dropwise a solution of DMSO (440 mg, 5.6 mmol) in dry CH_2Cl_2 (7.0 mL) at -70 °C. After the mixture had been stirred for 20 min at -70 °C, a solution of alcohol 7^7 (404 mg, 2.8 mmol) in dry CH₂Cl₂ (5.0 mL) was added dropwise, and stirring was continued for 15 min at -70 °C. Triethylamine (1.9 mL, 14 mmol) was added slowly to the reaction mixture, which was stirred for 10 min at -70 °C, warmed to 0 °C and further stirred for 2 h. Water was added to the mixture, and the organic phase was extracted with CH₂Cl₂. The organic extract was washed with brine, dried with MgSO₄, and concentrated in vacuo, giving a aldehyde that was used for the next step without further purification. By analogy with the synthesis of 1, the aldehyde (1.03 g, 7.0 mmol) was treated with NaH (202 mg, 8.4 mmol) and diethyl ethoxycarbonylmethylphosphonate (1.88 g, 8.4 mmol) in dry CH₂Cl₂ at 0 °C, and the resulting mixture was stirred for 10 min at 0 °C. Flash column chromatography [hexane-ethyl acetate (3:1)] of the reaction mixture afforded ester (E)-3 (679 mg, 45%). An oil; IR (film): 1710 cm⁻¹ (C=O); ¹H NMR (AL3): δ 1.29 (t, 3H, J = 7.2 Hz, CH₃), 2.35 (d, 1H, J = 3.3 Hz, H-3'), 2.42 (d, 1H, J=6.3 Hz, H-3'), 2.70–2.77 (m, 1H, H-2'), 4.21 (q, 2H, J=7.2 Hz, OCH₂), 6.19 (d, 1H, J=15.6 Hz, H-2), 6.78 (dd, 1H, *J*=15.6 Hz, 7.8, H-3), 6.95–7.05, 7.21–7.29 (m, 5H, Ph); ¹³C NMR (AL4): δ 14.3 (q, CH₃), 35.9 (t, C-3'), 39.6 (d, C-2'), 60.4 (t, OCH₂), 120.3, 122.7, 128.8 (3d, 5C in Ph), 122.5 (d, C-2), 146.7 (d, C-3), 152.9 (s, C in Ph), 165.6 (s, C-1); EI-MS *m*/*z* 217 (M⁺, 16%), 172 (6), 144 (100), 112 (13), 104 (13), 91 (6), 84 (14), 77 (20), 51 (5); HRMS calcd for C₁₅H₁₅NO₂: 217.1103. Found: 217.1104.

4.2.4. Ethyl (E)-3-(1-tritylaziridin-2-yl)acrylate (E)-5. By analogy with the synthesis of 1, aldehyde 8^8 (4.0 g, 12.8 mmol) was treated with NaH (460 mg, 19 mmol) and diethyl ethoxycarbonylmethylphosphonate (4.3 g, 19 mmol) in dry CH₂Cl₂ at 0 °C, and the resulting mixture was stirred for 10 min at 0 °C. Flash column chromatography [hexane-ethyl acetate (9:1)] of the reaction mixture afforded ester (E)-5 (4.26 g, 87%). Colorless crystals; mp 82–83 °C (hexane–ethyl acetate); IR (CHCl₃): 1710 cm⁻ (C=O); ¹H NMR (AL4): δ 1.29 (t, 3H, J=7.3 Hz, CH₃), 1.48 (d, 1H, J = 6.4 Hz, H-3'), 1.80–1.85 (m, 1H, H-2'), 1.92 (d, 1H, J=2.4 Hz, H-3'), 4.20 (q, 2H, J=7.3 Hz, OCH₂), 6.04 (d, 1H, J=15.6 Hz, H-2), 6.94 (dd, 1H, J=15.6, 8.0 Hz, H-3), 7.18–7.45 (m, 9H, Ph), 7.47 (d, 6H, J =

1.2 Hz, Ph); ¹³C NMR (AL4): δ 14.4 (q, CH₃), 30.8 (t, C-3'), 33.2 (d, C-2'), 60.3 (t, OCH₂), 74.4 (s, CPh₃) 121.9 (d, C-2), 127.2, 127.4, 129.0 (3d, 15C in Ph), 143.8 (s, 3C in Ph), 149.0 (d, C-3), 166.0 (s, C-1); EI-MS *m*/*z* 383 (M⁺, 0.1%), 257 (4), 243 (100), 228 (8), 215 (4), 180 (3), 165 (52), 154 (2), 115 (2), 91 (3), 77 (4). Anal. Calcd for C₂₆H₂₅NO₂: C, 81.43; H, 6.57; N, 3.65%. Found: C, 81.30; H, 6.50; N, 3.59%.

4.2.5. Ethyl (E)-3-(1-benzoylaziridin-2-yl)acrylate (E)-4. To a solution of 5 (897 mg, 2.34 mmol) in CHCl₃ (2.3 mL) and MeOH (1.8 mL) was added dropwise trifluoroacetic acid (3.5 mL) at 0 °C. After the mixture had been stirred for 30 min at 0 °C, water was added to the mixture, and the organic phase was extracted with CHCl₃. The organic extract was washed with sat. aqueous NaHCO3 solution and brine, dried with Na₂SO₄, and concentrated in vacuo. To a solution of the residue (236 mg) in CHCl₃ (3 mL) was added triethylamine (0.47 mL) and then benzoic anhydride (378 mg, 1.67 mmol) at 0 °C. After the mixture had been stirred for 2 h at 0 °C, water was added to the mixture, the organic phase was extracted with CHCl₃. The extract was subjected to the same workup as used for the synthesis of **1**. The residue was subjected to flash column chromatography [hexane-ethyl acetate (3:1)] to yield ester (E)-4 (334 mg, 58% from **5**). A colorless oil; IR (film): 1710 cm^{-1} (C=O); ¹H NMR (AL4): δ 1.31 (t, 3H, J = 7.2 Hz, CH₃), 2.43 (d, 1H, J=3.2 Hz, H-3'), 2.85 (d, 1H, J=5.6 Hz, H-3'), 3.11-3.17 (m, 1H, H-2[']), 4.22 (q, 2H, J=7.2 Hz, OCH₂), 6.21 (d, 1H, J=16.0 Hz, H-2), 6.74 (dd, 1H, J=16.0, 8.0 Hz, H-3) 7.45 (t, 2H, J=7.6 Hz, Ph), 7.54–7.59 (m, 1H, Ph), 7.99 (d, 2H, J = 7.6 Hz, Ph); ¹³C NMR (AL4): δ 14.3 (q, CH₃), 33.6 (t, C-3'), 38.0 (d, C-2'), 60.7 (t, OCH₂), 124.4 (d, C-2), 128.4, 129.0, 132.9 (3d, 5C in Ph), 132.3 (s, C in Ph), 144.0 (d, C-3), 165.3 (s, C-1), 177.9 (s, NC=O); EI-MS m/z 245 (M⁺, 7%), 200 (2), 140 (3), 117 (20), 105 (100), 95 (2), 77 (22), 51 (3); HRMS calcd for C₁₄H₁₅NO₃: 245.1052. Found: 245.1053.

4.3. Irradiation of acrylonitrile 1

4.3.1. (2Z,2'RS,3'RS,4'SR,2''SR)-3-[1-Benzyl-4-(1-benzylaziridin-2-yl)-3-cyanopyrrolidin-2-yl]acrylonitrile 9A and (2Z,2'RS,3'RS,4'SR,2''RS)-3-[1-Benzyl-4-(1-benzylaziridin-2-yl)-3-cyanopyrrolidin-2-yl]acrylonitrile 9B. A solution of (*Z*)-1 (733 mg, 3.98 mmol) in acetonitrile (66 mL) was irradiated with a low-pressure mercury lamp in a quartz test tube (conversion 83%) for 6.5 h at rt. After removal of the solvent, flash column chromatography [hexane–ethyl acetate (7:3)] of the residue afforded dimers **9A** (197 mg, 32%) and **9B** (83.8 mg, 14%).⁹

Compound **9A**. Colorless crystals, mp 113–114 °C (hexane/ ethyl acetate); IR (CHCl₃): 2240, 2230 cm⁻¹ (C \equiv N); ¹H NMR (LA): δ 1.65 (d, 1H, J=6.1 Hz, H-3"), 1.75 (d, 1H, J=3.4 Hz, H-3"), 1.84–1.89 (m, 1H, H-2"), 1.96–2.02 (m, 1H, H-4'), 2.45 (dd, 1H, J=10.1, 7.9 Hz, H-5'), 2.73 (dd, 1H, J=10.1, 4.3 Hz, H-5'), 3.31, 3.50 (each d, 2H, J= 12.8 Hz, 1"-CH₂Ph), 3.36 (dd, 1H, J=8.5, 7.0 Hz, H-3'), 3.40, 3.69 (each d, 2H, J=13.4 Hz, 1'-CH₂Ph), 3.72 (dd, 1H, J=9, 7.0 Hz, H-2'), 5.53 (dd, 1H, J=11.0, 0.6 Hz, H-2), 6.59 (dd, 1H, J=11.0, 9.2 Hz, H-3), 7.16–7.33 (m, 10H, Ph); ¹³C NMR (LA): δ 33.9 (t, C-3"), 37.5 (d, C-3'), 40.5 (d, C-2"), 42.5 (d, C-4'), 56.0 (t, C-5'), 57.1 (t, 1'-CH₂Ph), 64.5 (t, 1"-CH₂Ph), 65.0 (d, C-2'), 103.6 (d, C-2), 114.9, 116.9 (2s, C-1, CN), 127.3, 127.4, 128.29, 128.31, 128.4, 128.5 (6d, 10C in Ph), 137.4, 138.6 (2s, 2C in Ph), 151.9 (d, C-3); EI-MS *m*/*z* 368 (M⁺, 0.9%), 277 (4), 261 (12), 210 (23), 158 (8), 120 (27), 91 (100), 65 (5). Anal. Calcd for $C_{24}H_{24}N_4$: C, 78.23; H, 6.57; N, 15.20%. Found: C, 78.17; H, 6.63; N, 15.10%.

Compound 9B. Colorless crystals, mp 58-60 °C (hexane/ ethyl acetate); IR (CHCl₃): 2260, 22 $\hat{4}0$ cm⁻¹ (C \equiv N); ¹H NMR (LA): δ 1.48 (d, 1H, J=6.1 Hz, H-3"), 1.68 (d, 1H, J=3.4 Hz, H-3"), 1.85–1.89 (m, 1H, H-2"), 2.11–2.18 (m, 1H, H-4'), 2.49 (dd, 1H, J=10, 9.2 Hz, H-5'), 2.98 (ddd, 1H, J=10, 5.2, 4.9 Hz, H-5'), 3.07, 4.03 (each d, 2H, J=13.4 Hz, 1''-CH₂Ph), 3.40, 3.83 (each d, 2H, J=13.4 Hz, 1'-CH₂Ph), 3.46 (dd, 1H, J=8, 6.1 Hz, H-3'), 3.75 (dd, 1H, J=9.2, 6.1 Hz, H-2', 5.59 (dd, 1H, J=11.0, 0.6 Hz, H-2), 6.65 (dd, 1H, J = 11.0, 9.2 Hz, H-3), 7.25–7.33 (m, 10H, Ph); ¹³C NMR (LA): δ 33.4 (t, C-3"), 38.8 (d, C-3'), 40.0 (d, C-2"), 43.1 (d, C-4'), 54.4 (t, C-5'), 57.3 (t, 1'-CH₂Ph), 64.2 (t, 1"-CH₂Ph), 65.3 (d, C-2'), 104.0 (d, C-2), 114.8, 117.4 (2s, C-1, CN), 127.1, 127.5, 128.1, 128.38, 128.43, 128.44 (6d, 10C in Ph), 137.5, 138.6 (2s, 2C in Ph), 151.9 (d, C-3); EI-MS *m/z* 368 (M⁺, 0.8%), 277 (4), 261 (15), 210 (31), 158 (6), 120 (31), 91 (100), 65 (5); HRMS calcd for C₂₄H₂₄N₄: 368.2001. Found: 368.2007.

4.3.2. Triplet sensitization of 1. A solution of (Z)-1 (794 mg, 4.31 mmol) in acetone (80 mL) was irradiated with a high-pressure mercury lamp in a Pyrex test tube (conversion 58%) for 20 h at rt. After removal of the solvent, flash column chromatography [hexane–ethyl acetate (7:3)] of the residue afforded (*E*)-1 (295 mg, 64%⁹).

4.4. General procedure for the irradiation of acrylonitrile 1 with various alkenes

A 0.060 mol L⁻¹ solution of (*E*)- or (*Z*)-**1** in dry acetonitrile with 10 equiv of alkene was irradiated with a low-pressure mercury lamp in a quartz test tube at 0 °C. After removal of the solvent, flash column chromatography afforded the adducts. The results are summarized in Table 1.

4.4.1. (2*E*,2^{*t*}*RS*,3^{*t*}*RS*)-3-(1-Benzyl-3-cyanopyrrolidin-2yl)acrylonitrile (*E*)-10a. An oil; IR (CHCl₃): 2240 cm⁻¹ (C \equiv N); ¹H NMR (AL4): δ 2.04–2.20, 2.21–2.30 (each m, 2H, H₂-4^{*t*}), 2.37–2.43 (m with td character, 1H, *J*=9.5, 7 Hz, H-5^{*t*}), 3.13 (ddd, 1H, *J*=9.5, 7.0, 2.9 Hz, H-5^{*t*}), 3.18– 3.25 (m, 1H, H-3^{*t*}), 3.34–3.39 (m, 1H, overlapped with d at δ 3.36, H-2^{*t*}), 3.36, 3.86 (each d, 2H, *J*=13.6 Hz, 1^{*t*}-CH₂Ph), 5.77 (dd, 1H, *J*=16.1, 1.1 Hz, H-2), 6.78 (dd, 1H, *J*=16.1, 7.3 Hz, H-3), 7.24–7.38 (m, 5H, Ph); ¹³C NMR (AL4): δ 28.5 (t, C-4^{*t*}), 33.4 (d, C-3^{*t*}), 51.6 (t, C-5^{*t*}), 57.5 (t, 1^{*t*}-CH₂Ph), 65.5 (d, C-2^{*t*}), 103.7 (d, C-2), 116.2, 118.8 (2s, C-1, CN), 127.2, 128.17, 128.20 (3d, 5C in Ph), 137.1 (s, C in Ph), 151.1 (d, C-3); EI-MS *m/z* 237 (M⁺, 30%), 197 (6), 184 (17), 160 (5), 146 (8), 91 (100), 65 (9); HRMS calcd for C₁₅H₁₅N₃: 237.1266. Found: 237.1271.

4.4.2. (2*E*,2'*RS*,3'*SR*)-3-(1-Benzyl-3-cyanopyrrolidin-2yl)acrylonitrile (*E*)-10b. An oil; IR (CHCl₃): 2220 cm⁻¹ (C \equiv N); ¹H NMR (AL4): δ 2.07–2.16, 2.20–2.30 (each m, 2H, H₂-4'), 2.47–2.55 (m with q character, 1H, J=9 Hz, H-5'), 2.82 (ddd, 1H, J=10.3, 7.7, 5.9 Hz, H-3'), 3.08 (ddd, 1H, J=9.5, 8, 2.9 Hz, H-5'), 3.32 (t, 1H, J=7.7 Hz, H-2'), 3.37, 3.86 (each d, 2H, J=12.8 Hz, 1'-CH₂Ph), 5.77 (dd,

111, J = 9.5, 8, 2.9 Hz, Hz), 5.52 (t, HI, J = 7.7 Hz, Hz), 3.37, 3.86 (each d, 2H, J = 12.8 Hz, 1'-CH₂Ph), 5.77 (dd, 1H, J = 16.1, 0.7 Hz, H-2), 6.58 (dd, 1H, J = 16.1, 7.7 Hz, H-3), 7.23–7.38 (m, 5H, Ph); ¹³C NMR (AL4): δ 27.8 (t, C-4'), 34.0 (d, C-3'), 52.0 (t, C-5'), 57.9 (t, 1'-CH₂Ph), 69.6 (d, C-2'), 103.5 (d, C-2), 116.0, 119.8 (2s, C-1, CN), 127.5, 128.3, 128.4 (3d, 5C in Ph), 137.0 (s, C in Ph), 151.9 (d, C-3); EI-MS *m*/*z* 237 (M⁺, 27%), 197 (6), 184 (16), 160 (6), 146 (10), 91 (100), 65 (9); HRMS calcd for C₁₅H₁₅N₃: 237.1266. Found: 237.1270.

4.4.3. (2Z,2'RS,3'RS)-3-(1-Benzyl-3-cyanopyrrolidin-2vl)acrylonitrile (Z)-10a. Colorless crystals; mp 105-106 °C (hexane-ethyl acetate); IR (CHCl₃): 2230, 2210 cm⁻¹ (C \equiv N); ¹H NMR (LA): δ 2.16–2.24, 2.25– 2.30 (each m, 2H, H₂-4'), 2.31–2.36, 3.12–3.16 (each m, 2H, H₂-5'), 3.29 (ddd, 1H, *J*=8.8, 7.3, 5.5 Hz, H-3'), 3.43, 3.83 (each d, 2H, J = 13.4 Hz, 1'-CH₂Ph), 3.68 (dd, 1H, J = 9.2, 7.3 Hz, H-2'), 5.59 (dd, 1H, J = 11.0, 0.6 Hz, H-2), 6.61 (dd, 1H, J = 11.0, 9.2 Hz, H-3), 7.24–7.36 (5H, m, Ph); ¹³C NMR (LA): δ 28.5 (t, C-4'), 33.3 (d, C-3'), 51.8 (t, C-5'), 57.4 (t, 1'-CH₂Ph), 64.9 (d, C-2'), 103.8 (d, C-2), 114.9, 119.3 (2s, C-1, CN), 127.5, 128.4, 128.6 (3d, 5C in Ph), 137.5 (s, C in Ph), 151.9 (d, C-3); EI-MS *m*/*z* 237 (M⁺, 35%), 197 (5), 184 (20), 160 (7), 146 (10), 91 (100), 65 (9). Anal. Calcd for C₁₅H₁₅N₃: C, 75.92; H, 6.37; N, 17.71%. Found: C, 75.71; H, 6.48; N, 17.61%.

4.4.4. (2*Z*,2*′RS*,3*′SR*)-3-(1-Benzyl-3-cyanopyrrolidin-2-yl)acrylonitrile (*Z*)-10b. A colorless oil; IR (CHCl₃): 2260, 2240 cm⁻¹ (C≡N); ¹H NMR (LA): δ 2.12–2.19, 2.26–2.35 (each m, 2H, H₂-4^{*i*}), 2.48–2.55 (m with q character, 1H, *J*=9 Hz, H-5^{*i*}), 2.85 (ddd, 1H, *J*=10.1, 8.9, 6.7 Hz, H-3^{*i*}), 3.05–3.10 (m with td character, 1H, *J*=9, 3 Hz, H-5^{*i*}), 3.44, 3.83 (each d, 2H, *J*=12.8 Hz, 1^{*i*}-CH₂Ph), 3.75 (dd, 1H, *J*=9.5, 8.9 Hz, H-2^{*i*}), 5.56 (d, 1H, *J*=11.0 Hz, H-2), 6.28 (dd, 1H, *J*=11.0, 9.5 Hz, H-3), 7.25–7.33 (m, 5H, Ph); ¹³C NMR (LA): δ 27.8 (t, C-4^{*i*}), 33.6 (d, C-3^{*i*}), 52.1 (t, C-5^{*i*}), 57.8 (t, 1^{*i*}-CH₂Ph), 68.3 (d, C-2^{*i*}), 104.2 (d, C-2), 114.7, 119.5 (2s, C-1, CN), 127.6, 128.4, 128.7 (3d, 5C in Ph), 137.5 (s, C in Ph), 151.2 (d, C-3); EI-MS *m/z* 237 (M⁺, 35%), 197 (7), 184 (17), 160 (8), 146 (14), 91 (100), 65 (10); HRMS calcd for C₁₅H₁₅N₃: 237.1266. Found: 237.1270.

4.4.5. Methyl (2RS,3RS)-1-benzyl-2-[(*E*)-2-cyanovinyl]pyrrolidine-3-carboxylate (*E*)-11a. An oil; IR (CHCl₃): 2210 (C \equiv N), 1725 cm⁻¹ (C=O); ¹H NMR (AL4): δ 1.95– 2.14, 2.15–2.26 (each m, 2H, H₂-4), 2.40–2.47 (m with td character, 1H, *J*=9.5, 6 Hz, H-5), 3.04–3.10 (m with ddd character, 1H, *J*=9, 7.5, 2 Hz, H-5), 3.20–3.28 (m with q character, 1H, *J*=9 Hz, H-3), 3.41, 3.80 (each d, 2H, *J*= 13.2 Hz, 1-CH₂Ph), 3.51 (ddd, 1H, *J*=9.2, 7.0, 1.1 Hz, H-2), 3.68 (s, 3H, OCH₃), 5.53 (dd, 1H, *J*=16.1, 1.1 Hz, H-2[']), 6.78 (dd, 1H, *J*=16.1, 7.0 Hz, H-1[']), 7.24–7.35 (m, 5H, Ph); ¹³C NMR (AL3): δ 26.7 (t, C-4), 48.1 (d, C-3), 52.0 (q, OCH₃), 52.4 (t, C-5), 58.1 (t, 1-CH₂Ph), 66.0 (d, C-2), 101.6 (d, C-2[']), 116.9 (s, CN), 127.2, 128.3, 128.4 (3d, 5C in Ph), 137.9 (s, C in Ph), 152.8 (d, C-1[']), 171.8 (s, CO₂); EI-MS *m*/*z* 270 (M⁺, 18%), 230 (15), 211 (22), 179 (51), 91 (100), 65 (11); HRMS calcd for $C_{16}H_{18}N_2O_2{:}$ 270.1368. Found: 270.1371.

4.4.6. Methyl (2RS,3SR)-1-benzyl-2-[(E)-2-cyanovinyl]pyrrolidine-3-carboxylate (E)-11b. An oil; IR (CHCl₃): 2210 (C \equiv N), 1725 cm⁻¹ (C=O); ¹H NMR (AL4): δ 2.01– 2.14 (m, 2H, H₂-4), 2.34–2.42 (m with q character, 1H, J =9 Hz, H-5), 2.81 (ddd, 1H, J=9.9, 7.7, 5.5 Hz, H-3), 2.98-3.03 (m with ddd character, 1H, J=9.5, 7.3, 2.9 Hz, H-5), 3.30, 3.85 (each d, 2H, J = 12.8 Hz, $1-CH_2$ Ph), 3.34-3.39 (m with t character, 1H, J=7 Hz, 2-H), 3.72 (s, 3H, OCH₃), 5.69 (dd, 1H, J = 16.3, 1.1 Hz, H-2'), 6.70 (dd, 1H, J = 16.3, 7.2 Hz, H-1'), 7.24–7.35 (m, 5H, Ph); 13 C NMR (AL4): δ 27.2 (t, C-4), 49.2 (d, C-3), 52.2 (q, OCH₃), 52.7 (t, C-5), 58.5 (t, 1-CH₂Ph), 68.6 (d, C-2), 101.3 (d, C-2'), 116.9 (s, CN), 127.2, 128.3, 128.5 (3d, 5C in Ph), 138.1 (s, C in Ph), 155.0 (d, C-1[']), 173.5 (s, CO₂); EI-MS m/z 270 (M⁺, 17%), 230 (27), 211 (26), 179 (75), 91 (100), 65 (11); HRMS calcd for C₁₆H₁₈N₂O₂: 270.1368. Found: 270.1364.

4.4.7. Methyl (2RS,3RS)-1-benzyl-2-[(Z)-2-cyanovinyl]pyrrolidine-3-carboxylate (Z)-11a. An oil; bp 160 °C at 0.40 mm Hg; IR (CHCl₃): 2220 (C \equiv N) and 1730 cm⁻¹ (C=O); ¹H NMR (AL4): δ 2.01–2.09, 2.23–2.89 (each m, 2H, H₂-4), 2.45 (dt, 1H, J=9.5, 7.0 Hz, H-5), 3.04–3.09 (m, 1H, H-5), 3.29-3.37 (m with q character, 1H, J=9 Hz, H-3), 3.54, 3.76 (each d, 2H, J = 13.2 Hz, 1-CH₂Ph), 3.66 (s, 3H, OCH_3), 3.86–3.91 (m with dd character, 1H, J=9.5, 9.2 Hz, H-2), 5.35 (dd, 1H, J = 11.0, 0.7 Hz, H-2'), 6.42 (dd, 1H, J = 11.0, 9.5 Hz, H-1'), 7.29–7.31 (5H, m, Ph); ¹³C NMR (AL3): δ 27.1 (t, C-4), 47.7 (d, C-3), 51.8 (q, OCH₃), 52.4 (t, C-5), 57.7 (t, 1-CH₂Ph), 65.2 (d, C-2), 101.0 (d, C-2'), 115.3 (s, CN), 127.2, 128.2, 128.7 (3d, 5C in Ph), 138.1 (s, C in Ph), 153.3 (d, C-1'), 172.6 (s, CO₂); EI-MS *m*/*z* 270 (M⁺, 33%), 230 (16), 211 (24), 184 (15), 179 (62), 91 (100), 65 (8). Anal. Calcd for C₁₆H₁₈N₂O₂: C, 71.09; H, 6.71; N, 10.36%. Found: C, 70.94; H, 6.70; N, 10.30%.

4.4.8. Methyl (2RS,3SR)-1-benzyl-2-[(Z)-2-cyanovinyl]pyrrolidine-3-carboxylate (Z)-11b. Colorless crystals; mp 45–48 °C; IR (CHCl₃): 2220 (C \equiv N), 1730 cm⁻¹ (C=O); ¹H NMR (AL4): δ 2.05–2.23 (m, 2H, H₂-4), 2.37–2.45 (m, 1H, H-5), 2.83 (ddd, 1H, J = 10.6, 8.4, 5.5 Hz, H-3), 3.00– 3.05 (m with ddd character, 1H, J=9.5, 8.1, 2 Hz, H-5), 3.40, 3.80 (each d, 2H, J = 13.2 Hz, $1-CH_2$ Ph), 3.61-3.67 (m with t character, 1H, J=9 Hz, H-2), 3.75 (s, 3H, OCH₃), 5.42 (dd, 1H, J = 11.0, 0.7 Hz, H-2'), 6.38 (dd, 1H, J = 11.0,9.2 Hz, H-1'), 7.25–7.32 (m, 5H, Ph); 13 C NMR (AL4): δ 26.8 (t, C-4), 49.0 (d, C-3), 52.3 (q, OCH₃), 53.1 (t, C-5), 58.4 (t, 1-CH₂Ph), 68.3 (d, C-2), 101.8 (d, C-2'), 115.2 (s, CN), 127.1, 128.1, 128.6 (3d, 5C in Ph), 138.2 (s, C in Ph), 154.3 (d, C-1^{\prime}), 173.0 (s, CO₂); EI-MS m/z 270 (M⁺, 23%), 230 (34), 211 (29), 179 (100), 91 (82), 65 (9); HRMS calcd for C₁₆H₂₈N₂O₂: 270.1368. Found: 270.1367.

4.4.9. *tert*-Butyl (2*RS*,3*RS*)-1-benzyl-2-[(*Z*)-2-cyanovinyl]pyrrolidine-3-carboxylate 12a. Colorless crystals; mp 54–55 °C; IR (CHCl₃): 2230 (C \equiv N), 1725 cm⁻¹ (C=O); ¹H NMR (LA): δ 1.42 (s, 9H, CMe₃), 1.96–2.03, 2.16–2.26 (each m, 2H, H₂-4), 2.46 (dt, 1H, *J*=9.2, 7.0 Hz, H-5), 3.02 (ddd, 1H, *J*=9.2, 8, 2 Hz, H-5), 3.18–3.24 (m with q character, 1H, *J*=9 Hz, H-3), 3.55, 3.75 (each d, 2H, *J*=13.4 Hz, 1-CH₂Ph), 3.88–3.92 (m with t character, 1H, $J=9.5 \text{ Hz}, 2-\text{H}, 5.34 \text{ (dd, 1H, } J=11.0, 0.9 \text{ Hz}, \text{H-2'}\text{)}, 6.45 \text{ (dd, 1H, } J=11.0, 10 \text{ Hz}, \text{H-1'}\text{)}, 7.22-7.31 \text{ (m, 5H, Ph)}; {}^{13}\text{C}$ NMR (LA): δ 27.0 (t, C-4), 28.1 (q, CMe_3), 48.5 (d, C-3), 52.4 (t, C-5), 57.7 (t, 1-CH_2Ph), 65.2 (d, C-2), 81.1 (s, CMe_3), 100.7 (d, C-2'), 115.5 (s, CN), 127.2, 128.3, 128.7 (3d, 5C in Ph), 138.4 (s, C in Ph), 153.7 (d, C-1'), 171.2 (s, CO_2); EI-MS *m*/*z* 312 (M⁺, 11%), 255 (21), 239 (17), 221 (19), 211 (11), 184 (9), 165 (37), 133 (14), 91 (100), 41 (9); HRMS calcd for C₁₉H₂₄N₂O₂: 312.1838. Found: 312.1834.

4.4.10. tert-Butyl (2RS,3SR)-1-benzyl-2-[(Z)-2-cyanovinyl]pyrrolidine-3-carboxylate 12b. Colorless needles; mp 78-79 °C (hexane/ethyl acetate); IR (CHCl₃): 2230 $(C \equiv N)$, 1725 cm⁻¹ (C=O); ¹H NMR (LA): δ 1.48 (s, 9H, CMe₃), 1.99–2.08, 2.12–2.18 (each m, 2H, H₂-4), 2.37–2.43 (m with q character, 1H, J=9 Hz, H-5), 2.74 (ddd, 1H, J=10.3, 8, 5 Hz, H-3), 2.97-3.02 (m with ddd character, 1H, J=9, 8, 2 Hz, H-5), 3.38, 3.82 (each d, 2H, J=13.1 Hz, 1-CH₂Ph), 3.57–3.61 (m with dd character, 1H, J=9, 8 Hz, H-2), 5.40 (dd, 1H, J = 11.0, 0.6 Hz, H-2'), 6.37 (dd, 1H, J = 11.0, 9.5 Hz, H-1'), 7.21–7.31 (m, 5H, Ph); ¹³C NMR (LA): δ 26.6 (t, C-4), 27.9 (q, CMe₃), 50.2 (d, C-3), 53.2 (t, C-5), 58.3 (t, 1-CH₂Ph), 68.3 (d, C-2), 81.4 (s, CMe₃), 101.6 (d, C-2'), 115.6 (s, CN), 127.2, 128.2, 128.7 (3d, 5C in Ph), 138.5 (s, C in Ph), 154.9 (d, C-1[']), 172.0 (s, CO₂); EI-MS m/z 312 (M⁺, 4%), 255 (39), 239 (15), 216 (10), 165 (48), 91 (100), 41 (6). Anal. Calcd for C₁₉H₂₄N₂O₂: C, 73.05; H, 7.74; N, 8.97%. Found: C, 73.14; H, 7.81; N, 8.92%.

4.4.11. (2Z,1'RS,2'RS,5'SR)-3-(3-Benzyl-8-oxo-3-azabicyclo[3.3.0]oct-2-yl)acrylonitrile 13. Colorless crystals; mp 91-92 °C (hexane/ethyl acetate); IR (CHCl₃): 2220 $(C \equiv N)$, 1730 cm⁻¹ (C=O); ¹H NMR (AL4): δ 1.80–1.89, 2.03-2.14 (each m, 2H, H2-6'), 2.21-2.30 (m with dddd character, 1H, J=18, 8, 5, 1 Hz, H-7'), 2.35-2.46 (m, 1H, H-7'), 2.54 (dd, 1H, J=9.8, 8 Hz, H-4'), 2.78–2.83 (m with t character, 1H, J=8 Hz, H-1[']), 2.89 (dd, 1H, J=9.8, 1.4 Hz, H-4'), 2.90–2.98 (m, 1H, H-5'), 3.17, 3.79 (each d, 2H, J=13.8 Hz, 3'-CH₂Ph), 3.68 (dd, 1H, J=9.8, 7.3 Hz, H-2'), 5.51 (dd, 1H, J=11.0, 0.7 Hz, H-2), 6.46 (dd, 1H, J=11.0, 9.8 Hz, H-3), 7.22–7.32 (5H, m, Ph); 13 C NMR (AL3): δ 28.3 (t, C-6'), 38.0 (d, C-5'), 39.4 (t, C-7'), 55.2 (d, C-1'), 57.9, 61.0 (2t, C-4', 1-CH₂Ph), 68.3 (d, C-2'), 102.1 (d, C-2), 115.3 (s, C-1), 127.0, 128.1, 128.2 (3d, 5C in Ph), 138.2 (s, C in Ph), 153.0 (d, C-3), 217.9 (s, C-8'); EI-MS m/z 266 (M⁺, 33%), 226 (5), 210 (20), 184 (13), 175 (16), 91 (100), 65 (9). Anal. Calcd for C₁₇H₁₈N₂O: C, 76.66; H, 6.81; N, 10.52%. Found: C, 76.71; H, 6.86; N, 10.46%.

4.4.12. (2Z,1^{*i*}*RS*,2^{*i*}*RS*,5^{*i*}*SR*)-3-(3-Benzyl-6,8-dioxo-7phenyl-3,7-diazabicyclo[3.3.0]oct-2-yl)acrylonitrile 14. Colorless crystals; mp 49–51 °C; IR (CHCl₃): 2230 (C \equiv N), 1720 cm⁻¹ (C=O); ¹H NMR (LA): δ 2.56 (dd, 1H, *J*=10.1, 7.9 Hz, H-4^{*i*}), 3.33, 3.83 (each d, 2H, *J*= 13.4 Hz, 3^{*i*}-CH₂Ph), 3.36 (td, 1H, *J*=7.9, 0.6 Hz, H-5^{*i*}), 3.47 (d, 1H, *J*=10.1 Hz, H-4^{*i*}), 3.54 (t, 1H, *J*=7.9 Hz, H-1^{*i*}), 3.79 (dd, 1H, *J*=9.8, 7.9 Hz, H-2^{*i*}), 5.59 (dd, 1H, *J*= 11.0, 0.6 Hz, H-2), 6.45 (dd, 1H, *J*=11.0, 9.8 Hz, H-3), 7.20–7.23, 7.24–7.32, 7.39–7.43, 7.47–7.51 (4m, 10H, Ph); ¹³C NMR (LA): δ 43.4 (d, C-5^{*i*}), 48.8 (d, C-1^{*i*}), 55.9 (t, C-4^{*i*}), 56.8 (t, 3^{*i*}-CH₂Ph), 66.4 (d, C-2^{*i*}), 103.5 (d, C-2), 115.2 (s, C-1), 126.3, 127.6, 128.4, 128.5, 128.8, 129.2 (6d, 10C in Ph), 131.7, 136.8 (2s, 2C in Ph), 151.1 (d, C-3), 174.4, 177.4 (2s, C-6', C-8'); EI-MS m/z 357 (M⁺, 55%), 317 (21), 266 (21), 184 (19), 119 (8), 91 (100), 65 (8); HRMS calcd for C₂₂H₁₉N₃O₂: 357.1477. Found: 357.1479.

4.4.13. Methyl (*Z*)-1-benzyl-2-(2-cyanovinyl)-3-pyrroline-3-carboxylate 15. An oil; bp 160 °C at 0.20 mm Hg; IR (CHCl₃): 2230 (C=N), 1720 cm⁻¹ (C=O); ¹H NMR (LA): δ 3.50 (ddd, 1H, *J*=17, 5, 2 Hz, H-5), 3.74 (s, 3H, OCH₃), 3.74, 3.97 (each d, 2H, *J*=13.4 Hz, 1-CH₂Ph), 3.87 (ddd, 1H, *J*=17, 5.5, 2 Hz, H-5), 4.82–4.87 (m, 1H, 2-H), 5.39 (dd, 1H, *J*=10.7, 0.6 Hz, H-2'), 6.37 (dd, 1H, *J*=10.7, 9.2 Hz, H-1'), 6.89 (q, 1H, *J*=2.1 Hz, H-4), 7.23–7.33 (m, 5H, Ph); ¹³C NMR (LA): δ 51.7 (q, OCH₃), 57.5 (t, 1-CH₂Ph), 59.0 (t, C-5), 68.8 (d, C-2), 100.7 (d, C-2'), 115.7 (s, CN), 127.3, 128.4, 128.6 (3d, 5C in Ph), 133.5 (s, C-3), 138.3 (s, C in Ph), 141.8 (d, C-4), 153.2 (d, C-1'), 163.0 (s, CO₂); EI-MS *m*/*z* 268 (M⁺, 8%), 216 (17), 177 (15), 91 (100), 65 (14); HRMS calcd for C₁₆H₁₆N₂O₂: 268.1212. Found: 268.1218.

4.5. Thermal reactions of nitrile 1 with alkenes

By analogy with the photoreactions of (Z)-1, a 0.060 mol L^{-1} solution of (Z)-1 in xylene with 10 equiv of *tert*-butyl acrylate or *N*-phenylmaleimide was heated under reflux. Flash column chromatography afforded the adducts. The results are summarized in Table 1.

4.6. Irradiation of ethyl acrylate 2

4.6.1. Ethyl (2E,2'RS,3'SR,4'SR)-3-[1-benzyl-4-(1-benzyl-aziridin-2-yl)-3-ethoxycarbonylpyrrolidin-2-yl]acrylate 16A and ethyl (2E,2'RS,3'SR,4'SR)-3-[1-benzyl-4-(1-benzylaziridin-2-yl)-3-ethoxycarbonylpyrrolidin-2-yl]acryl-ate 16B. By analogy with the photolysis of 1, a solution of (*E*)-2 (51.8 mg, 0.223 mmol) in acetonitrile was irradiated (conversion 98%) for 2 h at rt. Preparative TLC [hexane-ethyl acetate-diethylamine (9:1:0.5)] of the reaction mixture afforded dimers 16A (9.8 mg, 19%⁹) and 16B (3.3 mg, 7%⁹).

Compound **16A**. An oil; IR (CHCl₃): 1720 cm^{-1} (C=O); ¹H NMR (LA): δ 1.22, 1.31 (2t, 6H, J=7.0 Hz, 2CH₃), 1.36 (d, 1H, J = 6.1 Hz, H-3"), 1.63–1.67 (m, 2H, H-2", H-3"), 2.16-2.22 (m, 1H, H-4'), 2.38 (dd, 1H, J=9.7, 8 Hz, H-5'), 2.49 (dd, 1H, J=8.2, 5.2 Hz, H-3'), 2.77 (dd, 1H, J=9.7, 2.4 Hz, H-5'), 3.14, 3.88 (each d, 2H, J=13.1 Hz, 1"-CH₂Ph), 3.23, 3.56 (each d, 2H, J = 12.8 Hz, 1'-CH₂Ph), $3.25 (m, 1H, H-2'), 4.21, 4.22 (2q, 4H, J=7.0 Hz, 20CH_2),$ 6.00 (dd, 1H, J = 15.6, 0.6 Hz, H-2), 6.80 (dd, 1H, J = 15.6, 0.6 Hz, H-2)7.9 Hz, H-3), 7.22–7.33 (m, 10H, 2Ph); 13 C NMR (LA): δ 14.2, 14.3 (2q, 2CH₃), 33.2 (t, C-3"), 42.8 (d, C-2"), 43.5 (d, C-4'), 54.2 (d, C-3'), 55.6 (t, C-5'), 58.0 (t, 1"-CH₂Ph), 60.4, 60.8 (2t, 2OCH₂), 64.7 (t, 1'-CH₂Ph), 69.0 (d, C-2'), 123.2 (d, C-2), 127.0, 127.1, 128.26, 128.34, 128.5 (5d, 10C in Ph), 138.7, 139.2 (2s, 2C in Ph), 148.2 (d, C-3), 166.2, 173.3 $(2s, C-1, 3'-CO_2);$ EI-MS m/z 462 (M⁺, 2%), 417 (6), 342 (41), 282 (17), 233 (17), 120 (7), 91 (100); HRMS calcd for C₂₈H₃₄N₂O₄: 462.2518. Found: 462.2527.

Compound **16B**. An oil; IR (CHCl₃): 1730 cm⁻¹ (C=O); ¹H NMR (LA): δ 1.22, 1.30 (2t, 6H, *J*=7.0 Hz, 2CH₃), 1.44 (d, 1H, *J*=6.4 Hz, H-3"), 1.68–1.73 (m, 2H, H-2", H-3"), 1.99-2.05 (m, 1H, H-4'), 2.40 (dd, 1H, J=9.8, 7.9 Hz, H-5'), 2.62 (dd, 1H, J=8.5, 5.8 Hz, H-3'), 2.73 (dd, 1H, J=9.8, 2.7 Hz, H-5'), 3.16, 3.86 (each d, 2H, J=13.4 Hz, 1''-CH₂Ph), 3.19, 3.50 (each d, 2H, J = 12.8 Hz, 1'-CH₂Ph), 3.34 (m with t character, 1H, J=8 Hz, H-2'), 4.11, 4.20 $(2q, 4H, J=7.0 \text{ Hz}, 20\text{CH}_2), 6.05 \text{ (dd, 1H, } J=15.6, 0.6 \text{ Hz},$ H-2), 6.87 (dd, 1H, J = 15.6, 7.9 Hz, H-3), 7.15–7.31 (m, 10H, 2Ph); ¹³C NMR (LA): δ 14.17, 14.23 (2q, 2CH₃), 33.0 (t, C-3"), 43.5 (d, C-2"), 44.5 (d, C-4'), 53.5 (d, C-3'), 57.0 (t, C-5'), 57.8 (t, 1"-CH₂Ph), 60.4, 60.8 (2t, 2OCH₂), 64.7 (t, 1'-CH₂Ph), 68.5 (d, C-2'), 123.5 (d, C-2), 127.0, 127.1, 128.2, 128.3, 128.5 (5d, 10C in Ph), 138.6, 138.9 (2s, 2C in Ph), 148.2 (d, C-3), 166.2, 172.9 (2s, C-1, 3'-CO₂); EI-MS *m*/*z* 462 (M⁺, 2%), 417 (12), 342 (100), 282 (20), 233 (23), 120 (6), 91 (77); HRMS calcd for C₂₈H₃₄N₂O₄: 462.2518. Found: 462.2516.

4.6.2. Triplet sensitization of 2. By analogy with the photolysis of **1**, a solution of (*E*)-**2** (424 mg, 1.83 mmol) in acetone (31 mL) was irradiated with a high-pressure mercury lamp in a Pyrex test tube (conversion 84%) for 11.5 h at rt. After removal of the solvent, flash column chromatography [hexane–ethyl acetate (3:2)] of the residue afforded (*Z*)-**2** (67 mg, $25\%^9$).

4.6.3. Ethyl (2E,2'RS,3'RS)-**3**-(**1**-benzyl-**3**-cyanopyrrolidin-**2**-yl)acrylate **17a** and ethyl (2E,2'RS,3'SR)-**3**-(**1**benzyl-**3**-cyanopyrrolidin-**2**-yl)acrylate **17b**. By analogy with the photoreactions of **1**, a 0.060 mol L⁻¹ solution of (*E*)-**2** in dry acetonitrile with 10 equiv of acrylonitrile was irradiated. Preparative TLC [hexane–ethyl acetate (3:1)] afforded the adducts. The results are summarized in Table 2.

Compound 17a. An oil; bp 180 °C at 0.20 mm Hg (decomp.); IR (CHCl₃): 2250 (C≡N), 1715 cm (C=O); ¹H NMR (LA): δ 1.31 (t, 3H, J=7.0 Hz, CH₃), 2.13-2.27 (m, 2H, H₂-4'), 2.28-2.34 (m with dt character, 1H, J=9, 8 Hz, H-5', 3.07-3.12 (m with ddd character, 1H, J=9, 8, 3 Hz, H-5'), 3.16–3.21 (m with ddd character, 1H, J=8.5, 7, 6.1 Hz, H-3', 3.27, 3.92 (each d, 2H, J=13.1 Hz, 1'-CH₂Ph), 3.27–3.31 (m, 1H, H-2'), 4.22 (q, 2H, J =7.0 Hz, OCH₂), 6.14 (dd, 1H, J = 15.6, 0.9 Hz, H-2), 6.98 (dd, 1H, J = 15.6, 8.2 Hz, H-3), 7.22–7.33 (m, 5H, Ph); ¹³C NMR (LA): δ 14.2 (q, CH₃), 28.4 (t, C-4'), 33.7 (d, C-3'), 51.3 (t, C-5'), 57.1 (t, 1'-CH₂Ph), 60.7 (t, OCH₂), 65.8 (d, C-2'), 119.6 (s, CN), 126.2 (d, C-2), 127.3, 128.3, 128.5 (3d, 5C in Ph), 137.9 (s, C in Ph), 144.0 (d, C-3), 165.3 (s, C-1); EI-MS m/z 284 (M⁺, 10%), 255 (11), 239 (10), 211 (35), 193 (68), 158 (38), 140 (14), 112 (19), 91 (100); HRMS calcd for C₁₇H₂₀N₂O₂: 284.1525. Found: 284.1529.

Compound **17b.** An oil; IR (CHCl₃): 2240 (C=N), 1715 cm⁻¹ (C=O); ¹H NMR (LA): δ 1.31 (t, 3H, J= 7.0 Hz, CH₃), 2.05–2.12, 2.26–2.30 (2m, 2H, H₂-4'), 2.38– 2.44 (m with q character, 1H, J=9 Hz, H-5'), 2.84 (ddd, 1H, J=10.7, 8.2, 6.4 Hz, H-3'), 3.03 (ddd, 1H, J=9.5, 8.2, 2.7 Hz, H-5'), 3.25, 3.93 (each d, 2H, J=12.8 Hz, 1'-CH₂Ph), 3.27 (t, 1H, J=8.2 Hz, 2'-H), 4.22 (q, 2H, J= 7.0 Hz, OCH₂), 6.20 (dd, 1H, J=15.9, 0.6 Hz, H-2), 6.79 (dd, 1H, J=15.9, 8.2 Hz, H-3), 7.23–7.33 (m, 5H, Ph); ¹³C NMR (LA): δ 14.2 (q, CH₃), 27.7 (t, C-4'), 33.8 (d, C-3'), 51.8 (t, C-5'), 57.6 (t, 1'-CH₂Ph), 60.7 (t, OCH₂), 69.5 (d, C-2'), 120.5 (s, CN), 125.6 (d, C-2), 127.4, 128.4, 128.6 (3d, 5C in Ph), 137.7 (s, C in Ph), 145.1 (d, C-3), 165.5 (s, C-1); EI-MS m/z 284 (M⁺, 5%), 255 (11), 239 (15), 211 (25), 193 (67), 158 (27), 140 (12), 112 (15), 91 (100); HRMS calcd for C₁₇H₂₀N₂O₂: 284.1525. Found: 284.1523.

4.7. Reaction of N-phenylaziridine (E)-3

4.7.1. Ethyl (2E,2'RS,3'RS)-**3**-(**3**-cyano-1-phenylpyrrolidin-2-yl)acrylate 18a and ethyl (2E,2'RS,3'SR)-**3**-(**3**-cyano-1-phenylpyrrolidin-2-yl)acrylate 18b. By analogy with the photoreactions of (Z)-1, a 0.060 mol L⁻¹ solution of (E)-**3** in dry acetonitrile with 10 equiv of acrylonitrile was irradiated. Preparative TLC [SiO₂; hexane–ethyl acetate (3:1)] afforded the adducts. The results are summarized in Table 2.

Compound 18a. An oil; IR (CHCl₃): 2240 (C \equiv N), 1720 cm⁻¹ (C=O); ¹H NMR (AL3): δ 1.28 (t, 3H, J= 7.2 Hz, CH₃), 2.26–2.41, 2.44–2.53 (2m, 2H, H₂-4'), 3.28 (ddd, 1H, J = 12.0, 7.8, 6.3 Hz, H-3'), 3.34-3.43 (m with td character, 1H, J=9, 7 Hz, H-5'), 3.60-3.67 (m with td character, 1H, J=9, 2 Hz, H-5'), 4.19 (q, 2H, J=7.2 Hz, OCH_2), 4.55–4.60 (m, 1H, H-2'), 6.00 (dd, 1H, J=15.6, 1.5 Hz, H-2), 6.51–6.54 (m with d character, 2H, J=7.9 Hz, H-2", H-6"), 6.75–6.80 (m with t character, 1H, J=7.3 Hz, H-4''), 7.04 (dd, 1H, J=15.6, 4.8 Hz, H-3), 7.21-7.26 (m, 2H, H-3", H-5"); ¹³C NMR (AL4): δ 14.3 (q, CH₃), 27.9 (t, C-4'), 33.1 (d, C-3'), 47.0 (t, C-5'), 59.9 (d, C-2'), 60.8 (t, OCH₂), 112.1 (d, C-2", C-6"), 117.5 (s, CN), 117.7 (d, C-4"), 125.3 (d, C-2), 129.2 (d, C-3", C-5"), 142.4 (d, C-3), 145.4 (s, C-1"), 165.1 (s, C-1); EI-MS *m/z* 270 (M⁺, 68%), 241 (70), 225 (34), 197 (50), 171 (18), 144 (100), 112 (16), 104 (11), 84 (13), 77 (24); HRMS calcd for C₁₇H₂₀N₂O₂: 270.1368. Found: 270.1371.

Compound 18b. An oil; IR (CHCl₃): 2240 (C \equiv N), 1720 cm⁻¹ (C=O); ¹H NMR (AL3): δ 1.27 (t, 3H, J= 7.2 Hz, CH₃), 2.32–2.39 (m, 2H, H₂-4'), 3.06–3.11 (m, 1H, H-3'), 3.57–3.72 (m, 2H, H₂-5'), 4.18 (q, 2H, J=7.2 Hz, OCH₂), 4.58–4.62 (m, 1H, H-2'), 5.99 (dd, 1H, J=15.3, 1.7 Hz, H-2), 6.54–6.58 (m with d character, 2H, J=9 Hz, H-2'', H-6''), 6.76–6.82 (m with t character, 1H, J=7.3 Hz, H-4''), 6.86 (dd, 1H, J=15.3, 4.8 Hz, H-3), 7.20–7.28 (m, 2H, H-3", H-5"); ¹³C NMR (AL4): δ 14.3 (q, CH₃), 27.8 (t, C-4'), 34.7 (d, C-3'), 47.2 (t, C-5'), 60.9 (t, OCH₂), 63.3 (d, C-2'), 112.6 (d, C-2", C-6"), 117.8 (d, C-4"), 119.7 (s, CN), 124.0 (d, C-2), 129.2 (d, C-3", C-5"), 144.3 (d, C-3), 145.6 (s, C-1"), 165.4 (s, C-1); EI-MS *m*/*z* 270 (M⁺, 67%), 241 (72), 225 (32), 197 (53), 171 (24), 144 (100), 112 (15), 104 (12), 84 (13), 77 (28); HRMS calcd for $C_{17}H_{20}N_2O_2$: 270.1368. Found: 270.1365.

4.7.2. Ethyl 2,5-dihydro-1*H***-1-benzazepine-5-carboxylate 19.** A solution of (*E*)-**3** (100 mg, 0.46 mmol) in xylene (7.6 mL) with 10 equiv of 3,4-dihydro-2*H*-pyrane (386 mg, 4.6 mmol) was heated under reflux for 1.5 h (conversion 98%). After removal of the solvent, preparative TLC [hexane–ethyl acetate (3:1)] of the residue afforded benzazepine **19** (41.2 mg, 42%⁹). An oil; IR (CHCl₃): 3340 (N–H), 1720 cm⁻¹ (C=O); ¹H NMR (AL4): δ 1.22 (t, 3H, *J*=7.2 Hz, CH₃), 3.30 (br s, 1H, NH), 3.66–3.73 (m with ddd character, 1H, *J*=17.2 Hz, H-2), 3.75– 3.82 (m with d character, 1H, *J*=17.2 Hz, H-2), 4.19 (q, 2H, J=7.2 Hz, OCH₂), 4.42–4.44 (m with d character, 1H, J= 7.2 Hz, H-5), 5.61–5.66 (m, 1H, H-3), 5.97–6.04 (m, 1H, H-4), 6.87 (dd, 1H, J=7.6, 1 Hz, H-9), 6.96 (td, 1H, J=7.6, 1.2 Hz, H-7), 7.04 (dd, 1H, J=7.6, 1.6 Hz, H-6), 7.16 (td, 2H, J=7.6, 1.6 Hz, H-8); ¹³C NMR (AL4): δ 14.3 (q, CH₃), 48.4 (t, C-2), 49.2 (d, C-5), 61.0 (t, OCH₂), 121.8 (d, C-9), 122.5 (d, C-7), 124.3 (d, C-4), 128.1 (d, C-8), 128.6 (d, C-6), 129.1 (d, C-3), 134.3 (s, C-5a), 147.9 (s, C-9a), 172.5 (s, 5-CO); EI-MS *m*/*z* 217 (M⁺, 28%), 188 (3), 172 (5), 144 (100), 127 (4), 115 (7), 72 (2); HRMS calcd for C₁₃H₁₅NO₂: 217.1103. Found: 217.1099.

4.8. Reaction of N-benzoylaziridine (E)-4

4.8.1. Ethyl (2E,4E)-5-benzamido-2,4-pentadienoate 20. By analogy with the photoreactions of 1, a solution of (E)-4 (124 mg, 0.51 mmol) in dry acetonitrile (8.5 mL) with 10 equiv of acrylonitrile (270 mg, 5.1 mmol) was irradiated for 5 h at rt (conversion 74%). After removal of the solvent, preparative TLC [hexane-ethyl acetate (3:1)] of the residue afforded dienoate **20** (27.9 mg, 30%⁹). Colorless plates; mp 124–127 °C (hexane/ethyl acetate); IR (CHCl₃): 1690 cm⁻ (C=O); ¹H NMR (AL4): δ 1.27 (t, 3H, J=7.2 Hz, CH₃), 4.19 (q, 2H, J=7.2 Hz, OCH₂), 5.74 (d, 1H, J=15.2 Hz, H-2), 6.09 (dd, 1H, J = 14.0, 11.6 Hz, H-4), 7.31 (dd, 1H, J=15.2, 11.6 Hz, H-3), 7.45 (t, 2H, J=7.2 Hz, H-3', H-5), 7.49–7.54 (m with d character, 1H, J = 14.0 Hz, H-2), 7.56 (t, 1H, J=7.2 Hz, H-4'), 7.86 (d, 2H, J=7.2 Hz, H-2', H-6'), 8.78 (br d, 1H, J = 11 Hz, NH); ¹³C NMR (AL4): δ 14.3 (q, CH₃), 60.2 (t, OCH₂), 111.3, 111.8 (2d, C-2, C-4), 127.3, 128.5 (2d, 4C in Ph), 132.5, 133.2 (2d, C-5, C in Ph), 132.8 (s, C in Ph), 143.4 (d, C-3), 164.7, 167.2 (2s, C-1, CONH); EI-MS *m/z* 245 (M⁺, 19%), 200 (4), 140 (7), 105 (100), 77 (26), 51 (4); HRMS calcd for $C_{14}H_{15}NO_3$: 245.1052. Found: 245.1046.

4.8.2. Thermal reactions of (*E*)-**4.** A solution of (*E*)-**4** (300 mg, 1.22 mmol) in xylene (20 mL) with 10 equiv of *tert*-butyl acrylate (1.54 g, 12 mmol) was heated under reflux for 3.5 h (conversion 83%). After removal of the solvent, flash column chromatography [hexane–ethyl acetate (6:1)] of the residue afforded pyridine 21^{12} (9.1 mg, $4\%^9$) and a mixture of isomers of dienoate **20** (134 mg).

4.9. Reaction of *N*-trityl aziridine (*E*)-5

4.9.1. Ethyl (2E,2'RS,3'RS)-3-(3-cyano-1-tritylpyrrolidin-2-yl)acrylate 22. By analogy with the photoreactions of (Z)-1, a 0.060 mol L⁻¹ solution of (E)-5 in dry acetonitrile with 10 equiv of acrylonitrile was irradiated. Preparative TLC [hexane-ethyl acetate (5:1)] afforded the adduct 22. The results are summarized in Table 2. An oil; IR (CHCl₃): 2240 (C≡N), 1710 cm⁻¹ (C=O); ¹H NMR (AL4): δ 1.27–1.34 (m, 1H, H-3') 1.36 (t, 3H, J=7.2 Hz, CH₃), 1.62–1.70, 1.75–1.87 (2m, 2H, H₂-4'), 3.07 (td, 1H, J=13, 8.4 Hz, H-5'), 3.48 (ddd, 1H, J=13, 10.0, 4.0 Hz, H-5'), 4.19 (t, 1H, J = 6.4 Hz, H-2') 4.28 (q, 2H, J = 7.2 Hz, OCH_2), 6.34 (d, 1H, J=15.6 Hz, H-2), 7.09 (dd, 1H, J= 15.6, 6.4 Hz, H-3), 7.18–7.57 (m, 15H, Tr); ¹³C NMR (AL4): δ 14.4 (q, CH₃), 29.1 (t, C-4'), 31.3 (d, C-3'), 48.3 (t, C-5'), 60.8 (t, OCH₂), 63.0 (d, C-2'), 78.0 (s, CPh₃), 118.9 (s, CN), 123.7 (d, C-2), 126.6 (d, 3C in Ph), 127.9 (d, 6C in Ph), 128.8 (d, 6C in Ph), 143.8 (s, 3C in Ph), 144.8 (d, C-3),

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165.8 (s, C-1); EI-MS m/z 436 (M⁺, 0.4%), 359 (1), 243 (100), 165 (2); HRMS calcd for C₂₉H₂₈N₂O₂: 436.2151. Found: 436.2150.

4.9.2. Ethyl (2E,2'RS,3'RS)-3-(3-methoxycarbonyl-1-tritylpyrrolidin-2-yl)acrylate 23. By analogy with the photoreactions of (Z)-1, a solution of (E)-5 in dry acetonitrile with 10 equiv of methyl acrylate was irradiated. Preparative TLC [hexane-ethyl acetate (7:1)] afforded the adduct 23. The results are summarized in Table 2. Colorless crystals; mp 134-135 °C (hexane/ethyl acetate); IR (CHCl₃): 1720 (C=O), 1710 cm⁻¹ (C=O); ¹H NMR (AL4): δ 1.33 (t, 3H, J=7.1 Hz, OCH₂CH₃), 1.34–1.40, 1.86–1.97 (2m, 2H, H_2 -4'), 1.59 (dt, 1H, J=10.5, 8.3 Hz, H-3'), 2.97-3.06, 3.41-3.49 (m, 2H, H₂-5'), 3.49 (s, 3H, OCH₃), 4.19–4.23 (m, 1H, overlapping with q at δ 4.23, H-2') 4.23 (q, 2H, J=7.1 Hz, OCH₂), 6.14 (d, 1H, J=15.6 Hz, H-2), 6.86 (dd, 1H, J = 15.6, 6.4 Hz, H-3), 7.12– 7.29 (m, 9H, Tr), 7.56 (d, 6H, J=7.6 Hz, Tr); ¹³C NMR (AL4): δ 14.3 (q, OCH₂CH₃), 26.6 (t, C-4'), 47.6 (d, C-3'), 48.3 (t, C-5'), 51.5 (q, OCH₃), 60.4 (t, OCH₂), 63.4 (d, C-2'), 78.0 (s, CPh₃), 122.1 (d, C-2), 126.3 (d, 3C in Ph), 127.7 (d, 6C in Ph), 129.0 (d, 6C in Ph), 144.2 (s, 3C in Ph), 146.6 (d, C-3), 166.2 (s, C-1), 171.6 (s, CO₂CH₃); EI-MS m/z 469 $(M^+, 1\%), 392 (3), 243 (100), 228 (4), 198 (2), 165 (23),$ 154 (2), 91 (2); HRMS calcd for C₃₀H₃₁NO₄: 469.2253. Found: 469.2252.

4.10. Application to the synthesis of (\pm) -isoretronecanol 27

4.10.1. Ethyl (2E,2'RS,3'RS)-3-(3-methoxycarbonylpyrrolidin-2-yl)acrylate 24. To a solution of **23** (430 mg, 0.86 mmol) in chloroform (0.7 mL) and methanol (0.7 mL) was trifluoroacetic acid (1.3 mL) at rt. After being stirred for 1 h at rt, the reaction mixture was extracted with water (2×3 mL). The aqueous phase was neutralized with aqueous saturated NaHCO₃ and extracted with chloroform (3×5 mL). The organic phase was washed with brine, dried with MgSO₄, and concentrated in vacuo to yield **24** (168 mg, 83%).

An oil; IR (CHCl₃): 3510 (N–H), 1710 cm⁻¹ (C=O); ¹H NMR (AL4): δ 1.28 (t, 3H, J=7.2 Hz, OCH₂CH₃), 1.77 (brs, 1H, NH), 2.01–2.20 (m, 2H, H₂-4'), 2.91–2.99 (m with ddd-character, 1H, J=11, 8.3, 7.8 Hz, H-5'), 3.11–3.17 (m, 1H, H-3'), 3.29 (ddd, 1H, J=11.2, 8.6, 4.4 Hz, H-5'), 3.64 (s, 3H, OCH₃), 3.87–3.92 (m, 1H, H-2') 4.19 (q, 2H, J= 7.2 Hz, OCH₂), 5.99 (dd, 1H, J=15.6, 1.5 Hz, H-2), 6.89 (dd, 1H, J=15.6, 6.3 Hz, H-3); ¹³C NMR (AL4): δ 14.3 (q, OCH₂CH₃), 29.3 (t, C-4'), 46.3 (t, C-5'), 48.5 (d, C-3'), 51.7 (q, OCH₃), 60.4 (t, OCH₂), 62.8 (d, C-2'), 122.3 (d, C-2), 144.5 (d, C-3), 165.7 (s, C-1), 173.4 (s, CO₂CH₃); EI-MS m/z 227 (M⁺, 21%), 198 (100), 181 (40), 154 (41), 128 (38), 116 (80), 100 (48), 56 (42); HRMS calcd for C₁₁H₁₇NO₄: 227.1158. Found: 227.1160.

4.10.2. Ethyl (2'RS,3'RS)-**3**-(**3**-methoxycarbonylpyrrolidin-**2**-yl)propanate **25**. A solution of **24** (18.1 mg, 0.08 mmol) in ethyl acetate (0.5 mL) with 10% Pd/C (5.2 mg) under hydrogen was stirred for 21 h at rt. The reaction mixture was filtered with celite, and the filtrate was concentrated in vacuo, giving a residue that was subjected to

NH-silica gel column chromatography [hexane–ethyl acetate (1:5)] to afford **25** (11.7 mg, 64%).

An oil; IR (CHCl₃): 3410 (N–H), 1720 cm⁻¹ (C=O); ¹H NMR (AL4): δ 1.25 (t, 3H, J=7.2 Hz, OCH₂CH₃), 1.62–1.85 (m, 3H, NH and H₂-3), 1.93–2.10 (m, 2H, H₂-4'), 2.44–2.50 (m, 2H, H₂-2), 2.79–2.87 (m, 1H, H-5'), 2.92–2.97 (m, 1H, H-3'), 3.04–3.11 (m, 1H, H-2'), 3.18–3.25 (m, 1H, H-5'), 3.68 (s, 3H, OCH₃), 4.13 (q, 2H, J=7.2 Hz, OCH₂); ¹³C NMR (AL4): δ 14.4 (q, OCH₂CH₃), 26.4 (t, C-3), 30.3 (t, C-4'), 32.5 (t, C-2), 46.4 (t, C-5'), 47.6 (d, C-3'), 51.5 (q, OCH₃), 60.4 (t, OCH₂), 63.0 (d, C-2'), 173.0, 174.9 (2s, C-1, CO₂CH₃); EI-MS *m*/*z* 229 (M⁺, 4%), 183 (34), 155 (29), 152 (24), 128 (100), 97 (76), 69 (26); HRMS calcd for C₁₁H₁₉NO₄: 229.1314. Found: 229.1314.

4.10.3. Methyl (4RS,5RS)-8-oxo-1-azabicyclo[3.3.0]oct-4-ylcarboxylate 26. A solution of 25 (14.6 mg, 0.065 mmol) in toluene (1.0 mL) was refluxed for 15.5 h. The reaction mixture was concentrated in vacuo, giving a residue that was subjected to flush column chromatography [ethyl acetate] to afford 26^5 (10.3 mg, 87%).

An oil; IR (CHCl₃): 1730 (C=O), 1695 cm⁻¹ (C=O); ¹H NMR (AL4): δ 1.69–1.78 (m, 1H, H-6), 2.16–2.43 (m, 4H, H₂-3, H-6, H-7), 2.61–2.70 (m, 1H, H-7), 3.02 (td, 1H, *J*= 7.1, 3.4 Hz, H-4), 3.04–3.11 (m, 1H, H-2), 3.70 (s, 3H, OCH₃), 3.82 (td, 1H, *J*=11.2, 7.6 Hz, H-2), 4.16 (q, 1H, *J*= 7 Hz, H-5); ¹³C NMR (AL4): δ 22.6 (t, C-6), 30.4 (t, C-3), 34.0 (t, C-7), 41.3 (t, C-2), 45.5 (d, C-4), 51.9 (q, OCH₃), 63.2 (d, C-5), 172.6, 175.2 (2s, C-8, CO₂CH₃); EI-MS *m*/*z* 183 (M⁺, 37%), 183 (34), 155 (45), 152 (26), 97 (100), 69 (44); HRMS calcd for C₉H₁₃NO₃: 183.0895. Found: 183.0892.

Compound **26**.^{5b} ¹H NMR (250 MHz): δ 1.5–1.75 (m, 1H), 2.0–2.4 (m, 4H), 2.45–2.7 (m, 1H), 2.85–3.1 (m, 2H), 3.6 (s, 3H), 3.5–3.8 (m, 1H), 4.08 (q, 1H, J=7 Hz); ¹³C NMR (62.5 MHz): δ 22.3, 30.2, 33.6, 41.0, 45.2, 51.8, 63.1, 172.9, 175.4.

Acknowledgements

This work was partially supported by the Grant-in-Aid for Scientific Research (C) (No. 13672239) administered by Japan Society for the Promotion of Science and by Bio Venture Center of Meiji Pharmaceutical University. The authors wish to thank the staff of the Analysis Center of Meiji Pharmaceutical University for performing the elemental analysis and measurements of LA-NMR (HMBC and NOESY spectra) (Miss S. Kubota) and mass spectra (Miss T. Koseki). We are also grateful to Mr. N. Goi for his technical assistance.

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Tetrahedron

Tetrahedron 60 (2004) 10899-10906

Density functional study of the lithiation of cyclic vinyl ethers in solution

Zhiqing Yan and John F. Sebastian*

Department of Chemistry and Biochemistry, Miami University, Oxford, OH 45056, USA

Received 13 May 2004; revised 10 September 2004; accepted 13 September 2004

Available online 2 October 2004

Abstract—Lithiation of three cyclic vinyl ethers—2,3-dihydrofuran, 3,4-dihydro-2*H*-pyran, and 2,3-dihydrooxepin in ethereal solution are investigated at the density functional theory level of B3LYP/6-31++G(d,p). Several solvation models were used, including the microsolvation model, the continuum models such as the Onsager model, the polarized continuum models, and the mixed discrete-continuum model. Both the microsolvation and the mixed discrete-continuum model gave results consistent with experiments. Theoretical calculations also indicate that lithiation of 2,3-dihydrooxepin undergoes allylic lithiation concomitant with ring opening.

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1. Introduction

The synthetic application of α -lithiated cyclic vinyl ethers in organic chemistry has been of interest since at least 1951. These lithiated compounds have been mainly used as acyl anion synthons and as precursors of α -alkoxyalkenylcuprates in Kocienski's metallate rearrangement.¹ Lithiation of mid-sized cyclic vinyl ethers at vinylic versus allylic positions has been an active project in our group. Computational investigations of the lithiation of cyclic vinyl ethers in the gas phase at Hartree-Fock (geometry optimization) and second-order Møller-Plesset perturbation theory (single point) levels have suggested that in most cases, the vinylic positions are energetically favored.² However, using exchange-correlation functionals of B3LYP in combination with the mid-sized basis set, 6-31 + + G(d,p), we have shown that allylic lithiation is favored in the case of oxete, the 4-membered ring system, which is in contrast with previous studies.³ The density functional theory (DFT) method-B3LYP has been widely used in most of the recent computational organolithium studies and has proved to be significantly more efficient than some of the post-HF methods (such as MP2 level) of comparable accuracy.⁴ It has also been successfully used in investigations of ether solvent effects involving lithium enolates, the results of which agree well with more expensive MP4 level calculations.⁵ The inclusion of electron correlation within the geometry optimization increases the accuracy of the optimized structures and therefore the relative energy preferences. Herein we wish to summarize our theoretical work of the ether solvated lithiation of 2,3-dihydrofuran (DHF)—the 5-membered ring system,⁶ 3,4-dihydro-2*H*-pyran (DHP)—the 6-membered ring system,⁷ and 2,3-dihydrooxepin (DHOP)—the 7-membered ring system with two conjugated double bonds (Fig. 1).



Figure 1. Cyclic vinyl ethers included in this study.

The solvation models used here are microsolvation, in which short-range solvent interaction is examined, and the continuum reaction field models, including the Onsager model, the polarized continuum model (PCM, including the newer CPCM–PCM model using the polarizable conductor calculation model and IEFPCM–PCM model using the integral equation formalism model), and the isodensity polarized continuum model (IPCM), where the long-range solvation is considered. We then combined both short-range and long-range solvation into the mixed discrete-continuum solvation model (mixed solvation model). In the current study, both the microsolvation model and the mixed discrete-continuum model seem better in reproducing the experimental results. It is well-known that vinylic lithiation is preferred experimentally for DHF and DHP,⁸ and it is

Keywords: Density functional theory; Microsolvation; Continuum solvation; Mixed discrete-continuum solvation; Transition states.

^{*} Corresponding author. Tel.: +1 513 529 3337; fax: +1 513 529 5715; e-mail: sebastjf@muohio.edu

^{0040–4020/\$ -} see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.035



Scheme 1.

generally believed that both the electronegative vinyl ether oxygen and precomplexation affect lithiation regioselectivity. Our DFT-B3LYP calculations incorporated with different solvation models reproduce the vinylic lithiation for DHF in all cases. For DHP, all of the continuum models give allylic lithiation in contrast with experiment, but the microsolvation model predicts vinylic lithiation with a small relative energy preference of only 0.17 kcal/mol. However, when combining the discrete microsolvation and the continuum solvation we obtained more reasonable results, indicating long-range solvation also affects the outcome of the lithiation. We have also shown previously in experiments that lithiation of DHOP with 6-lithiated-3,4-dihydro-2H-pyran in THF leads to ring-opening to give a trienolate product,⁹ but the detailed mechanism of this reaction was unclear. Our computational work described herein with all

solvation models, except for the continuum IPCM model, suggests allylic lithiation with a direct ring-opening mechanism for DHOP.

2. Methods of calculations

The DFT method we used here, B3LYP, includes a combination of Becke's three-parameter nonlocal exchange functional¹⁰ with the correlation functional of Lee–Yang–Parr.¹¹ The solvent used in the SCRF calculations is diethylether which is available in the Gaussian 98w program package.¹² In the previous DFT studies of the oxete system, the use of water as the 'microsolvent' resulted in unnecessary hydrogen bonding.³ Limited by computational efficiency and cost, one molecule of dimethyl ether seems to

Table 1. Relative energies (kcal/mol) obtained for DHF, DHP and DHOP in the gas phase and in solution with different solvation models ($\Delta E = E_{\text{vinylic}} - E_{\text{allylic}}$)

Molecule	Models	Reactant ^a	Vinylic transition state	Allylic transition state	Vinylic product	Allylic product	ΔE (TS)	ΔE (products)
DHF	Gas phase	0.00	20.22	22.32	-3.28	1.86	-2.10	-5.14
	Onsager model ^b	0.00	25.12	28.14	0.56	7.16	-3.01	-6.60
	PCM model ^b	0.00	25.62	29.04	-2.38	3.22	-3.42	-5.60
	CPCM model	0.00	25.79	29.30	-2.03	3.39	-3.51	-5.43
	IEFPCM model	0.00	26.01	29.36	-1.91	3.76	-3.35	-5.68
	IPCM model ^b	0.00	15.66	26.24	-1.33	-0.22	-10.58	-1.11
	Microsolvation ^c	0.00	15.28	19.06	-7.41	-4.11	-3.78	-3.30
	PCM (mixed)	0.00	19.81	24.84	-7.61	-1.72	-5.02	-5.89
	CPCM (mixed)	0.00	19.88	24.72	-7.61	-1.65	-4.84	-5.95
	IEFPCM (mixed)	0.00	19.86	24.67	-7.38	-1.60	-4.81	-5.78
	IPCM (mixed)	0.00	19.49	26.01	-8.16	-0.51	-6.52	-7.64
DHP	Gas phase	0.00	22.94	20.39	0.02	-1.64	2.54	1.66
	Onsager model ^b	0.00	28.52	26.90	4.97	4.78	1.62	0.19
	PCM model ^b	0.00	27.97	27.49	1.17	0.60	0.47	0.57
	CPCM model	0.00	28.12	27.78	1.42	0.84	0.34	0.58
	IEFPCM model	0.00	28.43	27.75	1.58	0.96	0.68	0.62
	IPCM model ^b	0.00	27.35	26.06	-2.53	-4.19	1.29	1.66
	Microsolvation ^c	0.00	17.82	17.99	-4.14	-7.71	-0.17	3.57
	PCM (mixed)	0.00	22.44	24.18	-3.86	-4.28	-1.74	0.42
	CPCM (mixed)	0.00	22.64	24.32	-3.68	-4.04	-1.68	0.36
	IEFPCM (mixed)	0.00	22.68	24.21	-3.44	-4.05	-1.53	0.60
	IPCM (mixed)	0.00	23.40	24.82	-2.26	-3.12	-1.42	0.86
DHOP ^d	Gas phase	0.00	21.92	11.59	-1.33	-49.19	10.33	47.86
	Onsager model ^b	0.00	26.06	16.61	1.25	-44.65	9.45	45.90
	PCM model ^b	0.00	9.88	1.89	-17.24	-60.85	7.98	43.61
	CPCM model	0.00	10.28	2.30	-16.83	-60.53	7.98	43.69
	IEFPCM model	0.00	10.38	2.13	-16.79	-60.65	8.25	43.85
	IPCM model ^b	0.00	0.43	1.37	-17.63	-60.53	-0.94	42.89
	Microsolvation ^c	0.00	16.51	8.64	-5.33	-52.17	7.88	46.83
	PCM (mixed)	0.00	21.22	14.78	-4.62	-48.18	6.44	43.57
	CPCM (mixed)	0.00	21.66	15.41	-4.32	-48.24	6.24	43.92
	IEFPCM (mixed)	0.00	21.38	14.90	-4.60	-48.36	6.48	43.76
	IPCM (mixed)	0.00	22.26	15.43	-3.54	-47.46	6.82	43.92

^a Reactants in each model are set to be 0 in energy, they are not necessarily the same in different models.

^b Calculations with diethyl ether as the bulk solvent.

^d In the case of DHOP, allylic transition state and product are referred to the acyclic process.

^c Calculations with one dimethyl ether molecule explicitly expressed in the system.

be the simplest but reasonable analog solvent for the microsolvation model (Scheme 1) even though lithium may coordinate to two or more solvent molecules. Our results indicate that such a model can represent the trend of microsolvation efficiently. In the mixed discrete-continuum solvation, the continuum models were used for single point calculations on the microsolvation optimized geometries.

All the calculations were performed using either the GAUSSIAN 94 program package or the GAUSSIAN 98w (Windows version) program.¹³ The standard 6-31 + +G(d,p) basis set was used (available in both programs). All stationary points were characterized as minima (with no imaginary frequency) or saddle points (with one and only one imaginary frequency) by harmonic vibrational frequency analysis at the same level. All of the transition states of the microsolvation model were located and optimized by both the synchronous transit-guided quasi-Newton method with three molecule specifications $(QST3)^{14}$ and the Berny algorithm (TS),¹⁵ and verified to be connecting the corresponding minima (reactant and product) by a proceeding IRC calculation (six points on each side of the transition structure along the reaction path with a step size of 0.1 au) using the same basis set and theory level. All relative energies reported are of internal energies and zero-point energy (ZPE) corrections are included. All the structural parameters were obtained with CS Chem 3D PRO version 6.0.

3. Results and discussion

As expected, in all three microsolvation modeled lithiation reactions, the three components of the reactant—methyl lithium, dimethyl ether and the cyclic vinyl ether tend to form a 'pre-complex'. The pre-complex is energetically favored because the lone pairs of electrons of the two oxygen atoms (one in the cyclic vinyl ether, the other in the solvent) interact with the lithium cation.

For the lithiation reaction of DHF, the 5-membered ring system, with all the models that we used in the gas phase and ethereal solutions, vinylic lithiation is favored over the corresponding allylic process (Table 1, Figs. 2 and 3). The inclusion of solvation (long-range, short-range and combined) increases ΔE (TS).

For the lithiation reaction of DHP, the 6-membered ring system, previous gas phase geometry optimization at the Hartree–Fock level reproduced the experimental result that vinylic lithiation is favored over the corresponding allylic process by a calculated ΔE (TS) of -2.7 kcal/mol. However, with the inclusion of correlation energy, the MP2 level single point calculation predicts the allylic process.^{2,16} All continuum models and even the gas phase model at the current DFT level also predict the allylic lithiation. With the microsolvation model, the current DFT calculation gives a small ΔE (TS) of -0.17 kcal/mol, favoring the vinylic



Figure 2. Progress of reaction diagram for deprotonation of DHF resulting from microsolvation geometry optimizations (B3LYP/6-31 + +G(d,p), relative energies are in kcal/mol).



Figure 3. DFT optimized geometries of dimethyl ether microsolvated DHF system.



Progress of Reaction —

-

Figure 4. Progress of reaction diagram for deprotonation of DHP resulting from microsolvation geometry optimizations (B3LYP/6-31 + G(d,p), relative energies are in kcal/mol).





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Figure 5. DFT optimized geometries of dimethyl ether microsolvated DHP system.

H²⁷

H28

C1

7



Figure 6. Progress of reaction diagram for deprotonation of DHOP resulting from microsolvation geometry optimizations (B3LYP/6-31 + +G(d,p), relative energies are in kcal/mol).



Figure 7. DFT optimized geometries of dimethyl ether microsolvated DHOP system.

lithiation (Table 1, Figs. 4 and 5); a more reasonable ΔE (TS) of around -1.5 kcal/mol was obtained by using the mixed solvation model. Since basis set deficiency has been very likely excluded in previous studies,¹⁶ this could suggest that the short-range solvation must be included to successfully reproduce the experimental results.

For the lithiation reaction of DHOP, the 7-membered ring system, things are more complicated. First of all, at the Hartree-Fock level with the same basis set, the allylic lithiated product in the gas phase (optimized as energy minimum) was calculated as a cyclic structure. However, when we tried to calculate the same system at MP2 and B3LYP level even with a basis set as small as 3-21G, both failed to give the cyclic form but yielded an open-chain trienolate product instead, indicating ring-opening during the gas phase lithiation process. We then 'froze' the C9–O2 bond to hold the cyclic form of the allylic lithiation product, but the calculations would not converge in a reasonable period of time as long as electronic correlation is included. The same thing happened in the microsolvation model at the current level (17 in Figs. 6 and 7). We noticed that after reasonable rotation about the carbon-carbon single bonds our calculated allylic product lead to the actual trienolate product of previous experiments.⁹ The microsolvation model indicates that the formation of the incipient trienolate structure is favored over the cyclic vinylic process by 7.88 kcal/mol in ΔE (TS). All of the reaction field solvation models at the current DFT level gave the same energetic trend of preference, except for the IPCM model (Table 1).

Scheme 2 shows three possible pathways of this reaction. Our calculation results suggest that the lithiation reaction of DHOP follows an E2 type mechanism (Scheme 2), since allylic lithiation is energetically favored and the cyclic form of the allylic product could not be formed as a minimum.

The conformations of all the allylic species are more puckered than the vinylic ones, which has also been observed in the previous studies of oxete lithiation.⁵ The lithium cation is sitting atop the ring in the allylic processes, which suggests electronic delocalization. In the acyclic form of the allylic product (trienolate) of DHOP lithiation (17), the lithium cation only interacts with the terminal double bond. The isolated anions (gas phase) for each system are also calculated at the same theory level. Without the lithium cation, the isolated vinyl anion of DHF is 3.1 kcal/mol lower in energy than the isolated allyl anion whereas the vinyl anion of DHP is, in fact, 8.67 kcal/mol higher in energy than the allyl anion. In the case of DHOP anions, the cyclic and acyclic allyl anions are 23.36 and 44.14 kcal/mol, respectively, lower in energy than the vinyl one.

All three cyclic vinyl ethers have two hydrogen atoms attached to the allylic carbon atom. Our calculations with



Scheme 2.

the microsolvation model indicated that they are equivalent at the current DFT level of theory: the energy differences between the two allylic transition structures are 6×10^{-4} kcal/ mol for DHF, 6×10^{-4} kcal/mol for DHP and 9×10^{-6} kcal/ mol for DHOP, and the energy differences between the two allylic products are 0.01 kcal/mol for DHF, 0.18 kcal/mol for DHP and 0.08 kcal/mol for DHOP, all of which are negligible in this study.

Comparing the corresponding vinylic angles in the reactants and vinylic lithiation products of DHF (\angle O2–C12–C10 in **2** and **6**), DHP (\angle O8–C9–C10 in **8** and **12**) and DHOP (\angle O3–C4–C5 in **14** and **18**), the microsolvation calculations suggest vinyl compressions of 7.5, 8.09 and 8.08°, respectively. The allylic expansions (comparing \angle C4– C10–C12 in **2** and **5**, \angle C9–C10–C11 in **8** and **11**, \angle C6– C7–C8 in **14** and **17**) are 0.3° for DHF and 5.84° for DHOP but – 1.6° for DHP. The atomic charges were determined by Natural Population Analysis (NPA), based on the geometry optimized in the same solvation model.

The charges of the migrating hydrogen atoms in all the transition structures are low (+0.20-0.25 electrons) compared to the corresponding hydrogen atoms in the reactants and products, suggesting that the transition structures are multi-center processes, which agrees with previous HF level optimizations.²

4. Conclusions

In summary, we have successfully reproduced the experimental results of the lithiation reactions of DHF and DHP in ether solution using the DFT method (B3LYP/6-31 + + (d,p)). The inclusion of short-range solvation (microsolvation) is necessary for predicting the experimentally consistent regioselectivity of lithiation. The continuum solvation, on the other hand, also affects the lithiation reaction to some extent, especially in the case of DHP. The newer CPCM and IEFPCM models give very similar results to the PCM model in all three systems. In our case, the IPCM does not seem to be a good solvation model even though an isodensity surface is a more natural and intuitive

shape for the solute cavity. We also examined the mechanism of the lithiation of DHOP, and the theoretical results suggest an allylic ring-opening lithiation via a direct E2 type pathway. The experimental confirmation of this mechanism will be published elsewhere.

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Tetrahedron

Tetrahedron 60 (2004) 10907-10914

3-Methyl-4*H***-[1,2,4]-oxadiazol-5-one:** a versatile synthon for protecting monosubstituted acetamidines

Alan E. Moormann,^{*} Jane L. Wang, Katherine E. Palmquist, Michele A. Promo, Jeffery S. Snyder, Jeffrey A. Scholten, Mark A. Massa, James A. Sikorski[†] and R. Keith Webber

Pfizer Inc., 700 Chesterfield Parkway West, Chesterfield MO 63017, USA

Received 28 May 2004; revised 9 September 2004; accepted 10 September 2004

Available online 29 September 2004

Abstract—The utilization of 3-methyl-4H-[1,2,4]-oxadiazol-5-one as a versatile protected acetamidine is demonstrated through employment in a variety of synthetic sequences. The potassium salt (**2a**) or the neutral form (**2b**) is alternatively shown to be superior for various synthetic reactions (i.e., alkylation, Michael addition, Mitsunobu) to incorporate side chains for further synthesis. The 3-methyl-4H-[1,2,4]-oxadiazol-5-one moiety was found to be stable to acid or base under non-aqueous conditions. It was also found to be stable to many reagents commonly used for organic synthesis. Despite this stability, the free acetamidine may be released by mild reduction including Lindlar hydrogenation or dissolving metal reductions. Alternatively, the hydroxyl amidine may be formed via alkaline hydrolysis. © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

Amidine groups are found in many medicinal drugs, for example, cardiovascular, anti-diabetic, antibacterial, antiprotozoal, anthelmintic, anti-inflammatory, central nervous system, antineoplastic and others.^{1,2} N-monosubstituted acetamidines have been useful as arginine mimetics and they have been utilized in nitric oxide synthase (NOS) inhibitors in particular.^{3,4} The basic nature of the amidine and its facile hydrolysis to the corresponding amide makes the synthesis and purification of compounds containing them troublesome. For our needs we sought a versatile method to prepare highly functionalized N-monosubstituted acetamidines. This investigation led to the introduction of the acetamidine in a protected form that enabled facile synthetic manipulations.

A survey of the literature revealed that oxadiazol-5-ones have been previously described as precursors to amidines.^{5a-c,6a-d} These prior investigations did not explore the utility of this conversion as a component of a synthetic sequence. The work described here explores the synthetic utility of the use of 3-methyl-4H-[1,2,4]-oxadiazol-5-one as a protected acetamidine. We have found this heterocycle to be stable to many synthetic conditions and have used it in extended synthetic schemes. The mild reaction conditions for the selective generation of acetamidines and hydroxy-acetamidines is described as well.

2. Results and discussion

The utility of 3-methyl-4*H*-[1,2,4]-oxadiazol-5-one is dependent upon a consistent route for preparation. We utilized a procedure described by Hett et al. (Scheme 1), which is suitable for the kilogram scale synthesis of the potassium salt 2a.⁷ A simple bubbling of HCl through an ether suspension of 2a affords the protonated form of the heterocycle 2b.

Scheme 1.

Keywords: Protected amidine; 3-Methyl-4*H*-[1,2,4]-oxadiazol-5-one.

^{*} Corresponding author. Tel.: +1 636 247 7553; fax: +1 636 247 5400; e-mail: alan.e.moormann@pfizer.com

⁺ Current address: AtheroGenics, Inc., 8995 Westside Parkway, Alpharetta, GA 30004, USA. Tel: +1 678 336 2733; fax: +1 678 336 2501.



Scheme 2.

Schemes 2-4 examine the reactivity of the potassium salt **2a** with activated halides. These schemes also show the feasibility of unmasking the acetamidine in the presence of other sensitive groups and the stability of the oxadiazolone ring to further synthetic manipulations.

The potassium salt 2a readily reacts with benzyl bromides to form the benzyl adducts (Scheme 2). The heterocycles 4aand 4b were subsequently hydrolyzed in aqueous hydroxide to afford the *N*-hydroxy amidines 5a and 5b. Heating 4awith Lindlar's catalyst in MeOH at reflux or with zinc in HOAc at 60 °C reduced the N–O bond to afford the acetamidine.

Mono-alkylation of the allylic dibromide 7 was accomplished by limiting the amount of **2a** (Scheme 3). This produced a mixture favoring the mono-alkylated derivative **8**, along with a small amount of the di-alkylated derivative that was separable by chromatography. The oxadiazolinone **8** was further elaborated to the phthalimide protected amino derivative **9** using potassium phthalimide. The phthalimide was removed with hydrazine to yield the free amine while

retaining the protected amidine **10**. Alternatively, sonication of **9** in the presence of zinc and HOAc released the acetamidine **11** without altering the phthalimide. In both deprotections the double bond was retained.

A facile reaction of the α -chloroketoester 12 with 2a afforded the α -ketoester 13 (Scheme 4). Deprotection of 13 via hydrogenation with Lindlar's catalyst in MeOH yielded the acetamidine 14 and a small amount of the imidazole 15. The mixture was separated and it was determined that 14 slowly cyclizes to form the imidazole 15.

The protonated form of the heterocycle, **2b**, was found to undergo the Mitsunobu reaction with dihydroxy-olefinic alcohol **16** in a preferential reaction of the allylic alcohol to the alkyl alcohol (Scheme 5). The reaction afforded a mixture of N and O-alkylated products **17** and **18**. These isomers were separable by chromatography and the structures confirmed by NMR. Only the protonated form **2b** was successful in this reaction. Attempts to use the potassium salt **2a** directly or via in situ generation of **2b**, failed to afford the Mitsunobu product. Although the alkyl



Scheme 3.





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alcohols were found to be much less reactive than allylic alcohols, a mixture of O and N- alkylated products of the alkyl alcohol was observed when a large excess of Mitsunobu reagents were utilized along with extended reaction time.

After exploring the reactivity of 2a and 2b, several fluoroolefins were synthesized to demonstrate the flexibility and chemical stability of the oxadiaxolinone when incorporated into a synthetic sequence. Schemes 6–8 exemplify the use of a Michael addition, alkylation and Mitsunobu reaction, respectively.

100% crude



Scheme 6.



100% crude



27

Scheme 7.



Scheme 8.

The Michael addition was successful with **2b** to afford the saturated aldehyde **19** (Scheme 6). As with the Mitsunobu reaction only the protonated form successfully produced product. The aldehyde **19** was further elaborated using Wittig conditions to the fluoro-olefinic ester **21** in high yield with no alteration of the heterocycle. The oxadiazolinone was further shown to be stable during reduction of the ester group in **21** with Red-Al to afford the alcohol **22**. The alcohol **22** was then converted to the bromide **23**, using PPh₃/CBr₄ in CH₂Cl₂, to set up for further potential elaboration.

A selective alkylation of an allyl mesylate in the presence the alkyl mesylate utilizing the potassium salt **2a** afforded **29** (Scheme 7). Further elaboration to the iodo-olefin **30** afforded an analog of **23** where the amidine precursor is transposed in relation to the halide.

The one carbon shorter homolog of **29** was prepared using a Mitsunobu reaction to yield **36** (Scheme 8). Further elaboration as for **30** afforded the corresponding homolog **37**.

3. Conclusions

3-Methyl-4*H*-[1,2,4]-oxadiazol-5-one **2** was demonstrated to be selective and versatile for the incorporation of the acetamidine in a protected form. The oxadiazolone ring was stable and robust, through multiple synthetic manipulations. Furthermore, the incorporated oxadiazolone produced products, which could be purified by silica chromatography and often by recrystallization. Catalytic reduction using Lindler's catalyst or zinc/HOAc selectively reduced the N–O bond, releasing the acetamidine in the presence of olefins and other reducible groups. The released acetamidine was often isolated pure. When purification was required, simple reverse-phase chromatography afforded clean product.

Only the protonated form of the oxadiazolone 2b participated in Michael additions and Mitsunobu reactions while the salt 2a participated in alkylation reactions. Both the Mitsunobu and alkylation reactions were selective for the allylic versus alkyl sites. Weak nucleophilic hydrides reduced a distal carbonyl to an alcohol without altering the oxadiazolinone. The oxadiazolinone ring was found to be stable to mild acidic conditions, but was hydrolyzed to afford the *N*-hydroxyamidine in 1 N NaOH. Hydrazine reacted with phthalimide **8** to release the amine **34** in the presence of the oxadiazolone ring.

4. Experimental

4.1. General

¹H NMR spectra were recorded on either a Varian Unity Plus 300 (300 MHz) or a Varian Unity Inova 400 (400 MHz) spectrometer. All NMR spectra are 400 MHz unless stated otherwise. All proton chemical shifts are recorded in ppm (δ) relative to trimethylsilane. All fluorine chemical shifts are recorded in ppm (δ) relative to a reference of C₆H₅CF₃ in benzene (Varian standard sample), which has a ¹⁹F chemical shift of -64 ppm, with an error of <1 ppm. Column chromatography was performed using either Biotage Flash 40 or 12 system. Reverse phase chromatography was performed on a Gilson semi-preparative HPLC with a YMC Combiprep ODS-A semi-prep column eluting with acetonitrile/water (0.01% TFA) at 20 mL/min. Mass spectra were obtained on a HP series 1100MSD, and high-resolution mass spectra were obtained with a PerSeptive Biosystems Mariner TOF. All solvents and reagents were purchased from Sigma-Aldrich.

4.1.1. Preparation of 3-methyl-1,2,4-oxadiaxolin-5-one (**2b**). Oxadiazolone **2a**⁷ (30.0 g, 0.217 mol) was suspended in Et₂O (150 mL) and cooled in an ice bath. HCl gas was bubbled through the slurry until the exotherm ceased. The mixture was stirred for 2 h, the solid filtered and the filtrate concentrated. The residue was dried under high vacuum to yield 15.5 g (71.4%) of **2b**. ¹H NMR (DMSO-d₆): δ 1.82 (s, 3H); ¹³C NMR (CDCl₃): δ 11.8, 162.5, 168.2; MS(CI) calcd for C₃H₄N₂O₂: [M+H] 101. Found: [M+H] 101; Anal. calcd for C₃H₄N₂O₂: C, 36.01; H, 4.03; N, 27.99. Found: C, 35.76; H, 3.99; N, 28.26.

4.1.2. 4-Benzyl-3-methyl-1,2,4-oxadiazol-5(4H)-one (4a). Benzyl bromide (1.13 g, 6.59 mmol) in acetone (15 mL) was added at ambient temperature to a suspension of 2a (1 g, 7.25 mmol), and tetrabutylammonium bromide (0.21 g, 0.66 mmol) in acetone (20 mL). The mixture was stirred 18 h, then quenched with brine and EtOAc. The organic layer was separated and the aqueous layer was extracted three times with EtOAc. The combined organic layers were washed with brine and dried over Na₂SO₄. The filtrate was concentrated and the product was dried under vacuum to yield 1.12 g (89%) 4a as a white solid, mp 64-65 °C. ¹H NMR (CDCl₃): δ 2.12 (s, 3H), 4.74 (s, 2H), 7.26 (m, 2H), 7.36 (m, 3H); 13 C NMR (CDCl₃): δ 10.9, 46.0, 127.6, 129.0, 129.5, 134.2, 156.8, 159.9; HRMS calcd for C₁₀H₁₀N₂O₂: [M+H] 191.0821. Found: [M+H]191.0822; Anal. calcd for C₁₀H₁₀N₂O₂: C, 63.15; H, 5.39; N, 14.73. Found: C, 63.22; H, 5.36; N, 14.79.

4.1.3. 4-(4-Bromobenzyl)-3-methyl-1,2,4-oxadiazol-5(4*H***)-one (4b). 4-Bromobenzyl bromide (1.65 g, 6.59 mmol) was allowed to react as in 4a** to yielded 1.60 g (90%) of **4b** as a white solid, mp 108–109 °C. ¹H NMR (CDCl₃): δ 2.14 (s, 3H), 4.69 (s, 2H), 7.13 (d, 2H), 7.50 (d, 2H); ¹³C NMR (CDCl₃): δ 10.9, 45.4, 123.5, 129.3, 132.7, 132.8, 149.5, 160.1; HRMS calcd for C₁₀H₉N₂O₂Br: [M+H] 268.9926. Found: [M+H] 268.9923; Anal. calcd for C₁₀H₉N₂O₂Br: C, 44.63; H, 3.37; N, 10.41. Found: C, 44.76; H, 3.40; N, 10.37.

4.1.4. 3-Methyl-4-(4-nitrobenzyl)-1,2,4-oxadiazol-5(4*H***)one (4c). 4-Nitrobenzyl bromide (1.42 g, 6.59 mmol) was allowed to react as in 4a** to yielded 1.35 g (87%) of **4c** as a white solid, mp 150–152 °C. ¹H NMR (CDCl₃): δ 2.17 (s, 3H), 4.85 (s, 2H), 7.45 (d, 2H), 8.25 (d, 2H); ¹³C NMR (CDCl₃): δ 10.9, 45.2, 124.8, 128.5, 141.7, 156.2; HRMS calcd for C₁₀H₉N₃O₄: [M+H] 236.1963. Found: [M+H] 236.1959; Anal. calcd for C₁₀H₉N₃O₄: C, 51.07; H, 3.86; N, 17.87. Found: C, 51.19; H, 3.88; N, 17.83.

4.1.5. (1Z)-N-Benzyl-N'-Hydroxyethanimidamide (5a). **4a** (190 mg, 1.0 mmol) was suspended in 5% NaOH

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(2 mL) at 60 °C for 3 h. The mixture was neutralized with 1 N HCl to pH 7–8. The solid was collected, washed with cold water several times and air dried to yield 100 mg (61%) of **5a** as a white solid, mp 138–139 °C. ¹H NMR (CD₃OD): δ 1.76 (s, 3H), 4.35 (s, 2H), 7.26 (m, 5H); ¹³C NMR (CD₃OD): δ 13.5, 45.5, 114, 126.5, 127.2, 128.4; HRMS calcd For C₉H₁₂N₂O: [M+H] 165.1028. Found: [M+H] 165.0987; Anal. calcd for C₉H₁₂N₂O: C, 65.83; H, 7.37; N, 17.05. Found: C, 65.83; H, 7.39; N, 17.06.

4.1.6. (1*Z*)-*N*-(**4-Bromobenzyl**)-*N*'-hydroxyethanimidamide (5b). **4b** (270 mg, 1.0 mmol) was allowed to react as in **5a** to give 200 mg (83%) of **5b** as a white solid, mp 129– 130 °C. ¹H NMR (CD₃OD): δ 1.74 (s, 3H), 4.31 (s, 2H), 7.22 (d, 2H), 7.47 (d, 2H); ¹³C NMR (CD₃OD): δ 13.5, 44.5, 120.1, 120.5, 128.4, 131.5; Anal. calcd for C₉H₁₁BrN₂O: C, 44.47; H, 4.56; N, 11.52. Found: C, 44.69; H, 5.59; N, 11.66.

4.1.7. Preparation of (iminoethyl)benzylamine (6). Method A: Oxadiazolone **4a** (380 mg, 2.0 mmol) was combined with MeOH (2 mL), HOAc (2 mL), water (2 mL) and zinc dust (570 mg, 8.7 mmol). The mixture was heated at 60 °C for 4 h. LCMS indicated product formation with no starting material. The reaction mixture was filtered and the filtrate concentrated. The crude product was purified by Reverse phase HPLC (0–100% acetonitrile with 0.01% HOAc) isolated as the HOAc salt to yield 300 mg (72%) of **6** as a clear oil. ¹H NMR (CD₃OD): δ 1.86 (s, 3H), 4.46 (s, 2H), 7.37 (m, 5H); ¹³C NMR (CDCl₃): δ 17.7, 46.0, 127.6, 127.9, 128.2, 128.8, 134.7, 164.0; HRMS calcd for C₉H₁₂N₂: [M+H] 149.1079. Found: [M+H] 149.1041.

Method B: Oxadiazolone **4a** (190 mg, 1.0 mmol) and Lindlar's catalyst Pd–CaCO₃ (290 mg) were combined in MeOH (10 mL). The mixture was heated at 60 °C for 2 h. LCMS indicated a new product with **4a** consumed. The reaction mixture was filtered and washed with MeOH. The filtrate was concentrated to dryness to yield **6** quantitively.

4.1.8. 4-[(2E)-4-Bromobut-2-enyl]-3-methyl-1,2,4-oxadiazol-5(4H)-one (8). Trans-1,4-dibromo-2-butene (7) (50 g, 0.23 mol) was dissolved in acetone (500 mL). Oxadiazolone 2a (16 g, 0.12 mol) was added, followed by tetrabutylammonium bromide (3.9 g, 0.012 mol). The reaction mixture was stirred at ambient temperature for 18 h, diluted with brine and the product was extracted into EtOAc. The organic extract was washed with brine, dried over MgSO₄, filtered and concentrated to a yellow semisolid residue. The product 8 was extracted from the residue into CH₂Cl₂ and concentrated to an oily residue. This residue was triturated with hot hexane to remove unreacted 7, then chromatographed on silica, eluting with EtOAc/ hexane to yield 14.2 g (50%) of 8 as a yellow oil. ¹H NMR (300 MHz, CDCl₃): δ 2.2 (s, 3H), 3.9 (d, 2H), 4.2 (d, 2H), 5.7 (d of t, 1H), 5.9 (d of t, 1H).

4.1.9. 2-[(*2E*)-**4-**(**3-**Methyl-**5-**oxo-**1**,**2**,**4-**oxadiazol-**4**(5*H*)-**y**])**but-2-enyl]-1***H***-isoindole-1**,**3**(2*H*)-dione (9). Bromide **8** (2 g, 8.58 mmol), tetrabutylammonium bromide (0.26 g, 0.86 mmol) and potassium phthalimide (1.9 g, 10.3 mmol) were dissolved in acetone (20 mL) and stirred for 2 h at ambient temperature. The solid was filtered and washed

with H_2O (50 mL), and the filtrate was extracted with EtOAc. The organic layer was dried over MgSO₄, and concentrated to yield 1.32 g (52%) of **9** as a yellow solid. ¹H NMR (300 MHz, CDCl₃): δ 2.21 (s, 3H), 4.06 (d, 2H), 4.32 (d, 2H), 5.59 (m, 2H), 7.71 (m, 2H) 7.89 (m, 2H); LC–MS calc for C₁₅H₁₃N₃O₄: [M+H] 300. Found: [M+H] 300; Anal calcd for C₁₅H₁₃N₃O₄: C, 60.20; H, 4.38; N, 14.04. Found: C, 59.98; H, 4.34; N, 14.10.

4.1.10. *N*-Benzylethanimidamide (10). Phthalimide 9 (150 mg, 0.5 mmol) was dissolved/suspended in EtOH (3.0 mL). Hydrazine hydrate (50 mg, 1.0 mmol) was added and the mixture was stirred at reflux for 30 min. The reaction mixture was concentrated and the residue dissolved in dilute K₂CO₃. The insoluble material was filtered and the filtrate was chromatographed using reverse phase to yield 10 mg of **10** (7.5%) isolated as the TFA salt. ¹H NMR (MeOD): δ 2.25 (s, 3H), 3.55 (dd, *J*=6.4, 1.2 Hz, 2H), 4.29, 4.28 and 4.27, 4.270 (d of d, *J*=5.2, 1.2 Hz, 2H), 5.78–5.71 (m, 1H), 5.97–5.90 (m, 1H); ¹⁹F NMR (MeOD): δ –77.37; HRMS calcd for C₇H₁₁N₃O₂: [M+H] 170.0924. Found: [M+H] 170.0916.

4.1.11. *N*-[(2*E*)-4-(1,3-Dioxo-1,3-dihydro-2*H*-isoindol-2yl)but-2-enyl]ethanimidamide (11). Phthalimide **9** (150 mg, 0.5 mmol) and zinc (101 mg; 1.5 mmol) were dissolved/suspended in HOAc (10 mL) and sonicated for 10 min. The resulting mixture was filtered and concentrated. The residue was chromatographed using reverse phase chromatography to yield 96 mg (18%) of **11** isolated as the TFA salt. ¹H NMR (MeOD): δ 2.23 (s, 3H), 4.04 (dd, *J* = 11.2, 6.4 Hz, 1H), 4.23 (dd, *J*=5.2, 1.2 Hz, 2H), 4.31 (d of d, *J*=16.0, 4.8 Hz, 1H), 5.83–5.69 (m, 2H), 7.73–7.51 (m, 4H); ¹⁹F NMR (MeOD): δ –77.38; HRMS calcd for C₁₄H₁₅N₃O₂: [M+H] 258.1164. Found: [M+H] 258.1152.

4.1.12. Ethyl **4-(3-methyl-5-oxo-1,2,4-oxadiazol-4(5***H***)yl)-3-oxobutanoate (13). Chloroester 12** (5.0 g, 0.03 mol) was added drop wise to a mixture of Cs₂CO₃ (13 g, 0.04 mol), NaI (50 mg) and **2a** (6 g, 0.04 mol) dissolved/ suspended in DMSO (40 mL), and stirred for 3 h at ambient temperature. The reaction was quenched with NH₄Cl solution and the product extracted into EtOAc. The residue was chromatographed over silica, eluting with EtOAc/ hexane to yield 5.3 g (58%) of **13** as a yellow oil. ¹H NMR (CDCl₃): δ 1.33, (t, *J*=9.6 Hz, 3H), 2.20 (s, 3H), 3.60 (s, 2H), 4.25 (q, *J*=9.2 Hz, 2H), 4.63 (s, 2H); MS(CI) calcd for C₉H₁₂N₂O₅: [M+H] 229. Found: [M+H] 229; Anal. calcd for C₉H₁₂N₂O₅: C, 47.37; H, 5.30; N, 12.28. Found: C, 47.40; H, 5.46; N, 11.86.

4.1.13. Ethyl 4-(ethanimidoylamino)-3-oxobutanoate (14) and ethyl (2-methyl-1*H*-imidazol-5-yl)acetate (15). Ester 13 (80 mg; 0.35 mmol) was added to 5% Pd–C (10 mg) in EtOH (20 mL) and placed under 40 psi hydrogen gas for 4 h. The reaction mixture was filtered and concentrated at ambient temperature. The residue was chromatographed using reverse phase. Two products were isolated:

Ester 14 was the first component off the column. ¹H NMR (MeOD): δ 1.25 (t, J=7.2 Hz, 3H), 2.35 (s, 3H), 3.59
(s, 2H), 4.15 (q, J=9.6 Hz, 2H), 4.91 (s, 2H); HRMS calcd for C₈H₁₄N₂O₃: 187.1083, found: 187.1064.

Imidazole **15** was the second component off the column. ¹H NMR (MeOD): δ 1.24 (t, J=7.2 Hz, 3H), 2.35 (s, 3H), 3.58 (s, 2H), 4.14 (q, J=6.0 Hz, 2H), 6.70 (s, 1H); HRMS calcd for C₈H₁₂N₂O₂: [M+H] 169.0977. Found: [M+H] 169.0933.

Heating converted 14 to 15.

4.1.14. 4-[(2E)-5-Hydroxypent-2-enyl]-3-methyl-1,2,4oxadiazol-5(4H)-one (17) and (3E)-5-[(3-methyl-1,2,4oxadiazol-5-yl)oxy]pent-3-en-1-ol (18). Dihydroxy-olefin 16 (109 mg, 1.0 mmol), Ph₃P (262 mg, 1.0 mmol) and 2b (100 mg, 1.0 mmol) were dissolved in THF (5.0 mL). The reaction mixture was cooled in an ice bath before adding DEAD (174 mg, 1.0 mmol) drop wise. TLC (EtOAc) indicated that 16 was consumed and two new products were present. The reaction mixture was concentrated and chromatographed, eluting with EtOAc/hexane. The first compound to elute was identified as the O-alkylated derivative **18**, 13 mg (7%). ¹H NMR (CDCl₃): δ 2.23 (s, 3H), 2.32 (q, J = 15 Hz, 2H), 3.68 (t, J = 12.4 Hz, 2H), 4.15 (dd, J=6, $\hat{1}.2$ Hz, 2H), 5.55, (pent. J=5.6 Hz, 1H), 5.73, (pent, Hz, J=8.4 Hz, 1H); MS(CI) calcd for C₈H₁₂N₂O₃: [M+H] 185. Found: [M+H] 185; Anal. calcd for C₈H₁₂N₂O₃: C, 36.01; H, 4.03; N, 27.99. Found: C, 35.76; H, 3.99; N, 28.26.

The second product to elute was the N-alkylated derivative **17**, 26 mg (14%). ¹H NMR (CDCl₃): δ 2.25 (s, 3H), 2.36 (q, J=6.4 Hz, 2H), 3.70 (t, J=6.0 Hz, 2H), 4.89 (dd, J=6.0, 0.8 Hz, 2H), 5.80 (pent, J=6.4 Hz, 1H), 5.94 (pent, J=7.2 Hz, 1H); MS(CI) calcd for C₈H₁₂N₂O₃: [M+H] 185. Found: [M+H] 185; Anal. calcd for C₈H₁₂N₂O₃: C, 36.01; H, 4.03; N, 27.99. Found: C, 36.18; H, 4.14; N, 27.86.

Dihydroxy-olefin **16** (109 mg, 1.0 mmol) and **2b** (100 mg, 1 mol) were dissolved in THF (5.0 mL). Ph₃P–polymer (3.0 mmol/g) (500 mg, 1.5 mmol) was added and the mixture was slowly stirred while adding DEAD (174 mg, 1.0 mmol). The products were identical to the reaction with unbound Ph₃P: O-alkylated 34 mg (18%) of **18**, and N-alkylated 50 mg (27%) of **17**.

4.1.15. 3-(3-Methyl-5-oxo-1,2,4-oxadiazol-4(5*H***)-yl)propanal (19). Oxadiazolone 2b** (729 mg, 7.24 mmol) was dissolve in ethanol (15 mL) then combined with acrolein (0.53 mL, 7.24 mmol) and triethylamine (0.10 mL, 0.723 mmol) and stirred for 18 h. The reaction mixture was partitioned between H₂O/EtOAc. The organic layer was washed with brine, dried over MgSO₄ and concentrated producing 1.3 g of **19** (100% crude) as a pale orange oil. ¹H NMR (CDCl₃): δ 2.36 (s, 3H), 3.04 (t, *J*=6 Hz, 2H), 3.80 (t, *J*=6 Hz, 2H), 9.73 (s, 1H); ¹³C NMR (CDCl₃): δ 18.6, 40.8, 58.6, 95.1, 157.0, 198.9; MS(CI) calcd for C₆H₈N₂O₃: [M+H] 157. Found: [M+H] 157.

4.1.16. Ethyl (2E)-2-fluoro-5-(3-methyl-5-oxo-1,2,4-oxadiazol-4(5H)-yl)pent-2-enoate (21). Triethylfluorophosphonate (**20**) (52.9 g, 0.218 mol) and LiCl (10.1 g, 0.238 mol) were dissolved in THF (100 mL). DBU (33.2 g, 0.218 mol) was added drop wise over a 20 min. period. After stirring 1 h a deep orange-red color developed, where the mixture was cooled to -78 °C. Aldehyde 19 (31.2 g, 0.199 mol) dissolved in THF (60 mL) was added to the reaction mixture over a 45 min period. The mixture was stirred 4 h at -78 °C then quenched with saturated NH₄Cl solution. After warming to ambient temperature the mixture was diluted with H₂O (100 mL) and the product extracted into EtOAc. The organic layer was dried over MgSO4 and concentrated to yield 47.9 g (98%) of 21 as a dark amber oil. ¹H NMR (CDCl₃): δ 1.285 (t, J=6.8 Hz, 3H), 2.26 (s, 3H), 2.85 (q, J = 6.4 Hz, 2H), 3.64 (t, J = 7.2 Hz, 2H), 4.24 (q, J=7.2 Hz, 2H), 5.85 (dt, J=19.6, 8.8 Hz, 1H); ¹³C NMR (CDCl₃): δ 10.46, 14.21, 24.97, 25.02, 41.10, 41.14, 62.11, 117.01, 117.21, 148.11, 150.70, 156.55, 159.34, 160.46, 160.81; ¹⁹F NMR (CDCl₃): δ -117.43 (d, J=20.4 Hz); MS(CI) calcd for $C_{10}H_{13}FN_2O_4$: [M+H] 245. Found: [M+ H] 245.

4-[(3E)-4-Fluoro-5-hydroxypent-3-enyl]-3-4.1.17. methyl-1,2,4-oxadiazol-5(4H)-one (22). Ester 21 (6.2 g, 25.4 mmol) was dissolved in THF (100 mL) and cooled to -5 °C. Red-Al 68% in toluene (8.5 mL, 27.9 mmol) was added drop wise to the cooled solution. The reaction mixture was monitored by TLC every 0.5 h until the starting material was consumed. The reaction was quenched with 75 mL of saturated Rochelle's salt. The product was extracted into CH₂Cl₂ (2×150 mL), dried over MgSO₄ and concentrated. The residue was chromatographed over silica, eluting with EtOAc/hexane to yield 2.1 g (41%) of 22 as an oil. ¹H NMR (CDCl₃): δ 2.22 (s, 3H), 2.43 (q, J= 8.0 Hz, 2H), 3.62–3.56 (m, 2H), 4.12 (d, J=20 Hz, 2H), 5.14–5.05 (m, 1H); ¹³C NMR (CDCl₃): δ 10.66, 14.38, 24.16, 24.25, 42.35, 57.50, 60.63, 103.54, 103.77, 156.55, 159.80, 162.32, 171.46; ¹⁹F NMR (CDCl₃): δ – 106.69 (dd, J = 42.0, 30.4 Hz; MS(CI) calcd for C₈H₁₁FN₂O₃: [M+H] 203. Found: [M+H] 203; Anal. calcd for C₈H₁₁FN₂O₃·0.1 H₂O: C, 47.10; H, 5.53; N, 13.73. Found: C, 46.96; H, 5.41; N, 13.40.

4.1.18. 4-[(3E)-5-Bromo-4-fluoropent-3-enyl]-3-methyl-1,2,4-oxadiazol-5(4H)-one (23). Alcohol 22 (800 mg, 3.98 mmol) and CBr₄ (3.28 g, 9.9 mmol) were dissolved in CH₂Cl₂ and cooled to -5 °C. Ph₃P-polymer (3.0 mmol/ g) (3.98 g, 12.0 mmol) was added to the reaction mixture and stirred for 18 h. The reaction mixture was filtered and concentrated. The residue was chromatographed over silica, eluting with EtOAc/hexane to yield 600 mg (57%) of 23 as an oil. ¹H NMR (CDCl₃): δ 2.28 (s, 3H), 2.46 (q, J= 12.0 Hz, 2H), 3.68 (t, J = 6.8 Hz, 2H), 4.10 (d, J = 22.8 Hz, 2H), 5.35–5.26 (m, 1H);); 13 C NMR (CDCl₃): δ 10.67, 25.13, 27.85, 31.97, 41.66, 41.92, 105.47, 105.70, 156.25, 169.32; ¹⁹F NMR (CDCl₃): δ -105.27 (dd, J=44.0, 22.4 Hz); MS(CI) calcd for C₈H₁₀BrFN₂O₂: [M+H] 265 and 267. Found: [M+H] 265 and 267; Anal. calcd for C₈H₁₀BrFN₂O₂·0.05 EtOAc: C, 36.55; H, 3.89; N, 10.40; F, 7.05; Br, 29.65. Found: C, 36.92; H, 3.89; N, 10.35; F, 6.69: Br, 29.75.

4.1.19. Ethyl (2*E*)-5-{[*tert*-butyl(dimethyl)silyl]oxy}-2-fluoropent-2-enoate (25a) and ethyl (2*Z*)-5-{[*tert*-butyl-(dimethyl)silyl]oxy}-2-fluoropent-2-enoate (25b). NaH (60% suspension in mineral oil) (1.7 g, 72.0 mmol) was

washed with hexane to remove the mineral oil, toluene (200 mL), **20** (17.2 g, 71.0 mmol) and **24**⁸ (13.2 g; 0.07 mol) dissolved in toluene (50 mL) was reacted at 0 °C as described for Compound **21** to yield 5.9 g (30.5%) of **25a**. ¹H NMR (CDCl₃): δ 0.04 (s, 6H), 0.87 (s, 9H), 1.33 (t, *J*=7.2 Hz, 3H), 2.75–2.69 (m, 2H), 3.68 (dt, *J*=0.8, 6.0 Hz, 2H), 4.28 (q, *J*=7.2 Hz, 2H), 6.19 (dt, *J*=33.6, 7.6 Hz, 1H); ¹⁹F NMR (CDCl₃): δ -121.67 (d, *J*=23.2 Hz); MS(CI) calcd for C₁₃H₂₅O₃FSi: [M+H] 277. Found: [M+H] 277.

A minor component was isolated and purified and identified as **25b** the *Z*-isomer: ¹H NMR (CDCl₃): δ 0.037 (s, 6H), 0.87 (s, 9H), 1.330 (t, *J*=7.2 Hz, 3H), 2.47–2.42 (m, 2H), 3.68 (dt, *J*=0.8, 8.0 Hz, 2H), 4.28 (q, *J*=7.2 Hz, 2H), 6.00 (dt, *J*=21.2, 8.0 Hz, 1H). ¹⁹F NMR (CDCl₃): δ – 129.79 (d, *J*=33.2 Hz); MS(CI) calcd for C₁₃H₂₅O₃FSi: [M+H] 277. Found: [M+H] 277.

4.1.20. (2*E*)-5-{[*tert*-Butyl(dimethyl)silyl]oxy}-2-fluoropent-2-en-1-ol (26). Ester 25a (6.0 g, 21.7 mmol) dissolved in THF (70 mL), Red-Al 68% in toluene (9.2 mL, 30.0 mmol) was reacted at -5 °C as described for Compound 22 to yield 4.0 g (79%) of 26. ¹H NMR (CDCl₃): δ 0.04 (s, 6H), 0.87 (s, 9H), 2.29–2.21 (m, 3H, 1 exchangeable), 3.63 (dt, *J*=1.2, 8.0 Hz, 2H), 4.20 (d, *J*=26.0 Hz, 2H), 5.20 (dt, *J*=26.0, 12.0 Hz, 1H); ¹⁹F NMR (CDCl₃): δ -109.63 (q, 28.4 Hz); MS(CI) calcd for C₁₁H₂₃FO₂Si: [M+H] 235. Found: [M+H] 235; Anal. calcd for C₁₁H₂₃FO₂Si: C, 56.37; H, 9.89. Found: C, 56.38; H, 10.19.

4.1.21. (2*E*)-2-Fluoropent-2-ene-1,5-diol (27). Alcohol 26 (131.0 g, 0.56 mol) was dissolved in EtOH (1.5 l) and concn HCl (15 mL) was added and the mixture was stirred for 0.25 h. TLC (20% EtOAc/hexane) indicated that the starting material was consumed. The mixture was concentrated and the residue was azeotroped twice with toluene (500 mL) to produce 71.8 g (100% crude) of **27** as an oil. ¹H NMR (CDCl₃): δ 2.30–2.23 (m, 2H), 3.64 (dt, *J*=0.8, 8.0 Hz, 2H), 4.18 (d, *J*=29.6 Hz, 2H), 5.21 (dt, *J*=27.0, 11.6 Hz, 1H); ¹⁹F NMR (CDCl₃): δ -108.16 (q, *J*=30.8 Hz); MS(CI) calcd for C₅H₉O₂F: [M+H] 121. Found: [M+H] 121.

4.1.22. (2E)-2-Fluoro-5-[(methylsulfonyl)oxy]pent-2envl methanesulfonate (28). Dihydroxyolefin 27 (71.8 g, 0.56 mol) and Et₃N (141.4 g/195 mL, 1.4 mol) were dissolved in CH₂Cl₂ (1.01) and cooled in an ice bath. Ms_2O (243.4 g, 1.4 mol) was dissolved in CH_2Cl_2 (0.5 l) and added drop wise. The reaction mixture was stirred 15 min when TLC (60% EtOAc/hexane) indicated that the starting material was consumed. The reaction mixture was partitioned between H₂O/CH₂Cl₂. The organic layer was washed with H₂O, dried over MgSO₄ and concentrated to yield 164.9 g (100% crude) of **28** as a crystalline solid. ¹H NMR (CDCl₃): δ 2.55 (q, J = 10.0 Hz, 2H), 3.02 (s, 3H), 3.07 (s, 3H), 4.25 (dt, J=1.2, 8.4 Hz, 2H), 4.82 (d, J=27.6 Hz, 2H), 5.50 (dt, J=24.8, 11.2 Hz, 1H); ¹⁹F NMR (CDCl₃): δ -107.91 (q, J=27.2 Hz); MS(CI) calcd for $C_7H_{13}O_6S_2F$: [M+NH₄] 294. Found: [M+NH₄] 294.

4.1.23. (*3E*)-**4**-Fluoro-5-(3-methyl-5-oxo-1,2,4-oxadiazol-**4**(5*H*)-yl)pent-3-enyl methanesulfonate (29). Dimesylate **28** (164.9 g, 0.55 mol) dissolved in MEK (0.6 l) was added to a slurry of **2a** (84.8 g, 0.61 mol) in MEK (0.6 l) and heated at reflux 6.0 h where TLC (60% EtOAc/hexane) indicated that **28** was still present. Additional **2a** (17 g) was added and the mixture was stirred at reflux for an additional 2.0 h at which time the starting material was consumed. The reaction mixture was cooled and filtered and the solid washed with MEK. The filtrate was concentrated to yield 204 g (100% crude) of **29** as a crystalline solid. ¹H NMR (CDCl₃): δ 2.29 (s, 3H), 2.63 (q, J=6.4 Hz, 2H), 3.01 (s, 3H), 4.26 (dt, J=0.8, 6.4 Hz, 2H), 4.31 (d, J=19.6 Hz, 2H), 5.409 (dt, J=19.6, 8.4 Hz, 1H); ¹⁹F NMR (CDCl₃): δ -106.89 (q, J=21.2 Hz); MS(CI) calcd for C₉H₁₃N₂O₅SF: [M+H] 281. Found: [M+H] 281.

4.1.24. 4-[(2E)-2-Fluoro-5-iodopent-2-enyl]-3-methyl-**1,2,4-oxadiazol-5(4H)-one (30).** Mesylate **29** (204.8 g, 0.56 mol) was dissolved in MEK (1.21) and solid NaI (167 g, 1.12 mol) was added to the mixture and stirred 18 h at which time TLC (60% EtOAc/hexane) indicated that the starting material was consumed. The reaction mixture was partitioned between an equal volume of H₂O and the aqueous layer was washed with Et₂O then CH₂Cl₂ (500 mL each). The combined organic layers were washed with 1% $Na_2S_2O_3$, dried over MgSO₄ and concentrated. The residue was chromatographed over silica, eluting with EtOAc/ hexane to yield 87.4 g (50%) of **30** as a crystalline solid. 1 H NMR (CDCl₃): δ 2.30 (s, 3H), 2.74 (q, J=7.2 Hz, 2H), 3.20 (dt, J=0.8, 6.8 Hz, 2H), 4.31 (d, J=19.6 Hz, 2H), 5.39 (dt, J=19.6, 8.4 Hz, 1H); ¹⁹F NMR (CDCl₃): δ -108.31 (q, J = 21.2 Hz; MS(CI) calcd for C₈H₁₀FIN₂O₂: [M+H] 313. Found: [M+H] 313; Anal. calcd for $C_8H_{10}FIN_2O_2$; C, 30.79; H, 3.23; N, 8.98; I, 40.66. Found: C, 30.75; H, 2.99; N, 8.92; I, 40.41.

4.1.25. Ethyl (2*E*)-4-{[*tert*-butyl(dimethyl)silyl]oxy}-2-fluorobut-2-enoate (32a) and ethyl (2*Z*)-4-{[*tert*-butyl(dimethyl)silyl]oxy}-2-fluorobut-2-enoate (32b). NaH (60% suspension in mineral oil) (3.9 g, 0.10 mol), THF (300 mL), **20** (25 g, 0.10 mol) and **31**⁹ (17.4 g; 0.1 mol) was reacted at 0 °C as described for Compound **21** to yield 8.9 g (34%) of **32a**. ¹H NMR (CDCl₃): δ 0.08 (s, 6H), 0.88 (s, 9H), 1.25 (t, *J*=7.2 Hz, 3H), 4.28 (q, *J*=7.2 Hz, 2H), 4.64 (dd, *J*=3.6, 5.6 Hz, 2H), 6.01 (dt, *J*=19.6, 5.6 Hz, 1H); ¹⁹F NMR (CDCl₃): δ -123.95 (dt, *J*=20.8, 3.2 Hz); MS(CI) calcd for C₁₂H₂₃O₃FSi: [M+H] 263. Found: [M+H] 263.

A minor component was isolated and purified and identified as the *Z*-isomer **32b**: ¹H NMR (CDCl₃): δ 0.08 (s, 6H), 0.88 (s, 9H), 1.25 (t, *J*=7.2 Hz, 3H), 4.28 (q, *J*=7.2 Hz, 2H), 4.41 (dd, *J*=6.4, 1.2 Hz, 2H), 6.19 (dt, *J*=33.6, 6.4 Hz, 1H); ¹⁹F NMR (CDCl₃): δ -127.28 (dt, *J*=35.2, 3.6 Hz); MS (CI) calcd for C₁₂H₂₃O₃FSi: [M+H] 263. Found: [M+H] 263.

4.1.26. (2*E*)-4-{[*tert*-butyl(dimethyl)silyl]oxy}-2-fluorobut-2-en-1-ol (33). Ester 32a (8.7 g, 33.0 mmol) was dissolved in THF (100 mL) and cooled in an ice bath. LiBH₄ 2.0 M in THF (33 mL, 66.0 mmol) was added and the mixture stored at -40 °C for 18 h. The reaction was quenched with H₂O. A solid formed which was filtered and washed with THF. The filtrate was concentrated and chromatographed over silica, eluting with EtOAc/hexane to yield 4.5 g (62%) of 33. ¹H NMR (CDCl₃): δ 0.08 (s, 6H), 0.88 (s, 9H), 4.20 (dd, J=6.8, 2.4 Hz, 2H), 4.24 (d, J=19.6 Hz, 2H), 5.38 (dt, J=20.4, 7.2 Hz, 1H); ¹⁹F NMR (CDCl₃): δ -109.28 (q, J=22.0 Hz); MS(CI) calcd for C₁₀H₂₁O₂FSi: [M+H] 221. Found: [M+H] 221; Anal. calcd for C₁₀H₂₁O₂FSi: C, 54.51; H, 9.61. Found: C, 54.44; H, 9.78.

4.1.27. 4-((2E)-4-{[tert-Butyl(dimethyl)silyl]oxy}-2-fluorobut-2-enyl)-3-methyl-1,2,4-oxadiazol-5(4H)-one (34). Alcohol 33 (25.5 g, 0.115 mol), 2b (11.3 g, 0.1 mol) and triphenylphosphine (29.7 g, 0.11 mol) were dissolved in THF (500 mL) and cooled in an ice bath. DEAD (19.1 g/ 17.3 mL, 0.11 mol) was added drop wise to the reaction mixture. The reaction progress was monitored by TLC (60% EtOAc/hexane). When 33 was consumed the reaction mixture was concentrated and the residue triturated with Et₂O. The filtrate was concentrated and chromatographed over silica, eluting with EtOAc/hexane to yield 10.4 g (31%) of **34**. ¹H NMR (CDCl₃): δ 0.08 (s, 6H), 0.88 (s, 9H), 2.28 (s, 3H), 4.31 (dd, J=6.4, 2.8 Hz, 2H), 4.44 (d, J=19.2 Hz, 2H), 5.53 (dt, J = 20.0, 6.4 Hz, 1H); ¹⁹F NMR (CDCl₃): δ -109.06 (q, J=20.8 Hz); MS(CI) calcd for C₁₃H₂₃N₂O₃FSi: [M+H] 303. Found: [M+H] 303; Anal calcd for C13H23N2O3FSi: C, 51.63; H, 7.67; N, 9.26. Found: C, 51.84; H, 7.82; N, 8.95.

4.1.28. 4-[(*2E*)-**2-**Fluoro-**4-**hydroxybut-**2-**enyl]-**3-**methyl-**1,2,4-oxadiazol-5(4***H***)-one (35**). Oxadiazolone **34** (10.4 g, 34.0 mmol) was dissolved in EtOH (100 mL) and concn HCl (1.0 mL) was added and the mixture stirred for 1 h. TLC (60% EtOAc/hexane) indicated that **34** was consumed. The mixture was concentrated to dryness to yield 6.9 g (100% crude) of **35**. ¹H NMR (CDCl₃): δ 2.31 (s, 3H), 4.25 (dd, *J*=6.8, 2.8 Hz, 2H), 4.44 (d, *J*=21.2 Hz, 2H), 5.65 (dt, *J*=20.0, 7.2 Hz, 1H); ¹⁹F NMR (CDCl₃): δ -107.88 (q, *J*=21.6 Hz); MS(CI) calcd for C₇H₉FN₂O₃: [M+H] 189. Found: [M+H] 189.

4.1.29. (2*E*)-3-Fluoro-4-(3-methyl-5-oxo-1,2,4-oxadiazol-**4**(5*H*)-yl)but-2-enyl methanesulfonate (36). Alcohol 35 (6.9 g, 34.0 mmol) and Et₃N (4.0 g/5.5 mL, 40.0 mmol) were dissolved in CH₂Cl₂ (100 mL). Ms₂O (6.9 g, 40.0 mol) was added drop wise 30 min. TLC (60% EtOAc/hexane) indicated that **35** was consumed. The reaction mixture was poured into H₂O and the layers separated. The organic layer was washed with saturated NaHCO₃, dried over MgSO₄ and concentrated to yield 8.3 g (92%) of **36** as a crystalline solid. ¹H NMR (CDCl₃): δ 2.31 (s, 3H), 3.06 (s, 3H), 4.42 (d, *J*= 20.0 Hz, 2H), 4.92 (dd, *J*=8.0, 1.2 Hz, 2H), 5.67 (dt, *J*= 8.7, 8.0 Hz, 1H); ¹⁹F NMR (CDCl₃): δ -101.10 (q, *J*= 19.6 Hz); MS(CI) calcd for C₈H₁₁FN₂O₅S: [M+H] 267. Found: [M+H] 267; Anal. calcd for C₈H₁₁FN₂O₅S: C, 36.09; H, 4.16; N, 10.52. Found: C, 36.43; H, 4.06; N, 10.41. 4.1.30. 4-[(2*E*)-2-Fluoro-4-iodobut-2-enyl]-3-methyl-**1,2,4-oxadiazol-5(4H)-one (37).** Mesylate **36** (8.3 g, 32.0 mmol) and NaI (9.8 g, 65.0 mmol) were stirred for 30 min in MEK. TLC (40% EtOAc/hexane) indicated that 36 was consumed. The reaction mixture was partitioned between Et_2O/H_2O . The aqueous layer was washed with Et₂O and the combined organic layers were washed with 1% Na₂S₂O₃, dried over MgSO₄ and concentrated. Chromatographed over silica eluting with EtOAc/hexane to yield 4.5 g (47%) of **37** as a crystalline solid. ¹H NMR (CDCl₃): δ 2.31 (s, 3H), 3.98 (dd, J=9.6, 0.8 Hz, 2H), 4.34 (d, J=19.6 Hz, 2H), 5.78 (dt, J=16.8, 7.6 Hz, 1H); ¹⁹F NMR (CDCl₃): $\delta - 106.08$ (q, J = 19.2 Hz); MS(CI) calcd for C₇H₈FIN₂O₂: [M+H] 299. Found: [M+H] 299; Anal. calcd for C₇H₈FIN₂O₂: C, 28.21; H, 2.71; N, 9.40. Found: C, 28.36; H, 3.04; N, 9.37.

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Tetrahedron

Tetrahedron 60 (2004) 10915-10920

Synthesis of new unsymmetrical optically active (s)-(+)-naproxen dendrimers

Srinagesh Kumar Potluri,* A. Raghu Ramulu and M. Pardhasaradhi

Gas Based Chemicals and Organic Intermediates Division, Indian Institute of Chemical Technology, Hyderabad 500 007, India

Received 29 July 2004; revised 7 September 2004; accepted 9 September 2004

Available online 30 September 2004

Abstract—Preparation of new unsymmetrical optically active (s)-(+)-naproxen dendrimers containing 2-hydroxymethyl-1,4-butanediol and 2,2-bis(hydroxymethyl)-1,4-butanediol cores is described. These polyarylester dendrimers are unsymmetrical with respect to chain lengths and aid in studying controlled drug delivery based on differential enzymatic cleavage. Synthesis of a new acid dendritic wedge containing (s)-(+)-naproxen is also reported.

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1. Introduction

Dendrimers are macromolecules with well-defined and perfectly branched architecture, synthesized in an iterative sequence of reactions.^{1–5} Unprecedented control over structural unit positioning renders these materials a wide range of applications in the areas of drug delivery, catalysis, nanostructured materials, liquid crystals, electrochemical devices, and adhesives.^{6–12} Typically, dendrimers with a combination of a discrete number of functionalities in one molecule and with high densities of local active groups have attracted a lot of attention from medicinal chemists.¹³ The low polydispersity provided by dendritic systems has favored their use in biomedical applications because of reproducible pharmacokinetic behavior.¹⁴

Owing to the advantages of dendritic therapeutics^{15–21} and the promising characteristics of novel polyarylester dendrimers²² in our initial in vitro studies, we were inspired to undertake a systematic study by introducing optically active drug molecules on the periphery of unsymmetrical dendrimers. Dendrimers built on unsymmetric cores by virtue of their unequal chain lengths are likely to get hydrolyzed by the enzymes at different rates. The differential rates of hydrolysis would release the dendrimer bound drugs gradually, thus sustaining the availability of the drug for longer periods. In addition, ester bonds are characterized by their susceptibility to hydrolysis, an attractive feature for biodegradable systems.

From the academic standpoint, study of chiral dendrimers should enable insights upon the impact of chirality on the topology of the macromolecular system in particular and molecular chirality in general. As a part of our program directed towards the development of efficient synthetic procedures for the preparation^{23,24} and application of dendritic macromolecules for sustained drug release and catalysis,²³ we have given considerable attention to extending the studies to the synthesis of chiral dendrimers. Therefore, we now report the synthesis of unsymmetrical



Scheme 1.

Keywords: Dendrimers; Convergent approach; Dendritic wedge; Unsymmetrical; Polyaryl ester; Trichloroethyl protection.

^{*} Corresponding author. Present address: Department of Chemistry, Southern Methodist University, Fondren Science Bldg, 3215 Daniel Avenue, Dallas, TX 75275, USA. Tel.: +1 214 768 4392; fax: +1 214 768 4089; e-mail: spotluri@mail.smu.edu

^{0040–4020/\$ -} see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.033





polyester dendritic scaffolds with a non-steroidal antiinflammatory, analgesic and antipyretic drug, s-(+)naproxen, on the periphery.

2. Results and discussion

By implementing a convergent approach, synthesis of new optically active naproxen dendrimers based on 2-hydroxymethyl-1,4-butanediol (5) and 2,2-bis(hydroxymethyl)-1,4-butanediol (6) was accomplished.^{23,24} To begin the synthesis, 2,2,2-trichloroethyl-3,5-dihydroxybenzoate (1) was condensed with *s*-(+)-naproxen (2) using DPTS/DCC to afford the G1-ester dendritic wedge (3) in 68% yield (Scheme 1). The trichloroethylester of (3) was then removed selectively in the presence of other ester groups using zinc and acetic acid to provide the acid wedge (4) in 93% yield. Surprisingly, coupling of two relatively small dendrons (4) and bisphenol (1) to prepare higher generation wedges was difficult to achieve in good yields and purity.

In order to facilitate the assignments of ¹H NMR and ¹³C NMR δ values in the complex dendritic structures, *s*-(+)-naproxen was reacted with triol (5) and tetrol (6) in the presence of DPTS/DCC to give the first generation dendrimers (7) and (8) in 63 and 60% yields, respectively (Scheme 2).

To prepare the second-generation dendrimers, the focal point acid of the convergent dendron (4) was reacted with

 H_2



the cores (5) and (6) using DPTS/DCC to give dendrimers (9) and (10) in 65 and 66% yields, respectively (Scheme 3).

In the absence of higher generation dendritic wedge, an alternative strategy was used in the preparation of third generation dendrimers (13) and (14). To achieve this, compounds (11) and (12), previously reported by us,²² derived from the cores (5) and (6) were employed.

Accordingly, the phenolic intermediates **11** and **12** were reacted with acid dendron (**4**) in the presence of DCC/DPTS to give third-generation dendrimers **13** and **14** in 20 and 14% yields, respectively (Scheme 4).

Low yields of dendrimers 13 and 14 may be assumed to be originating from some kind of intramolecular transesterification reactions occurring since the reactions were performed for extended periods of time (6 days). A large number of other minor products at various stages of esterification were isolated predominantly as mixtures. In addition, compounds having relatively higher R_f values when compared to the required product were also observed. A remarkable aspect of the present work is the use of phenolic cores that are more reactive when compared to the aliphatic cores.

The present study corroborates our previous work,²² that steric crowding could be circumvented by a fine combination of convergent approach with a little divergent synthesis. This strategy particularly helped us in achieving



the synthesis of higher molecular weight dendrimers of third generation.

Characterization of the ester dendrimers was performed using a combination of ¹H NMR and ¹³C NMR spectroscopy and mass spectrometry techniques. The large $R_{\rm f}$ difference between the partially and completely reacted alcoholic/phenolic groups of the core facilitated easy purification of the dendrimers. To establish the extent of reaction and the generation number from the ¹H NMR spectra, the integration of resonances due to ethereal hydrogens of the core (δ : 4.4) was compared with the methoxy hydrogens (δ : 3.8). Furthermore, the discrete resonances arising from aromatic and benzylic hydrogens were invaluable in detecting impurities and defects. MS techniques (FAB and MALDI) in combination with elemental analysis and NMR were used to establish the molecular weight and purity of the dendritic wedges and dendrimers.

Chiroptical property studies of the dendrimers were performed using the protocol reported by Lartigue et al.²⁵ The specific rotation values of higher generation dendrimers were similar (Fig. 1). On the other hand, molar rotation



Scheme 4.



Figure 1.





increased with generation number and multiplicity of functional groups (Fig. 2). However, by dividing the molar rotation values with the number of stereogenic units, the optical rotations of higher generation dendrimers remained just about constant (Fig. 3). These observations suggest that the peripheral naproxen groups appear not to interact with each other and are conformationally more flexible for direct interaction with enzymes. In contrast, the first generation dendrimers showed lower specific rotations when compared to s(+) naproxen. This discrepancy may be attributed to back folding of peripheral groups towards the core thus resulting in some kind of internal compensation. However, in the absence of molecular modeling studies the justification presented here is an assumption. To explain the topology and conformational flexibility of the dendrimers possessing chiral peripheral surface groups,



some researchers²⁶⁻³⁰ have used this kind of justification based on optical rotation.

3. Conclusion

The requirement for controlled drug release based on differential rates of hydrolysis has inspired the design and preparation of the chiral dendrimers having (s)-(+)-naproxen groups on the periphery. In vitro enzymatic hydrolysis of our dendrimers has shown some promising results. Furthermore, biological evaluation of these dendrimers with emphasis on biocompatibility of the by-products of hydrolysis will follow. Nonetheless, the dendritic systems exemplified in this paper seem to have immense potential due to their polyvalency and well-defined structures. From the synthetic standpoint, the new dendritic wedges and the route presented here demonstrate feasibility for the introduction of other diverse biodegradable polyesters and other drug molecules.

4. Experimental

4.1. General directions

¹H NMR (200 MHz) and ¹³C NMR (50 MHz) spectra were recorded on Varian FT (Gemini) instrument, using tetramethyl silane (TMS) as the internal standard. The chemical shifts are expressed in δ scale. The abbreviations such as s, d, t, m, b refers to singlet, doublet, triplet, multiplet and broad, respectively. The trisubstituted benzene and napthyl rings are abbreviated as Ar and Naph, respectively. Mass spectra were recorded on VG Micromass 7070 H (EI), Autospec (FAB), and Kratos Kompact SEQ (MALDI-TOF). Optical rotations were measured on JASCO DIP-370 digital polarimeter. Elemental analysis was recorded on a Perkin-Elmer 240C-CHN analyzer. The melting points reported are uncorrected and were determined using Buchi 525 instrument. Solvents were purified and dried before use. Completion of the reaction and purity of the synthesized compounds were checked by TLC performed on silica gel (acmes) plates, using iodine and H₂SO₄ for visualizing the spots.

4.1.1. 2,2,2-Trichlorethyl-3,5-di[1-(6-methoxy-2-naphthyl) ethyl carbonyloxy]benzoate (3). To a solution of (s)-(+)-naproxen (2) (15.60 g, 0.067 mol), 2,2,2-trichloroethyl-3,5-dihydroxy benzoate (1) (6.44 g, 0.022 mol) in dry dichloromethane (50 ml) was added 4-(dimethylamino)pyridinium *p*-toluenesulphonate (DPTS) (1.32 g, 0.004 mol). The contents were stirred at 25 °C for 15 min under nitrogen atmosphere. Dicyclohexylcarbodiimide (DCC) (13.96 g, 0.067 mol) was then added and stirring continued at room temperature for 24 h, during this time a precipitate of dicyclohexyl urea appeared. The reaction mixture filtered and the filtrate evaporated to dryness under reduced pressure, the crude product was column chromatographed (SiO_2) eluting with initially hexane and then a mixture of EtOAc/hexane (4:1) gave 3 (10.9 g) as a white solid in 68% yield. Mp 108–110 °C. ¹H NMR (CDCl₃) δ : 1.68 (d, 6H, J=7.0 Hz; 2×CH₃CH), 3.90 (s, 6H, 2× OCH_3), 4.05 (q, 2H, J=7.0 Hz, $2 \times CHCH_3$), 4.89 (s, 2H,

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CH₂CCl₃), and 6.98–7.75 (m, 15H: $3 \times \text{ArH}$, $2 \times \text{NaphH}$). ¹³C NMR (CDCl₃) δ : 18.39, 45.49, 55.30, 74.54, 76.38, 105.70, 119.17, 120.45, 120.74, 125.93, 126.13, 127.90, 128.99, 129.30, 130.59, 133.90, 134.52, 151.30, 157.86, 163.42, and 172.41. MS (FAB); *m*-nitrobenzylalcohol (matrix); *m*/*z* (%): 710 (M⁺, 61), 663 (41), 307 (100). Anal. Calcd for C₃₇H₃₁O₈Cl₃: C, 62.59; H, 4.40. Found: C, 62.5; H, 4.36. [α]²⁵_D = +45.6 (*c* 1.0, CHCl₃).

4.1.2. 3,5-Di[(1-(6-methoxy-2-naphthyl)ethylcarbonyloxy)] benzoic acid (4). To a solution of (3) (3.0 g, 4.22 mmol) in THF (25 ml) was added glacial acetic acid (5 ml) and the solution was stirred at room temperature for 15 min under nitrogen atmosphere. Zinc dust (0.780 g, 0.012 mol) was then added and the contents were stirred vigorously at room temperature for 2 h. The reaction mixture was filtered and the filtrate poured into water (100 ml) and extracted with diethyl ether (2×100 ml). The combined extracts were washed with water, dried over anhydrous Na₂SO₄ and evaporated to dryness under reduced pressure. The crude product was column chromatographed (SiO_2) eluting with initially benzene and then chloroform to give 4 and it was further purified by recrystallization from benzene as a white solid (2.27 g) in 93% yield. Mp 104-106 °C. ¹H NMR (CDCl₃) δ : 1.68 (d, 6H, J=7.0 Hz, 2× CH₃CH), 3.90 (s, 6H, $2 \times OCH_3$), 4.06 (q, 2H, J = 7.0 Hz, $2 \times CHCH_3$), and 6.98–7.75 (br m, 15H: $3 \times ArH$, $2 \times$ Naph*H*). ¹³C NMR (CDCl₃) δ: 18.34, 45.45, 55.22, 105.74, 119.06, 120.53, 125.90, 126.12, 127.44, 128.25, 128.97, 129.27, 131.19, 133.87, 134.51, 151.16, 157.79, 169.78, and 172.38. MS (FAB); m-nitrobenzylalcohol (matrix); m/z: 578 (M⁺, 16), 212 (4), 185 (100). Anal. Calcd for C₃₅H₃₀O₈: C, 72.65; H, 5.23. Found C, 72.60; H, 5.19. $[\alpha]_D^{25} = +97.5$ (c 1.0, CHCl₃).

4.2. General procedure

4.2.1. First generation dendrimer (7). The title compound was prepared from triol (5) (400 mg, 3.3 mmol) and (s)-(+)-naproxen (2) (3.05 g, 13.2 mmol), DPTS (194 mg, 660 µmol), and DCC (2.74 g, 13.3 mmol) following the procedure as described for compound 3. After 24 h, the crude product was column chromatographed (SiO₂), eluting with initially chloroform and then a mixture of EtOAC/ $CHCl_3$ (1:9) to give 7 (1.57 g) as a white solid in 63% yield. Mp 166–170 °C. ¹H NMR (CDCl₃) δ : 1.34 (q, 2H, J= 4.7 Hz, CHCH₂), 1.49 (d, 9H, J = 7.0 Hz, $3 \times CH_3$ CH), 1.81 (m, 1H, CHCH₂), 3.70 (m, 3H, 3×CHCH₃), 3.80–3.92 [(m, 15H: 6H, 3×CH₂O; 9H, 3×OCH₃)], and 7.02–7.68 (m, 18H, 3×NaphH). ¹³C NMR (CDCl₃) δ: 18.12, 27.17, 34.65, 45.28, 55.16, 62.03, 63.73, 63.85, 105.62, 118.58, 125.82, 126.03, 127.06, 128.84, 129.16, 133.63, 135.35, 157.61, 174.10, and 174.21. MS (FAB); *m*-nitrobenzylalcohol (matrix); m/z: (%): 779 [(M+23)⁺, 4], 756 (M⁺, 16), 185 (100). Anal. Calcd for C₄₇H₄₈O₉: C, 74.58; H, 6.39. Found: C, 74.53; H, 6.30. $[\alpha]_D^{25} = +37.5$ (*c* 1.0, CHCl₃).

4.2.2. First generation dendrimer (8). This compound was prepared from 2,2-bis(hydroxymethyl)-1,4-butanediol (6) (0.150 g, 1.0 mmol), s-(+)-naproxen (2) (1.150 g, 5.0 mmol), DPTS (0.058 g, 0.2 mmol) and DCC (1.030 g, 5.0 mmol) by following the above general procedure. After 24 h, the crude product was column chromatographed

(SiO₂) eluting with initially hexane and then benzene to give **8** (0.60 g) as a white solid in 60% yields. Mp 124–126 °C. ¹H NMR (CDCl₃) δ : 1.23 (br s, 2H, CCH₂), 1.40 (dd, 12H, J=7.0, 7.0 Hz, 4×CH₃CH), 3.48–3.72 [m, 12H: (8H, 4×CH₂O), (4H, 4×CHCH₃)], 3.84 (two s, 12H, 4× OCH₃), and 6.69–7.60 (m, 24H, 4×NaphH). ¹³C NMR (CDCl₃) δ : 17.79, 18.16, 29.33, 40.55, 45.16, 55.09, 60.13, 63.85, 105.54, 118.87, 125.82, 125.89, 126.00, 127.08, 128.75, 129.16, 133.60, 135.03, 135.31, 157.56, 173.56, and 173.98. MS (FAB); *m*-nitrobenzyl alcohol (matrix); *m*/*z* (%): 1022 [(M+23)⁺, 8], 999 (M⁺, 52), 307 (100). Anal. Calcd for C₆₂H₆₂O₁₂: C, 74.53; H, 6.25. Found: C, 74.58; H, 6.19. [α]_D²⁵ = +33.6 (*c* 1.0, CHCl₃).

4.2.3. Second generation dendrimer (9). The title compound was prepared from triol (5) (0.050 g, 0.41 mmol), G1-acid wedge (4) (0.963 g, 1.6 mmol), DPTS (0.024 g, 0.08 mmol) and DCC (0.343 g, 1.6 mmol) by following the above general procedure. After 84 h, the crude product was column chromatographed (SiO₂) eluting with initially chloroform and then a mixture of EtOAC/ $CHCl_3$ (1:19) to give (9) (0.493 g) as a white solid in 65% yield. Mp 88–92 °C. ¹H NMR (CDCl₃) δ: 1.27 (s, 2H, CHC H_2), 1.63 (d, 18H, J=7.0, 7.0 Hz, 6×C H_3 CH), 1.80 $(m, 1H, CHCH_2), 3.88 (s, 18H, 6 \times OCH_3), 4.03 (m, 6H, 6 \times$ CHCH₃), 4.33 (m, 6H, 3×CH₂O), and 6.93–7.73 [m, 45H: (9H, $3 \times \text{Ar}H$), (36H, $6 \times \text{Naph}H$)]. ¹³C NMR (CDCl₃) δ : 18.37, 29.67, 35.02, 45.43, 55.27, 62.96, 64.68, 105.63, 119.10, 120.01, 120.10, 125.96, 126.15, 127.44, 128.93, 129.28, 131.69, 133.85, 134.57, 151.11, 151.14, 157.78, 164.49, 164.59, and 172.38. MS (MALDI); m/z (%): 1842 $[(M+39)^+, 100], 1825 [(M+23)^+, 64], 1802 (M^+, 60).$ Anal. Calcd for C₁₁₀H₉₆O₂₄: C, 73.32; H 5.37. Found: C, 73.26; H 5.43. $[\alpha]_D^{25} = +92.0$ (*c* 0.4, CHCl₃).

4.2.4. Second generation dendrimer (10). The title compound was prepared from tetrol (6) (100 mg, 0.66 mmol), G1-acid wedge (4) (1.926 g, 3.3 mmol), DPTS (0.034 g, 0.12 mmol) and DCC (0.658 g, 3.2 mmol) by following the general procedure. After 5 days, the crude product was column chromatographed (SiO₂) eluting initially with CHCl₃ and then a mixture of EtOAC/CHCl₃ (1:19) to give dendrimer (10) (1.050 g) in 66% yield as a white solid. Mp 145–149 °C. ¹H NMR (CDCl₃) δ : 1.25 (s, 2H, CCH₂), 1.60 (d, 24H, J=7.0 Hz, $8 \times CH_3$ CH), 3.83 (two s, 24H, $8 \times \text{OCH}_3$), 3.98 (q, 8H, J = 7.0 Hz, $8 \times$ $CHCH_3$), 4.20 (s, 6H, 3× CH_2O), 4.30 (br s, 2H, CH_2O), and 6.85-7.65 [m, 60H: (12H, 4×ArH; 48H, 8×NaphH]. ¹³C NMR (CDCl₃) δ: 18.33, 29.68, 41.17, 45.44, 55.25, 61.95, 64.68, 105.69, 119.06, 119.97, 120.30, 121.01, 125.98, 126.16, 127.43, 128.96, 129.28, 131.27, 133.86, 134.60, 151.12, 151.22, 157.77, 164.12, and 172.29. MS (MALDI); m/z (%): 2432 [(M+39)⁺, 26], 2416 [(M+23)⁺, 44], 2393 (M⁺, 100). Anal. Calcd for C₁₄₆H₁₂₆O₃₂: C, 73.29; H, 5.31. Found C, 73.19; H, 5.41. $[\alpha]_D^{25} = +86.0$ (c 1.0, CHCl₃).

4.2.5. Third generation dendrimer (13). The title compound was prepared from (**11**) (100 mg, 189 μ mol), acid wedge (**4**) (0.875 g, 1.514 mmol), DPTS (0.11 mg, 37 μ mol) and DCC (311 mg, 1.514 mmol) by following the general procedure. After 6 days, the crude product was column chromatographed (SiO₂) eluting with initially

CHCl₃ and then a mixture of EtOAC/CHCl₃ (1:19) to give **13** (0.150 g) as white solid in 20% yield. Mp 83–86 °C. ¹H NMR (CDCl₃) δ : 1.27 (s, 2H, CHCH₂), 1.65 (d, 37H, J= 7.0 Hz, 12×CH₃CH, CHCH₂), 3.85 (s, 36H, 12×OCH₃), 4.02 (m, 12H, 12×CHCH₃), 4.48 (m, 6H, 3×CH₂O), and 7.00–7.70 [m, 99H: (27H, 9×ArH); (72H, 12×NaphH)]. ¹³C NMR (CDCl₃) δ : 18.40, 24.48, 32.26, 45.47, 55.26, 63.86, 65.89, 105.70, 119.12, 120.18, 120.35, 120.46, 120.66, 125.92, 126.18, 127.49, 128.96, 129.28, 130.76, 132.05, 133.89, 134.51, 150.90, 151.29, 157.82, 162.69, 164.38, and 172.34. MS (MALDI); m/z (%): 3892 (M⁺, 7). Anal. Calcd for: C₂₃₆H₁₉₂O₅₄. C, 72.83; H, 4.97. Found: C, 72.63; H, 5.01. $[\alpha]_D^{25} = +93.0$ (*c* 1.0, CHCl₃).

4.2.6. Third generation dendrimer (14). The title compound was prepared from (12) (100 mg, 144 µmol), acid wedge (4) (0.832 g, 1.44 mmol), DPTS (0.008 g, 0.028 mmol) and DCC (0.296 g, 1.44 mmol) by following the above general procedure. After 6 days, the crude product was column chromatographed (SiO_2) eluting with initially CHCl₃ and then a mixture of EtOAC/CHCl₃ (1:19) to give (14) (0.124 g) as a white solid in 14% yield. Mp 115–120 $^{\circ}$ C. ¹H NMR (CDCl₃) δ : 1.27 (s, 2H, CCH₂), 1.58 (d, 48H, J= 7.0 Hz, $16 \times CH_3$ CH), 3.81 (s, 48H, $16 \times OCH_3$), 3.93 (q, $16H, J = 7.0 Hz, 16 \times CHCH_3), 4.48 (b, 8H, 4 \times CH_2O), and$ 6.96–7.63 [m, 132H: (36H, 12×ArH); (96H, 16×NaphH)]. ¹³C NMR (CDCl₃) δ: 18.40, 29.68, 32.02, 45.45, 55.24, 62.02, 65.06, 105.70, 119.10, 20.40, 120.47, 120.68, 125.94, 126.18, 127.48, 128.96, 130.79, 131.55, 133.87, 134.54, 150.79, 150.92, 151.27, 157.80, 162.60, 164.06, and 172.34. Anal. Calcd for C₃₁₄H₂₅₄O₇₂: C, 72.82; H, 4.94. Found: C, 72.62; H, 5.03. $[\alpha]_D^{25} = +92.0$ (*c* 1.0, CHCl₃).

Acknowledgements

We are grateful to UGC—New Delhi for providing fellowships to S. K. P. and A. R. R. We thank CSIR—New Delhi and IICT—Hyderabad for providing the facilities to work.

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Tetrahedron

Tetrahedron 60 (2004) 10921-10926

Dearomatization of furans via [2,3]-Still–Wittig rearrangement

Patrick A. Caruana and Alison J. Frontier*

Department of Chemistry, University of Rochester, 414 Hutchison Hall, Rochester, NY 14627, USA

Received 18 June 2004; revised 7 September 2004; accepted 9 September 2004

Available online 7 October 2004

Abstract—Furans and benzofurans of type 1 were dearomatized via the [2,3]-Still–Wittig rearrangement. Enol ethers 2 could be isolated or isomerized to the corresponding furans 3. The substitution pattern at the homofuranylic position had a strong influence on reaction behavior. Benzofurans rearranged with the greatest efficiency, and employment of a 3-substituted benzofuran (1; $R' = CH_3$) allowed the creation of a quaternary carbon center.

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1. Introduction

The [2,3]-Wittig rearrangement is a six-electron sigmatropic process driven by the conversion of a carbon anion to an oxygen anion (Eq. 1).¹ The reaction is stereospecific with respect to the anionic carbon (inversion of stereochemistry,² nonbonded interactions in the transition state can lead to highly stereoselective olefin geometry,³ and chirality transfer from the carbon–oxygen stereogenic center to the new carbon–carbon bond is efficient.^{1a} These features confer upon the rearrangement a high level of synthetic utility, which has figured in a number of impressive applications to natural product synthesis.⁴ An aromatic π -system can also participate in the [2,3]-rearrangement, producing a highly unstable intermediate that typically undergoes immediate rearomatization in situ (see Eq. 2).⁴ However, [2,3]-Wittig variants effecting permanent destruction of an aromatic system are more elusive. The Sommelet-Hauser rearrangement of nitrogen and sulfur ylides can dearomatize benzene derivatives,⁶ but no examples of the dearomatization of aromatic heterocycles via [2,3]-sigmatropic rearrangement have been reported. Such a reaction could have the potential to create synthetically useful heterocyclic products with unusual architecture.



Keywords: [2,3]-Sigmatropic rearrangement; Furans; Heterocycles; Dearomatization.



2. Results and discussion

Initial studies arose from our attempts to convert 2substituted furans into 2,3-disubstituted furans, via a [2,3]rearrangement pathway analogous to the reaction shown in Eq. 1. Carbanions were generated from alkylstannanes 1^8 and submitted to Still–Wittig rearrangement conditions (Table 1).^{3a} We expected to isolate the rearranged and rearomatized furyl alcohol **3b**, following spontaneous isomerization of unstable-looking intermediate **2b**. However, **2b** was surprisingly robust, surviving aqueous workup and purification by flash chromatography on silica gel. To our knowledge, intermediates analogous to **2** have never been successfully isolated from the [2,3]-Wittig rearrangement of aromatic heterocycles, and we were inspired to conduct a more detailed examination of this interesting transformation.

The [2,3]-rearrangement of anions derived from a range of stannanes 1 was carried out, and the results are presented in Table 1. Reactions proceeded with varying degrees of efficiency, giving enol ethers 2 in 20–80% yield, along with undesired 2-methoxymethyl furans 4 and 2-hydroxymethyl furans 5. The furan analog of 1f also underwent [2,3]-rearrangement.⁹ Treatment of enol ethers 2a-2c, 2e, and 2f

^{*} Corresponding author. Tel./fax: +1 585 2752568; e-mail: frontier@chem.rochester.edu

^{0040–4020/\$ -} see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.029





Entry	Stannane 1	Reaction conditions ^a	Ratio (2:4:5:3) ^b	Product	Yield ^c
1	SnBu ₃ 1a	<i>n</i> -BuLi, THF, -78 °C then H ₂ O	7:7:0:1	H 2a ^d	20
2	CH ₃ CH ₃ Ib SnBu ₃	<i>n</i> -BuLi, THF, -78 °C then H ₂ O	4:1:0:trace		50
3	O O SnBu ₃	<i>n</i> -BuLi, THF, $-78 \text{ °C} \rightarrow -50 \text{ °C}$ then CD ₃ OD	2:1:0:0		66
4	O SnBu ₃ 1d	n-BuLi, THF, −78 °C, then H ₂ O	4 only	o dd	88
5	SnBu ₃ 1e	<i>n</i> -BuLi, THF, -78 °C, then H ₂ O	1:2:0:2	CH 2e	_
6	SnBu ₃ 1e	<i>n</i> -BuLi, THF, −78 °C then 10% TFA/MeOH	0:1:0:1.5	OH 3e	56
7	SnBu ₃ 1f	MeLi, THF, - 78 °C, then MeOH	2.5:1:0:—	OH 2f	65 ^f

^a 1.0 equiv of alkyllithium. ^b Determined by ¹H NMR.

^c Isolated.

^d Isolated as a 5:1 mixture of 2a:3a; a small amount of rearomatization occurred during purification.

² A single isomer (¹H and ¹³C NMR); geometry not known.

^f See Ref. 11.

with 10% trifluoroacetic acid in methanol or THF effected rearomatization to give furyl alcohols 3^{10}

Substitution at the homofuranylic position of the system retarded rearrangement. Substrates with a primary or secondary carbon at this position rearranged at -78 °C (entries 2 and 3). Cyclohexyl derivative 1c (tertiary homofuranylic carbon) required warming to -50 °C to achieve best results, and 1d (quaternary carbon) failed to rearrange under any conditions.

Benzofuran stannane 1e underwent smooth [2,3]-rearrangement, but partial rearomatization to 3e could not be avoided (entry 5). As **2e** and **3e** proved difficult to separate, a one-pot rearrangement/isomerization process was developed to convert 1e directly into 3e (entry 6). Finally, rearrangement of stannane **1f** proceeded without incident to give enol ether **2f**, with efficient formation of a new quaternary center.¹¹ It is significant that this version of the rearrangement allows the creation of intermediates that are unable to rearomatize.

Methyl ethers 4, resulting from protodestannylation, were often present in significant quantities.¹² To determine whether a proton transfer event was competing with the [2,3]-sigmatropic rearrangement, the reaction mixture was quenched with CD₃OD and D₂O. ¹H NMR analysis of methyl ethers 4c and 4d showed a mixture of -OCH₃ and -OCH₂D peaks, indicating that some extent of inter- or

intramolecular proton transfer does occur prior to the external quench.¹³ Alcohol **5** is presumably the result of an α -elimination pathway, as similar processes have been observed by others.¹⁴ However, reaction mixtures never contained significant amounts of products resulting from the (potentially competitive) [1,2]-Wittig rearrangement pathway.^{5a}

Product ratios were largely unaffected by changes in reaction temperature (identical results for -90 and -78 °C) or structure of alkyllithium (*n*-BuLi, *s*-BuLi, *t*-BuLi, MeLi). Tetrahydrofuran gave better results than ether or dimethoxyethane, solvents reported to effect slower and faster Sn-Li transmetallation, respectively.¹⁵ We sought to increase the reactivity of the carbanion by conventional means, but addition of lithium solvators (HMPA and TMEDA), and counterion exchange $(Li \rightarrow K)$ had no effect. Lithium salts (LiCl¹⁶ and LiClO₄) were added to prevent postulated chelation of the organolithium to the furan oxygen, a situation that would create unfavorable geometry for rearrangement. Unfortunately, these salts seemed to interfere with transmetallation, and little rearrangement was observed. To our frustration, basic substrate architecture seemed to influence the rearrangement efficiency of furans **1a–1f** more profoundly than any of the reaction conditions we examined.

A set of optimization experiments on benzofuran **1e** was more fruitful (Table 2). Excess alkyllithium led to higher yields of rearranged product **3**,¹⁷ and methyllithium gave cleaner product mixtures, simplifying purification. Alternatives to tin–lithium exchange for generating the intermediate carbanion were also explored. Treatment of the TMS methyl ether analog **1i**¹⁸ with *n*-BuLi at low temperature led to the unexpected formation of 2-hydroxy-*trans*-cinnamaldehyde in 22% yield (entry 7).¹⁹ Isolation of this product provides additional evidence for the undesired transfer of a proton from the furanylic position. Finally, attempts to generate the desired anion via reductive desulfurization of the phenylsulfide derivative²⁰ led to decomposition (entry 8).

Surprisingly, the heterocyclic substrates shown in Figure 1 did not undergo [2,3]-rearrangement under a variety of

Table 2. Optimization of benzofuran rearrangement



Entry	Х	Reaction conditions ^a	Yield (3)
1	SnBu ₃	<i>n</i> -BuLi (1.1 equiv)	56
2	SnBu ₃	n-BuLi (3.0 equiv)	64
3	SnBu ₃	n-BuLi (5.0 equiv)	77
4	SnBu ₃	MeLi (1.0 equiv)	25
5	SnBu ₃	MeLi (2.0 equiv)	80
6	SnBu ₃	MeLi (3.0 equiv)	68
7	SiMe ₃	<i>n</i> -BuLi (5.0 equiv)	b
8	SPh	LiNp ^c	0

^a Reactions were run in THF at -78 °C.

^b Product was 2-hydroxy-trans-cinnamaldehyde.

^c Lithium naphthalenide.



Figure 1. Substrates that did not undergo rearrangement.

conditions. Methyl ether **4** was the major product observed when rearrangement was attempted.

3. Conclusion

In summary, we have accomplished the dearomatization of furan and benzofuran substrates via the [2,3]-Still–Wittig rearrangement with varying degrees of efficiency. Many of the enol ether products could be isolated by flash chromatography or subjected to acidic conditions to give 2,3-disubstituted furans **3**. It was also possible to form quaternary centers in this process, to give products with no available rearomatization pathway. Although the substrate scope of the rearrangement was somewhat disappointing, the rearrangement showed promising synthetic utility for benzofuran derivatives.

4. Experimental

Reactions were done in oven-dried glassware under an argon atmosphere unless otherwise noted. Diethyl ether and tetrahydrofuran (THF) were freshly distilled from sodium/ benzophenone under nitrogen. Dichloromethane and 1,2 dimethoxyethane were freshly distilled over calcium hydride. Tetramethylethylenediamine (TMEDA) and hexa-methylphosphoramide (HMPA) were also freshly distilled over calcium hydride. Methyl lithium, *n*-butyl lithium, *s*-butyl lithium, and *t*-butyl lithium were titrated by dropwise addition to diphenylacetic acid (200 mg in 5 mL THF) under argon. Furfural was distilled under vacuum and stored in the refrigerator. Lithium chloride was stored in a nitrogen-filled glove box before use, and lithium perchlorate was dried at 60 °C for 2 h.

Column chromatography was performed on EM Science silica gel 60 (230–400) mesh. Visualization was done with *p*-anis aldehyde stain and a UV lamp. Proton and carbon NMR spectra were collected on 400 MHz spectrometers at ambient temperature. Chemical shifts δ are reported relative to CDCl₃. High-resolution mass spectra were measured by the Chemistry Instrumentation Center of the Department of Chemistry at the University of Buffalo.

2-Hydroxymethyl furans $1e^{21}$, $1f^{22}$, $1g^{23}$ (for R = H; R = Me comes from entry 6), and $1h^{24}$ were prepared according to literature methods. ICH₂SnBu₃ was also prepared according to a literature procedure.²⁵

4.1. Preparation of 2-hydroxymethyl furan

Sodium borohydride (0.39 g, 10.4 mmol) was added to a solution of furfural (1.0 g, 10.4 mmol) in methanol at 0 °C. After 30 min the reaction was concentrated and dissolved in

ether. The organic layer was washed with water $(1 \times)$ and the aqueous layer was then washed with ether $(2 \times)$. The combined organic extracts were washed with brine, dried over anhydrous magnesium sulfate, and concentrated to give a clear, faint yellow oil in essentially quantitative yield. Spectroscopic data were consistent with those previously reported in the literature (see www.aldrich.com).

4.2. General procedure for the synthesis of furanyl alcohols (precursors to 1b, 1c, 1d)

A solution of furfural (1.0 equiv) in diethyl ether (0.33 M) was cooled to -30 °C. The corresponding Grignard reagent (1.0 equiv) was added and TLC was used to judge completion. The reaction was then quenched with saturated ammonium chloride and warmed to room temperature. After diluting with ether, the organic layer was washed with water and brine. The organic phase was then dried with anhydrous magnesium sulfate and concentrated. Silica gel chromatography (4:1 hexane:ethyl acetate) afforded light yellow oils in essentially quantitative yield. Spectroscopic data were consistent with those previously reported in the literature: **1b**;²⁶ **1c**;²⁷ **1d**.²⁸

4.3. General procedure for the synthesis of stannanes 1a-1h

KH (30% dispersion in mineral oil, 2.0 equiv) was washed with hexane $(2 \times)$ under an argon atmosphere by magnetic stirring and subsequent removal of the hexane via syringe. THF (0.16 mmol/1 mL) was then added and the suspension was cooled to 0 °C. 2-Furan hydroxy methyl compounds (1.0 equiv) were then added dropwise in THF (1.7 mmol/ 1 mL), and the reaction was allowed to warm to room temperature for 2 h. The resulting mixture was cooled again to 0 °C and ICH₂SnBu₃ in THF (3.4 mmol/1 mL) was added dropwise. The mixture was then allowed to warm to room temperature and stirred overnight. Saturated ammonium chloride was used to quench the reaction, followed by dilution with ether. The layers were separated and the aqueous phase was washed one more time with ether. The combined organic extracts were dried with anhydrous magnesium sulfate and concentrated. Silica gel chromatography (100% hexane or hexane \rightarrow hexane/ethyl acetate) afforded light yellow oils in 50-70% yield. Stannyl methyl ether products were identified by ¹H NMR.²⁹

4.3.1. Preparation of 3-hydroxymethyl 2-methyl furan (**3a**). Stannane **1a** (0.35 g, 0.77 mmol) was dissolved in THF (5 mL) and cooled to -78 °C. *n*-BuLi (1.6 M in hexanes, 0.48 mL, 0.77 mmol) was then added dropwise and the reaction was stirred for 20 min. H₂O (0.3 mL) was then added and the solution was allowed to warm to room temperature. The reaction mixture was then diluted with 70 mL ether, dried with anhydrous magnesium sulfate, and concentrated to a crude oil. Silica gel chromatography (92:8 dichloromethane:diethyl ether) afforded 13 mg of a light yellow oil **2a**:**3a**=5:1 as assigned by ¹H NMR (22%). (**2a**) ¹H NMR (400 MHz, CDCl₃) δ 6.53 (m, 1H), 5.19 (s, 1H), 4.68 (s, 1H), 4.27 (s, 1H), 3.66 (m, 3H), 1.76 (br. s, 1H).

Treatment of the product with 10% HCl (aq) or cat. AcOH

in CDCl₃ gave furan $3a^{5c}$ after aqueous workup and silica gel column purification.

4.3.2. Preparation of 3-hydroxymethyl 2-ethyl furan (3b). Stannane 1b (0.37 g, 0.78 mmol) was dissolved in THF (5.0 mL) and cooled to -78 °C. *n*-BuLi (1.6 M in hexanes, 0.49 mL, 0.78 mmol) was then added dropwise and the reaction was stirred for 30 min. H₂O (0.5 mL) was then added and the solution was allowed to warm to room temperature. The reaction mixture was then diluted with 70 mL ether, dried with anhydrous magnesium sulfate, and concentrated to a crude oil. Silica gel chromatography (dichloromethane \rightarrow 92:8 dichloromethane:diethyl ether) afforded 56 mg (57%) of **2b** as a light yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 6.53 (m, 1H), 5.22–5.17 (m, 2H), 3.76-3.70 (m, 3H), 1.68 (dd, J=7.2, 1.7 Hz, 3H). Treatment of the product with 10% HCl (aq) in CDCl₃ gave furan $3b^{30}$ after aqueous workup and silica gel column purification. ¹H NMR (400 MHz, CDCl₃) δ 7.27 (d, J= 1.8 Hz), 6.37 (d, J=1.7 Hz), 4.49 (s, 2H), 2.66 (q, J=7.6 Hz), 1.29 (br. s, 1H), 1.22 (t, J = 7.6 Hz).

4.3.3. Enol ether (2c). Stannane **1c** (0.095 g, 0.18 mmol) was dissolved in THF (1.2 mL) and cooled to -78 °C. *n*-BuLi (1.6 M in hexanes, 0.11 mL, 0.18 mmol) was then added dropwise and the reaction was stirred for 20 min at -78 °C. After warming to -50 °C for 10 min, 0.3 mL CD₃OD was added and the solution was allowed to warm to room temperature. The reaction mixture was then diluted with ether and washed with water $(1 \times)$. The aqueous layer was washed with ether $(1 \times)$ and the combined ether extracts were washed with brine $(1 \times)$. The organic phase was then dried over anhydrous magnesium sulfate and concentrated. Silica gel chromatography (dichloromethane) afforded 22 mg (65%) of 2c as a colorless oil. IR (thin film) 3366, 2922, 2850, 1694, 1615, 1448, 1144 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.53 (m, 1H), 5.18 (m, 1H), 5.05 (d, J=9.4 Hz, 1H), 3.77–3.60 (m, 3H), 2.01 (m, 1H), 1.59–1.57 (m, 5H), 1.28–0.88 (m, 5); 13 C NMR (75 MHz, CDCl₃) δ 153.5, 145.4, 109.6, 103.4, 64.8, 45.6, 36.3, 34.7, 33.8, 25.9; HRMS (EI) m/z 194.13069 [(M⁺); Calcd for C₁₂H₁₈O₂ 194.1301].

4.3.4. Enol ether (2e). Stannane 1f (0.18 g, 0.35 mmol) was dissolved in THF (2.8 mL) and cooled to -78 °C. *n*-BuLi (1.6 M in hexanes, 0.24 mL, 0.35 mmol) was then added dropwise and the reaction was stirred for 20 min. H₂O (0.5 mL) was then added and the solution was allowed to warm to room temperature. The reaction mixture was then diluted with ether and washed with water $(1 \times)$. The aqueous layer was washed with ether $(1 \times)$ and the combined ether extracts were washed with brine $(1 \times)$. The organic phase was then dried over anhydrous magnesium sulfate and concentrated. Silica gel chromatography (two columns were run: 80:20 dichloromethane:ether, then 9:1 dichloromethane:ether) afforded 9 mg of a pure sample of 2e as a colorless oil. IR (thin film) 3370, 2929, 2875, 1684, 1609, 1477, 1461, 1367, 1331, 1155, 936, 827, 750, cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.28 (d, J= 7.3 Hz, 1H), 7.22 (t, J=7.6 Hz, 1H), 6.99 (t, J=7.5 Hz, 1H), 6.91 (d, J = 8 Hz), 4.81 (d, J = 2.3 Hz, 1H), 4.38 (d, J =2.2 Hz, 1H), 4.11 (br. s, 1H), 3.87 (m, 2H), 1.77 (t, J =

6.4 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 162.3, 157.8, 128.8, 126.6, 124.3, 121.9, 109.4, 85.7, 65.7, 46.8.

4.3.5. 2-Methyl-3-hydroxymethyl benzofuran (3e). Stannane **1e** (0.18 g, 0.35 mmol) was dissolved in THF (2.8 mL) and cooled to -78 °C. MeLi (1.6 M in diethyl ether, 0.47 mL, 0.70 mmol) was then added dropwise and the reaction was stirred for 30 min. MeOH (0.4 mL), followed by 0.03 mL TFA was then added and the solution was allowed to warm to room temperature. The reaction mixture was then diluted with ether and washed with water (1×). The aqueous layer was washed with ether (1×), and the combined ether extracts were washed with saturated sodium bicarbonate (1×) and brine (1×). The organic phase was then dried over anhydrous magnesium sulfate and concentrated. Silica gel chromatography (dichloromethane \rightarrow 9:1 dichloromethane:ether) afforded 47 mg (80%) of **3e**³¹ as a white solid.

4.3.6. Enol ether (2f). Stannane 1f (3.65 g, 7.00 mmol) was dissolved in THF (46 mL) and cooled to -78 °C. MeLi (1.6 M in diethyl ether, 13.0 mL, 21.0 mmol) was then added dropwise and the reaction was stirred for 20 min at -78 °C. MeOH (6.0 mL) was then added and the solution was allowed to warm to room temperature. The reaction mixture was then diluted with ether and washed with water $(1 \times)$. The aqueous layer was washed with ether $(1 \times)$ and the combined ether extracts were washed with brine $(1 \times)$. The organic phase was then dried over anhydrous magnesium sulfate and concentrated. Silica gel chromatography (4:1 hexane:ethylacetate) afforded 0.984 (80%) of 2f as a faint yellow oil. IR (thin film) 3390, 2967, 2928, 2868, 1684, 1609, 1476, 1462, 1240, 1172, 1105, 1040, 936, 827, 750 cm^{-1} ; ¹H NMR (400 MHz, CDCl₃) δ 7.26–7.18 (m, 2H), 7.01 (t, J=7.3 Hz, 1H), 6.94 (d, J=8.0 Hz, 1H), 4.79 (d, J=2.7 Hz, 1H), 4.27 (d, J=2.7 Hz, 1H), 3.69-3.59 (m,2H), 1.67 (t, J=7.0 Hz, 1H), 1.45 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 167.8, 156.8, 131.3, 128.6, 123.0, 122.0, 109.4, 84.0, 70.5, 49.7, 23.3; HRMS (EI) m/z 176.08308 [(M^+); Calcd for C₁₁H₁₂O₂ 176.0832].

General notes. Enol ether 2c also isomerized to the corresponding furan 3c under similar acidic conditions in moderate yield. All methyl ethers 4 were identified according to ¹H NMR (characteristic signal at 3.2 ppm (s, 3H)).

Acknowledgements

We thank the University of Rochester for support of this research. We are grateful to Dr. Alice Bergmann (director of the Chemistry Instrumentation Center at the University of Buffalo) for performing HRMS analysis.

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Tetrahedron

Tetrahedron 60 (2004) 10927-10941

Conformational analysis of six-membered ring dioxaphosphinanes. Part 1: Anancomeric thiophosphates

Javier Hernández,[†] Rafael Ramos,[‡] Noé Sastre, Rocío Meza,[§] Herbert Hommer,[¶] Magali Salas and Barbara Gordillo^{*}

Departamento de Química, Centro de Investigación y de Estudios Avanzados del Instituto Politécnico Nacional, Apartado Postal 14-740, Mexico, D F., Mexico

Received 27 March 2004; revised 2 September 2004; accepted 9 September 2004

Available online 1 October 2004

Abstract—A study of the conformation of a series of anancomeric axial and equatorial 2-aryloxy-2-thio-1,3,2 λ^5 -dioxaphosphinanes 2–12 in solution and solid state is reported. In accord to the stereoelectronic theory, aryl thiophosphates substituted with electron-withdrawing (EW) groups will tend to occupy axial positions in chair ring conformations due to the stabilizing *endo*-anomeric ($n_{\pi}O-\sigma_{P-X}^{-}$) hyperconjugative interaction. The antiperiplanar orientation of the orbitals involved in the stereoelectronic interaction is a requirement that is fulfiled in the axial series of compounds when the ring adopts a chair conformation. Therefore, in the equatorial series of thiophosphates, the axial seeking characteristics of aryloxy-EW groups might render the molecule with distortion of the chair conformation. An opposite trend is anticipated for the less axial seeking aryl thiophosphates substituted with electron releasing (ER) groups. A detailed analysis of the ${}^{3}J_{HH}$, ${}^{3}J_{PH}$ and ${}^{3}J_{CP}$ coupling constants allowed us to conclude that there is no contribution of high energy twist-boat conformations in the equatorial thiophosphates in both configurations. X-ray geometrical analysis of bond distances and bond angles supports clearly the participation of hyperconjugative *endo*-anomeric ($n_{\pi}O-\sigma_{P-OAr}^{-}$) effect in the stabilization of axial series of compounds and the participation of *endo*-anomeric ($n_{\pi}O-\sigma_{P-OAr}^{-}$) effect in the stabilization of axial series of compounds and the participation of the equatorial thiophosphates in chair conformation of the equatorial thiophosphates in chair conformations. \mathbb{C} 2004 Published by Elsevier Ltd.

1. Introduction

Six-membered ring thiophosphates are interesting systems from the conformational point of view since there are several electronic effects, as the *endo-* and *exo*-anomeric effects, that can play an important role in the conformational preferences of the substituents on phosphorus.^{1–3} Steric effects in thiophosphates are not very important as in the monosubstituted cyclohexanes⁴ or as in the closest heterocyclic system, the 2-substituted-1,3-dioxanes.⁵ The decreased 1,3-*syn* axial steric hindrance of the substituent with the 4,6-hydrogens of the 1,3,2-dioxaphosphinane ring

0040–4020/\$ - see front matter @ 2004 Published by Elsevier Ltd. doi:10.1016/j.tet.2004.09.025

is due to the longer P–O (1.58 Å) *endo*-cyclic bond distances as compared to C–O in 1,3-dioxanes (1.42 Å) or C–C in cyclohexane (1.54 Å),⁶ as well as the propensity of the *endo*-cyclic oxygens to adopt sp² hybridization instead of sp³ that renders the ring with more flexibility than the corresponding 1,3-dioxane or cyclohexane.⁷ As observed in the pentacoordinative chemistry of phosphorus compounds,⁸ electronegative substituents in phosphates and thiophosphates will tend to adopt axial orientations, and electron-donating substituents, equatorial ones (Scheme 1).⁹

$$\sum_{O} \sum_{O} \sum_{O$$

X = electronegative atom or group



Y = electron releasing atom or group

Scheme 1.

Keywords: Dioxaphosphinanes; Thiophosphates; Conformation; NMR; LFER correlations; X-ray crystal structures.

^{*} Corresponding author. Tel.: +525 5061 3729; fax: +525 5747 7113; e-mail: ggordill@cinvestav.mx

[†] Present address: Centro de Investigación en Alimentación y Desarrollo A. C. Hermosillo Sonora, Mexico.

^{*} Present address: Unidad de Servicios de Apoyo en Resolución Analítica Universidad Veracruzana, Xalapa Veracruz, Mexico.

[§] Present address: Departamento Ciencias Básicas Ingeniería y Tecnología Universidad Autónoma de Tlaxcala, Tlaxcala, Mexico.

[¶] Present address: Degussa Construction Polymers GmbH, D-83308 Trostberg, Germany.



Scheme 2.

These preferences rely on stereoelectronic grounds, the *endo*-anomeric $(n_{\pi}O-\sigma_{P-X}^*)$ effect favoring the axial conformer when the X-substituent is electronegative (such as halogen, OR, etc.), because the acceptor σ^* antibonding orbital of an electronegative substituent is lower-lying than the antibonding orbital of an electron-releasing substituent, therefore giving rise to a more stabilizing 2 electrons/2 orbitals interaction (Scheme 2a).^{3,10} On the contrary, the



Scheme 3.



Scheme 4.

exo-anomeric $(n_{\pi}Y-\sigma_{P-O\ endo-cyclic}^{*})$ hyperconjugative interaction will promote that an electron releasing substituent, such as an amine, tend to occupy the equatorial position in a dioxaphosphinane ring (Scheme 2b).⁷ The equatorial isomer might also be favored by the $n_{\pi}O-\sigma_{P=S}^{*}$ endo-hyperconjugative intraction.¹¹

From experimental point of view, van Nuffel et al.¹² have proved the participation of stereoelectronic interactions in the axial conformational preference of several phosphates and their thio derivatives, through the shortening of the *endo*-cyclic P–O bond and the lengthening of the P–X bond that is based on the resonance hybrids shown in Scheme 2. It is worth of mention that since the interconversion barrier of chair-to-twist or chair-to-boat conformations in 1,3,2dioxaphosphinanes is very low (0.5–3.5 kcal/mol),¹ the stabilizing *endo*- or *exo*-anomeric interactions and/or the steric hindrance when the substituents are bulky, can force the molecule to adopt a twist-boat conformation in preference to the chair (Scheme 3).⁷

Reported in this work is the conformational analysis in solution and solid state of a series of axial and equatorial anancomeric aryl thiophosphates substituted with electronwithdrawing (EW) and electron-releasing (ER) groups (2–12), (Scheme 4). Spectroscopic data, as ¹H, ¹³C and ³¹P NMR are correlated with Hammett constants (σ_p), ¹³ within the context of LFER theory.¹⁴ We analyzed thoroughly the X-ray geometrical parameters of two axial and two equatorial compounds substituted with EW or ER groups to account for the participation of the *endo*-anomeric ($n_{\pi}O-\sigma_{P-OAr}^{*}$) and ($n_{\pi}O-\sigma_{P=S}^{*}$) stabilizing interactions.

2. Results

The synthesis of both series of aryl thiophosphates 2-12-ax and 2-12-eq was accomplished through the stereoselective formation of the phosphite intermediates from the phosphochloridite and aryl alcohols as shown in Scheme 5. Due to the epimerization of the equatorial phosphite intermediate in the presence of an excess of phenol,¹⁵ in route A, the *p*-X-substituted phenol was slowly added to the phosphorochloridite (1). On the contrary, in route B the thermodynamically more stable axial phosphite was obtained by addition of 1 to the corresponding *p*-X-substituted phenol.



 $\mathsf{X}=\mathsf{NO}_2,\,\mathsf{CN},\,\mathsf{CHO},\,\mathsf{Br},\,\mathsf{CI},\,\mathsf{NHCOCH}_3,\,\mathsf{H},\,\mathsf{C}_6\mathsf{H}_5,\,\mathsf{CH}_3,\,\mathsf{OCH}_3,\,\mathsf{NH}_2$

Chemical shifts (δ)			$^{1}\mathrm{H}$		³¹ P
Compound	H _{4,6a}	H _{5a}	H _{5e}	H _{7,8}	
$2-ax (X=NO_2)$	4.76	1.86	1.90	1.42	54.3
3-ax (X = CN)	4.73	1.85	1.87	1.43	54.5
4-ax (X=CHO)	4.75	1.83	1.85	1.42	55.1
5-ax (X=Br)	4.75	1.82	1.87	1.42	55.5
6-ax (X=Cl)	4.75	1.83	1.90	1.45	55.6
7-ax (X = NHCOCH ₃)	4.77	1.86	1.88	1.44	55.9
8- ax (X=H)	4.79	1.82	1.88	1.43	55.8
9-ax (X = C_6H_5)	4.81	1.83	1.88	1.44	55.8
10 - ax (X = CH ₃)	4.77	1.82	1.84	1.41	56.2
11- ax (X=OCH ₃)	4.77	1.81	1.86	1.42	56.7
$12-ax (X = NH_2)$	a	a	а	a	56.8

Table 1. Selected ¹H and ³¹P NMR chemical shifts (in ppm) for axial aryl thiophosphates 2–12 in CDCl₃

^a Undetermined (mixed with the equatorial epimer).

Table 2. Selected ¹H and ³¹P NMR chemical shifts (in ppm) for equatorial aryl thiophosphates 2–12 in CDCl₃

Chemical shifts (δ)		¹ H					
Compound	H _{4,6a}	H _{5a}	H _{5e}	H _{7,8}			
$2-eq (X=NO_2)$	4.85	1.77	1.90	1.41	59.2		
3-eq (X=CN)	4.84	1.79	1.87	1.41	59.3		
4-eq (X=CHO)	4.79	1.54	1.69	1.34	59.5		
5 - eq (X=Br)	4.76	1.64	1.77	1.32	60.0		
6-eq (X=Cl)	4.83	1.71	1.87	1.39	60.1		
7 - eq (X=NHCOCH ₃)	4.80	1.75	1.85	1.38	60.4		
8-eq $(X=H)$	4.83	1.70	1.89	1.39	60.4		
9 -eq (X = C_6H_5)	4.78	1.67	1.77	1.34	60.4		
10 -eq (X = CH_3)	4.80	1.59	1.72	1.34	60.9		
11-eq $(X = OCH_3)$	4.73	1.63	1.75	1.31	61.0		
12 - eq (X = NH ₂)	4.52	1.68	1.82	1.38	61.3		

Table 3. ¹H NMR backbone coupling constants (in Hz) for axial aryl thiophosphates 2–12. First-order analysis in CDCl₃ at 27 °C

Compound/coupling constant	${}^{3}J_{\rm H4,6aH5a}$	${}^{3}J_{\rm H4,6aH5e}$	${}^{3}J_{\rm H4,6aH7,8}$	$^{2}J_{\mathrm{H5aH5e}}$	${}^{3}J_{\mathrm{H4,6aP}}$	$^{4}J_{ m H5aP}$	${}^{4}J_{\mathrm{H5eP}}$	$^4J_{ m H7,8P}$
$2-ax (X = NO_2)$	11.3	3.0	6.2	14.6	2.6	1.0	2.6	2.3
3-ax (X=CN)	10.5	2.5	6.4	14.8	1.2	1.5	1.3	2.5
4-ax (X=CHO)	8.0	2.7	6.2	13.0	1.2	а	1.5	2.2
5-ax (X=Br)	10.8	2.8	6.2	14.8	1.2	1.0	2.7	2.2
6-ax (X=Cl)	11.1	3.0	6.6	14.1	1.1	1.1	1.3	2.1
7 - ax (X = NHCOCH ₃)	12.9	2.9	6.2	13.0	1.3	1.0	1.8	2.3
8- ax (X=H)	10.8	2.8	6.6	14.4	1.3	1.1	2.6	2.6
9 - ax (X = C ₆ H ₅)	9.9	2.9	6.2	13.8	1.2	1.0	2.9	2.2
10 - ax (X = CH ₃)	10.7	3.0	6.2	14.5	1.2	1.1	2.7	2.2
11 - ax (X=OCH ₃)	10.8	3.3	6.2	14.5	1.2	1.0	3.0	2.2
12 - ax (X = NH ₂)	b	b	b	b	b	b	b	b

^a The signal was not observed.

^b Undetermined (mixed with its equatorial epimer).

The thermal equilibration of equatorial phosphites to the axial epimers was achieved by heating them in toluene (route C). The equatorial phosphite intermediates substituted with EW groups tend to epimerize to the axial ones in shorter heating times than those substituted with ER groups. The equatorial or axial phosphites were reacted stereospecifically with sulfur to produce the thiophosphates 2-12-eq or 2-12-ax under the conditions shown in Scheme 5. We observed that equatorial aryl thiophosphates were obtained in a cleaner manner, without their configurational isomers, if the reaction was maintained under vigorously toluene reflux, particularly in the case of aryl-EW thiophosphates.

The conformational analysis of the axial and equatorial

series of thiophosphates (2–12) was assessed by analysis of their spectral NMR characteristics. The chemical shifts (δ) of ¹H are reported in Tables 1 and 2. The correct assignment of the signals and the backbone coupling constants in the ¹H NMR (Tables 3 and 4) were obtained through the first-order analysis of the spectra, along with homo and heteronuclear decoupling experiments. The complete assignment of ¹³C signals (Tables 5 and 6), was achieved by means of ¹H, ¹³C correlated 2D NMR spectra, as well as inverse ¹H-detected HMQC and HMBC experiments. Two-dimensional techniques were particularly useful for the assignment of ¹³C NMR signals of the configurational *p*-phenyl thiophosphates (9-*ax* and 9-*eq*). The participation of the stereoelectronic interactions was analyzed in the solid state through

Table 4.	H NMR backbone	coupling constants	(in Hz) fo	or equatorial ar	yl thiophosphates	2–12. Fi	irst-order analy	ysis in CI	DCl ₃ at 27 °	С
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Compound/coupling constant	${}^{3}J_{\rm H4,6aH5a}$	${}^{3}J_{\rm H4,6aH5e}$	³ J _{H4,6aH7,8}	$^{2}J_{\rm H5aH5e}$	${}^{3}J_{\mathrm{H4,6aP}}$	${}^{4}J_{\mathrm{H5aP}}$	${}^{4}J_{\rm H5eP}$	$^4J_{ m H7,8P}$
2 - eq (X=NO ₂)	10.5	2.6	6.3	14.6	1.2	1.0	2.8	2.2
3-eq(X=CN)	11.2	2.7	6.6	14.5	3.3	a	3.6	2.0
4-eq (X=CHO)	10.4	2.7	6.3	14.5	2.8	1.8	2.3	2.3
5-eq(X=Br)	10.6	2.8	5.9	14.5	2.6	а	2.3	2.0
6-eq (X=Cl)	11.2	3.0	6.6	14.0	2.5	а	2.3	2.0
7-eq (X=NHCOCH ₃)	10.8	2.3	6.2	14.1	2.6	а	2.3	2.4
8-eq (X=H)	10.9	2.6	6.3	14.5	2.3	a	2.3	2.3
9-eq $(X = C_6 H_5)$	9.9	2.9	6.2	14.2	3.0	a	2.3	2.0
$10-eq (X = CH_3)$	11.2	2.6	6.6	14.5	2.6	2.3	2.6	2.6
11-eq $(X = OCH_3)$	10.9	2.6	6.3	14.5	2.6	a	2.6	2.0
$12-eq (X=NH_2)$	11.2	2.6	6.6	14.5	3.3	а	2.6	2.0

^a The signal was not observed.

Table 5. Selected ¹³C NMR signal assignments in axial aryl thiophosphates 2–12^a

Compound	C _{4,6}	C ₅	C _{7,8}	C _i	C_o
2 - ax (X=NO ₂)	77.7 (9.1)	40.6 (4.9)	22.4 (9.6)	156.2 (6.1)	121.3 (5.4)
3-ax (X=CN)	77.1 (9.3)	40.3 (5.2)	22.3 (9.9)	154.3 (6.2)	121.3 (5.2)
4-ax (X = CHO)	76.7 (8.9)	40.2 (5.6)	22.1 (10.0)	155.4 (5.5)	120.7 (5.5)
5- ax (X = Br)	77.2 (8.9)	40.8 (4.8)	22.4 (9.6)	150.6 (6.4)	122.6 (4.9)
6-ax (X = Cl)	76.7 (10.4)	40.6 (4.9)	22.6 (9.5)	149.5 (6.3)	121.7 (5.2)
7 - ax (X = NHCOCH ₃)	76.8 (9.5)	42.7 (5.0)	22.5 (9.8)	147.5 (6.2)	121.9 (5.0)
8- ax (X=H)	76.2 (8.8)	39.9 (4.4)	21.9 (9.9)	150.5 (7.0)	119.8 (5.5)
9 - ax (X = C ₆ H ₅)	77.1 (8.9)	40.7 (4.7)	22.5 (9.6)	150.8 (6.5)	121.0 (5.0)
10 - ax (X=CH ₃)	76.9 (8.9)	40.7 (4.6)	22.4 (9.9)	148.8 (6.2)	120.4 (4.8)
11- ax (X=OCH ₃)	76.9 (8.9)	40.7 (4.8)	22.4 (9.5)	144.9 (7.2)	121.7 (4.8)
12 - ax (X=NH ₂)	76.2 (9.3)	40.5 (4.7)	22.3 (9.8)	143.8 (7.8)	121.1 (5.2)

^a Chemical shifts (δ) in ppm from TMS in CDCl₃. In parentheses J_{CP} in Hz.

Table 6. Selected ¹³C NMR signal assignments in equatorial aryl thiophosphates 2–12^a

Compound	C _{4,6}	C ₅	C _{7,8}	C _i	C_o	
$2-eq (X=NO_2)$	76.1 (5.5)	40.8 (5.5)	22.1 (8.9)	155.0 (7.7)	122.1 (4.4)	
3-eq(X=CN)	76.0 (5.5)	40.9 (6.6)	22.1 (8.9)	152.6 (6.6)	122.4 (4.4)	
4-eq (X=CHO)	75.9 (5.4)	40.8 (5.8)	22.1 (10.0)	155.0 (6.0)	122.0 (4.4)	
5- eq (X=Br)	75.6 (5.5)	40.9 (5.5)	22.0 (10.0)	149.5 (8.9)	123.1 (4.5)	
6-eq (X=Cl)	76.1 (5.7)	41.3 (5.1)	22.5 (9.5)	149.4 (8.0)	123.1 (5.0)	
7- eq (X=NHCOCH ₃)	75.8 (5.5)	41.1 (5.5)	22.4 (9.5)	149.6 (8.1)	121.7 (4.6)	
8-eq $(X=H)$	75.4 (5.5)	41.0 (4.4)	22.1 (10.0)	150.3 (7.7)	121.1 (4.4)	
9 -eq (X = C_6H_5)	75.5 (5.5)	41.0 (5.5)	22.1 (10.0)	149.9 (7.7)	121.5 (5.5)	
10 - <i>eq</i> (X = CH_3)	75.3 (5.5)	41.0 (5.5)	22.1 (10.0)	148.3 (7.7)	121.0 (5.5)	
11-eq $(X = OCH_3)$	75.6 (5.5)	40.9 (5.6)	22.0 (10.0)	144.0 (7.7)	122.0 (4.4)	
12 - eq (X=NH ₂)	75.8 (5.5)	40.8 (5.5)	21.9 (9.9)	142.6 (8.8)	121.8 (4.4)	

^a Chemical shifts (δ) in ppm from TMS in CDCl₃. In parentheses J_{CP} in Hz.

the X-ray diffraction analysis of four thiophosphates, two of the axial series $[2-ax (X=NO_2), and 10-ax (X=CH_3)]$ and two of the equatorial $[2-eq (X=NO_2) and 11-eq (X=OCH_3)]$. These molecules are examples of aryl thiophosphates with EW or ER groups in both configurations.

3. Discussion

3.1. NMR analysis

As observed in Tables 1 and 2, in the axial series of thiophosphates (2–12), the proton H_{5a} is slightly upfield shifted (ca. 0.05 ppm) than the H_{5e} , this trend is also observed for the equatorial epimers (2–12). The chemical shift of protons $H_{4,6a}$ for the axial thiophosphates is at around 4.75 ppm meanwhile the same protons in the

equatorial series appear at around 4.85 ppm. ¹³C NMR chemical shifts of carbons in the heterocycle are scarcely sensitive to configuration, however a slight downfield shift (1.3 ppm in average) is observed for $C_{4,6}\xspace$ for the axial compounds, thus not giving evidence of a γ -gauche effect (see discussion below). The coupling constants ${}^{2}J_{CP}$ of C_{4,6} for the axial thiophosphates are 3-5 Hz larger than the equatorial ones (Tables 5 and 6). The conformational analysis of the complete series of thiophosphates was performed with the coupling constants ${}^{3}J_{\text{HH}}$, ${}^{3}J_{\text{HP}}$ and ${}^{3}J_{\text{CP}}$ (Tables 3–6). Heteronuclear ${}^{1}\text{H}\{{}^{31}\text{P}\}$ decoupling experiments led to simplification of the signals, therefore vicinal ${}^{3}J_{\rm HP}$ were easily obtained by direct comparison with the ¹H NMR signals of the undecoupled spectra. For compounds (2-11)-ax the irradiation at 1.41-1.45 ppm (CH₃'s at C_4 , C_6) led to decoupling of the methine ($H_{4,6a}$) multiplet to an apparent doublet of double doublet (ddd) with ${}^{3}J_{\rm HH} = 9.9-11.3$ Hz (*anti*), ${}^{3}J_{\rm HH} = 2.5-3.3$ Hz (*gauche*) and ${}^{3}J_{\rm HP} = 1.2-2.6$ Hz, these coupling constants are consistent with a chair conformation.^{2,16} A similar experiment was performed for the compounds (2-12)-eq, irradiation at 1.31-1.41 ppm led to a double of triplets (dt) for the methine (H_{4a}) with ${}^{3}J_{HH} = 9.9 - 11.2$ Hz (*anti*), ${}^{3}J_{HH} = 2.6 - 3.0$ Hz (*gauche*) and ${}^{3}J_{HP} = 2.3 - 3.6$ Hz, here also values of coupling constants suggest that in solution the series of equatorial thiophosphates are in chair conformation. On the other hand, the ${}^{3}J_{C5P}$ and ${}^{3}J_{C7,8P}$ coupling constants obtained from the ${}^{13}C$ NMR spectra of compounds (2–12) are in the range of 4.4-5.7 and 8.9-10.0 Hz, respectively, for both, axial and equatorial, series of thiophosphates (Tables 5 and 6) suggesting also a chair conformation for the six-membered ring in solution.^{2,17} The fact that the ${}^{3}J_{CP}$ is not showing dependence on configuration, support the argument given by Quin¹⁸ and Nifantiev¹⁹ on the uses of ${}^{3}J_{CP}$ as the most useful C–P coupling constant to analyze the conformation of dioxaphosphinanes (note that in an ideal chair conformation the dioxaphosphinane dihedral angles $\omega_{C5-C4-O3-P}$ and $\omega_{C5-C6-O1-P}$ are 60° meanwhile $\omega_{C8-C6-O1-P}$ and $\omega_{C7-C4-O3-P}$ are 180°).

3.2. Electronic effects

In phosphorus compounds with aromatic rings, it has been demonstrated that EW or ER groups in the *para* position of the ring, causes polarization of charge density inducing changes in chemical shifts and coupling constants on the phosphorus and linked atoms.²⁰ Depending upon the aromatic organophosphorus compound, the so-called substituent-induced chemical shifts (SCS) exhibits in ³¹P NMR one of the two possible tendencies, one that correlates ³¹P chemical shifts directly with Hammett (σ_p) constants and the other that correlates them reversibly.²¹ The factors that determine the tendency are the electronegativity of the α -atoms directly linked to phosphorus and the phosphorus

hybridization,²² and it has been observed frequently that these two factors cannot be separated for to understand the behavior.²³ Thus as expected, an EW group in aryl phosphazanes²⁴ provoke dishielding of the phosphorus nucleus resulting in a downfield shift of ³¹P NMR signal. Nevertheless, the opposite behavior is observed for cinnamyl phosphonates²⁵ and aryl phosphates.^{9a} The fundamental difference between these three types of organophosphorus compounds is that in phosphonates or phosphates the aromatic ring is in a group linked to phosphorus through a single bond; however, in aryl phosphazenes the aromatic ring is in a group linked to phosphorus through a double bond. From this observation it is evident that the transfer of charge density from the aromatic ring to phosphorus or vice versa follows a different mechanism when the transmission is through a σ or through a π bond.²⁶ In this work we found that the ³¹P NMR signals of arylthiophosphates are reversibly correlated with $\sigma_{\rm p}$ being the correlation factor R = 0.979 for the axial series of thiophosphates and R = 0.982 for the equatorial. From the slope of the lines m = -1.871 for the axial series and m = -1.631 for the equatorial it can be deduced that the phosphorus of the axial isomers is more sensitive to charge polarization of the aromatic ring than the phosphorus of the equatorial ones. The reversible correlation may be explained by a change in bond distances and bond angles involving α -bonded atoms to phosphorus and the changes in torsion angles with β -atoms as proposed by Gorenstein for phosphates.²⁷ We will discuss this point in the structural analysis below. On the other hand, neither the ¹H nor the ¹³C chemical shifts of the dioxaphosphinane ring correlate well with the Hammett $(\sigma_{\rm p})$ constants. However, a good normal correlation of ¹³C chemical shifts of the C_{ipso} with σ_p was found (R=0.926 for the axial and R=0.911 for the equatorial), Cipso of NO2 substituted arylthiophosphate being downfield shifted from Cipso of OCH3 aryl thiophosphate, as shown in Tables 5 and 6.



Figure 1. ORTEP drawing of ax-2-p-nitrophenoxy-2-thio-cis-4,6-dimethyl-1,3,2 λ ⁵-dioxaphosphinane (2-ax), molecule A and B.



Figure 2. ORTEP drawing of eq-2-p-nitrophenoxy-2-thio-cis-4,6-dimethyl-1,3,2 λ^5 -dioxaphosphinane (2-eq).

Based on the grounds of the stereoelectronic theory, EW groups will show more propensity to occupy axial orientations than ER groups (Scheme 2), therefore a detailed comparison of the structural parameters of the two axial 2-ax (X = NO₂), and 10-ax (X = CH₃) and the two equatorial 2-eq $(X=NO_2)$ and 11-eq $(X=OCH_3)$ thiophosphates not only led us to account for the participation of stereoelectronic interactions in the conformation adopted for the molecules but also for the reversible correlation found in the chemical shift of the signals of ³¹P NMR with σ_p . We also analyzed the steric compression of the aryloxy group in the axial thiophosphates and the flexibility of the 1,3,2dioxaphosphinane ring for to relieve the intramolecular van der Waals repulsive compression of the nonbonded substituents through compromise between the intraannular torsional strain and the Baeyer strain, as proposed for highly hindered thiophosphoramidates.

3.3. Structural analysis

The ORTEP drawings obtained from the X-ray analysis of **2**-*ax*, **2**-*eq*, **10**-*ax* and **11**-*eq* are shown in Figures 1–4. Data collection and refinement parameters, bond distances, bond angles, and torsion angles are provided in Tables 7–10.

Thiophosphates 2-*ax*, 2-*eq* and 11-*eq* crystallized in the monoclinic space group $P2_1/n$, $P2_1/c$ and $P2_1/a$ correspondingly, whereas 10-*ax* crystallized in the orthorhombic *P* space group $P2_1$ *n b*. In the case of compound 2-*ax*, two molecules were found in the asymmetric unit. Molecules A and B are not in equivalent positions, and it is interesting to note that the change of torsion angle P2–O10–C11–C12 by 74° brings with it a change in bond angle C4–O3–P2 (from 120° in molecule A to 117° in molecule B) that speaks about the high flexibility of the dioxaphosphinane ring in the OPO region ascribed to the propensity of the *endo*-cyclic oxygens to change from sp³ to sp² hybridization.⁷

3.4. Torsion and bond angles

The geometry at the phosphorus center is tetrahedral for all

the compounds; the sum of the four angles at the phosphorus goes from 436.04 to 439.71° (see Table 11). However, it is notable that equatorial thiophosphates are closer than the axial to the expected ideal tetrahedral angle (436°). The dioxaphosphinane ring lacks of perpendicular symmetry, the oxygens are almost flat edges [the internal COP angles are of around 119° for 2-*ax*, 2-*eq* and 10-*ax* and around 116° for 11-*eq*] and the phosphorus is a puckery end [the OPO angles are of around 106° for 2-*ax*, 2-*eq* and 10-*ax* and 104° for 11-*eq*]. There is ring flattening in the OPO region for both the axial and equatorial thiophosphates [torsion angles:



Figure 3. ORTEP drawing of ax-2-p-methylphenoxy-2-thio-cis-4,6-dimethyl-1,3,2 λ ⁵-dioxaphosphinane (10-ax).



Figure 4. ORTEP drawing of eq-2-p-methoxyphenoxy-2-thio-cis-4,6-dimethyl-1,3,2 λ^5 -dioxaphosphinane (11-eq).

 (ω_{POCC5}) are in the range of 44.6–55.8°; and the internal (ω_{OPOC}) are in the range of 33.1–52.7°] being the flattening less severe for **11**-*eq* than all others. The OPO ring flattening has also been found in analog phosphates.^{9a}

The Baeyer and Pitzer strain^{4,29} are summarized in Table 11 for compounds 2-*ax*, 2-*eq*, 10-*ax* and 11-*eq* [calculated as the average of the internal bond angles (θ), and torsion angles (ω) of the dioxaphosphinane ring, respectively]. Taking into consideration that the bond angle in a molecule free of Baeyer strain (propane) is 112.4°, the molecule with the highest Baeyer strain is 2-*eq* (113.1°). Contrary to what is expected, the intraannular torsion angles also decrease for 2-*eq* more than for any other compound (51.7–54.3° vs

Table 7. X-ray crystal data for compounds 2, 10-ax and 11- eq^a

47.5°) leading to considerable Pitzer strain. As a consequence, there is not a very good agreement between the values of $\cos \omega$ and $-\cos \theta/(1 + \cos \theta)$ for this molecule (0.68 vs 0.65) as for the others, indicating that the compromise between bond angles and torsion angles imposed by the constraint of the dioxaphosphinane ring which leads to the minimum strain in **2**-*eq*, is not perfect. We have observed that for thiophosphoramidates in conformations other than chair, the compromise between $\cos \omega$ and $-\cos \theta/(1 + \cos \theta)$ is not fulfilled either.⁷ Our interpretation of this result is that due to the axial seeking characteristics of the *p*-NO₂-phenoxy substituent, the molecule will tend to deform out of an ideal chair that otherwise in the case of **2**-*eq* obligates the aryloxy group to

	2 -ax	2 - <i>eq</i>	10 - <i>ax</i>	11 -eq
Formula	C11H14NO5PS	C11H14NO5PS	C12H12O2PS	C12H12O4PS
Fw	303.26	303.28	272.29	288.30
Crystal system	Monoclinic	Monoclinic	Orthorombic P	Monoclinic
Crystal size (mm)	$0.12 \times 0.18 \times 0.24$	$0.54 \times 0.45 \times 0.39$	$0.50 \times 0.50 \times 0.30$	$0.33 \times 0.30 \times 0.18$
Space group	$P2_1/n$	$P2_1/c$	$P2_1 n b$	$P2_1/a$
Radiation	ΜοΚα	ΜοΚα	ΜοΚα	ΜοΚα
Wavelength (Å)	0.71073	0.71073	0.71073	0.71073
a (Å)	8.04550(10)	6.856(1)	9.7597(10)	7.9454(10)
$b(\mathbf{A})$	26.9886(4)	22.911(1)	11.257(2)	20.1215(10)
$c(\mathbf{A})$	13.06750(10)	9.128(1)	12.9736(10)	9.4777(10)
α (deg)	90.00	90.00	90.00	90.00
β (deg)	95.805(6)	91.3(11)	90.00	104.407(10)
γ (deg)	90.00	90.00	90.00	90.00
$V(Å^3)$	2822.88(6)	1433.3(10)	1425.4(4)	1467.6(7)
Z	8	4	4	4
$2\theta_{\rm max}$ (deg)	4.36-49.94	11-12	4.80-53.94	11-12
$\rho_{\text{calc}} (\text{mg m}^{-3})$	1.427	1.41	1.269	1.30
Absorption coefficient	0.357	0.351	0.333	0.333
(mm^{-1})				
No. of reflections collected	5315	3475	1645	2101
No. of independent	4947	3122	1645	1792
reflections				
No. of observed reflections	2142	2348	1298	1285
$R_1 [F > 4\sigma(F)]$	0.0380	0.037	0.0338	0.0327
wR_2	0.1002	0.047	0.0961	0.0395
R_1 (all data)	0.1587	0.057	0.0500	0.0549
wR_2	0.1377	0.048	0.1060	0.0423
GOF on F^2	0.963	1.028	1.034	1.111
Max. shift for final cycle of	0.000	0.0004	0.000	0.0002
least-squares				
Δ/σ	0.000	0.000	0.001	0.000
Max. peak in final difference syntheses $(e/Å^3)$	0.223	0.33	0.344	0.17

^a Standard deviations are in parentheses.

Tabla	8	Selected	bond	langths	(Å)	for 2	10 av	and	11 aa ^a
rable	о.	Selected	bona	lenguis	(A)	101 2,	10- <i>ax</i>	anu	11 -eq

	2-ax molecule A	2-ax molecule B	2 - <i>eq</i>	10 -ax	11 -eq
O1-P2	1.559(3)	1.560(2)	1.5732(12)	1.565(3)	1.572(2)
O3-P2	1.562(3)	1.574(3)	1.5691(13)	1.568(2)	1.567(2)
O10-P2	1.605(3)	1.604(3)	1.5889(14)	1.601(3)	1.578(2)
S9-P2	1.8952(15)	1.8946(16)	1.9093(7)	1.9009(13)	1.9091(10)
O1-C6	1.476(6)	1.478(4)	1.466(2)	1.500(5)	1.472(3)
O3–C4	1.481(4)	1.470(5)	1.469(2)	1.508(6)	1.470(4)
C4–C5	1.515(5)	1.515(5)	1.504(3)	1.493(7)	1.494(4)
C5-C6	1.509(6)	1.512(5)	1.497(3)	1.519(7)	1.504(4)
C6–C8	1.505(5)	1.505(5)	1.502(3)	1.487(7)	1.498(4)
C4–C7	1.506(6)	1.513(5)	1.498(3)	1.511(7)	1.509(5)
C11-O10	1.403(4)	1.395(4)	1.403(2)	1.407(4)	1.406(3)

^a Standard deviations in parentheses.

Table 9. Selected bond angles (θ) in deg for 2, 10-ax and 11-eq^a

	2-ax molecule A	2 - <i>ax</i> molecule B	2 - <i>eq</i>	10 - <i>ax</i>	11 -eq
O1-P2-O3	106.49(14)	105.92(14)	105.72(7)	105.78(14)	104.06(11)
O1-P2-O10	98.72(15)	99.94(14)	103.39(7)	105.33(15)	102.53(11)
01-C6-C5	107.5(3)	108.6(3)	109.71(14)	106.9(3)	109.0(2)
O3-C4-C5	109.7(3)	109.0(3)	108.20(14)	110.0(3)	109.2(2)
O3-P2-O10	105.24(14)	104.81(14)	102.39(7)	100.33(15)	98.50(11)
O1-P2-S9	114.64(12)	114.83(11)	116.24(6)	113.65(11)	115.76(8)
O3-P2-S9	112.68(12)	113.82(12)	116.23(6)	114.77(12)	116.90(9)
C4-O3-P2	120.2(2)	117.0(2)	120.33(11)	120.6(3)	116.48(18)
C6O1P2	118.2(2)	119.3(2)	122.57(11)	118.0(3)	116.49(16)
C6-C5-C4	112.7(3)	113.5(3)	112.27(14)	111.8(4)	114.7(2)
P2010C11	126.5(2)	124.2(2)	121.4(1)	122.2(2)	124.34(17)
O10-C11-C12	116.9(3)	115.9(3)	120.10(16)	121.8(3)	118.3(3)
O10-C11-C16	121.3(4)	121.9(3)	117.67(16)	117.0(3)	120.2(3)
S9-P2-O10	117.58(11)	116.01(11)	111.24(5)	115.58(11)	116.61(9)

^a Standard deviations in parentheses.

Table 10. Selected torsion angles (ω) in deg. for **2**, **10**-*ax* and **11**-*eq*^{a,b}

	2-ax molecule A	2-ax molecule B	2 - <i>eq</i>	10 - <i>ax</i>	11 -eq	
S9-P2-O3-C4	-165.36	-174.17	93.03	-165.97	76.35	
O10-P2-O3-C4	65.30	58.07	-145.52	69.46	-157.93	
O1-P2-O3-C4	-38.85	-47.07	-37.58	-39.85	-52.66	
O1-C6-C5-C4	61.66	56.05	57.24	61.99	54.84	
S9-P2-O1-C6	169.23	172.06	-97.54	170.43	-77.19	
O10-P2-O1-C6	-64.92	-63.10	140.29	-62.08	154.73	
O3-P2-O1-C6	43.90	45.56	33.06	43.62	52.50	
P2-O3-C4-C5	47.03	55.30	50.56	49.23	55.77	
P2-O3-C4-C7	170.86	179.08	177.98	171.63	179.03	
P2O1C6C8	-179.80	-173.55	-169.53	179.79	-178.82	
P2O1C6C5	-56.48	-51.43	-44.60	-56.37	-55.20	
C11-O10-P2-O1	-179.36	-176.28	-61.19	-82.74	98.19	
C11-O10-P2-O3	70.80	74.18	48.53	167.59	-155.26	
C11-O10-P2-S9	-55.59	-52.24	173.35	43.57	-29.35	
C12-C11-O10-P2	-124.54	132.16	80.42	59.32	112.70	
C16-C11-O10-P2	58.92	-50.53	-102.95	-123.37	-72.75	
O3-C4-C5-C6	-57.14	-58.85	-61.68	-58.90	-55.09	
O10-P2-O3-C4	65.30	58.07	-145.52	69.46	-157.93	
O1-P2-O3-C4	-38.85	-47.07	-37.58	-39.85	-52.66	
P2O1C6C5	-56.48	-51.43	-44.60	-56.37	-55.20	

^a Standard deviations in parentheses. ^b Right-hand rule.²⁸

be in the equatorial position. This argument might be supported by the fact that the aryloxy group in 2-eq has torsion angles $[\omega_{O10PO1C6} = 140.3^{\circ} \text{ and } \omega_{O10PO3C4} = 145.5^{\circ}]$ that are almost 40° away from the 180° (expected for the group in equatorial orientation) and pointing towards a pseudo-axial position.

3.5. Stereoelectronic interactions

Several years ago, Gorenstein³⁰ coined the term 'gauche NMR effect' to support the observation that in molecules with gauche segments all the atoms that conform the segment tend to be shielded, therefore upfield shifted. In

	11 -eq	2 -eq	10 - <i>ax</i>	2 - <i>ax</i>	
				Molecule A	Molecule B
Geometry at phosphorus ^a	437.56	436.04	438.40	439.71	438.63
Baeyer strain ^b	111.66	113.13	112.18	112.47	112.22
Pitzer strain ^c	54.34	47.46	51.66	50.84	52.38
$\cos \omega$	0.58	0.68	0.62	0.63	0.61
$-\cos\theta/(1+\cos\theta)$	0.58	0.65	0.61	0.62	0.61

Table 11. Structural properties of 2, 10-ax and 11-eq

^a Calculated as the sum of the bond angles (O1P2O3), [OP2O10(mean)], [OP2S9(mean)] and (S9P2O10) in deg.

^b Calculated as the average value of the bond angles (O1P2O3), (O1C6C5), (O3C4C5), (C4O3P2), (C6C5C4), and (C6O1P2) in deg.

^c Calculated as the average absolute value of the torsion angles (O1P2O3C4), (O1C6C5C4), (O3P2O1C6), (O3C4C5C6), (P2O3C4C5), and (P2O1C6C5) in deg.

cyclic six-membered ring dioxaphosphinanes the ³¹P NMR signal of axial substituted compounds is normally upfield shifted from the equatorial,² thus demonstrating the participation of a gauche NMR effect. The ground, in which this gauche NMR effect relies is the decrease in the intraannular OPO bond of the dioxaphosphinanes, and it was nicely demonstrated by Gorenstein²⁷ that at least in phosphates there is a correlation between the OPO bond angle and the ³¹P NMR chemical shift. We thought of the possibility to explain the observed reversibility of ³¹P NMR shifts with $\sigma_{\rm p}$ or substituent-induced chemical shifts (SCS) with the changes in OPO bond angles, however unfortunately we found the opposite, that means that the compounds of the same series of thiophosphates, axial or equatorial, with smaller intraannular OPO bond are downfield shifted than those with larger OPO bond angles. For example, the OPO angle for 2-ax is 106.2° and for 10-ax is 105.78° and their ³¹P NMR are shifted to 54.3 and 56.2 ppm, respectively; by the same taken, the OPO bond angle for **2**-eq is 105.72° and for **11**-eq is 104.06° and their ³¹P NMR signals are shifted to 59.2 and 61.0 ppm, respectively. The other factor that can be disregarded from our data is that the shielding of the ³¹P NMR signal is due to a $d\pi$ -p π interaction involving the aryloxy group^{31,32} because in such case, we would expect that an ER group as p-OCH₃ would enhance the orbital overlap causing an upfield shift of the signal and an EW group as p-NO₂ a downfield shift. Alternatively the reversibility found in SCS may be explained by an effect of a compensation of charge density to the ³¹P nucleus given by the assistance of the free electron pairs of the endo-cyclic oxygens of the dioxaphosphinane ring, when an EW group is substituted in the para position of the phenyl ring of the aromatic thiophosphate, as shown in Scheme 6 (structure A). This hypothesis is supported by the fact that analog phosphinanes³³ where compensation of charge density on phosphorus by *endo*-cyclic α -atoms is not

possible, show a normal SCS trend (Scheme 6, structures B and C).

It is worthwhile to note that the transfer of charge density by endo-cyclic oxygens to phosphorus give rise to the known attractive *endo*-anomeric $n_{\pi}O-\sigma_{P-OAr}^{*}$ hyperconjugative interaction for axial thiophosphates that stabilize the chair conformation with the aryl-EW group more than with the aryl-ER. On the other hand, the attractive endo-anomeric $n_{\pi}O-\sigma_{P=S}^{*}$ hyperconjugative interaction for equatorial thiophosphates stabilizes the P=S group in the axial position of a chair conformation more than P=O in analog equatorial phosphates.³⁴ Indeed, by doing an individual examination of the bond lengths (Table 8), we observed that the data indicate clearly that both endo-hyperconjugative interactions do participate in the stabilization of these anancomeric thiophosphates (see Scheme 2). In particular, for 2-ax we found that endo-cyclic O1-P2 or O3-P2 bonds are shorter [1.560 Å (mean between molecule A and B) and 1.568 Å (mean between molecule A and B), respectively] than the exo-cyclic O10-P [1.605 Å (mean between molecule A and B)]. For 10-ax we also found evidence of the $n_{\pi}O-\sigma^*_{P-OAr}$ endo-hyperconjugative interaction since the O1-P2 or O3-P2 bonds (1.565 and 1.568 Å, respectively) are shorter than O10-P2 (1.601 Å). The fact that the shortening of the *endo*-cyclic O–P bonds in 2-ax (X=NO₂) is more pronounced than in 10-ax (X=Me) [1.564 Å (mean) for 2-ax vs 1.567 Å (mean) for 10-ax] and the lengthening of the O10-P2 bond is also more important for **2**-ax than for **10**-ax [1.605 Å for **2**-ax vs 1.601 Å for **10**-ax] is in agreement with the increase in the axial seeking characteristics of aryloxy substituted with EW as compared with ER groups. On the other hand, for the equatorial thiophosphates 2-eq $(X=NO_2)$ and 11-eq $(X=OCH_3)$ the anomeric $n_{\pi}O - \sigma_{P=S}^{*}$ endo-hyperconjugative interaction is supported by the shortening of the endo-cyclic O1-P2 or



X= H, ${}^{31}P=51.1 \text{ ppm}$ X= NO₂, ${}^{31}P=54.2 \text{ ppm}$

X = H, ${}^{31}P = 52.8 \text{ ppm}$ $X = NO_2$, ${}^{31}P = 56.0 \text{ ppm}$

(A)

O3–P2 bonds [1.572 Å (mean) for 2-eq and 1.570 Å (mean) for 11-eq], although somewhat less than in the axial isomers, and slight lengthening of the P=S bonds (1.909 Å for 2-eq or 11-eq vs 1.895 and 1.901 Å for 2-ax and 10-ax, respectively). It is clear that this stereoelectronic mechanism of stabilization of the equatorial thiophosphates is less important than the axial one, because in the equilibria of mobile aryl thiophosphates the axial conformer always predominates over the equatorial.^{1,2} This might be a result of the bonding electron repulsion caused by the lone pairs on sulfur when it is in the axial position.¹¹

4. Conclusions

The conformational analysis of a series of axial and equatorial anancomeric *p*-X-aryloxy thiophosphates is analyzed in terms of the electronic characteristics of the *p*-substituent. The coupling constants ${}^{3}J_{HH}$, ${}^{3}J_{HP}$ and ${}^{3}J_{CP}$ and X-ray structures suggest that in solution and solid state, the 1,3,2-dioxaphosphinane ring is in a chair conformation regardless of configuration. We observed that there is a reverse substituent-induced chemical shifts on the ${}^{31}P$ NMR signals being the effect more pronounced for axial than for equatorial thiophosphates. These results could be explained by the compensation of charge density on phosphorus by the lone pairs on *endo*-cyclic oxygens of the dioxaphosphinane ring for thiophosphates substituted with aryl EW groups.

The structural analysis performed by X-ray on four thiophosphates, two axial and two equatorial, led to the conclusion that in all cases, the dioxaphosphinane ring is flatten in the OPO region and is lacking of perpendicular symmetry being the *endo*-cyclic oxygens flat regions of the ring and the phosphorus atom a puckery end. A thoroughful analysis of bond distances allowed to support the participation of the anomeric $n_{\pi}O-\sigma^*_{P-OAr}$ *endo*-hyperconjugative interaction for to stabilization of axial thiophosphates and the anomeric $n_{\pi}O-\sigma^*_{P=S}$ *endo*-hyperconjugative interaction for to stabilization equatorial thiophosphates, both in chair conformations.

5. Experimental

Melting points are uncorrected. Nuclear magnetic resonance spectra were recorded on Jeol Eclipse 270 and Bruker Avance 300 spectrometers in CDCl_3 (δ 7.26, ¹H; δ 77.0, ¹³C), ¹H at 270.2 and 300.1 MHz, ¹³C at 67.8 and 75.5 MHz, and ³¹P at 109.3 and 121.5 MHz, respectively. Phosphorus NMR spectra are reported in ppm downfield (+) from 85% H₃PO₄ used as external standard. Mass spectra (EI) were measured on a Hewlett Packard 5989A spectrometer using electron impact (EI) at 70 eV. The reactions were performed under an atmosphere of nitrogen in oven-dried glassware. Solvents and solutions were transferred by syringe-septum and cannula techniques. Toluene was of reagent grade and was dried and distilled immediately before use from sodium/benzophenone. Triethylamine was dried and distilled from LiAlH₄. Products were purified by flash column chromatography on silica gel 230-400 mesh. Yields are given for isolated products. Galbraith Laboratories, Inc., Knoxville, TN

performed microanalyses of configurational isomers 3-9. Microanalyses of configurational isomers 2, 10, 11 and 12-eq were recorded in Thermo Finnigan Flash 1112 analyzer.

n-Hexane was used for recrystallization of all samples, affording crystals suitable for X-ray diffraction analysis. Crystallographic work was performed in an Enraf-Nonius CAD-4 diffractometer. Data collection: CAD-4 Software.³⁵ Cell refinement: CAD-4 Software. Data reduction for the axial structures **2**-*ax* and **10**-*ax*: WINGX,³⁶ solved by direct methods SHELXS97³⁷ and refined with SHELXL97.³⁸ Data reduction for the equatorial structures **2**-*eq* and **11**-*eq*: CRYSTALS,³⁹ solved and refined with CRYSTALS.³⁹ Molecular graphics: CAMERON⁴⁰ and dihedral angles: PLATON.⁴¹ Crystallographic data for structures have been deposited at Cambridge Crystallographic Data Center and the deposition numbers are: CCDC 234720 for **2**-*ax*, CCDC 234719 for **10**-*ax*, CCDC 235613 for **2**-*eq*, and CCDC 235614 for **11**-*eq*.

5.1. General procedure for the preparation of intermediates arylphosphites

Route A. In a three-necked 250 mL flask, fitted with a stirbar, and rubber septa, were placed 3.64 mmol of p-Xphenol, and 45 mL of dry toluene. The solution was stirred at room temperature until the *p*-X-phenol was solubilized (in the case of some p-X-phenols as p-acetamido, and p-amino, 10 mL of acetonitrile was added in order to solubilize them), then 3.64 mmol of the phosphorochloridite 1, followed by 3.64 mmol of triethylamine were added at once, via syringe, resulting in precipitation of triethylammonium chloride. The suspension was stirred for 5 min and the solid was filtered off through a filter tipped cannula. The solid was washed two times with 15 mL of dry toluene collecting the filtrate in a round-bottomed flask. The solvent was then removed under vacuum without heating to avoid epimerization of the equatorial phosphites to the axial ones. In all cases, products were yellowish oils.

Route B. In a three-necked 250 mL flask, fitted with dropping funnel, a stirbar, and rubber septa, were placed 3.64 mmol of p-X-phenol, 3.64 mmol of triethylamine, and 45 mL of dry ethyl ether. The mixture was stirred at room temperature for 30 min and 3.64 mmol of the phosphoro-chloridite **1** was added dropwise via syringe maintaining the stirring for additional 30 min. The solid in the resulting suspension was filtered off through a filter tipped cannula and the filtrate added to a lateral outlet round-bottomed flask equipped with a stirbar, rubber septa and reflux condenser. The solution was heated under reflux for 2 h and after cooling, the solvent was removed in a rotary evaporator. In all cases, products were yellowish oils.

Route C. In a round-bottomed 250 mL flask, fitted with reflux condenser, a stirbar, and rubber septa, were placed 3.64 mmol of the equatorial phosphites (obtained from route A) and 45 mL of dry toluene. The solution was stirred under reflux for 12-48 h until the epimerization to the axial isomers was complete. The epimerization process was followed by ³¹P NMR (equatorial phosphites are downfield shifted than axial phosphites by around 5 ppm, see

Scheme 5). Phosphites substituted with electron releasing groups (ERG) took longer for to epimerize than those substituted with electron-withdrawing groups (EWG). After cooling, the solvent was removed under vacuum in a rotary evaporator.

5.2. General procedure for the preparation of aryl thiophosphates 2–12

In a round-bottomed 100 mL flask, fitted with a reflux condenser, a stirbar, and rubber septa, were placed 2.21 mmol of elemental sulfur. A solution of 2.21 mmol of equatorial phosphite (obtained from route A) or axial phosphite (obtained from route B or C) in 60 mL of dry toluene was added to the flask and the resulting suspension was stirred under reflux for 24 h. After cooling, the suspension was concentrated under vacuum and the residue washed with an aqueous solution of 10% sodium bicarbonate. The product was extracted with methylene chloride and the organic layer dried over sodium sulfate. The solvent was removed in a rotary evaporator and the crude product was purified by flash chromatography using hexane/ethyl acetate as eluent.

5.2.1. *ax*-2-Chloro-*cis*-4,6-dimethyl-1,3,2- λ^3 -dioxaphosphinane (1). This compound was obtained from a mixture of *meso*- and *rac*-pentanediols and phosphorus trichloride by the stereoselective approach reported by us.⁴²

5.2.2. ax-2-p-Nitrophenoxy-2-thio-cis-4,6-dimethyl-1,3, $2\lambda^{3}$ -dioxaphosphinane (2-ax). According to the general procedure described above, 1.0 g (3.69 mmol) of axial *p*-nitrophenyl phosphite (route B), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 75:25) gave 1.0 g of yellow crystals (90% yield) of mp 111–112 °C. ¹H NMR δ 1.42 (d, J=2.3 Hz, 6H), 1.86 (m, J=14.6, 11.3, 1.0 Hz, 1H), 1.90 (m, J=14.6, 3.0, 2.6 Hz, 1H), 4.76 (m, J=11.3, 6.2, 3.0, 2.6 Hz, 2H), 7.36 (dd, ${}^{3}J = 8.9$ Hz, ${}^{4}J_{HP} = 1.2$ Hz, 2H), 8.24 (d, ${}^{3}J = 8.9$ Hz, 2H); ${}^{13}C$ NMR δ 22.4 (d), 40.6 (d), 77.7 (d), 121.3 (d, C_o), 126.0 (s, C_m), 145.1 (s, C_p), 156.2 (d, C_i); ³¹P NMR δ 54.3. Mass spectrum (m/z) 303 (M^+) , 262 (M^+-41) , 236 $(M^+ - 67)$, 218 $(M^+ - 85)$, 171 $(M^+ - 132)$, 149 $(M^+ - 67)$ 154), 123 $(M^+ - 180)$, 97 $(M^+ - 206)$, 69 $(M^+ - 234)$, 41 $(M^+ - 262)$. Anal. Calcd for $C_{11}H_{14}O_5PNS$: C, 43.57; H, 4.65. Found: C, 43.67; H, 4.85.

5.2.3. *eq*-2-*p*-Nitrophenoxy-2-thio-*cis*-4,6-dimethyl-1,3, 2λ⁵-dioxaphosphinane (2-*eq*). According to the general procedure described above, 1.5 g (5.54 mmol) of equatorial *p*-nitrophenyl phosphite (route A), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 75:25) gave 1.52 g of pale yellow crystals (91% yield) of mp 90–91 °C. ¹H NMR δ 1.41(d, J=2.2 Hz, 6H), 1.77 (m, J=14.6, 10.5, 1.0 Hz, 1H), 1.90 (m, J=14.6, 2.6, 2.8 Hz, 1H), 4.85 (m, J=10.5, 6.3, 2.6, 1.2 Hz, 2H), 7.35 (dd, ³J=9.2 Hz, ⁴ J_{HP} =1.6 Hz, 2H), 8.23 (d, ³J=8.9 Hz, 2H); ¹³C NMR δ 22.1 (d), 40.8 (d), 76.1 (d), 122.1 (d, C_o), 125.4 (s, C_m), 145.1 (s, C_p), 155.0 (d, C_i); ³¹P NMR δ 59.2. Mass spectrum (*m*/*z*) 303 (M⁺), 262 (M⁺-41), 236 (M⁺-67), 218 (M⁺-85), 205 (M⁺-98), 171 (M⁺-132), 149 (M⁺-154), 123 (M⁺-180), 97 (M⁺-206), 69

 $(M^+ - 234)$, 41 $(M^+ - 262)$. Anal. Calcd for $C_{11}H_{14}O_5PNS$: C, 43.57; H, 4.65. Found: C, 43.80; H, 4.51.

5.2.4. ax-2-p-Cyanophenoxy-2-thio-cis-4,6-dimethyl-1,3, $2\lambda^{3}$ -dioxaphosphinane (3-ax). According to the general procedure described above, 1.0 g (3.98 mmol) of axial p-cyanophenyl phosphite (route B), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 85:15) gave 1.04 g of the product as pale yellow crystals (93% yield) of mp 110–111 °C. ¹H NMR δ 1.43 (d, J=2.5 Hz, 6H), 1.85 (m, J=14.8, 10.5, 1.5 Hz, 1H), 1.87 (m, J=14.8, 2.5, 1.3 Hz, 1H), 4.73 (m, J=10.5, 6.4, 2.5, (iii, J = 14.6, 2.6, 1.5 Hz, HI), 4.75 (iii, J = 16.6, 6.4, 2.5, 1.2 Hz, 2H), 7.30 (dd, ${}^{3}J = 9.1$ Hz, ${}^{4}J_{HP} = 1.3$ Hz, 2H), 7.65 (d, ${}^{3}J = 9.1$ Hz, 2H); ${}^{13}C$ NMR δ 22.3 (d), 40.3 (d), 77.1 (d), 108.8 (s, CN), 118.3 (s, C_p), 121.3 (d, C_o), 134.1 (s, C_m), 154.3 (d, C_i); ³¹P NMR δ 54.5. Mass spectrum (*m*/*z*) 283 (M^+) , 242 $(M^+ - 41)$, 216 $(M^+ - 67)$, 197 $(M^+ - 86)$, 165 $(M^+ - 118), 149 (M^+ - 134), 119 (M^+ - 164), 90 (M^+ - 164))$ 193), 69 $(M^+ - 234)$, 41 $(M^+ - 242)$. Anal. Calcd for C₁₂H₁₄O₃PNS: C, 50.88; H, 4.98. Found: C, 50.91; H, 5.16.

5.2.5. eq-2-p-Cyanophenoxy-2-thio-cis-4,6-dimethyl-1,3, $2\lambda^{5}$ -dioxaphosphinane (3-eq). According to the general procedure described above, 1.5 g (5.97 mmol) of equatorial *p*-cyanophenyl phosphite (route A), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 80:20) gave 1.52 g of the product as pale yellow crystals (90% yield) of mp 106–107 °C. ¹H NMR δ 1.41 (d, J = 2.0 Hz, 6H), 1.79 (m, J = 14.5, 11.2 Hz, 1H), 1.87 (m, J=14.5, 2.7, 3.6 Hz, 1H), 4.84 (m, J=11.2, 6.6, 2.7, 3.3 Hz, 2H), 8.26 (d, ${}^{3}J = 8.6$ Hz, 2H), 8.57 (d, ${}^{3}J = 8.9$ Hz, 2H); ${}^{13}C$ NMR δ 22.1 (d), 40.9 (d), 76.0 (d), 109.4 (s, CN), 118.1 (s, C_p), 122.4 (s, C_o), 133.8 (s, C_m), 152.6 (d, C_i); ³¹P NMR δ 59.3. Mass spectrum (m/z) 283 (M^+) , 242 $(M^+ - 41)$, 216 $(M^+ - 67)$, 197 $(M^+ - 86)$, 165 $(M^+ - 118)$, 149 $(M^+ - 67)$ 134), 119 (M⁺ - 164), 90 (M⁺ - 193), 69 (M⁺ - 234), 41 $(M^+ - 242)$. Anal. Calcd for $C_{12}H_{14}O_3PNS$: C, 50.88; H, 4.98. Found: C, 51.02; H, 5.21.

5.2.6. *ax-2-p*-Formylphenoxy-2-thio-*cis*-4,6-dimethyl-**1,3,2** λ^{5} -dioxaphosphinane (4-*ax*). According to the general procedure described above, 1.0 g (3.94 mmol) of axial *p*-formylphenyl phosphite (route B), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 60:40) gave 1.01 g of the product as oil (90% yield). ¹H NMR δ 1.42 (d, *J*=2.2 Hz, 6H), 1.83 (m, *J*= 13.0, 8.0 Hz, 1H), 1.85 (m, *J*=13.0, 2.7, 1.5 Hz, 1H), 4.75 (m, *J*=8.0, 6.2, 2.7, 1.2 Hz, 2H), 7.35 (dd, ³*J*=8.6 Hz, ⁴*J*_{HP}=1.5 Hz, 2H), 7.87 (d, ³*J*=8.6 Hz, 2H), 9.94 (s, CHO); ¹³C NMR δ 22.1 (d), 40.2 (d), 76.7 (d), 120.7 (d, *C*_o), 131.7 (d, *C*_m), 133.3 (s, *C*_p), 155.4 (d, *C*_i), 190.8 (s, CHO); ³¹P NMR δ 55.1. Mass spectrum (*m*/*z*) 286 (M⁺), 245 (M⁺ – 41), 219 (M⁺ – 67), 199 (M⁺ – 87), 167 (M⁺ – 119), 149 (M⁺ – 137), 138 (M⁺ – 148), 121 (M⁺ – 165), 97 (M⁺ – 189), 69 (M⁺ – 217), 41 (M⁺ – 245). Anal. Calcd for C₁₂H₁₅O₄PS: C, 50.35; H, 5.28. Found: C, 50.46; H, 5.40.

5.2.7. *eq-2-p*-Formylphenoxy-2-thio-*cis*-4,6-dimethyl-1, 3,2 λ^5 -dioxaphosphinane (4-*eq*). According to the general procedure described above, 1.0 g (3.94 mmol) of equatorial *p*-formylphenyl phosphite (route A), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 60:40) gave 1.02 g of brown crystals (91% yield)

of mp 55–57 °C. ¹H NMR δ 1.34 (d, J=2.3 Hz, 6H), 1.54 (m, J=14.5, 10.4, 1.8 Hz, 1H), 1.69 (m, J=14.5, 2.7, 2.3 Hz, 1H), 4.79 (m, J=10.4, 6.3, 2.7, 2.8 Hz, 2H), 7.30 (m, 2H), 7.80 (m, 2H), 9.91 (s, CHO); ¹³C NMR δ 22.1 (d), 40.8 (d), 75.9 (d), 122.0 (d, C_o), 131.4 (d, C_m), 133.6 (s, C_p), 155.0 (d, C_i), 190.8 (s, CHO); ³¹P NMR δ 59.5. Mass spectrum (m/z) 286 (M⁺), 219 (M⁺ -67), 199 (M⁺ -87), 169 (M⁺ -117), 149 (M⁺ -137), 138 (M⁺ -148), 122 (M⁺ -164), 85 (M⁺ -201), 69 (M⁺ -217), 41 (M⁺ - 245). Anal. Calcd for C₁₂H₁₅O₄PS: C, 50.35; H, 5.28. Found: C, 50.35; H, 5.41.

5.2.8. ax-2-p-Bromophenoxy-2-thio-cis-4,6-dimethyl-1, $3,2\lambda^3$ -dioxaphosphinane (5-ax). According to the general procedure described above, 1.0 g (3.27 mmol) of axial *p*-bromophenyl phosphite (route B), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 85:15) gave 1.03 g of white crystals (93% yield) of mp 132–133 °C. ¹H NMR δ 1.42 (d, J = 2.2 Hz, 6H), 1.82 (m, J=14.8, 10.8, 1.0 Hz, 1H), 1.87 (m, J=14.8, 2.8, 1.0 Hz)2.7 Hz, 1H), 4.75 (m, J=10.8, 6.2, 2.8, 1.2 Hz, 2H), 7.19 $(dd, {}^{3}J=8.9 \text{ Hz}, {}^{4}J_{\text{HP}}=1.4 \text{ Hz}, 2\text{H}), 7.49 (d, {}^{3}J=8.9 \text{ Hz},$ 2H); ¹³C NMR δ 22.4 (d), 40.8 (d), 77.2 (d), 118.3 (s, C_p), 122.6 (d, C_o), 133.1 (s, C_m), 150.6 (d, C_i); ³¹P NMR δ 55.5 (56.0). Mass spectrum (m/z) 338 $(M^+ + 1)$, 296 $(M^+ - 41)$, 270 (M^+ -67), 188 (M^+ -149), 172 (M^+ -165), 149 $(M^+ - 188)$, 69 $(M^+ - 268)$, 41 $(M^+ - 296)$. Anal. Calcd for C₁₁H₁₄O₃PSBr: C, 39.19; H, 4.19. Found: C, 39.17; H, 4.16.

5.2.9. eq-2-p-Bromophenoxy-2-thio-cis-4,6-dimethyl-1, **3,2\lambda^{5}-dioxaphosphinane** (5-eq). According to the general procedure described above, 1.5 g (4.91 mmol) of equatorial p-bromophenyl phosphite (route A), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 80:20) gave 1.54 g of white crystals (90% yield) of mp 64–65 °C. ¹H NMR δ 1.32 (d, J=2.0 Hz, 6H), 1.64 (m, J=14.5, 10.6 Hz, 1H), 1.77 (m, J=14.5, 2.8, 2.3 Hz, 1H), 4.76 (m, J = 10.9, 6.3, 2.6, 2.3 Hz, 2H), 7.01 (dd, ${}^{3}J =$ 8.6 Hz, ${}^{4}J_{\text{HP}} = 1.3$ Hz, 2H), 7.38 (d, ${}^{3}J = 8.6$ Hz, 2H); ${}^{13}\text{C}$ NMR δ 22.0 (d), 40.9 (d), 75.6 (d), 118.6 (s, C_p), 123.1 (d, C_o , 132.5 (s, C_m), 149.5 (d, C_i); ³¹P NMR δ 60.0. Mass spectrum (m/z) 338 $(M^+ + 1)$, 305 $(M^+ - 32)$, 252 $(M^+ - 32)$ 85), 190 (M^+ – 147), 172 (M^+ – 165), 149 (M^+ – 188), 101 (M^+ - 236), 69 (M^+ - 268), 41 (M^+ - 296). Anal. Calcd for C₁₁H₁₄O₃PSBr: C, 39.19; H, 4.19. Found: C, 39.21: H. 4.28.

5.2.10. *ax-2-p*-Chlorophenoxy-2-thio-*cis*-4,6-dimethyl-**1,3,2** λ^5 -dioxaphosphinane (6-*ax*). According to the general procedure described above, 2.8 g (10.73 mmol) of axial *p*-chlorophenyl phosphite (route A-to-C), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 85:15) gave 0.72 g of white crystals (23% yield) of mp 126–128 °C. ¹H NMR δ 1.45 (d, *J*=2.1 Hz, 6H), 1.83 (m, *J*=14.1, 11.1, 1.1 Hz, 1H), 1.90 (m, *J*=14.1, 3.0, 1.3 Hz, 1H), 4.75 (m, *J*=11.1, 6.6, 3.0, 1.1 Hz, 2H), 7.12 (d, ³*J*=8.8 Hz, 2H), 7.30 (d, ³*J*=8.8 Hz, 2H); ¹³C NMR δ 22.6 (d), 40.6 (d), 76.7 (d), 121.7 (d, *C*_o), 129.8 (s, *C*_p), 130.5 (s, *C*_m), 149.5 (d, *C*_i); ³¹P NMR δ 55.6. Mass spectrum (*m*/*z*) 292 (M⁺), 224 (M⁺ – 68), 149 (M⁺ – 143), 128 (M⁺ – 164), 99 (M⁺ – 193), 69 (M⁺ – 154), 41 (M⁺ – 251). Anal. Calcd for $C_{11}H_{14}O_3PSC1$: C, 45.14; H, 4.82. Found: C, 45.27; H, 5.04.

5.2.11. eq-2-p-Chlorophenoxy-2-thio-cis-4,6-dimethyl-1, **3.2\lambda^5-dioxaphosphinane** (6-eq). According to the general procedure described above, 2.8 g (10.73 mmol) of equatorial *p*-chlorophenyl phosphite (route A), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 90:10) gave 1.54 g of white crystals (50% yield) of mp 59–60 °C. ¹H NMR δ 1.39 (d, J=2.0 Hz, 6H), 1.71 (m, J=14.0, 11.2 Hz, 1H), 1.87 (m, J=14.0, 3.0, 2.3 Hz, 1H), 4.83 (m, J = 11.2, 6.6, 3.0, 2.5 Hz, 2H), 7.16 (d, ${}^{3}J =$ 8.6 Hz, 2H), 7.38 (d, ${}^{3}J$ = 8.6 Hz, 2H); ${}^{13}C$ NMR δ 22.5 (d), 41.3 (d), 76.1 (d), 122.9 (d, C_p), 123.1 (d, C_o), 130.1 (d, C_m), 149.4 (d, C_i); ³¹P NMR δ 60.1. Mass spectrum (*m/z*) 292 (M^+) , 224 $(M^+ - 68)$, 149 $(M^+ - 143)$, 128 $(M^+ - 164)$, 99 $(M^+ - 193)$, 69 $(M^+ - 154)$, 41 $(M^+ - 251)$. Anal. Calcd for C₁₁H₁₄O₃PSCI: C, 45.14; H, 4.82. Found: C, 45.17; H, 4.98.

5.2.12. ax-2-p-Acetamidophenoxy-2-thio-cis-4,6-dimethyl-1,3,2 λ^5 -dioxaphosphinane (7-ax). According to the general procedure described above, 3.0 g (10.73 mmol) of axial *p*-acetamidophenyl phosphite (route A-to-C), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 85:15) gave 0.75 g of white crystals (22% yield) of mp 188–190 °C. ¹H NMR δ 1.44 (d, J= 2.3 Hz, 6H), 1.86 (m, J=13.0, 12.9, 1.0 Hz, 1H), 1.88 (m, J=13.0, 2.9, 1.8 Hz, 1H), 2.79 (s, 3H), 4.77 (m, J=12.9, 6.2, 2.9, 1.3 Hz, 2H), 7.12 (d, ${}^{3}J=8.9$ Hz, 2H), 7.49 (d, ${}^{3}J=$ 8.9 Hz, 2H), 8.56 (s, 1H); ¹³C NMR δ 22.5 (d), 24.7 (s), 42.7 (d), 76.8 (d), 121.9 (d, C_o), 122.6 (s, C_p), 135.1 (s, C_m), 147.5 (d, C_i), 168.4 (s); ³¹P NMR δ 55.9. Mass spectrum (m/z) 315 (M^+) , 273 (M^+-42) , 205 (M^+-110) , 187 $(M^+ - 128)$, 125 $(M^+ - 190)$, 108 $(M^+ - 207)$, 69 $(M^+ - 190)$ 246), 43 (M^+ – 272). Anal. Calcd for C₁₃H₁₈O₄PNS: C, 49.52; H, 5.75. Found: C, 49.64; H, 5.79.

5.2.13. *eq*-2*-p*-Acetamidophenoxy-2-thio-*cis*-4,6-dimethyl-**1**,3,2λ⁵-dioxaphosphinane (7-*eq*). According to the general procedure described above, 3.0 g (10.73 mmol) of equatorial *p*-acetamidophenyl phosphite (route A), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 70:30) gave 1.54 g of pale yellow crystals (90% yield) of mp 102–104 °C. ¹H NMR δ 1.38 (d, *J*=2.4 Hz, 6H), 1.75 (m, *J*=14.1, 10.8 Hz, 1H), 1.85 (m, *J*=14.1, 2.3, 2.3 Hz, 1H), 2.14 (s, 3H), 4.80 (m, *J*=10.8, 6.2, 2.3, 2.6 Hz, 2H), 7.10 (dd, ³*J*=8.6 Hz, ⁴*J*_{HP}=1.3 Hz, 2H), 7.46 (d, ³*J*= 8.6 Hz, 2H), 7.64 (s, 1H); ¹³C NMR δ 22.4 (d), 24.7 (s), 41.1 (d), 75.8 (d), 121.0 (s, C_{*p*}), 121.7 (d, C_{*o*}), 135.5 (s, C_{*m*}), 149.6 (d, C_{*i*}), 168.6 (s); ³¹P NMR δ 60.4. Mass spectrum (*m*/*z*) 315 (M⁺), 273 (M⁺-42), 229 (M⁺-86), 205 (M⁺-110), 187 (M⁺-128), 125 (M⁺-190), 108 (M⁺-207), 69 (M⁺-246), 43 (M⁺-272). Anal. Calcd for C₁₃H₁₈O₄PNS: C, 49.52; H, 5.75. Found: C, 49.64; H, 5.92.

5.2.14. *ax*-2-Phenoxy-2-thio-*cis*-4,6-dimethyl-1,3,2 λ^{5} dioxaphosphinane (8-*ax*).¹⁵ According to the general procedure described above, 1.0 g (4.42 mmol) of axial phenyl phosphite (route B), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 95:5) gave 1.05 g of white crystals (92% yield) of mp 120–122 °C. ¹H NMR δ 1.43 (d, *J*=2.6 Hz, 6H), 1.82 (m, *J*=13.8, 9.9,

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1.0 Hz, 1H), 1.88 (m, J=14.4, 2.8, 2.6 Hz, 1H), 4.79 (m, J=10.8, 6.6, 2.8, 1.3 Hz, 2H), 7.19 (m, 3H), 7.35 (m, ${}^{3}J$ =8.9 Hz, 2H); 13 C NMR δ 21.9 (d), 39.9 (d), 76.2 (d), 119.8 (d, C_o), 125.5 (s, C_p), 129.5 (s, C_m), 150.5 (d, C_i); 31 P NMR δ 55.8.

5.2.15. *eq*-2-Phenoxy-2-thio-*cis*-4,6-dimethyl-1,3,2 λ^5 -dioxaphosphinane (8-*eq*).¹⁵ According to the general procedure described above, 3.0 g (13.27 mmol) of equatorial phenyl phosphite (route A), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 98:2) gave 3.18 g of white crystals (93% yield) of mp 55–56 °C. ¹H NMR δ 1.39 (d, J=2.3 Hz, 6H), 1.70 (m, J=14.5, 10.9 Hz, 1H), 1.89 (m, J=14.5, 2.6, 2.3 Hz, 1H), 4.83 (m, J=10.9, 6.3, 2.6, 2.3 Hz, 2H), 7.20 (m, 3H), 7.35 (m, 2H); ¹³C NMR δ 22.1 (d), 41.0 (d), 75.4 (d), 121.1 (d, C_o), 125.5 (s, C_p), 129.5 (s, C_m), 150.3 (d, C_i); ³¹P NMR δ 60.4.

5.2.16. ax-2-p-Phenylphenoxy-2-thio-cis-4,6-dimethyl-1,3,2 λ^{5} -dioxaphosphinane (9-ax). According to the general procedure described above, 3.5 g (11.59 mmol) of axial *p*-phenylphenyl phosphite (route B, or the sequence A-to-C), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 98:2) gave 3.48 g of white powder (90% yield) of mp 145–146 °C. ¹H NMR δ 1.44 (d, J = 2.2 Hz, 6H), 1.83 (m, J = 13.8, 9.9, 1.0 Hz, 1H), 1.88 (m, J=13.8, 2.9, 2.9 Hz, 1H), 4.81 (m, J=9.9, 6.2, 2.9, 1.2 Hz, 2H), 7.27 (m, 2H), 7.35 (m, 1H), 7.44 (m, 2H), 7.59 (m, 4H); ¹³C NMR δ 22.5 (d), 40.7 (d), 77.1 (d), 121.0 (d, C_o), 127.4 (s, C_m), 127.8 (s, C_{p'}), 128.7 (s, C_{o'}), 129.2 (s, C_{m'}), 138.5 (s, C_p), 140.5 (s, C_{i'}), 150.8 (d, C_i); ³¹P NMR δ 55.8. Mass spectrum (*m*/*z*) 334 (M⁺), 266 (M⁺ - 68), 186 (M⁺ - 148), 170 $(M^+ - 164)$, 141 $(M^+ - 193)$, 91 $(M^+ - 243)$, 69 $(M^+ - 265)$, 28 $(M^+ - 306)$. Anal. Calcd for C₁₇H₁₉O₃PS: C, 61.07; H, 5.73. Found: C, 61.05; H, 5.44.

5.2.17. eq-2-p-Phenylphenoxy-2-thio-cis-4,6-dimethyl-1,3,2 λ^3 -dioxaphosphinane (9-eq). According to the general procedure described above, 2.5 g (8.27 mmol) of equatorial *p*-phenylphenyl phosphite (route A), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 98:2) gave 2.51 g of white powder (91% yield) of mp 165–166 °C. ¹H NMR δ 1.34 (d, J=2.0 Hz, 6H), 1.67 (m, J = 14.2, 9.9 Hz, 1H), 1.77 (m, J = 14.2, 2.9, 2.3 Hz, 1H), 4.78 (m, J=9.9, 6.2, 2.9, 3.0 Hz, 2H), 7.19 (m, 2H), 7.28 (m, 1H), 7.36 (m, 2H), 7.48 (m, 4H); ¹³C NMR δ 22.1 (d), 41.0 (d), 75.5 (d), 121.5 (d, C_o), 127.0 (s, C_{m'}), 127.3 (s, $C_{p'}$), 128.2 (s, $C_{o'}$), 128.8 (s, C_m), 138.6 (d, C_p), 140.2 (s, $C_{i'}$), 149.9 (d, C_i); ³¹P NMR δ 60.4. Mass spectrum (*m*/*z*) $334 (M^+)$, 266 (M⁺-68), 186 (M⁺-148), 170 (M⁺-164), 141 (M⁺ - 193), 115 (M⁺ - 219), 85 (M⁺ - 249), 69 $(M^+ - 265)$, 41 $(M^+ - 293)$. Anal. Calcd for $C_{17}H_{19}O_3PS$: C, 61.07; H, 5.73. Found: C, 61.28; H, 6.00.

5.2.18. *ax-2-p*-Methylphenoxy-2-thio-*cis*-4,6-dimethyl-**1,3,2** λ^5 -dioxaphosphinane (10-*ax*). According to the general procedure described above, 1.5 g (6.25 mmol) of axial *p*-methylphenyl phosphite (route B, or the sequence A-to-C), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 80:20) gave 1.03 g of white crystals (89% yield) of mp 138–139 °C. ¹H NMR δ 1.41 (d, J=2.2 Hz, 6H), 1.82 (m, J=14.5, 10.7, 1.1 Hz, 1H), 1.84 (m, J=14.5, 3.0, 2.7 Hz, 1H), 2.32 (s, CH₃), 4.77 (m, J=10.7, 6.2, 3.0, 1.2 Hz, 2H), 7.06 (dd, ${}^{3}J=8.5$ Hz, ${}^{4}J_{\rm HP}=1.5$ Hz, 2H), 7.17 (d, ${}^{3}J=8.5$ Hz, 2H); 13 C NMR δ 20.9 (s, CH₃), 22.4 (d), 40.7 (d), 76.9 (d), 120.4 (s, C_p), 130.6 (d, C_o), 135.3 (s, C_m), 148.8 (d, C_i); 31 P NMR δ 56.2. Mass spectrum (*m*/*z*) 272 (M⁺), 204 (M⁺ - 68), 186 (M⁺ - 86), 149 (M⁺ - 123), 124 (M⁺ - 148), 108 (M⁺ - 164), 91 (M⁺ - 181), 69 (M⁺ - 203), 43 (M⁺ - 229), 41 (M⁺ - 231). Anal. Calcd for C₁₂H₁₇O₃PS: C, 52.93; H, 6.29. Found: C, 53.03; H, 6.40.

5.2.19. eq-2-p-Methylphenoxy-2-thio-cis-4,6-dimethyl- $1,3,2\lambda^5$ -dioxaphosphinane (10-eq). According to the general procedure described above, 3.5 g (14.58 mmol) of equatorial p-methylphenyl phosphite (route A), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 80:20) gave 3.6 g of white crystals (91% yield) of mp 82–83 °C. ¹H NMR δ 1.34 (d, J=2.6 Hz, 6H), 1.59 (m, J = 14.5, 11.2, 2.3 Hz, 1H), 1.72 (m, J = 14.5, 2.6, 2.6 Hz, 1H), 2.27 (s, CH₃), 4.80 (m, J = 10.9, 6.3, 2.6, 2.6 Hz, 2H), 7.03 (dd, ${}^{3}J=8.6$ Hz, ${}^{4}J_{HP}=2.0$ Hz, 2H), 7.08 (d, ${}^{3}J=$ 8.6 Hz, 2H); ¹³C NMR δ 20.8 (s, CH₃), 22.1 (d), 41.0 (d), 75.3 (d), 121.0 (d, C_o), 129.9 (d, C_o), 135.1 (s, C_m), 148.3 (d, C_i); ³¹P NMR δ 60.9. Mass spectrum (*m*/*z*) 272 (M⁺), 204 (M⁺-68), 186 (M⁺-86), 149 (M⁺-123), 124 (M⁺-148), 108 (M^+ – 164), 91 (M^+ – 181), 69 (M^+ – 203), 43 $(M^+ - 229)$, 41 $(M^+ - 231)$. Anal. Calcd for $C_{12}H_{17}O_3PS$: C, 52.93; H, 6.29. Found: C, 52.73; H, 6.41.

5.2.20. *ax-2-p*-Methoxyphenoxy-2-thio-*cis*-4,6-dimethyl-**1,3,2** λ^5 -dioxaphosphinane (11-*ax*). According to the general procedure described above, 3.0 g (11.71 mmol) of axial *p*-methoxyphenyl phosphite (route B, or the sequence A-to-C), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 85:15) gave 2.93 g of white powder (92% yield) of mp 94–96 °C. ¹H NMR δ 1.42 (d, *J*= 2.2 Hz, 6H), 1.81 (m, *J*=14.5, 10.8, 1.0 Hz, 1H), 1.86 (m, *J*=14.5, 3.3, 3.0 Hz, 1H), 3.78 (s, OCH₃), 4.77 (m, *J*=10.8, 6.2, 3.3, 1.2 Hz, 2H), 6.78 (dd, ³*J*=8.8 Hz, ⁴*J*_{HP}=1.5 Hz, 2H), 6.87 (d, ³*J*=8.8 Hz, 2H); ¹³C NMR δ 22.4 (d), 40.7 (d), 56.0 (s, OCH₃), 76.9 (d), 115.0 (s, C_m), 121.7 (d, C_o), 144.9 (d, C_i), 157.29 (d, C_p); ³¹P NMR δ 56.7. Mass spectrum (*m*/*z*) 288 (M⁺), 220 (M⁺-68), 202 (M⁺-86), 140 (M⁺-148), 124 (M⁺-164), 95 (M⁺-193), 69 (M⁺-219), 41 (M⁺-247). Anal. Calcd for C₁₂H₁₇O₄PS: C, 49.99; H, 5.94. Found: C, 50.06; H, 6.11.

5.2.21. eq-2-p-Methoxyphenoxy-2-thio-cis-4,6-dimethyl-1,3,2 λ^{5} -dioxaphosphinane (11-eq). According to the general procedure described above, 2.5 g (9.76 mmol) of equatorial p-methoxyphenyl phosphite (route A), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 80:20) gave 1.54 g of white crystals (90% yield) of mp 93–94 °C. ¹H NMR δ 1.31 (d, J=2.0 Hz, 6H), 1.63 (m, J=14.5, 10.9 Hz, 1H), 1.75 (m, J=14.5, 2.6, 2.6 Hz, 1H), 3.71 (s, OCH₃), 4.73 (m, J = 10.9, 6.3, 2.6, 2.6 Hz, 2H), 6.77 (dd, ${}^{3}J = 8.9$ Hz, ${}^{4}J_{HP} = 1.6$ Hz, 2H), 7.04 (d, ${}^{3}J = 8.9$ Hz, 2H); ${}^{13}C$ NMR δ 22.0 (d), 40.9 (d), 55.6 (s, OCH₃), 75.6 (d), 114.3 (s, C_m), 122.0 (d, C_o), 144.0 (d, C_i), 156.9 (d, C_p); ³¹P NMR δ 61.0. Mass spectrum (*m*/*z*) 288 (M^+) , 220 $(M^+ - 68)$, 202 $(M^+ - 86)$, 170 $(M^+ - 118)$, 140 $(M^+ - 148)$, 124 $(M^+ - 164)$, 119 $(M^+ - 169)$, 95 $(M^+ - 193)$, 69 $(M^+ - 219)$, 41 $(M^+ - 247)$. Anal. Calcd for $C_{12}H_{17}O_4PS$: C, 49.99; H, 5.94. Found: C, 49.81; H, 6.29.

5.2.22. *ax-2-p*-Aminophenoxy-2-thio-*cis*-4,6-dimethyl-**1,3,2λ⁵-dioxaphosphinane** (12-*ax*). A mixture of 1.3 g (11.86 mmol) of *p*-aminophenyl phosphites (*ax/eq* 7:93) was obtained through route B. The mixture was maintained at room temperature because heating led to decomposition of the intermediate phosphites. The mixture of phosphites was treated with elemental sulfur according to the general procedure described above. Flash chromatography (hexanes/ethyl acetate 35:65) was unsuccessful to purify the axial isomer, therefore it was analyzed as a mixture. ¹³C NMR δ 22.3 (d), 40.5 (d), 76.2 (d), 116.5 (s, C_m), 121.1 (d, C_o), 144.8 (d, C_i), 143.8 (s, C_p); ³¹P NMR δ 56.8.

5.2.23. *eq*-2-*p*-Aminophenoxy-2-thio-*cis*-4,6-dimethyl-**1**,3,2λ⁵-dioxaphosphinane (12-*eq*). According to the general procedure described above, 4.0 g (23.73 mmol) of equatorial *p*-aminophenyl phosphite (route A), was treated with elemental sulfur. Flash chromatography (hexanes/ethyl acetate 35:65) gave 1.5 g of brown crystals (23% yield) of mp 93–94 °C. ¹H NMR δ 1.38 (d, *J*=2.0 Hz, 6H), 1.68 (m, *J*=14.5, 11.2 Hz, 1H), 1.82 (m, *J*=14.5, 2.6, 2.6 Hz, 1H), 3.25 (br, NH₂), 4.52 (m, *J*=11.2, 6.6, 2.6, 3.3 Hz, 2H), 6.64 (m, 2H), 6.98 (m, 2H); ¹³C NMR δ 21.9 (d), 40.8 (d), 75.8 (d), 115.4 (s, C_m), 121.8 (d, C_o), 142.6 (d, C_i), 142.6 (s, C_p); ³¹P NMR δ 61.3. Mass spectrum (*m*/*z*) 274 (M⁺ + 1), 205 (M⁺ - 68), 187 (M⁺ - 86), 125 (M⁺ - 148), 109 (M⁺ -164), 94 (M⁺ - 179), 69 (M⁺ - 204), 41 (M⁺ - 232). Anal. Calcd for C₁₁H₁₆O₃PNS: C, 48.35; H, 5.90. Found: C, 48.52; H, 6.04.

Acknowledgements

We thank CONACyT (México) for financial support (Grant 32221-E). We are grateful to Geiser Cuellar for the elemental analysis of some of the compounds.

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Tetrahedron

Tetrahedron 60 (2004) 10943-10948

The facile preparation of alkenyl metathesis synthons

Travis W. Baughman, John C. Sworen and Kenneth B. Wagener*

The George and Josephine Butler Polymer Research Laboratory, Department of Chemistry, University of Florida, PO Box 117200, Gainesville, FL 32605, USA

Received 27 April 2004; revised 3 September 2004; accepted 9 September 2004

Available online 1 October 2004

Dedicated to Professor Robert H. Grubbs on the occasion of his receiving the Tetrahedron Prize for Creativity in Organic Chemistry

Abstract—We report synthetic methodology allowing the preparation of any length alkenyl halide from inexpensive starting reagents. Standard organic transformations were used to prepare straight chain α -olefin halides in excellent overall yields with no detectable olefin isomerization and full recovery of any unreacted starting material. Reported transformations can be used for the selective incorporation of pure α -olefin metathesis sites in highly functionalized molecules.

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1. Introduction

Olefin metathesis has emerged as one of the primary methods for mild carbon–carbon bond formation in organic synthesis, having been widely used across many areas of chemical research. Functional group tolerant ruthenium based catalysts permit the metathesis of highly functiona-lized compounds that were previously incompatible with traditional catalysts.¹ The development of these stable and reactive catalysts, such as Grubbs' second-generation complex, has expanded metathesis' utility in polymer chemistry^{2,3} and more recently in small molecule synthesis.^{4–8} Difficult cross-metathesis and ring-closing metathesis reactions can now be accomplished allowing access to large ring systems⁹ and functionalized polymer architectures.¹⁰ The application of efficient and mild transformations such as ring closing metathesis (RCM),¹¹ ring opening metathesis (ROM),¹² and cross-metathesis in small molecule and polymer chemistry.¹³

In general, RCM is valuable for selective and efficient ring closures of large functionalized natural products and supramolecularly organized substrates.¹⁴ Application of ROM in combination with CM effectively produces complex ring-opened cross-metathesis products from the coupling of cyclic and linear olefins; this method has been used to connect large molecules and is readily applied in

total synthesis.¹⁵ In addition, advances in CM and the understanding of olefin reactivity have stimulated the development of complex synthetic schemes where two or more olefins can be reacted regio- and stereoselectively forming only the target olefin in excellent yields.¹⁶ Acyclic diene metathesis (ADMET), a special form of CM yielding polymers, has been used in materials synthesis and industrial polymer modeling.² This rapid rise in olefin metathesis popularity, utility, and catalyst improvement has generated the need for inexpensive, high yielding α -olefin constructs and the means of incorporating metathesis sites into various complex architectures.

We now report three facile and inexpensive synthetic routes to a family of pure α -alkenyl halide metathesis synthons possessing exact methylene run lengths. These synthetic procedures and routine purification methods afford high yields of olefins with no detectable double bond isomerization. The mild transformations reported can also be used to place olefins in highly functionalized molecules without the need for expensive starting materials.

2. Results and discussion

Metathesis ideology has fueled the development of next generation catalyst systems for the production of specific, highly functionalized target molecules for medical and consumer use. Consequently, a large scale, high-yielding α -olefin synthesis is needed to produce molecules for subsequent substitution and metathesis chemistry. Before any metathesis can occur, olefins must be incorporated into substrates where mild and inexpensive techniques would be

Keywords: α -Olefin; Cross-metathesis; Alkenyl halide; ADMET; Metathesis.

^{*} Corresponding author. Tel.: +1 352 392 4666; fax: +1 352 392 9741; e-mail: wagener@chem.ufl.edu

^{0040–4020/\$ -} see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.021

preferred for the creation of target olefins. While wellknown literature techniques yield α -olefins directly or transform complex substrates into metathesis active compounds, unwanted side reactions occur leading to low yields.^{17,18} This makes recovery of pure starting materials difficult and expensive, while reaction products are sometimes extensively isomerized. These are common problems associated with α -olefin synthesis, and they often lead to structural defects when isomerized olefins are used in metathesis transformations.

Metathesis strategy in small molecule modification typically targets one specific regio- and stereoselective attachment of functionality to produce a high yield of a single molecule. While this facet of synthesis is important, controlling the number of methylene units between olefins is paramount when targeting specific ring size or exact structure. With this in mind, acyclic diene metathesis (ADMET) polymerization of symmetrical α, ω -diene monomers and ROMP of symmetrical cyclic olefins have become the polymerization methods of choice for the preparation of polymers with exact repeat units.

Substrate purity for metathesis reactions is essential, as structural flaws lead to low yielding ring closures, the isolation of the wrong metathesis products, or ill-defined polymer microstructure. Inclusion of internal and external olefins as starting materials in metathesis poses problems as methylene run lengths within product structure will be varied based on extent of olefin isomerization. In the case of small molecule synthesis, variable methylene content in the substrate yields different ring-sizes for RCM or incorrect methylene run lengths for ROM or CM reactions (Scheme 1). The inclusion of isomerized olefins in starting material synthesis creates ill-defined structures leading to numerous olefinic products.

The construction of ADMET monomers and isolation of exact polymer structures begins with the synthesis of pure α -olefins. Errors due to inclusion of isomerized olefins during polymer synthesis become multiplied with high olefin turnover, and ultimately these errors produce random polymer structures and ill-defined methylene run lengths between functional groups. These defects are detrimental for polymer modeling studies and lead to poor material properties relative to polymers obtained from purely α -olefins.¹⁹ We have addressed these problems starting with the inexpensive synthesis of 1-alkenyl bromides, reagents that

can be regarded as metathesis synthons in ADMET, crossmetathesis, and RCM chemistry (Scheme 1).

We have devised two routes to these synthons; the preparation of pure α, ω -alkenyl halides was performed either by the reduction of alkenyl esters or through the elimination of HBr from alkyl dibromides. Both routes are described herein, along with a procedure to generate long chain α -alkenyl halide synthons.

2.1. Alkenyl halide metathesis synthons from alkenyl esters

The key to success in generating ADMET, cross-metathesis and RCM dienes is found in locating a source of pure α -alkenyl halide bulk starting materials. Numerous sources of such starting materials exist based on alkenyl esters derived from fatty acids with no isomerization of the pendent olefin. These molecules can be easily reduced to their corresponding alcohols, and literature describing the reduction of various alkenyl esters extends to the 1940s including LAH reactions,^{20–23} titanium mediated reductions of esters,^{24,25} and polymethylhydrosilane reductions.²⁶ We have chosen the inexpensive conversion of an α -olefin ester or carboxylate to an alcohol using LAH in THF (Scheme 2). This method was chosen specifically for the reduction of 4-pentenyl ester as the byproduct ethanol could be easily removed via rotary evaporation.

These inexpensive reactions were done on the 100-gram scale in a large flask with sufficient volume for post reaction workup. Upon product isolation, compounds 1 and 2 were characterized via ¹H NMR and GC with no detectable olefin isomerization and above 90% purity. Both alkenyl alcohols were clean enough to move onto the next transformation without any further purification.

Conversion of these alkenyl alcohols to corresponding bromides was done using carbon tetrabromide and triphenyl phosphine. This reaction appears to be the most efficient route when considering all reasonable possibilities including the use of phosphorus tribromide,²⁷ bromine,²⁸ trifluoro aceticanhydride,²⁹ and various other conversions that first convert the alcohol to a good leaving group such as a mesylate^{30,31} or a silyl ether.³² The bromination with CBr₄ proceeded quickly and was complete almost as fast as all components could be mixed. The byproduct acid,



Scheme 1. Cross-metathesis, ring closing metathesis and the ADMET reaction.



Scheme 2. Alkenyl halide synthesis.

bromoform, does not isomerize the olefin and allows for isolation of clean α -olefin in good yields.

Synthesis of chlorine-functionalized α -olefins has been previously reported and is usually performed neat via addition of SOCl₂ to the liquid alcohol.^{21–23} We performed this conversion by adding the SOCl₂ to a solution of alkenyl alcohol **2** in pyridine acting as a solvent and an acid trap. Upon vacuum distillation, the alkenyl chloride **5** was obtained in good yield. Conversion of the chloride to the alkenyl iodide was performed by a Finkelstein reaction with sodium iodide in acetone.³³

2.2. Alkenyl halide metathesis synthons via selective dihalide elimination

A method of olefin incorporation within molecules was desired in addition to the production of inexpensive starting reagents for metathesis synthesis. Mild elimination conditions were developed allowing a second method of α -olefin production that could also permit olefin incorporation in functionalized molecules where application of substitution chemistry is unavailable. Literature methods designed to prepare alkenes and alkenyl halides involve the elimination of bromo and dibromo alkanes using hexamethylphophortriamide (HMPT).^{17,18} This route is expensive and potentially dangerous due to the highly carcinogenic compound HMPT. Harsh conditions and the highly reactive HMPT lead to a myriad of byproducts that interfere with purification and further, the high temperatures needed for

Br Br
$$HF$$
, toluene, 25°C 3

Scheme 3. A mild elimination route towards the synthesis of alkenyl bromide metathesis synthons.

this conversion (150 °C) can result in olefin isomerization in addition to halide elimination. One other disadvantage to this elimination chemistry is the inability to recover unreacted starting material from the complex reaction mixture. Recently, a potassium butoxide elimination has been reported affording alkenyl halides in good yield.³⁴ A milder, higher yielding room temperature route is presented in Scheme 3.

Simple KOtBu driven elimination in a THF/toluene solvent mixture produces the target molecules in ~65% yield. This elimination reaction is started at 0 °C and is allowed to warm to room temperature over 1 h. Upon quenching with aqueous acid, the dibromide/alkenyl bromide mixture can be easily purified affording the target molecule. The recovered dibromide can be recycled for further conversion to alkenyl bromide as necessary.

2.3. Synthesis of larger alkenyl bromide metathesis synthons

Up to this point, the incorporation of metathesis sites into target molecules has been limited by commercial availability and expense of alkenyl bromides. This is especially true when considering the preparation of longer chain alkenyl bromides, a problem that has been overcome by exploiting and extending the chemistry described in the previous sections. As an example of this strategy, we synthesized an alkenyl bromide containing 20 carbons, a molecule which is commercially unavailable.

The synthesis of extended alkenyl bromides began with the self-metathesis of 4 with Grubbs' first generation catalyst to afford 7 (Scheme 4). This dibromide was converted to 8 by exhaustive hydrogenation in a Parr bomb with Wilkinson's Rh catalyst under hydrogen pressure. The saturated 20-carbon dibromide 8 was then converted to the target

Scheme 4. The preparation of long chain alkenyl bromide metathesis synthons.

PC_{V3}

extended chain alkenyl bromide **9** using the same elimination procedure previously described for smaller alkenyl bromides. Pure compound **9** was obtained in good yield after recrystallization and column chromatography, and recovery of the pure starting reagents was accomplished during purification since very few side reactions accompany these mild transformations.

3. Conclusions

Three mild, inexpensive routes have been devised for the production of pure α -olefin containing halides as metathesis synthons. Using these methods, virtually any 1-alkenyl bromide can be made in high yields and devoid of olefin isomerization. Many of the reactions discussed here are either quantitative in nature, or the starting reagent can be easily recovered for further use.

4. Experimental

4.1. General information

All ¹H NMR (300 MHz) and ¹³C NMR (75 MHz) spectra were recorded in CDCl₃ on a Varian Associates Mercury 300 spectrometer. Chemical shifts are given in ppm and referenced to residual CHCl₃ at 7.27 ppm (¹H) and 77.23 ppm (¹³C) with 0.03 v/v% TMS as an internal standard. Splitting patterns are designated s, singlet; d, doublet; t, triplet; m, multiplet; br, broad. Gas chromatography was performed on a Shimadzu GC-17A chromatograph equipped with a RTX-5 (Restek Corp.) 15 m column and a HP-5 (Hewlett Packard) 25 m column with FID detection. Compounds were examined using mass spectrometry performed by The University of Florida Mass Spectrometry Services and elemental analysis performed by Atlantic Microlabs (Norcross, GA).

4.2. Materials

All starting materials were purchased from Aldrich except for acetic acid 4-pentenyl ester, which was supplied by TCI America. Grubbs' first generation [Ru] catalyst was synthesized as previously described by Grubbs et al.³⁵ Dry solvents were collected using an Aldrich keg system removing residual water by alumina filtration.

4.3. Synthesis of alkenyl alcohols

4.3.1. 4-Pentene-1-ol (1). Acetic acid pent-4-enyl ester (250 mL, 1.77 mol) was added dropwise over 2 h to a slurry of LAH (23.7 g, 0.625 mol) in diethyl ether (500 mL) at 0 °C. The solution was allowed to warm to room temperature over 1 h while stirring. The reaction was quenched via the sequential addition of 23 mL deionized water (23 mL), 15% (w/v) NaOH (23 mL), and deionized water (69 mL) waiting approximately 5 min between additions. The solution was allowed to stir the mixture became a bright white slurry. Additional ether (\sim 125 mL) was added and the solution was filtered, dried over MgSO₄, and concentrated to a colorless oil. Compound 1 was

obtained in 85% yield and 97% purity by GC. The 1 H NMR spectrum was consistent with the published spectrum.³⁶

4.3.2. 10-Undecene-1-ol (2). Zinc undecylenate (250 g. 0.579 mol) was added over 30 min via powder funnel to a stirred slurry of LAH (25.0 g, 0.659 mol) in dry THF (400 mL) in a 5 L round bottom flask at 0 °C. After the addition, the solution was allowed to warm to room temperature over 1 h while stirring. The reaction was quenched via addition of deionized water (25 mL), 15% (w/v) NaOH (25 mL), and more water (75 mL) waiting approximately 15 min between additions. The solution was allowed to stir until cool, and the reaction mix appeared as a white slurry. All precipitate was filtered, and the solution was concentrated to a turbid oil. The crude mixture was dissolved in ether (100%, v/v) and stirred over MgSO₄ (15 g) for 45 min. The solution was filtered and concentrated to afford compound 2 as a colorless oil in 87% yield (97% pure by GC). The ¹H NMR spectrum was consistent with the published spectrum.²⁴

4.4. Synthesis of alkenyl halides from alkenyl alcohols

4.4.1. 5-Bromo-1-pentene (3). A solution of **1** (100 g, 1.16 mol) and carbon tetratbromide (424 g, 1.27 mol) in dichloromethane (500 mL) was prepared in a 2 L flask and cooled to 0 °C. Triphenyl phosphine (333 g, 1.28 mol) was added via powder funnel in portions over 30 min with vigorous stirring. Upon addition of the phosphine, the colorless solution turned a pale brown color and was stirred for an additional 2 h at room temperature. The mixture was concentrated to a brown oil and quickly added to stirring hexane (1 L). The white precipitate was filtered, and the remaining solution was concentrated and fractionally distilled yielding compound **3** as a colorless oil (80%). The ¹H NMR spectrum was consistent with the published spectrum.³⁷

4.4.2. 11-Bromo-1-undecene (4). Using the procedure outlined for compound 3 (above), bromide 4 was prepared using alcohol 2 (150 g, 0.881 mol), carbon tetrabromide (323 g, 0.976 mol), triphenyl phosphine (255 g, 0.976 mol), and 800 mL of dichloromethane. Fractional distillation yielded compound 4 as a colorless liquid (95%). The ¹H NMR spectrum was consistent with the published spectrum.²⁸

4.4.3. 11-Chloro-1-undecene (5). In an argon purged 3 L round bottom flask, distilled thionyl chloride (259 g, 2.17 mol) was added dropwise over 1 h via cannula into a solution of **2** (200 g, 1.28 mol) in pyridine (50 mL). Upon compete addition, the reaction was heated to 50 °C for 2 h, cooled, and quenched via the addition of water (300 mL) and diethyl ether (300 mL) letting stir for 1 h. The remaining mixture was extracted, and the organic phase was washed with saturated NaHCO₃ (2×150 mL) and distilled water (100 mL). The solution was dried over MgSO₄, concentrated, and vacuum distilled to a colorless oil **5** (70%). The ¹H NMR spectrum was consistent with the published spectrum.³⁸

4.4.4. 11-Iodo-1-undecene (6). Sodium iodide (40.7 g, 0.271 mol) was added to distilled **5** (30.6 g, 0.162 mol) in

acetone (150 mL) and allowed to reflux for 3 days. The reaction mixture was then cooled and flooded with ether (300 mL). The precipitate was filtered and the ether solution was washed with water (2×100 mL). The organic layer was extracted, dried with MgSO₄, filtered, and concentrated to a colorless oil **6** (89%). The ¹H NMR spectrum was consistent with the published spectrum.³⁹

4.5. Synthesis of alkenyl bromides from dibromides

4.5.1. 5-Bromo-1-pentene (3). In a 1 L round bottom flask 1,5-dibromopentane (100 g, 0.434 mol) was dissolved in 450 mL of a 1:1 THF/toluene solution to favor the single eliminated product. The flask was cooled to 0 °C followed by the addition of solid KOtBu (73.0 g, 0.651 mol) over 30 min. After addition, the reaction was quenched using 1 M HCl (300 mL) and the organic layer was extracted, washed with saturated Na₂CO₃ (100 mL), and dried over magnesium sulfate. The solution was concentrated and distilled yielding 44 g of compound **3** (69%). The ¹H NMR spectrum was consistent with the published spectrum.³⁷

4.5.2. 1,20-Dibromo-eicos-10-ene (7). In an argon filled glove box, compound 4 (100 g, 429 mmol) was added to a 500 mL round bottom flask followed by Grubbs' catalyst (706 mg, 0.885 mmol, 500:1). The flask was heated at 35 °C for 24 h under a constant stream of argon. After 1 day, the flask was placed under vacuum (10 Torr) for an additional 48 h, cooled, and quenched with ethyl vinyl ether (5 mL). The crude reaction mixture was dissolved in toluene (200 mL) and precipitated into methanol (1000%, v/v) over the course of 30 min. The product was filtered as a white crystalline solid and washed with excess methanol yielding 75 g (80%). The following spectral properties were observed: ¹H NMR (CDCl₃): δ (ppm) 1.20–1.50 (br, 24H), 1.88 (q, 4H, $J_1 = 7.0$ Hz, $J_2 = 7.0$ Hz, CH_2CH_2Br), 2.01 (m, 4H, allylic CH₂), 3.42 (t, 4H, J = 6.7 Hz, CH₂Br), 5.40 (m, 2H, olefin); 13 C NMR (CDCl₃): δ (ppm) 28.4, 29.3, 29.7, 29.8, 32.8, 33.1, 34.4, 130.3; EI/HRMS: [M]⁺ calcd for C₂₀H₃₈Br₂: 438.1322, found: 438.1312.

4.5.3. 1,20-Dibromo-eicosane (8). In a 125 mL Parr bomb glass sleeve, compound 7 (35 g, 80 mmol) was dissolved in a minimal amount of toluene ($\sim 80 \text{ mL}$). Wilkinson's Rh hydrogenation catalyst (100 mg, 0.104 mmol) was added, and the bomb was charged with 800 psi of hydrogen. The reaction was allowed to proceed for 24 h at 50 °C. Additional toluene (200 mL) was added, and upon cooling to 0 °C, the product 8 crystallized out of solution and was collected by filtration. The filtrate was concentrated $(\sim 50\%)$, and the product was allowed to crystallize again. Upon isolation of the product from the second crystallization, both portions were combined and washed with cold toluene. Yield: 30 g (86%). The following spectral properties were observed: ¹H NMR (CDCl₃): δ (ppm) 1.28 (br, 28H), 1.42 (m, 4H, J = 6.7 Hz), 1.88 (q, 4H, $J_1 = 6.7$ Hz, $J_2 = 7.2 \text{ Hz}, CH_2CH_2Br), 3.42 (t, 4H, J = 6.9 \text{ Hz}, CH_2Br);$ ¹³C NMR (CDCl₃): δ (ppm) 28.4, 29.7, 29.8, 30.0, 33.1, 34.4; EI/HRMS: $[M]^+$ calcd for $C_{20}H_{40}Br_2$: 440.1478, found: 440.1616.

4.5.4. 20-Bromo-eicos-1-ene (9). In a 1 L round bottom flask compound 8 (50 g, 113 mmol) was dissolved in 2:1

THF/toluene mixture producing a 1 M solution. The mixture was cooled using an ice water bath, and potassium tert-butoxide (19.0 g, 170 mmol) was added in 2 g portions over 30 min. After addition, the reaction turned cloudy and was allowed to stir at 0 °C for 1 h. The reaction was quenched using water (100 mL) followed by 1 M HCl (100 mL). The organic layer was extracted and washed with 1 M HCl (50 mL), saturated Na₂CO₃ (50 mL), and 50 mL of water followed by drying with magnesium sulfate. The solution was concentrated yielding 38 g of crude material. Compound 9 was purified by room temperature recrystallization from 1-butanol (5 w/v%) followed by column chromatography using hexane. Compound 9 was collected as white solid. Yield: 24 g (60%). The following spectral properties were observed: ¹H NMR (CDCl₃): δ (ppm) 1.20–1.50 (br, 30H), 1.88 (q, 2H, J=7.2 Hz, CH₂CH₂Br), 2.01 (q, 2H, allylic CH₂), 3.42 (t, 2H, J = 6.9 Hz, CH₂Br), 5.09 (m, 2H, RHC= CH_2), 5.76 (m, 1H, RHC= CH_2); ¹³C NMR (CDCl₃): δ (ppm) 28.4, 29.3, 29.7, 29.8, 32.8, 33.1, 34.4, 114.3, 139.4; EI/HRMS: $[M]^+$ calcd for $C_{20}H_{39}Br$: 358.2235, found: 358.2246.

Acknowledgements

We would like to thank NSF, ARO, and the NASA Space Grant Consortium for funding. We would also like to thank and Timothy Hopkins and Ed Lehman for advice and synthetic work.

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Tetrahedron

Tetrahedron 60 (2004) 10949-10954

Enantiospecific synthesis of isomers of AES, a new environmentally friendly chelating agent

Petri M. Pihko,^{a,*} Terhi K. Rissa^a and Reijo Aksela^b

^aLaboratory of Organic Chemistry, Department of Chemical Technology, Helsinki University of Technology, P.O.B. 6100, FI-02015 HUT Espoo, Finland

^bKemira Oyj, Espoo Research Center, Luoteisrinne 2, FI-02271 Espoo, Finland

Received 20 July 2004; revised 24 August 2004; accepted 9 September 2004

Available online 1 October 2004

Abstract—Three four-step enantiospecific syntheses of different diastereomers of AES, a new biodegradable chelating agent, are described. The stereocenters in each of the isomers are accessible from L- and D-malic and aspartic acids. © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

Due to the increasing use of chelating agents, their environmental fate has become an important issue.¹ Approximately 50,000 tons of aminopolycarboxylates such as EDTA and DTPA are used annually; mainly in the textile, detergent and pulp and paper industries.² EDTA and DTPA have proven to be practically non-biodegradable in standard tests.³ For this reason, alternative chelating agents such as the aspartic acid derivatives ethylene diamine disuccinic acid (EDDS) and iminodisuccinic acid (IDS) have become more popular as biodegradable chelating agents, particularly for detergent applications.⁴

A series of novel diethanolamine derivatives such as aspartic acid ethoxysuccinate (AES) have recently been introduced as chelating agents suitable for pulp and paper applications (Fig. 1).⁵ Due to its higher biodegradability, lower nitrogen content and capacity to form inert complexes with iron and manganese ions, AES is a more environmentally friendly alternative to EDTA and DTPA in the pulp and paper industry.⁶ A non-selective but industrially viable route to AES via a lanthanum catalyzed Michael addition of diethanolamine to maleate has recently been described.⁷ The presence of three stereogenic centers in the AES molecule gives rise to stereoisomerism. When the pseudo-symmetrical nature of the AES molecule is taken into account, one can observe that the (*S*,*S*,*R*) and the (*R*,*S*,*S*) isomers are identical to each other, as are the (*S*,*R*,*R*) and the

(R,R,S) isomers. For this reason, there are only six possible isomers of AES, consisting of three pairs of enantiomers (Fig. 2).

Based on previous studies with the EDDS isomers,⁸ the different AES isomers are expected to have different biodegradability characteristics. For full characterization of these new chelating agents, we therefore needed access to all AES isomers. Herein, we present a simple and efficient protocol for the enantiospecific synthesis of AES.

2. Results and discussion

The three chiral centers of AES are, in principle, accessible from (R)- and (S)-isomers of the readily available malic and aspartic acids. Connecting these building blocks to form the AES framework, however, is not straightforward. There are numerous possible side-reactions including the formation of lactams and lactones as well as retro-Michael reactions.

Retrosynthetic analysis of the target molecule reveals two suitable synthetic strategies which use malic and aspartic





Keywords: Aldehydes; Amino acids; Chelating agents; Reductive amination; Stereoisomerism.

^{*} Corresponding author. Tel.: +358 9 451 2536; fax: +358 9 451 2538; e-mail: petri.pihko@hut.fi

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double O-alkylation

Scheme 1. Retrosynthetic analysis.

acid as starting materials: double *O*-alkylation and double *N*-alkylation (Scheme 1). Of the two disconnections, the double *N*-alkylation strategy appears more attractive since it would, at least in principle, allow the stepwise construction of the molecule by two consecutive *N*-alkylation steps. Thus, the (S,S,R) isomers could also be accessed by this method.

NH₂

NaO₂C

ĊO₂Na

2

We were also further discouraged from pursuing the O-alkylation strategy by our initial attempts at O-alkylation of (*S*)-malic acid dialkyl esters with tosylated diethanolamine derivatives (Scheme 2). Instead of the desired O-alkylation, the reaction invariably produced the corresponding morpholines derived from the tosylate **6**.

There are several reductive amination methods available which reliably give the monoalkylation product so we decided to use this approach next to perform double *N*-alkylation of the aspartate methyl ester using a suitable malic acid derived aldehyde.⁹ This approach would provide easy access to all AES isomers since the final reductive amination could then be performed with either of the enantiomeric aldehydes. However, we recognized that the



Scheme 2. Initial attempts at *O*-alkylation of dimethyl malate with diethanolamine-derived electrophiles.

requisite dialkylation reaction could be a difficult task given the low nucleophilicity of the monoalkylation product.

For the synthesis of the key aldehyde building block 9 (Scheme 3) and its enantiomer 16 (see Scheme 4), we initially used a procedure adapted from the previous synthesis of 9 by Samuelsson and co-workers.¹⁰ Thus, dimethyl (S)- or (R)-malate was alkylated with allyl bromide using freshly prepared Ag₂O as the bromide scavenger. We found that the amount of Ag₂O was critical for this transformation since use of more than 20 mol% excess of the reagent led to allylation of the solvent as well. After OsO₄-catalyzed dihydroxylation and subsequent cleavage of the resulting diol with sodium periodate, the aldehyde 9 was obtained in 60-65% overall yield from dimethyl malate. This procedure worked very efficiently on the small scale described by the authors (0.5 mmol). On scaleup, however, several modifications to the procedure were required. The aldehyde 9 is very prone to polymerization and decomposition, particularly when dry. Even traces of acids in deuterated chloroform were enough to completely destroy a batch of aldehyde 9 in a matter of hours! However, when pure, 9 could easily be stored in a freezer in a frozen cyclohexane matrix for several months without decomposition.



Scheme 3. Synthesis of aldehyde 9. Reagents and conditions: (a) allyl bromide (9 equiv), Ag_2O (1 equiv), toluene, rt, 2 h; (b) OsO_4 (2 mol%), *N*-methylmorpholine-*N*-oxide (NMO, 2 equiv), 3:1 THF/H₂O, 0 °C \rightarrow rt, 21 h; then NaIO₄ (2 equiv), 3:1 THF/H₂O, 80% from 5.

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Scheme 4. Synthesis of AES isomers. Reagents and conditions: (a) aldehyde 9 (2.6 equiv), aspartate 10 or 13 (1 equiv), NaBH₃CN (2.1 equiv), MeOH, 0 °C \rightarrow rt, 24 h; 40% 11, 48% 12 or 40% 14, 42% 15; (b) aldehyde 9 (3.65 equiv), aspartate 10 (1 equiv), NaBH₃CN (3 equiv), MeOH, 0 °C \rightarrow rt, 48 h, 80% 11; (c) aldehyde 9 or 16 (1.1 equiv), amine 12 (1 equiv), NaBH₃CN (1.1–1.5 equiv), formic acid (2 equiv), MeOH, 0 °C \rightarrow rt, 24 h, 40% 11 or 17 (55% 11, 59% 17 based on recovered 12); (d) 1 M aq. NaOH (6.8 equiv), 1:1 MeOH/THF, rt, 18 h, quant.

After considerable experimentation, we found that aldehyde **9** was obtained in 80% overall yield by filtering the crude reaction mixture through silica, concentration in vacuo, addition of brine followed by careful gradient extraction with Et_2O and CH_2Cl_2 . The first Et_2O extract had to be purified by filtration through a pad of silica, however, all subsequent extracts gave pure **9** upon concentration! If further purification was needed, **9** could readily be distilled in high vacuum using a Kugelrohr apparatus. With these modifications, multigram amounts of **9** and **16** could readily be prepared.

With the aldehydes **9** and **16** in hand, the reductive amination was explored. Initial experiments using Pd/C catalysis (2.2 equiv of aldehyde, 1 equiv **10**, EtOAc or EtOH solvents) gave complex reaction mixtures with only traces of the desired alkylation products. Very poor conversions were also obtained with sodium triacetoxyborohydride. However, sodium cyanoborohydride gave more satisfactory results. If all the reagents were added at the same time to a cooled mixture of 2.5 equiv of the aldehyde, 1 equiv of amine and 2 equiv of NaBH₃CN, substantial decomposition of the aldehyde was observed, even at 0 °C. However, with gradual addition of the amine and the cyanoborohydride over 1.5 h, and with 2 equiv of aldehyde, the monoalkylation product **12** was obtained in 63% yield and the dialkylation product **11** in 23% yield.

Better yields of the dialkylation product were obtained with 2.5 equiv of the aldehyde (40% 11, 48% 12). A more dramatic improvement in yield was realized when a further portion of the aldehyde (1.3 equiv) was added to the reaction mixture after 24 h. Using this procedure, the yield of the dialkylation product climbed to 80%. However,

purification of the product became progressively more difficult with increased amounts of the aldehyde. Thus, it was easier and more reliable to recycle the monoalkylated product **12** and to resubject it to the reaction conditions (in this case, 2 equiv of formic acid was added to the reaction mixture to facilitate the iminium ion formation). The overall yield of the dialkylated product after one cycle was 55%. The corresponding (S,R,S) isomer was synthesized in a similar manner using the (R)-aspartic acid dimethyl ester **13** as the starting material (Scheme 4). The hexamethylester **14** was obtained in 40% yield after one cycle, along with 42% of the monoalkylated product **15**.

To access the (S,S,R)-isomer, the (S,S)-tetraester **12** was treated with the (R)-malic acid derived aldehyde **16** (1.1 equiv). The (S,S,R) ester **17** was obtained in 40% yield (59% based on recovered **12**). All the hexamethyl esters were diastereomerically >95% pure according to NMR spectroscopic analysis (the ¹H and ¹³C NMR signals, although very close, are clearly distinct in all isomers).

Finally, the corresponding sodium salts of the AES isomers were readily obtained in quantitative yields by careful saponification of the products. Here, slow addition of a slight excess of NaOH was necessary to prevent unwanted retro-Michael reactions. Using an excess of NaOH led to the partial loss of one of the malic acid groups as the fumarate. However, no isomerization could be detected in the NMR spectra. The final products were judged to be >95% diastereomerically pure by ¹H and ¹³C NMR spectroscopy.

The three isomers of AES synthesized herein, the (S,S,S), the (S,R,S) and the (S,S,R) isomers, are each members of the three possible enantiomeric pairs of AES isomers. The remaining

isomers are thus enantiomeric to the ones described herein and accessible through identical procedures.

3. Conclusion

In summary, a concise and versatile synthesis of isomers of AES, a novel biodegradable chelating agent is described. The synthesis is applicable to the asymmetric synthesis of all isomers of AES in pure form and it is amenable to scaleup.

With the pure isomers **2–4** in hand, the differences in biodegradability and capacity as chelating agents can now be fully explored. These studies will be described in detail elsewhere.

4. Experimental

4.1. General methods

All reactions were carried out under an argon atmosphere in flame-dried glassware, unless otherwise noted. Nonaqueous reagents were transferred under argon via syringe or cannula and dried prior to use. THF was distilled from Na/benzophenone. CH₂Cl₂ was distilled over CaH₂. All synthetic intermediates were azeotropically dried with toluene prior to use. Other solvents and reagents were used as obtained from supplier, unless otherwise noted. Analytical TLC was performed using Merck silica gel F254 (230-400 mesh) plates and analyzed by UV light or by staining upon heating with vanillin solution (6 g vanillin, 5 mL conc. H₂SO₄, 3 mL glacial acetic acid, 250 mL EtOH) or with ninhydrin solution (1 g ninhydrin, 100 mL isopropanol, five drops glacial acetic acid). For silica gel chromatography, the flash chromatography technique was used, with Merck silica gel 60 (230-400 mesh) and p.a. grade solvents unless otherwise noted.

The ¹H NMR and ¹³C NMR spectra were recorded in CDCl₃ on a Bruker Avance 400 (¹H 399.98 MHz; ¹³C 100.59 MHz) spectrometer. The chemical shifts are reported in ppm relative to CHCl₃ (δ 7.26) for ¹H NMR and the residual CDCl₃ (δ 77.0) for ¹³C NMR spectra. IR spectra were recorded on a Perkin–Elmer Spectrum One spectrometer. Optical rotations were obtained with a Perkin–Elmer 343 polarimeter. High resolution mass spectrometric data were obtained by the University of Oulu Analytical Services on Micromass LCT spectrometer. The elemental analyses were performed at the Analytical Services of the Department of Chemical Technology, Laboratory of Organic Chemistry.

4.1.1. (*S*)-(2-Oxoethoxy)succinic acid, dimethyl ester 9 and (*R*)-(2-oxoethoxy)succinic acid, dimethyl ester 16. To a solution of dimethyl (*S*)-malate 5 (5.60 g, 34.6 mmol, 1 equiv) in toluene (60 mL) was added allyl bromide (27.0 mL, 312 mmol, 9 equiv) and silver (I) oxide (8.01 g, 34.6 mmol, 1 equiv). After stirring for 2 h 20 min at rt the mixture was filtered through Celite and the solvent was evaporated to give the crude product **8** (6.72 g, 96%) as a pale yellow oil. The crude product was used directly in the next reaction.

The corresponding (R)-isomer was synthesized according to the method for preparation of **8** described above, with the exception that 1.05 equiv of Ag₂O was used and the reaction time was 5 h. Compound **19** was obtained in quantitative yield. The NMR data of **6** and its enantiomer match those reported in the literature.

To an ice-cold solution of **8** (2.01 g, 9.94 mmol, 1 equiv) and *N*-methylmorpholine *N*-oxide monohydrate (2.26 g, 16.72 mmol, 2 equiv) in THF/H₂O 3:1 (62 mL) was added OsO_4 (2.5 wt% solution in *t*-BuOH, 2.1 mL, 0.167 mmol, 2.0 mol%). The reaction mixture was stirred for 3 h at 0 °C and then allowed to warm to rt, overnight. Solid sodium hydrogen sulfite (2.10 g) was then added and the mixture was stirred for an additional 30 min at rt. The mixture was filtered through a pad of silica (2×20 mL THF rinse) and the solvents were evaporated to give the crude diol intermediate.

The crude diol (9.94 mmol, 1 equiv) was dissolved in THF/H₂O 3:1 (107 mL) and sodium periodate (4.25 g, 19.9 mmol, 2 equiv) was added. The mixture was stirred at rt for 30 min and then filtered through silica $(2 \times 20 \text{ mL})$ THF rinse). After concentration, brine (100 mL) and Et₂O (100 mL) were added. The layers were separated and the aqueous phase was extracted with Et₂O (2×75 mL). The combined organic layers were dried (Na₂SO₄), filtered and concentrated, and the crude product was purified by dry flash chromatography (4×8 cm silica, EtOAc/hexanes stepwise gradient from 0 to 100% EtOAc, final flush with pure MeOH) to give a first batch, 0.44 g (22%), of the aldehyde. The aqueous layer was further extracted with CH_2Cl_2 (6×100 mL). These extracts were combined, dried (Na₂SO₄) and concentrated to give pure aldehyde 9 (1.27 g, 63%). Total yield of 9 1.71 g, 84% (80% from 5). 9: viscous, colorless oil, $[\alpha]_{D}^{22} = -62.8$ (c 0.9, CH₂Cl₂), lit.¹⁰ $[\alpha]_D^{22} = -44.1$ (c 0.5, CHCl₃). A reliable comparison of the rotation in CHCl₃ could not be made because the aldehyde decomposed relatively easily in dry CHCl₃! The NMR spectral data corresponded to those reported in the literature¹⁰ but were best recorded with a trace of added Et₂N.

The aldehyde **9** could be stored in a frozen cyclohexane matrix at -20 °C for several months without appreciable decomposition. If necessary, it could be purified by Kugelrohr distillation (0.1 mmHg, bp 80–90 °C).

The corresponding enantiomeric aldehyde **16** was prepared using the same procedure from dimethyl (*R*)-malate in 61% overall yield. Its NMR spectra were identical to that of **9**. $[\alpha]_D^{22} = +56.8 \ (c0.8, CH_2Cl_2).$

4.1.2. (2S,2'S',2''S)- $\{2-[[2'-(1'',2''-Bis-methoxycarbonyl-ethoxy)-ethyl]-(1',2'-bis-methoxycarbonyl-ethyl)$ $amino]-ethoxy}-succinic acid hexamethyl ester (11) and$ <math>(2S,2'S)-2-[2'-(1',2'-bis-methoxycarbonyl-ethylamino)ethoxy]-succinic acid tetramethyl ester (12). To a solution of aldehyde 9 (480 mg, 2.35 mmol, 2.35 equiv) in MeOH (1.8 mL) at 0 °C was added L-aspartic acid dimethyl ester hydrochloride 10 (197.6 mg, 1.00 mmol, 1 equiv) and NaBH₃CN (94.3 mg, 1.50 mmol, 1.5 equiv) in three equal portions at 30 min intervals. After the last addition, the

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resulting suspension was allowed to warm to rt over 18 h. The mixture was cooled to 0 °C and more 9 (265 mg, 1.30 mmol, 1.3 equiv) dissolved in MeOH (1.2 mL) was added, followed by NaBH₃CN (94.3 mg, 1.50 mmol, 1.5 equiv). The reaction mixture was allowed to gradually warm to rt over 5 h. After a total reaction time of 48 h, the mixture was filtered and concentrated. The residue was dissolved in CH₂Cl₂ (50 mL) and washed with sat. NaHCO₃ (20 mL). The aqueous layer was extracted with CH₂Cl₂ $(3 \times 15 \text{ mL})$. The combined organic layers were dried (Na₂SO₄) and concentrated, and the crude product was purified by flash chromatography (silica gel, 85% EtOAc in hexanes+1.5% triethylamine). Yield of 11 428 mg, 80%. **11:** colorless viscous oil, $[\alpha]_{D}^{22} = -82.7$ (*c*1.0, CH₂Cl₂); IR (film) 3634, 3462, 2998, 2955, 1737, 1438, 1371, 1279, 1169, 1128, 1002, 849, 782, 674 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 4.29 (dd, J=5.1, 7.5 Hz, 2H), 3.96 (t, J= 7.5 Hz, 1H), 3.77 (s, 6H), 3.70 (s, 9H), 3.67 (td, J=6.3, 9.7 Hz, 2H), 3.68 (s, 3H), 3.44 (td, J=6.2, 9.5 Hz, 2H), 2.90-2.74 (m, 8H), 2.73 (dd, J=7.5, 16.1 Hz, 1H), 2.58 (dd, J=7.5, 15.9 Hz, 1H); ¹³C NMR (CDCl₃, 400 MHz) δ 172.6, 171.8, 171.7, 170.5, 75.5, 70.8, 61.6, 52.2, 52.1 (2), 51.9, 51.7, 51.6, 37.6, 35.6; ESI MS calcd for $(M^+ + H)$ $C_{22}H_{36}NO_{14}$ 538.2136, found 538.2147, $\Delta = 2.1$ ppm.

An alternative procedure to access both amination products proceeded as follows: to a solution of aldehyde **9** (436 mg, 2.13 mmol, 2.6 equiv) in MeOH (1.8 mL) at 0 °C was added **10** (160 mg, 0.81 mmol, 1 equiv) and NaBH₃CN (107 mg, 1.70 mmol, 2.1 equiv), both in three equal portions at 45 min intervals. The resulting suspension was stirred for 3 h at 0 °C, after which time it was allowed to warm to rt and stirred overnight. The reaction mixture was then treated and purified as described above to afford **11** (152 mg, 35%) and **12** (143 mg, 51%). Resubjection of **12** to the reductive amination conditions with 2 equiv formic acid (see the procedure below) afforded **11** in 40% yield based on **12**, raising the total yield of **11** to 55% (based on **10**).

Compound **12**: colorless oil, $[\alpha]_{22}^{22} = -47.3$ (*c* 0.7, CH₂Cl₂); IR (film) 3338, 2955, 1739, 1661, 1438, 1367, 1279, 1198, 1171, 1132, 998, 850, 785 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 4.34 (dd, *J*=4.8, 8.1 Hz, 1H), 3.78 (s, 3H), 3.77 (ddd, *J*=3.8, 6.1, 10.4 Hz, 1H), 3.75 (s, 3H), 3.73 (s, 3H), 3.70 (s, 3H), 3.70 (t, *J*=5.9 Hz, 1H), 3.57 (ddd, *J*=3.8, 9.8 Hz, 3.8 Hz, 1H), 2.92–2.70 (m, 3H), 2.76 (dd, *J*=6.9, 15.2 Hz, 2H), 2.67 (dd, *J*=6.9, 16.1 Hz, 1H), 1.87 (s, 1H); ¹³C NMR (CDCl₃, 400 MHz) δ 173.8, 171.8, 171.3, 170.5, 75.3, 70.9, 57.6, 52.2, 52.1, 52.0, 51.8, 47.2, 37.7, 37.6; ESI MS calcd for (M⁺ + Na) C₁₄H₂₃NO₉Na 372.1271, found 372.1264, Δ =1.7 ppm.

4.1.3. (2S,2'R,2''S)-{2-[[2'-(1",2"-Bis-methoxycarbonylethoxy)-ethyl]-(1',2'-bis-methoxycarbonyl-ethyl)amino]-ethoxy}-succinic acid hexamethyl ester (14) and (2S,2'R)-2-[2'-(1',2'-bis-methoxycarbonyl-ethylamino)ethoxy]-succinic acid tetramethyl ester (15). The hexamethyl ester 14 was synthesized as described above for 11 and 12. Yield of 14: 40% and 15: 42%. 14: colorless viscous oil, $[\alpha]_D^{22} = +1.8$ (*c* 0.6, CH₂Cl₂); IR (film) 3634, 3465, 3000, 2955, 1737, 1438, 1370, 1276, 1170, 1128, 1002, 850, 782 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 4.28 (dd, J=5.3, 7.3 Hz, 2H), 3.94 (t, J=7.4 Hz, 1H), 3.76 (s, 6H), 3.71 (s, 3H), 3.70 (s, 6H), 3.68 (s, 3H), 3.66 (td, J =6.0, 9.8 Hz, 2H), 3.44 (td, J = 6.4, 9.6 Hz, 2H), 2.89–2.74 (m, 8H), 2.73 (dd, J=7.4, 16.0 Hz, 1H), 2.59 (dd, J=7.4, 16.0 Hz, 1H); 13 C NMR (CDCl₃, 400 MHz) δ 172.6, 171.8, 171.7, 170.5, 75.5, 70.6, 61.4, 52.14, 52.12 (2), 51.9, 51.7, 51.6, 37.6, 35.7; ESI MS calcd for $(M^+ + H) C_{22}H_{36}NO_{14}$ 538.2169, found 538.2136, $\Delta = 6.1$ ppm. 15: colorless oil, $[\alpha]_{\rm D}^{22} = -25.6$ (c0.9, CH₂Cl₂); IR (film) 3419, 2956, 1739, 1646, 1439, 1368, 1279, 1198, 1172, 1133, 1044, 999, 857, 784 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 4.29 (dd, J=4.8, 8.1 Hz, 1H), 3.77 (ddd, J=4.2, 5.5, 9.5 Hz, 1H), 3.74 (s, 3H), 3.71 (s, 3H), 3.69 (s, 3H), 3.66 (s, 3H), 3.66 (t, J =6.5 Hz, 1H), 3.52 (ddd, J=3.8, 7.5, 9.7 Hz, 1H), 2.90-2.63 (m, 5H), 2.64 (dd, J = 6.5, 16.0 Hz, 1H), 2.05 (s, 1H); ¹³C NMR (CDCl₃, 400 MHz) δ 173.7, 171.7, 171.3, 170.5, 75.4, 71.0, 57.6, 52.1, 52.0, 51.9, 51.7, 47.3, 37.7, 37.6; ESI MS calcd for $(M^+ + H) C_{14}H_{24}NO_9 350.1451$, found 350.1443, $\Delta = 2.4$ ppm.

4.2. General procedure for reductive amination of 12 with formic acid

A solution of aldehyde **16** (584 mg, 2.86 mmol, 1.1 equiv) and tetramethylester **12** (900 mg, 2.58 mmol, 1 equiv) in MeOH (5.0 mL) was cooled to 0 °C. NaBH₃CN (180 mg, 2.86 mmol, 1.1 equiv) and formic acid (0.19 mL, 5.0 mmol, 2 equiv) were added at 0 °C in one portion. The suspension was stirred for 3 h, after which time it was allowed to warm to rt and stirred overnight. The reaction mixture was concentrated and the residue was partitioned between CH_2Cl_2 (50 mL) and sat. NaHCO₃ (30 mL). The layers were separated and the aqueous layer was extracted with CH_2Cl_2 (3×30 mL). The combined organic layers were dried (Na₂SO₄), concentrated, and the crude product was purified by flash chromatography (85% EtOAc in hexane + 1–1.5% triethylamine) to give **17** (550 mg, 40%) and recovered **12** (170 mg, 19%).

This procedure was also used for the conversion of the reductive amination product **12** into **11**.

4.2.1. (2*S*,2^{*T*}*S*,2^{*T*}*R*)-{2-[[2'-(1^{*T*},2^{*T*}-Bis-methoxycarbonylethoxy)-ethyl]-(1',2'-bis-methoxycarbonyl-ethyl)amino]-ethoxy}-succinic acid hexamethyl ester (17). Pale yellow oil, $[\alpha]_D^{22} = -27.61$ (*c* 0.7, CH₂Cl₂); IR (film) 3456, 3155, 3002, 2955, 1736, 1438, 1370, 1280, 1171, 1129, 1002, 909, 732 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 4.29 (dd, *J*=3.0, 5.0 Hz, 1H), 4.27 (dd, *J*=3.0, 5.0 Hz, 1H), 3.94 (t, *J*=7.5 Hz, 1H), 3.76 (s, 6H), 3.71 (s, 3H), 3.70 (s, 6H), 3.67 (s, 3H), 3.71–3.63 (m, 2H), 3.46–3.40 (m, 2H), 2.90– 2.73 (m, 8H), 2.72 (dd, *J*=7.6, 16.2 Hz, 1H), 2.58 (dd, *J*= 7.3, 15.9 Hz, 1H); ¹³C NMR (CDCl₃, 400 MHz) δ 172.6, 171.8, 171.7, 171.6, 170.5, 75.5, 70.8, 70.6, 61.5, 52.2, 52.1 (2), 51.9, 51.6, 51.5, 37.6, 35.6; ESI MS calcd for (M⁺ + Na) C₂₂H₃₅NO₁₄Na 560.1955, found 560.1951, Δ =0.8 ppm.

4.3. General procedure for saponification of the hexamethyl esters

To a solution of hexamethyl ester **11** (133 mg, 0.25 mmol, 1 equiv) in 1:1 MeOH/THF-solution (1.2 mL) was added 1 M NaOH (1.70 mL, 1.70 mmol, 6.8 equiv). The mixture

was stirred at rt for 18 h and then concentrated to give crude **2** as a tan solid. The excess NaOH was removed by precipitation of the product from H₂O (0.5 mL) by adding ethanol (2 mL), affording **2** (153 mg, quant., calculated as $C_{16}H_{17}NO_{14}Na_6 \cdot H_2O \cdot (C_2H_5OH)_{0.5}$) as a white solid.

The same procedure was used for the synthesis of **3** and **4**, starting from **14** and **17**, respectively.

4.3.1. (2*S*,2*'S*,2*"S*)-2-{2'-[[2'-(1",2"-Dicarboxy-ethoxy)ethyl]-(1',2'-dicarboxy-ethyl)-amino]-ethoxy}-succinic acid, hexasodium salt (2). White powder, melting range 340–385 °C (dec.), $[\alpha]_D^{22} = -13.7$ (*c* 0.5, H₂O); IR (KBr) 3429, 2967, 1591, 1409, 1308, 1196, 1107, 881, 686 cm⁻¹; ¹H NMR (D₂O, 400 MHz) δ 4.11 (dd, *J*=3.2, 9.6 Hz, 2H), 3.67–3.51 (m, 5H), 2.90–2.76 (m, 4H), 2.64–2.34 (m, 8H); ¹³C NMR (D₂O, 400 MHz) δ 180.1, 179.4, 79.3, 67.4, 64.4, 50.9, 41.5, 37.3; ESI MS calcd for (M⁺ – Na) C₁₆H₁₇NO₁₄Na₅ 562.0138, found 562.0117, Δ =3.6 ppm. Anal. calcd for C₁₆H₁₇NO₁₄Na₆·H₂O·(C₂H₅OH)_{0.5}: C, 32.60; H, 3.54; N, 2.24. Found: C, 32.27; H, 3.24; N, 1.95.

4.3.2. (2*S*,2*'R*,2*''S*)-2-{2'-[[2'-(1",2"-Dicarboxy-ethoxy)ethyl]-(1',2'-dicarboxy-ethyl)-amino]-ethoxy}-succinic acid, hexasodium salt (3). White powder, melting range 320–370 °C (dec.), $[\alpha]_D^{22} = -2.8$ (*c* 0.8, H₂O); IR (KBr) 3419, 2180, 1586, 1409, 1308, 1197, 1108, 877 cm⁻¹; ¹H NMR (D₂O, 400 MHz) δ 4.06 (dd, *J*=3.4, 9.2 Hz, 2H), 3.70 (dd, *J*=5.5, 8.1 Hz, 1H), 3.64 (ddd, *J*=5.0, 8.7, 9.7 Hz, 2H), 3.78–3.42 (m, 2H), 2.77–2.52 (m, 9H), 2.61 (dd, *J*= 3.4, 15.4 Hz, 2H), 2.46 (dd, *J*=9.2 Hz, 15.4 Hz, 2H), 2.38 (ddd, *J*=5.4, 9.8, 15.3 Hz, 1H); ¹³C NMR (D₂O, 400 MHz) δ 180.1, 179.4, 79.1, 67.2, 63.8, 50.7, 41.5, 35.2; ESI MS calcd for (M⁺ +H) C₁₆H₁₈NO₁₄Na₆ 586.0113, found 586.0095, *Δ*=3.2 ppm. Anal. calcd for C₁₆H₁₇NO₁₄Na₆. H₂O·(C₂H₅OH)_{0.5}: C, 32.60; H, 3.54; N, 2.24. Found: C, 32.52; H, 3.39; N, 2.07.

4.3.3. (2*S*,2*'S*,2*"R*)-2-{2*'*-[[2*'*-(1*"*,2*"*-Dicarboxy-ethoxy)ethyl]-(1*'*,2*'*-dicarboxy-ethyl)-amino]-ethoxy}-succinic acid, hexasodium salt (4). White powder, melting range 300–310 °C (dec.), $[\alpha]_D^{22} = -9.7$ (*c* 0.7, H₂O); IR (KBr) 3434, 2970, 1603, 1385, 1307, 1193, 1105, 880, 683 cm⁻¹; ¹H NMR (D₂O, 400 MHz) δ 4.08 (dd, *J*=3.5, 6.0 Hz, 1H), 4.06 (dd, *J*=3.6, 5.9 Hz, 1H), 3.65 (dd, *J*=5.6, 8.5 Hz, 1H), 3.63–3.59 (m, 1H), 6.31 (t, *J*=6.3 Hz, 2H), 3.46 (td, *J*=5.7, 1.0 Hz, 1H), 2.82–2.69 (m, 4H), 2.58 (dd, *J*=4.4, 15.2 Hz, 1H), 2.57 (dd, *J*=6.5, 15.3 Hz, 1H), 2.57 (dd, *J*=6.2, 15.4 Hz, 1H), 2.44 (dd, *J*=9.7, 15.2 Hz, 1H), 2.44 (dd, *J*= 9.6, 15.3 Hz, 1H), 2.40 (dd, *J*=5.9, 15.5 Hz, 1H); ¹³C NMR (D₂O, 400 MHz) δ 180.8, 180.2, 180.1, 179.8, 179.4, 179.3, 79.3, 79.2, 67.5, 67.1, 63.9, 50.7, 50.6, 41.6, 41.5, 36.3; ESI MS calcd for $(M^+ + H) C_{16}H_{18}NO_{14}Na_6$ 586.0113, found 586.0123, $\Delta = 1.7$ ppm. Anal. calcd for $C_{16}H_{17}NO_{14}Na_6 \cdot (H_2O)_{2.5} \cdot (C_2H_5OH)_{0.5}$: C, 31.25; H, 3.86; N, 2.14. Found: C, 31.05; H, 3.74; N, 2.02.

Acknowledgements

We thank Dr. Jari Koivisto and Dr. Juho Helaja for NMR services, Ms. Riitta Ilmoniemi and Ms. Outi Vilamo for early contributions to the synthesis, and Dr. Melanie Clarke for valuable comments on the manuscript. Dr. Vesa Myllymäki deserves special thanks for keeping the project smoothly on track.

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Tetrahedron

Tetrahedron 60 (2004) 10955-10966

Dinucleotides containing two allyl groups by combinations of allyl phosphotriesters, 5-allyl-, 2'-O-allyl- and 2'-arabino-O-allyl uridine derivatives as substrates for ring-closing metathesis

Philip Børsting, Morten Freitag and Poul Nielsen*

Nucleic Acid Center,[†] Department of Chemistry, University of Southern Denmark, 5230 Odense M, Denmark

Received 9 July 2004; revised 24 August 2004; accepted 9 September 2004

Available online 30 September 2004

Abstract—Five different dinucleotides, each containing two allyl groups in various positions, were prepared and studied as substrates for ring-closing metathesis reactions. These dinucleotides were designed from appropriate nucleoside building blocks combining four different positions for the allyl group; the allyl phosphotriester linkage, 5-allyl-2'-deoxyuridine, and *ribo*- as well as *arabino*-configured 2'-*O*-allyluridine. Thus, convenient procedures for these building blocks were developed. From the dinucleotides, two new cyclic nucleotide structures were obtained; one connecting two adjacent nucleobase moieties and the other forming an unsaturated four-carbon linkage between the phosphate moiety and the adjacent pyrimidine nucleobase. The latter cyclic dinucleotide was also prepared with a saturated four-carbon linkage using a tandem ring-closing metathesis—hydrogenation procedure. This compound was found to be significantly more stable towards a nucleophilic ring-opening than its unsaturated counterpart.

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1. Introduction

The application of ring-closing olefin metathesis (RCM) for the preparation of medium and large rings has been a major tool in organic and bioorganic chemistry.¹ In particular, the introduction of highly efficient and functional group tolerant ruthenium-based catalysts by Grubbs and co-workers has made metathesis technology generally attractive.² A large number of heterocyclic rings have been constructed by RCM,^{1,3} and recently, the application of RCM for the synthesis of phosphorus heterocycles has been specifically reviewed.⁴ In a bioorganic context, cyclic peptide or peptide mimetic structures have been achieved by RCM reactions.⁵ In view of this, we have focused on the application of RCM in nucleic acid chemistry,^{6–10} and as a result, conformationally restricted bi- and tricyclic nucleoside monomers⁶ as well as di- and trinucleotides with large cyclic structures have been achieved.^{7–10} In addition, other research groups have recently prepared different nucleoside derivatives by RCM based strategies.¹¹

In the studies of nucleic acid chemical biology, a significant number of conformationally restricted nucleic acid frag-ments have been designed.¹² As a relatively new approach, however, the relationship between structure and function of nucleic acid secondary and tertiary structures¹³ has motivated the preparation of nucleic acids with covalent intra- and interstrand linkages.¹⁴ Sekine and co-workers have introduced a number of cyclic nucleotide and dinucleotide structures for mimicking nucleic acid secondary structures.¹⁵ In these cases, linkages have been established between the 2'-position and the nucleobase in the two adjacent nucleosides in a dinucleotide or between the nucleobase and the adjacent phosphate moiety.¹⁵ Recently, another example of a conformationally restricted dinucleotide for this purpose with a linkage between the phosphate and the adjacent 5'-position has been introduced.¹⁶ In all these cases, the covalent linkages have been obtained by conventional phosphoramidite chemistry,¹⁷ disulfide bond formation and/or peptide/amide bond formation.^{14–16} We have concentrated on the development of a general methodology based on RCM for the construction of this type of conformationally restricted nucleic acid fragments. Thus, we have synthesised a series of diastereomeric dinucleotides with phosphotriester internucleotide linkages containing seven-membered rings.⁷ These were prepared by an RCM reaction on a dinucleotide substrate in which two terminal double bonds have been introduced by a 5'-C-vinyl moiety and an allyl phosphotriester linkage.⁷ In a

Keywords: Ring-closing metathesis; Nucleosides; Dinucleotides; Conformational restriction; Nucleic acid secondary structures.

^{*} Corresponding author. Tel.: +45 65502565; fax: +45 66158780; e-mail: pon@chem.sdu.dk

[†] Nucleic Acid Center is funded by the Danish National Research Foundation for studies on nucleic acid chemical biology.

^{0040–4020/\$ -} see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.023

similar way, a ring connecting the phosphotriester linkage in a dinucleotide with a nucleobase has been demonstrated,^{8,9} and two adjacent phorphortriester linkages have been linked in a cyclic trinucleotide structure.⁸ Thus, a tandem RCM and hydrogenation procedure efficiently mediated the preparation of cyclic dinucleotides with a butylene nucleobase phosphotriester connection, which was unobtainable by a conventional hydrogenation method.⁹ In this paper, some of these results are reported with all experimental details, and the general study on exploring the scope of ringclosing metathesis in the construction of cyclic dinucleotides with large rings is continued. Thus, four different positions of terminal double bonds were investigated, the appropriate nucleoside building blocks were prepared, and a selection of the possible dinucleotides were synthesised and investigated as substrates for RCM. Hereby, two cyclic dinucleotide structures were obtained.

2. Results and discussion

Four different positions for the allyl groups were deduced for incorporation into dinucleotides; the allyl phosphotriester linkage **A**, 5-allyl-2'-deoxyuridine **B**, 2'-O-allyluridine **C**, and 2'-*arabino*-O-allyluridine **D** (Fig. 1). Among these, **A** and a 5-allyluridine counterpart of **B** have been applied in our earlier studies.^{7,8} From these four different allyl positions, a series of 12 different dinucleotides might be constructed; (5'-3') **AB**, **AC**, **AD**, **BB**, **BC**, **BD**, **CB**, **CC**, **CD**, **DB**, **DC** and **DD**, as **A** cannot logically be placed in the 3'-end of the dinucleotides. Nevertheless, we decided to make only a section of five of these dinucleotides representing all of the four different allyl positions in at least two dinucleotides each. Thus, **AB**, ⁹ **AD**, **BB**, **CB** and **CD** were produced and investigated as substrates for RCM reactions.

In order to make dinucleotides with an allyl phosphotriester linkage **A**, the phosphoramidite **1** (Scheme 1) was formed in two steps from thymidine as shown in the literature.^{7,18}



Figure 1. The four different positions for allyl groups in this study.



Scheme 1. Reagents and conditions: (a) DMTCl, AgNO₃, pyridine, 84%; (b) TBDMSCl, AgNO₃, pyridine, 86%; (c) *p*-TsOH, CH₂Cl₂, CH₃OH, 0 °C, 88%; (d) TBDMSCl, AgNO₃, pyridine, 42%; (e) CEOP(N(*i*-Pr)₂)₂, 4,5dicyanoimidazole, CH₂Cl₂, CH₃CN, 75%. DMT=4,4'-dimethoxytrityl, TBDMS=*tert*-butyldimethylsilyl, CE=2-cyanoethyl.

Hereby, the appropriate building block for incorporation of A into dinucleotides by standard phosphoramidite chemistry¹⁷ was formed. For the 5-allyluracil alternative \mathbf{B} , 5-allyl-2'-deoxyuridine 2 was synthesised from 2'-deoxyuridine in two steps using a known procedure based on the formation of an organomercuri-intermediate followed by a transmetallation with Li₂PdCl₄ and allylation with allylchloride in methanol.¹⁹ Subsequently, 2 was converted to the 3'-protected derivative 5 in three steps, that is, selective 5^{7} -O-tritylation to give 3, 3'-O-silylation to give 4, both reactions accelerated by silver nitrate, and finally acidic detritylation to give the product 5 in 64% yield over the three steps (Scheme 1). Hereby, the building block for incorporating **B** in the 3'-end of the dinucleotides was formed.⁹ Furthermore, **2** was selectively protected at the 5'-position as a silvl ether to give 6 and converted to the 3'-phosphoramidite 7 using a phosphordiamidite reagent and 4,5-dicyanoimidazole²⁰ as an activating reagent. This formed the building block for incorporating \mathbf{B} in the 5'-end of the dinucleotides.

The formation of 2'-O-allyl-ribonucleosides, C, is well described in the literature.^{21–24} Thus, Sproat et al. synthesised 2'-O-allyluridine²¹ via a neutral palladium(0) catalysed allylation method²⁵ combined with a 5',3'-TIPDS protection of the two alcohols and a 4-O-(2,6-dichlorophenyl) protection of the uracil moiety.²¹ Later, it was demonstrated that the allylation can be performed with a simpler and cheaper 3-*N*-(4-*tert*-butylbenzoyl) group as protection of the uracil moiety.²² Recently, also a 4-*O*-(2,6-dimethylphenyl) protection of the uracil has been successfully applied in the same reaction.²³ On the other hand, a conventional Williamson type allylation, that is, allyl bromide and sodium hydride, has been used with 5'-O-MMT-3'-O-TBDMS-uridine as substrate.²⁴ Thus, an allylation can be performed without a protecting group for the uracil, and we decided to attempt the same reaction with TIPDS-protected uridine **8** (Scheme 2). However, this led to only 20% of the 2'-O-allyl derivative due to a partial basic cleavage of the silyl ether moieties. Subsequently, we



Scheme 2. Reagents and conditions: (a) BzCl, Bu₄NBr, CH₂Cl₂, aq Na₂CO₃, 77%; (b) allylethylcarbonate, tris(benzylidenacetone)dipalladium(0), bis(diphenylphosphino)butane, THF, 92%, reflux; (c) i. TBAF, THF, ii. aq NH₃, CH₃OH, 95%; (d) TBDMSCl, AgNO₃, pyridine, 79%; (e) CEOP(Cl)N(*i*-Pr)₂, EtN(*i*-Pr)₂, OH₂Cl₂, 65%; (f) i. TMSCl, Et₃N, CH₂Cl₂, ii. BzCl, EtN(*i*-Pr)₂, pyridine, iii. *p*-TsOH, CH₂Cl₂, 81%; (g) as (b), (h) as (c), 75%, two steps; (i) TBDMSCl, imidazole, DMF, 83%; (j) 80% aq AcOH, 73%. TBDMS = *tert*-butyldimethylsilyl, TMS = trimethylsilyl, CE=2-cyanoethyl.

decided to apply the neutral palladium(0) catalysed allylation method²⁵ with **8**. Thus, in this first attempt, nucleobase protection was avoided and, as expected, only the 3-N,2'-O-dialkylated product, or even the 3-N-monoalkylated product, was obtained. Therefore, we decided to follow the procedure by Sproat and co-workers²² but with a conventional 3-N-benzoyl protection of the uracil instead of the 3-N-(4-tert-butylbenzoyl) group. Thus, the TIPDSprotected uridine 8 was treated with benzoyl chloride in a known phase transfer reaction with DCM/Na₂CO₃(aq) and tetrabutylammonium bromide as phase transfer catalyst to give **9** in 77% yield (Scheme 2).^{26,27} The palladium(0) catalysed allylation procedure²⁵ applied with **9** afforded **10** in 92% yield. Subsequently, desilylation with TBAF in THF followed by debenzoylation with 25% ammonia in methanol gave 2'-O-allyluridine 11 in 95% yield, that is a conveniently 67% overall yield from 8. Finally, 11 was reprotected as a silvl ether at the 5'-position to give 12 and converted to the 3'-phosphoramidite 13 by standard methods (Scheme 2). Hereby, the building block for incorporating C in the 5'-end of the dinucleotides was formed.

The formation of 2'-O-allyl-arabinouridine, **D**, has also been described in the literature.²⁸ However, only the conventional Williamson ether formation has been demonstrated using 5',3'-bis-O-tetrahydropyranyl protected arabinouridine without protection of the uracil moiety to afford the 2'-O-allyl product in 40% yield.²⁸ Therefore, we decided to use

the same procedure for allylation as applied with the *ribo*configured uridine. Thus, the known TIPDS-protected *arabino*-configured uridine 14^{29} was treated with benzoyl chloride using the same phase transfer conditions as before but in this case only 34% yield of the 3-N-benzoylated product 15 was obtained due to a formation of a significant amount of the 3-N,2'-O-dibenzoylated product. Therefore, an in situ trimethylsilyl protection of the 2'-O-position was accomplished before a conventional N-benzoylation to give, after mild acidic desilylation, 15 in 81% yield over the three steps (Scheme 2). The palladium based allylation^{22,25} afforded 16, and after complete deprotection, the target 2'-O-allyl-arabinouridine 17 was obtained in 76% yield from 15. Hereby, 17 has been obtained in approximately 53% overall yield from arabinouridine, that is, 45% from uridine counting the two step conversion of the 2'-configuration.³⁰ In comparison, the reported method,²⁸ albeit in fewer steps, gave 17 in only approximately 17% overall yield from uridine. Finally, 17 was reprotected as its *bis*-silyl ether 18 in 83% yield and selectively deprotected using mild acid to give 19 in 73% yield as the appropriate building block for incorporating **D** in the 3'-end of the dinucleotides.

The five different protected dinucleotides **20–24** were made by the same general method using standard phosphoramidite chemistry¹⁷ (Schemes 3–5). Thus, the appropriate 5'-alcohols and 3'-phosphoramidites were coupled with 1*H*tetrazole as the activator followed by oxidation with *t*BuOOH. This oxidation reagent was efficient and easily handled, and the alternative standard reagent iodine was avoided due to the presence of double bonds. The general procedure afforded the dinucleotides in 60% to 100% yields. In all cases, the expected mixtures of two phosphorus epimers were obtained in approximately equimolar ratios as estimated from ¹H and ³¹P NMR.

Subsequently, all the protected dinucleotides **20–24** were investigated as substrates for RCM reactions using 5–15 mol% of Grubbs second-generation catalyst **X** (Mes = 2,4,6-trimethylphenyl, Cy = cyclohexyl),² in dichloromethane or 1,2-dichloroethane under reflux. We only investigated this catalyst, as this is by far the most generally successful according to the literature^{1–4} as well as the most successfully applied in our lab for the synthesis of other dinucleotides with large rings.^{7–10} We did not make a thorough investigation of a range of solvents, as dichloromethane and 1,2-dichloroethane have been the only successful solvents in our previous studies on nucleoside and nucleotide substrates.^{6–10}



Of the five different dinucleotide substrates, however, neither **20**, **21** or **22** were good substrates for RCM reactions under these conditions (Scheme 3). In the case of **21** the substrate was converted to a highly polar material indicating polymerisation products, whereas both **20** and **22** could be re-isolated from the reaction mixtures. In the latter case,



Scheme 3. Reagents and conditions: (a) i. 1*H*-tetrazole, CH₂Cl₂, CH₃CN, ii. *t*-BuOOH, CH₂Cl₂, CH₃CN, toluene, 100%; (b) 1*H*-tetrazole, CH₃CN, ii. *t*-BuOOH, CH₂Cl₂, toluene, 85%; (c) 1*H*-tetrazole, CH₂Cl₂, CH₃CN, ii. *t*-BuOOH, CH₂Cl₂, CH₃CN, toluene, 78%; (d) 5–15 mol% **X**, CH₂Cl₂/ CICH₂CH₂Cl/THF, reflux. TBDMS=*tert*-butyldimethylsilyl, CE=2cyanoethyl.

however, solubility problems forced us to investigate other solvents; toluene, ethyl acetate, acetonitrile/dichloromethane and THF, and with the latter, traces of a cyclised product was indicated from MS. On the other hand, the dinucleotide **23** was slowly converted to a ring-closed product **25** (Scheme 4). The best result was obtained with 5 mol% of the catalyst in dichloromethane at reflux from which **25** was isolated in 23% yield as a mixture of four stereoisomers in an approximately 1:1:4:6 ratio as deduced from the ³¹P NMR spectrum. This indicates that both *E* and *Z*-configured products were obtained and also that the ratios are dependent on the configuration of the phosphorus and that the two isomers in the substrate reacted in different rates. Thus, 27% of the starting material was re-isolated with an approximately 1:10 ratio of phosphorus epimers. Nevertheless, the formation of a ring-closed product was also confirmed by MS showing the expected loss of ethylene, and by ¹H NMR demonstrating the conversion of terminal to internal double bonds.

Finally, the dinucleotide 24^9 turned out to be the most efficient substrate for RCM. Thus, a treatment with 5 mol% of the catalyst in dichloromethane afforded 26 in 58% yield as a mixture of two phosphorus epimers in an equimolar ratio as deduced from ³¹P NMR (Scheme 5). Again the product was also confirmed by MS and ¹H NMR showing the loss of ethylene and the conversion of terminal to internal double bonds. Thus, this RCM reaction was performed very smoothly, as it has also been demonstrated for its 3'-ribonucleosidic analogue.⁸ Thus, **26** was the most easily obtained of the five projected cyclic dinucleotides, and subsequently, 26 has been further investigated for its stability towards ammonia.⁹ Thus, ammonia is a standard reagent used in the final deprotection step in oligonucleotide synthesis, and as an allylic phosphotriester, 26 was expected to be labile towards nucleophiles. Thus, in an analytical experiment treating 26 with 32% aqueous ammonia for 24 h at room temperature, the allylic phosphotriester was found to react completely with ammonia at the allylic CH₂ group located next to the phosphate to give a zwitterionic ammonia adduct 27 as a dinucleotide with an achiral phosphordiester linkage in which also both TBDMS ethers were hydrolysed.⁹ In order to increase the stability of the phosphotriester, we decided to prepare the saturated analogue of 26. Thus, 24 was applied in a tandem RCM and hydrogenation reaction³¹ by performing the RCM reaction as described earlier followed by hydrogenation of the reaction mixture in a Parr-bomb with 1000 psi H₂ at 50 °C. This gave the cyclic dinucleotide 28 in 63% yield as an equimolar mixture of two phosphorus epimers.⁹ The protecting groups were subsequently removed by an acidic treatment to give the cyclic dinucleotide **29** in a quantitative yield. This dinucleotide was considerably more stable towards ammonia, as experiments showed that treatment with 32% NH₃(aq) at room temperature for 24 h resulted in only 10% conversion to the dinucleotide **30** as indicated by ³¹P NMR and MS. A harsher treatment with 32% NH₃(aq) at 55 °C for 5 days resulted, however, in a complete conversion to the ammonia adduct 30.

In summary, the present study has demonstrated that not all the five dinucleotide substrates were found to be substrates for RCM reactions. It could be argued, of course, that a longer range of catalysts and solvents should be included in the study. However, Grubbs second-generation catalyst X was in all cases tested by $us^{6-10,32}$ found to be superior to the first-generation catalyst ((Cy₃P)₂RuCl₂CHPh).² The recently commercialised Hoveyda-Grubbs second-generation catalyst³³ was not available, when most of the presented experiments were conducted. On the other hand, not even this catalyst has been found to be superior to X with dinucleotide substrates in our lab.³² In the case of **21** Grubbs first-generation catalyst has been attempted, but only polymerisation was observed once again. Concerning solvents, we have attempted toluene in other studies⁶⁻⁸ but this solvent has been incompatible with our nucleotide substrates. Furthermore, the major subject of this study has



Scheme 4. Reagents and conditions: (a) i. 1*H*-tetrazole, CH₃CN, ii. *t*-BuOOH, CH₃CN, toluene, 60%; (b) 5 mol% X, CH₂Cl₂, 23%, reflux. TBDMS = *tert*-butyldimethylsilyl, CE = 2-cyanoethyl.

been to explore and compare different potential positions for introducing allyl groups into nucleotide building blocks and to compare different dinucleotides as RCM substrates. Thus, if a given substrate reacts very slowly with **X** or seems to prefer a polymerisation metathesis instead of a ring-closing reaction, no perspectives were seen in exploring a long range of reaction conditions. Therefore, we must conclude that only two of the five substrates, **23** and **24** could be transferred into the envisioned cyclic dinucleotides. Thus, the cyclic structures **25** and **26/29** can be used in the design of oligonucleotides that are conformationally restricted and potentially useful for targeting and mimicking nucleic acid secondary structures. When comparing the four different building blocks **A**–**D** in Figure 1, all four have been very conveniently obtained and incorporated into dinucleotides. Thus, as an interesting spinoff from this study, very convenient procedures for preparing the two epimeric 2'-O-allyluridine **11** and 2'-Oallyl-arabinouridine **17** have been developed. Especially in the latter case, our procedure has surpassed the existing literature method. However, the efficient RCM reactions have been obtained only with **A** and **B**. Thus, taking in account our former studies,^{7,8} the allyl group on the phosphotriester linkage is very reactive towards the catalyst and very well-positioned for performing ring-closing reactions. The 2'-O-ribo position of the allyl group, **C**, on



Scheme 5. Reagents and conditions: (a) 1*H*-tetrazole, CH₃CN, then *t*-BuOOH, toluene, 64%; (b) 5 mol% X, CH₂Cl₂, reflux, 58%; (c) 32% aq NH₃; (d) From 24, 5 mol% X, CH₂Cl₂, reflux, then 1000 psi H₂, 50 °C, 63%; (e) 90% aq TFA, 100%. TBDMS = *tert*-butyldimethylsilyl.

the other hand, seems to be problematic. However, in other studies by us³² and others,^{11e} this position has been used to form cyclic nucleotide structures with large rings, and the reasons for the failure of 20 and 22 might be found in the particular conformational properties of these substrates. Similarly, the 2'-O-arabino position of the allyl group, **D**, has failed as demonstrated by the dinucleotides 21 and 22. Nevertheless, reactivity between this allyl group and the catalyst is proven by the fact, that 21 is converted by polymerisation, probably by cross metathesis. Therefore, the reason for 21 being a bad substrate for RCM should be found in its conformational limitations making it impossible for the two double bonds (and their ruthenium [2+ 2]adducts) to approach and react in an intramolecular reaction. The 5-position of the pyrimidine, **B**, seems to be a good position for reaction with the catalyst X, and the possibility of making the very large ring structure in compound 25, the largest of the projected rings in this study, might be partly driven by an ability to stack between the two pyrimidines. Furthermore, the planar geometry of these probably reduces any steric problems compared to the other positions for allyl groups. Nevertheless, also in this case, the reactivity is limited by geometry as illustrated by the fact that different reactivity was observed for the two phosphorus epimers. Thus, even though the selection of five dinucleotides that we choose for RCM studies led to the examination of each of the four allyl positions A-D, we cannot exclude the possibility that very efficient substrates and subsequent cyclic dinucleotides could be found with other combinations of A-D.

As evident from the present results, the butylene nucleobase to phosphotriester connection obtained in **29** has been efficiently obtained from the tandem RCM hydrogenation strategy. Further effort can now be put into separating the two phosphorus epimers of this compound and incorporating these into oligonucleotides. The stability of **29** towards ammonia supports these plans. Of the long range of possible cyclic dinucleotides investigated by this study, **29** is the most easily obtained and the obvious choice for further studies. Furthermore, the synthesis of other cyclic dinucleotides, constructed from allyl phosphotriester linkages and/or other positions for terminal alkene moieties^{10,32} are in progress.

3. Conclusion

Five different dinucleotides combining four nucleotide building blocks with different allyl substituents have been explored as substrates for RCM reactions. Two cyclic dinucleotide structures were obtained and the one further elaborated to give a stable butylene connection between a nucleobase and a phosphate internucleotide linkage. Even though not all projected cyclisations could be performed in practice, more general knowledge within the scopes and limitation of the RCM based methodology towards conformationally restricted nucleic acid fragments has been obtained. Thus, we envision a large potential for constructing a plethora of cyclic dinucleotide structures mimicking a large range of nucleic acid secondary structures, and we expect the present RCM methodology to be a general future tool in nucleic acid chemical biology.

4. Experimentals

4.1. General

All commercial reagents were used as supplied. When necessary, reactions were performed under an atmosphere of nitrogen. Column chromatography was carried out on glass columns using silica gel 60 (0.040-0.063 mm). NMR spectra were recorded on a Varian Gemini 2000 spectrometer. ¹H NMR spectra were recorded at 300 MHz, ¹³C NMR spectra at 75.5 MHz, and ³¹P NMR spectra at 121.5 MHz. Values for δ are in ppm relative to tetramethylsilane as internal standard or 85% H₃PO₄ as external standard. Fastatom bombardment mass spectra (FAB-MS) were recorded in positive ion mode on a Kratos MS50TC spectrometer and MALDI mass spectra were recorded on an Ionspec Ultima Fourier Transform mass spectrometer. Microanalyses were performed at The Microanalytical Laboratory, Department of Chemistry, University of Copenhagen. Assignments of NMR spectra are based on 2D spectra and follow standard nucleoside style; that is, the carbon next to the nucleobase is assigned C-1^{\prime}. For dinucleotides, the upper (5^{\prime}) nucleotide is depicted U1 or T1 and the lower nucleoside U2 or T2.

4.1.1. Preparation of 5-allyl-2'-deoxy-5'-O-dimethoxytrityluridine (3). 5-Allyl-2'-deoxyuridine 2^{19} (0.700 g, 2.61 mmol) was coevaporated twice with anhydrous pyridine and redissolved in anhydrous pyridine (40 mL) in a darkened flask. AgNO₃ (0.557 g, 3.28 mmol) was added and the mixture was stirred for 5 min. DMTCl (1.099 g, 3.24 mmol) was added and the reaction mixture was stirred for 4 h, concentrated under reduced pressure and coevaporated with toluene. The residue was dissolved in ethyl acetate (50 mL), washed with a saturated aqueous solution of NaHCO₃ (50 mL) and brine $(2 \times 50 \text{ mL})$, dried (Na₂SO₄) and concentrated under reduced pressure. The residue was purified by column chromatography (1–4% CH₃OH and 1% pyridine in CH_2Cl_2) to give the product as a white foam (1.280 g, 86%) (Found C, 68.59; H, 6.07; N, 4.75%; C₃₃H₃₄N₂O₇·1/2H₂O requires C, 68.38; H, 6.08; N, 4.83%); mp = 79–82 °C; $R_f 0.23$ (5% CH₃OH in CH₂Cl₂); ¹H NMR (CDCl₃) & 2.24–2.44 (2H, m, H-2'), 2.51–2.77 (2H, m, 5-CH₂), 3.33-3.43 (2H, m, H-5'), 3.78 (6H, s, OCH₃), 4.06 (1H, m, H-4'), 4.53 (1H, m, H-3'), 4.76–4.82 (2H, m, $CH=CH_2$), 5.60 (1H, m, $CH=CH_2$), 6.41 (1H, m, H-1'), 6.83 (4H, m, Ph), 7.16-7.41 (9H, m, Ph), 7.47 (1H, s, H-6), 9.33 (1H, s, NH); ¹³C NMR (CDCl₃) δ 30.5 (5-CH₂), 40.8 (C-2'), 55.2 (OCH₃), 63.5 (C-5'), 72.4 (C-3'), 84.8 (C-1'), 86.1 (Ph), 86.8 (C-4'), 113.23 (Ph), 113.71 (C-5), 116.65 (CH=CH₂), 125.3, 127.1, 127.9, 128.1, 128.2, 129.0, 130.1, 134.3, 135.4 (Ph), 135.4 (CH=CH₂), 136.1 (C-6), 144.3 (Ph), 150.4 (C-2), 158.6 (Ph), 163.1 (C-4); m/z FAB 570 (M).

4.1.2. Preparation of 5-allyl-3'*-O-tert***-butyldimethylsilyl-**2'**-deoxy-5'***-O***-dimethoxytrityluridine** (4). Compound 3 (0.490 g, 0.86 mmol) was dissolved in anhydrous pyridine (30 mL) in a darkened flask. AgNO₃ (0.165 g, 0.97 mmol) was added and the mixture was stirred for 5 min. TBDMSCI (0.145 g, 0.96 mmol) was added and the reaction mixture was stirred overnight, concentrated under reduced pressure and coevaporated with toluene. The residue was dissolved in ethyl acetate (50 mL), washed with a saturated aqueous

solution of NaHCO₃ (50 mL) and brine (50 mL), dried (Na_2SO_4) and concentrated under reduced pressure. The residue was purified by column chromatography (1-5%)CH₃OH and 1% pyridine in CH₂Cl₂) to give the product as a white foam (0.495, 86%) (Found C, 66.96; H, 7.09; N, 3.84%; C₃₉H₄₈N₂O₇Si H₂O requires C, 66.64; H, 7.16; N, 3.98%); mp = 60–64 °C; R_f 0.60 (5% CH₃OH in CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.02–0.16 (6H, m, Si(CH₃)₂), 0.81–0.97 (9H, m, C(CH₃)₃), 2.17–2.39 (2H, m, H-2[']), 2.60–2.88 (2H, m, 5-CH₂), 3.28–3.48 (2H, m, H-5[']), 3.82 (6H, s, OCH₃), 4.00 (1H, m, H-4'), 4.52 (1H, m, H-3'), 4.80–4.86 (2H, m, $CH=CH_2$), 5.67 (1H, m, $CH=CH_2$), 6.38 (1H, t, J=6.7 Hz, H-1'), 6.82 (4H, m, Ph), 7.16–7.51 (9H, m, Ph), 7.55 (1H, s, H-6), 8.98 (1H, s, NH); ¹³C NMR (CDCl₃) δ -4.9, -4.7 (Si(CH₃)₂), 17.9 (C(CH₃)₃), 25.7 (C(CH₃)₃), 30.6 (C-2') 41.3 (5-CH₂), 55.2 (OCH₃), 62.8 (C-5'), 72.1 (C-3'), 84.8 (C-1[']), 86.7 (Ph), 87.4 (C-4[']), 113.2 (Ph), 113.48 (C-5), $116.6 (CH = CH_2), 127.1, 127.7, 127.8, 127.9, 128.1, 128.2,$ 129.0, 129.1, 130.0, 134.4, 135.4 (Ph), 135.5 (CH=CH₂), 136.1 (C-6), 144.3 (Ph), 150.1 (C-2), 158.6 (Ph), 163.0 (C-4); *m/z* FAB 684 (M).

4.1.3. Preparation of 5-allyl-3'-O-tert-butyldimethylsilyl-2'-deoxyuridine (5). Compound 4 (0.450 g, 0.66 mmol) was dissolved in a mixture of CH₃OH and CH₂Cl₂ (2:3 v/v, 50 mL) and the solution was stirred at 0 °C. A solution of ptoluenesulfonic acid monohydrate (0.155 g, 0.82 mmol) in a mixture of CH₃OH and CH₂Cl₂ (2:3 v/v, 5 mL) was added over 5 min. The reaction mixture was stirred for 30 min and quenched by the addition of saturated aqueous ammonia (0.5 mL). The mixture was concentrated under reduced pressure and the residue was dissolved in CH₂Cl₂ (50 mL), washed with water $(3 \times 50 \text{ mL})$, dried (MgSO₄) and concentrated under reduced pressure. The residue was purified by column chromatography (1-3% CH₃OH in CH_2Cl_2) to give the product as a white foam (0.220 g, 88%); $R_{\rm f}$ 0.22 (5% CH₃OH in CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.01– 0.13 (6H, m, Si(CH₃)₂), 0.83–0.96 (9H, m, C(CH₃)₃), 2.19– 2.41 (2H, m, H-2'), 3.06-3.09 (2H, m, 5-CH₂), 3.72-3.76 (2H, m, H-5'), 3.92 (1H, m, H-4'), 4.50 (1H, m, H-3'), 5.11– 5.18 (2H, m, CH=CH₂), 5.85 (1H, m, CH=CH₂), 6.15 (1H, t, J=6.7 Hz, H-1'), 7.34 (1H, s, H-6), 9.01 (1H, s, NH);¹³C NMR (CHCl₃) δ -4.9, -4.7 Si(CH₃)₂), 18.0 $(C(CH_3)_3)$, 25.7 $(C(CH_3)_3)$, 30.5 (C-2'), 40.4 $(5-CH_2)$, 62.0 (C-5'), 71.6 (C-3'), 87.2 (C-1'), 87.6 (C-4'), 113.5 (C-5), 117.6 (CH=CH₂), 134.2 (CH=CH₂), 137.6 (C-6), 150.2 (C-2), 163.1 (C-4); HiRes MALDI FT-MS m/z (M+ Na) found/calcd 405.1828/405.1822.

4.1.4. Preparation of 5-allyl-5'-*O-tert***-butyldimethylsilyl-2'-deoxyuridine (6).** 5-Allyl-2'-deoxyuridine 2^{19} (0.979 g, 3.65 mmol) was coevaporated with anhydrous pyridine and redissolved in anhydrous pyridine (36 mL) in a darkened flask. AgNO₃ (0.503 g, 2.96 mmol) was added and the mixture was stirred for 5 min. TBDMSCl (0.235 g, 1.56 mmol) was added and the reaction mixture was stirred for 2 h. Another portion of TBDMSCl (0.339 g, 2.25 mmol) was added in 3 parts during 3 h, and the reaction was stirred for 5 h and quenched by the addition of CH₃OH (1 mL). The reaction mixture was concentrated under reduced pressure and coevaporated with toluene. The residue was dissolved in CH₂Cl₂ (80 mL) and washed with a saturated aqueous solution of NaHCO₃ (40 mL) and brine (40 mL), dried

 $(MgSO_4)$ and concentrated under reduced pressure. The residue was purified by dry column chromatography (2-4%) CH_3OH in CH_2Cl_2) to give the product as white foam (0.597 g, 42%) (Found C, 55.96; H, 8.02; N, 7.19%; C₁₈H₃₀N₂O₅Si · 1/4H₂O requires C, 55.86; H, 7.94; N, 7.23%); $R_{\rm f}$ 0.48 (10% CH₃OH in CH₂Cl₂); ¹H NMR (CDCl₃) δ 0.07–0.10 (6H, m, Si(CH₃)₂), 0.91 (9H, s, C(CH₃)₃), 2.08 (1H, m, H-2'), 2.40 (1H, m, H-2"), 3.05-3.07 (2H, m, 5-CH₂), 3.81–3.83 (2H, m, H-5[']), 4.04 (1H, m, H-4'), 4.40 (1H, m, H-3'), 5.06–5.12 (2H, m, CH=CH₂), 5.86 (1H, m, CH=CH₂), 6.34 (1H, m, H-1'), 7.42 (1H, s, H-6), 9.12 (1H, s, NH); ¹³C NMR (CDCl₃) δ -5.3, -5.1 (Si(CH₃)₂), 18.5 (C(CH₃)₃), 26.1 (C(CH₃)₃), 31.1 (C-2'), 41.1 (5-CH₂), 63.8 (C-5'), 72.7 (C-3'), 85.3 (C-1'), 87.2 (C-4'), 113.5 (C-5), 116.5 (CH=CH₂), 134.7 (CH=CH₂), 136.2 (C-6), 150.5 (C-2), 163.2 (C-4); HiRes MALDI FT-MS *m*/*z* (M+Na) found/calcd 405.1836/405.1816.

4.1.5. Preparation of 5-allyl-5'-O-(tert-butyldimethylsilyl)uridine-3'-O-(N,N-diisopropyl)-(2-cyanoethyl)phosphoramidite (7). Compound 6 (0.152 g, 0.40 mmol) was dissolved in anhydrous CH_2Cl_2 (2.5 mL) and a 0.5 M solution of 4,5-dicyanoimidazole (0.55 mL, 0.28 mmol) in CH₃CN was added. 2-Cyanoethyl-*N*,*N*,*N*',*N*'-tetraisopropylphosphordiamidite (0.126 g, 0.42 mmol) was added dropwise over 5 min and the reaction mixture was stirred at room temperature for 30 min. The mixture was diluted with CH_2Cl_2 (50 mL) and washed with a saturated aqueous solution of NaHCO₃ (25 mL) and brine (25 mL). The combined aqueous phases were extracted with CH₂Cl₂ (30 mL) and the combined organic phases were dried (Na₂SO₄) and concentrated under reduced pressure. The residue was purified by dry column chromatography (50-60% ethyl acetate and 1% triethylamine in petrol ether) to give the product as a colourless oil and an epimeric mixture (0.173 g, 75%); R_f 0.36, 0.49 (75% ethyl acetate in petrol ether); ¹H NMR (CDCl₃) δ 0.10–0.11 (6H, m, Si(CH₃)₂), 0.88–0.98 (9H, m, C(CH₃)₃), 1.17–1.22 (12H, m, CH(CH₃)), 2.01–2.11 (1H, m, H-2'), 2.39–2.55 (1H, m, H-2"), 2.61–2.66 (2H, m, CH₂CN), 3.06–3.08 (2H, m, 5-CH₂), 3.56–3.91 (6H, m, H-5['], CH₂OP, 2×CH(CH₃)), 4.09–4.18 (1H, m, H-4'), 4.48–4.54 (1H, m, H-3'), 5.06– 5.13 (2H, m, CH= CH_2), 5.80–5.91 (1H, m, CH= CH_2), 6.30–6.34 (1H, m, H-1'), 7.41, 7.45 (1H, 2 s, H-6), 8.27 (1H, br s, NH); ³¹P NMR (CDCl₃) δ 149.45, 149.89; *m/z* FAB 583 (M + H).

4.1.6. Preparation of 2'-O-allyl-3-N-benzoyl-3',5'-O,O-(1,1,3,3-tetraisopropyldisiloxan-1,3-diyl)uridine (10). Compound 9^{27} (0.496 g, 0.840 mmol) and allylethylcarbonate³⁴ (0.351 g, 2.70 mmol) was dissolved in anhydrous THF (4 mL). A solution of tris(dibenzylidenacetone)dipalladium(0) (0.012 g, 0.013 mmol, 1.5 mol%) and bis(diphenylphosphino)butane (0.038 g, 0.09 mmol, 10 mol%) in anhydrous THF (1 mL) was added. The reaction mixture was stirred at reflux for 45 min and then concentrated under reduced pressure. The residue was purified by column chromatography (4-50% ethyl acetate in petrol ether) to give the product as a white foam (0.487 g, 92%) (found: C, 59.19; H, 7.42; N, 4.43% C₃₁H₄₆N₂O₈Si₂ requires: C, 59.02; H, 7.35; N, 4.44%); R_f 0.71 (75% ethyl acetate in petrol ether); ¹H NMR (CDCl₃) δ 0.85–1.26 (28H, m, SiCH(CH₃)₂), 3.92 (1H, s, H-4'), 4.00 (1H, d, J = 14.0 Hz,

H-5'), 4.21–4.33 (5H, m, H-2', H-3', H-5", 2'-OCH₂), 5.15 (1H, dd, J=1.5, 12.4 Hz, CH= CH_2), 5.31 (1H, dd, J=1.5, 17 Hz, CH= CH_2), 5.79 (1H, d, J=8.2 Hz, H-5), 5.76 (1H, s, H-1'), 5.87 (1H, m, CH= CH_2), 7.48–7.95 (m, 5H, Ph), 8.04 (1H, d, J=8.2 Hz, H-6); ¹³C NMR (CDCl₃) δ 12.4, 12.8, 13.1, 13.4 (SiCH(CH₃)₂), 16.7, 16.8, 17.0, 17.1, 17.3, 17.3, 17.4, 17.5, 17.9 (SiCH(CH₃)₂), 59.4 (C-5'), 67.9 (C-3'), 71.2 (2'-OCH₂), 81.1 (C-4'), 82.0 (C-2'), 89.2 (C-1'), 101.45 (C-5), 117.4 (CH= CH_2), 129.2, 130.5, 131.3, 134.1 (Ph), 135.2 (CH= CH_2), 139.2 (C-6), 148.9 (C-2), 162.2 (C-4), 168.7 (C=O); m/z FAB 631 (M+H).

4.1.7. Preparation of 2'-O-allyluridine (11). Compound 10 (2.40 g, 3.80 mmol) was dissolved in THF (10 mL) and the solution was stirred at room temperature. A 1 M solution of TBAF in THF (8.6 mL, 8.6 mmol) was added over 5 min and the mixture was stirred for 30 min. A mixture of pyridine, CH₃OH and water (3:1:1 v/v, 10 mL) was added and the combined mixture was poured on 13 g amberlite IR-120[®] in pyridine, CH₃OH and water (3:1:1 v/v, 60 mL). The suspension was stirred for 30 min and filtered. The filtrate was concentrated under reduced pressure and the residue was dissolved in CH₃OH (50 mL) and added 25% aqueous ammonia (10 mL). The reaction mixture was stirred for 1 h 30 min, concentrated under reduced pressure and coevaporated with anhydrous ethanol. The residue was purified by dry column chromatography (0-10% CH₃OH in CH_2Cl_2) to give the product as a hygroscopic white foam (1.030 g, 95%); *R*_f 0.55 (10% CH₃OH in CH₂Cl₂); ¹H NMR (CD₃OD) § 3.71–3.89 (2H, m, H-5'), 3.90–4.02 (2H, m, H-2', H-4'), 4.20–4.24 (3H, m, H-3', 2'-OCH₂), 5.13–5.34 (2H, m, CH=CH₂), 5.68 (1H, d, J=7.9 Hz, H-5), 5.87-5.98 (1H, m, CH=CH₂), 5.95 (1H, d, J=3.8 Hz, H-1'), 8.07 (1H, d, J=7.9 Hz, H-6); ¹³C NMR (CD₃OD) δ 61.7 (C-5'), 70.0 (C-3'), 72.4 (2'-OCH₂), 82.5 (C-4'), 86.2 (C-2'), 89.2 (C-1'), 102.5 (C-5), 117.9 (CH=CH₂), 135.7 (CH=CH₂), 142.5 (C-6), 152.2 (C-2), 166.2 (C-4); HiRes MALDI FT-MS m/z (M+Na) found/calcd 307.0898/307.0901.

4.1.8. Preparation of 2'-O-allyl-5'-O-(tert-butyldimethylsilyl)uridine (12). Compound 11 (0.312 g, 1.10 mmol) was dissolved in anhydrous pyridine (9 mL). TBDMSCl (0.213 g, 1.41 mmol) and AgNO₃ (0.740 g, 4.36 mmol) were added, and the reaction mixture was stirred at room temperature for 24 h. The reaction was guenched by the addition of CH₃OH (1 mL) and the mixture was concentrated under reduced pressure and coevaporated with toluene. The residue was dissolved in ethyl acetate (75 mL) and washed with a saturated aqueous solution of NaHCO₃ (50 mL) and brine (50 mL). The aqueous phase was extracted with ethyl acetate (50 mL), and the combined organic phases were dried (Na₂SO₄) and concentrated under reduced pressure. The residue was purified by dry column chromatography (0-6% CH₃OH in CH₂Cl₂) to give the product as a white foam (0.344 g, 79%) (Found C, 54.61; H, 7.70; N, 6.81%; C₁₈H₃₀N₂O₆Si requires C, 54.25; H, 7.59; N, 7.03%); $R_{\rm f}$ 0.67 (10% CH₃OH in ethyl acetate); ¹H NMR $(CDCl_3) \delta 0.12 (6H, s, Si(CH_3)_2), 0.93 (9H s, C(CH_3)_3),$ 2.64 (1H, d, J=7.9 Hz, 3'-OH), 3.84–3.92 (2H, m, H-5'), 4.01–4.07 (2H, m, 2'-OCH₂), 4.20–4.24 (2H, m, H-2', H-4'), 4.40 (1H, m, H-3'), 5.23–5.33 (2H, m, CH=CH₂), 5.67 (1H, d, J=8.3 Hz, H-5), 5.89 (1H, m, CH=CH₂), 5.98 (1H, s, H-1[']), 8.06 (1H, d, J = 8.3 Hz, H-6), 9.07 (1H, s, NH); ¹³C

NMR (CDCl₃) δ -5.6 (Si(CH₃)₂), 18.4 (*C*(CH₃)₃), 25.9 (C(CH₃)₃), 61.3 (C-5'), 67.9 (C-3'), 71.4 (2'-OCH₂), 81.5 (C-4'), 84.6 (C-2'), 87.3 (C-1'), 102.0 (C-5), 118.7 (CH=CH₂), 133.3 (CH=CH₂), 140.0 (C-6), 150.1 (C-2), 163.3 (C-4); HiRes MALDI FT-MS *m*/*z* (M+Na) found/ calcd 421.1750/421.1765.

4.1.9. Preparation of 2'-O-allyl-5'-O-(*tert*-butyldimethylsilyl)uridine-3'-O-(N,N-diisopropyl)-(2-cyanoethyl)phosphoramidite (13). Compound 12 (0.123 g, 0.31 mmol) was dissolved in anhydrous CH₂Cl₂ (1.5 mL) and stirred at 0 °C. N,N-Diisopropylethylamine (0.30 mL, 0.227 g, 1.76 mmol) was added and chloro-N,N-diisopropylamino-(2-cyanoethyl)phosphine (0.20 g, 0.85 mmol) was added. The reaction mixture was stirred at 0 °C for 10 min and at room temperature for 5 h. The reaction mixture was cooled to 0 °C and another portion of chloro-N.N-diisopropylamino-(2-cyanoethyl)phosphine (0.20 g, 0.85 mmol) was added. The mixture was stirred at room temperature for 1.5 h and then quenched by the addition of water (1 mL). The mixture was diluted with ethyl acetate (10 mL) and washed with a 5% aqueous solution of NaHCO₃ (2× 25 mL) and brine (25 mL), dried (Na₂SO₄) and concentrated under reduced pressure. The residue was purified by column chromatography (0.5% triethylamine and 50% ethyl acetate in petrol ether) to give the product as yellow foam and an epimeric mixture (0.120 g, 65%); $R_{\rm f}$ 0.36 (75% ethyl acetate in petrol ether); ¹H NMR (CDCl₃) δ 0.10–0.13 (6H, m, Si(CH₃)₂), 0.94–0.97 (9H, m, C(CH₃)₃), 1.11–1.23 (12H, m, CH(CH₃)₂), 2.59–2.66 (2H, m, CH₂CN), 3.59–3.70 (4H, m, CH₂OP, CH(CH₃)₂), 3.70–3.84 (2H, m, H-5[']), 3.84–4.01 (2H, m, 2'-OCH₂), 4.17–4.32 (3H, m, H-2', H-3', H-4'), 5.16–5.33 (2H, m, CH=CH₂), 5.64–5.68 (1H, m, H-5), 5.85-6.92 (1H, m, CH=CH₂), 6.02-6.06 (1H, m, H-1'), 7.94–7.98 (2H, m, H-6); ³¹P NMR (CDCl₃) δ 150.91, 150.97; *m*/*z* FAB 599 (M+H).

4.1.10. Preparation of 3-N-benzoyl-1-(3',5'-0,0-(1,1,3,3tetraisopropyldisiloxan-1,3-diyl)-β-D-arabinofuranosyl) uracil (15). Compound 14²⁹ (5.33 g, 11.0 mmol) was dissolved in anhydrous CH₂Cl₂ (100 mL), stirred at 0 °C and added triethylamine (7.63 mL, 54.8 mmol). TMSCl (4.17 mL, 32.9 mmol) was added over 5 min and the reaction mixture was stirred at room temperature for 2 h. The mixture was cooled to 0 °C and added a 1.0 M aqueous solution of NaHCO₃ (100 mL). The organic phase was separated, dried (Na₂SO₄) and concentrated under reduced pressure. The residue was redissolved in anhydrous pyridine (50 mL) and stirred at 0 °C. N,N-diisopropylethylamin (5.62 mL, 32.9 mmol) was added and the mixture was stirred for 10 min. Benzoyl chloride (3.81 mL, 32.9 mmol) was added and the reaction mixture was stirred at room temperature for 17 h. The reaction mixture was cooled to 0 °C and a 1.0 M aqueous solution of NaHCO₃ (100mL) was added. The organic phase was dried (Na₂SO₄), concentrated under reduced pressure and coevaporated with toluene (3 \times 10 mL). The residue was redissolved in CH₂Cl₂ (100 mL) and stirred at 0 °C. A solution of p-toluenesulfonic acid (4.72 g, 27.4 mmol) in THF (50 mL) was added and the reaction mixture was stirred for 10 min. Triethylamine (3.05 mL, 21.9 mmol) was added and the mixture was stirred for 15 min. A 1.0 M aqueous solution of NaHCO₃ (100mL) was added, and the organic phase was separated, dried (Na₂SO₄) and concentrated under reduced pressure. The residue was purified by column chromatography (5–40% ethyl acetate in petrol ether) to give the product as a white foam (5.21 g, 81%); mp=50–53 °C; ¹H NMR (CDCl₃) δ 0.97–1.13 (28H, m, CH(CH₃)₂), 3.31 (1H, d, *J*=4.6 Hz, 2'-OH), 3.73 (1H, m, H-4'), 4.05 (1H, dd, *J*= 2.5, 13.5 Hz, H-5'), 4.12 (1H, dd, *J*=1.4, 13.5 Hz, H-5'), 4.16 (1H, t, *J*=8.6 Hz, H-3'), 4.42 (1H, m, H-2'), 5.75 (1H, d, *J*=8.2 Hz, H-5), 6.03 (1H, d, *J*=6.2 Hz, H-1'), 7.80 (1H, d, *J*=8.2 Hz, H-6), 7.93–7.42 (5H, m, Ph); ¹³C NMR (CDCl₃) δ 12.2, 12.8, 12.9, 13.3 (CH(CH₃)₂), 16.7, 16.8, 16.9, 17.1, 17.2, 17.3, 17.4 (CH(CH₃)₂), 60.0 (C-5'), 72.4 (C-3'), 75.4 (C-2'), 80.7 (C-4'), 101.3 (C-5), 128.3, 130.3, 131.3, 134.9 (Ph), 140.2 (C-6), 149.9 (C-2), 162.1 (C-4), 168.5 (C=O).

4.1.11. Preparation of $1-(2'-O-allyl-\beta-D-arabinofurano$ syl)uracil (17). Compound 15 (5.21 g, 8.82 mmol) was dissolved in anhydrous THF (35 mL) and added allylethylcarbonate³⁴ (1.51 mL, 13.2 mmol), tris(dibenzylideneacetone)dipalladium(0) (0.121 g,0.132 mmol) and 1,4-bis(diphenylphosphino)butane (0.376 g, 0.88 mmol). The reaction mixture was stirred at reflux for 3 h and then concentrated under reduced pressure. The crude compound 16 was dissolved in THF (50 mL) and a 1.0 M solution of TBAF in THF (19.4 mL, 19.4 mmol) was added over 5 min. The reaction mixture was stirred for 15 min and then concentrated under reduced pressure. The residue was dissolved in CH₃OH (35 mL) and the solution was stirred at 0 °C. NH₃(g) was bubbled through the solution for 45 min. The mixture was concentrated under reduced pressure and the residue was dissolved in a mixture of pyridine, CH₃OH and water (3:1:1 v/v, 80 mL). The mixture was stirred with Dowex[®] (50 W×2) for 2 h and the resin was removed by filtration. The filtrate was concentrated under reduced pressure and coevaporated with toluene. The residue was purified by column chromatography (0-7% CH₃OH in CH_2Cl_2) to give the product as white solid (1.90 g, 76%); mp = 167–168 °C; ¹H NMR (CD₃OD) δ 3.34–3.83 (2H, m, H-5'), 3.87 (1H, m, H-4'), 3.97 (1H, m, 2'-OCH₂), 4.10 (1H, m, H-2'), 4.15 (1H, m, 2'-OCH₂), 4.20 (1H, t, J=4.8 Hz, H-4'), 5.11–5.26 (2H, m, CH= CH_2), 5.69 (1H, d, J= 8.2 Hz, H-5), 5.97 (1H, m, CH=CH₂), 6.26 (1H, d, J= 5.2 Hz, H-1'), 7.85 (1H, d, J=8.2 Hz, H-6); ¹³C NMR (CD₃OD) δ 61.8 (C-5'), 72.6 (C-3'), 74.9 (2'-OCH₂), 84.6 (C-2'), 85.1 (C-4'), 85.7 (C-1'), 101.3 (C-5), 117.2 (CH=CH₂), 135.2 (CH=CH₂), 144.1 (C-6), 152.2 (C-2), 166.2 (C-4).

4.1.12. Preparation of 1-(2'-*O*-allyl-3',5'-*O*,*O*-bis(*tert*butyldimethylsilyl)-β-D-arabinofuranosyl)uracil (18). Compound 17 (0.200 g, 0.70 mmol) was dissolved in anhydrous DMF (7 mL) and added imidazole (0.480 g, 7.04 mmol) and TBDMSCl (0.530 g, 3.52 mmol). The reaction mixture was stirred for 24 h at room temperature, concentrated under reduced pressure and coevaporated with xylene (2.5 mL). The residue was dissolved in CH₂Cl₂ (20 mL), washed with water (2×20 mL), dried (Na₂SO₄) and concentrated under reduced pressure. The residue was purified by column chromatography (0–2% CH₃OH in CH₂Cl₂) to give the product as a colourless oil (0.298 g, 83%); ¹H NMR (CDCl₃) δ 0.09–0.13 (12H, m, Si(CH₃)₂), 0.87–0.93 (18H, m, C(CH₃)₃), 3.74–4.01 (6H, m, 2'-OCH₂, H-2', H-4', H-5'), 4.29 (1H, t, J=5.2 Hz, H-3'), 5.11–5.20 (2H, m, CH=CH₂), 5.65–5.81 (2H, m, CH=CH₂, H-5), 6.26 (1H, J=5.3 Hz, H-1'), 7.65 (1H, d, J=8.4 Hz, H-6), 8.91 (1H, br s, NH); ¹³C NMR (CDCl₃) δ – 5.5, – 5.4, –4.9, –4.5 (Si(CH₃)₂), 17.9, 18.4 (*C*(CH₃)₃), 25.5, 25.6, 25.9 (C(CH₃)₃), 61.0 (C-5'), 72.3 (C-3'), 73.6 (2'-OCH₂), 83.2, 83.7, 83.9 (C-1', C-2', C-3'), 101.2 (C-5), 117.6 (CH=CH₂), 133.3 (CH=CH₂), 141.9 (C-6), 150.3 (C-2), 163.2 (C-4); *m/z* MALDI (M+Na) 535.

4.1.13. Preparation of 1-(2'-O-allyl-3'-O-(tert-butyldimethylsilyl)-β-d-arabinofuranosyl)uracil (19). Compound 18 (0.701 g, 1.37 mmol) was dissolved in 80% aqueous acetic acid (20 mL) and the reaction mixture was stirred at 60 °C for 2 h 30 min. The reaction mixture was concentrated under reduced pressure and the residue was purified by column chromatography (0-2% CH₃OH in CH_2Cl_2) to give the product as a white foam (0.399 g, 73%); mp = 131–132 °C; ¹H NMR (CDCl₃) δ 0.11–0.16 (6H, m, Si(CH₃)₂), 0.87–0.94 (9H, m C(CH₃)₃), 2.23 (1H, m, 5'-OH), 3.78–3.98 (6H, m, 2'-OCH₂), 4.24 (1H, t, J =3.8 Hz, H-3'), 5.15–5.21 (2H, m, CH=CH₂), 5.71 (1H, d, J = 8.2 Hz, H-5), 5.75 (1H, m, CH=CH₂), 6.27 (1H, d, J =4.6 Hz, H-1[']), 7.62 (1H, d, J = 8.2 Hz, H-6), 8.71 (1H, br s, NH); ¹³C NMR (CDCl₃) δ -4.9, -4.6 (Si(CH₃)₂), 17.9 (C(CH₃)₃), 25.6 (C(CH₃)₃), 61.4 (C-5'), 72.3 (C-3'), 75.1 (2'-OCH₂), 83.3, 84.3, 84.4 (C-1', C-2', C-4'), 101.2 (C-5), 118.3 (CH=CH₂), 132.9 (CH=CH₂), 142.0 (C-6), 150.2 (C-2), 163.0 (C-4); HiRes MALDI FT-MS m/z (M+Na) found/calcd 421.1751/421.1765.

4.1.14. Preparation of (5-allyl-3'-O-tert-butyldimethylsilyl-2'-deoxyuridin-5'-yl) (2'-O-allyl-5'-O-tert-butyldimethylsilyluridin-3'-yl) 2-cyanoethylphosphate (20). Compound 13 (0.103 g, 0.17 mmol) and compound 5 (0.073 g, 0.19 mmol) were coevaporated with anhydrous CH₂Cl₂ and dissolved in anhydrous CH₂Cl₂ (6 mL). A 0.45 M solution of 1H-tetrazole in CH₃CN (1.9 mL, 0.86 mmol) was added and the reaction mixture was stirred for 1 h. The reaction was quenched by the addition of CH₃OH (1 mL) and the mixture was concentrated under reduced pressure. The residue (crude phosphite, $R_{\rm f}$ 0.41 (75% ethyl acetate in petrolether)) was dissolved in a mixture of CH₂Cl₂ and CH₃CN (3:1 v/v, 8 mL) and cooled to 0 °C. A 3 M solution of t-BuOOH in toluene (0.3 mL, 0.9 mmol) was added and the reaction mixture was stirred for 1 h at 0 °C and for 2 h at room temperature. The reaction was quenched by the addition of CH₃OH (1 mL) and the mixture was concentrated under reduced pressure. The residue was purified by column chromatography (50% ethyl acetate in petrol ether and 10% CH₃OH in CH₂Cl₂) to give the product as a white foam (0.156 g, 100%); $R_{\rm f}$ 0.27 (75%) ethyl acetate in petrol ether); ¹H NMR (CD₃CN) δ 0.10– 0.12 (12H, m, Si(CH₃)₂), 0.81–0.92 (18H, m, C(CH₃)₃), 2.18-2.26 (2H, m, T2-H-2'), 2.78-2.82 (4H, m, 5-CH₂, CH₂CN), 3.83–4.43 (12H, m, U1-H-5', U1-H-4', U1-H-2', U1-2'-OCH₂, T2-H-5', T2-H-4', T2-H-3', CH₂OP), 4.87-4.92 (1H, m, U1-H-3'), 5.01–5.29 (4H, m, $2 \times CH = CH_2$), 5.63-5.67 (1H, m, H-5), 5.78-5.94 (2H, m, $2 \times CH = CH_2$), 5.99–6.02 (1H, m, U1-H-1'), 6.15–6.21 (1H, m, T2-H-1'), 7.31 (1/2H, s, T2-H-6), 7.38 (1/2H, s, T2-H-6), 7.72-7.75 (1H, m, H-6), 8.91–8.95 (2H, br s, 2×NH); 31 P NMR (CD₃CN) δ -1.74, -1.49; m/z ESI 918 (M+Na), 896 (M+H).

4.1.15. Preparation of allyl (2'-O-allyl-3'-O-(tert-butyldimethylsilyl)arabinouridin-5'-yl) (5'-O-tert-butyldimethylsilylthymidin-3'-yl) phosphate (21). Compound **19** (0.081 g, 0.20 mmol) was coevaporated with anhydrous CH₃CN and dissolved in anhydrous CH₃CN (3 mL). A 0.45 M solution of 1H-tetrazole in CH₃CN (0.68 mL, 0.30 mmol) was added and the mixture for stirred for 10 min. A solution of 1 (0.252 g, 0.46 mmol) in anhydrous CH₃CN (3 mL) was added and the reaction mixture was stirred for 2 h. The reaction was quenched by the addition of CH₃OH (0.5 mL) and the reaction mixture was concentrated under reduced pressure and coevaporated with CH₂Cl₂. The residue was dissolved in anhydrous CH2Cl2 (4 mL) and cooled to 0 °C. A 3 M solution of t-BuOOH in toluene (0.14 mL, 0.41 mmol) was added dropwise and the reaction mixture was stirred at room temperature for 1 h. The reaction was quenched by the addition of CH₃OH (1 mL) and the mixture was concentrated under reduced pressure. The residue was purified by column chromatography (0-2%)CH₃OH in CH₂Cl₂) to give the product as a white foam and an epimeric mixture (0.148 g, 85%); mp = 70-71 °C; ¹H NMR (CDCl₃) δ 0.08–0.13 (12H, m, Si(CH₃)₂), 0.90–0.92 (18H, m, C(CH₃)₃), 1.90-1.94 (3H, br s, T1-CH₃), 2.00-2.06 (1H, m, T1-H-2'), 2.52-2.58 (1H, m, T1-H-2'), 3.90-4.28 (10H, m, U2-2'-OCH₂, U2-H-5', U2-H-4', U2-H-3', U2-H-2', T1-H-5', T1-H-4⁷), 4.56-4.61 (2H, m, CH₂OP), 4.98-5.02 (1H, m, T1-H-3'), 5.14-5.21 (2H, m, U2-CH=CH₂), 5.29-5.43 (2H, m, T1-CH=CH₂), 5.68-5.80 (2H, m, U2-CH=CH₂, U2-H-5), 5.95-6.01 (1H, m, T1-CH=CH₂), 6.28-6.32 (1H, m, U2-H-1'), 6.33-6.39 (1H, m, T1-H-1′), 7.48–7.52 (1H, m, T1-H-6), 7.61–7.69 (1H, m, U2-H-6), 9.53–9.69 (2H, m, 2×NH); $^{31}\mathrm{P}$ NMR (CDCl₃) δ -0.59; HiRes MALDI FT-MS m/z (M+Na) found/calcd 879.3420/879.3404.

4.1.16. Preparation of (2'-O-allyl-3'-O-(tert-butyldimethylsilyl)arabinouridin-5'-yl) (2'-O-allyl-5'-O-tertbutyldimethylsilyluridin-3'-yl) 2-cyanoethylphosphate (22). Compound 13 (0.140 g, 0.23 mmol) and compound **19** (0.111 g, 0.28 mmol) were coevaporated twice with anhydrous CH3CN and redissolved in anhydrous CH3CN (7 mL). A 0.45 M solution of 1*H*-tetrazole in CH₃CN (2.6 mL, 1.17 mmol) was added and the reaction mixture was stirred for 2 h. The reaction was quenched by the addition of CH₃OH (0.5 mL) and the mixture was concentrated under reduced pressure. The residue was dissolved in a mixture of CH2Cl2 and CH3CN (1:1 v/v, 16 mL) and stirred at 0 °C. A 3 M solution of t-BuOOH in toluene (0.38 mL, 1.14 mmol) was added and the reaction mixture was stirred at room temperature for 90 min. The reaction was quenched by the addition of CH₃OH (0.5 mL) and the mixture was concentrated under reduced pressure. The residue was purified by column chromatography (ethyl acetate) and precipitated twice from a mixture of ethyl acetate and petrol ether to give the product as a white powder and an epimeric mixture (0.181 g, 78%); $R_{\rm f}$ 0.27 (75% ethyl acetate in petrol ether); ¹H NMR (DMSO-d₆) δ 0.08–0.12 (12H, m, Si(CH₃)₂), 0.86–0.90 (18H, m, C(CH₃)₃), 2.92–2.96 (2H, m, CH₂CN), 3.77–4.28 (15H, m, U1-H-5', U1-H-4', U1-H-2', U2-H-5', U2-H-4', U2-H-3', U2-H-2', CH₂OP, U1-2'-OCH₂, U2-2'-OCH₂), 4.82-4.86 (1H, m, U1-H-3'), 5.09–5.25 (4H, m, 2×CH=CH₂), 5.59-5.68 (2H, m, 2×H-5), 5.70-5.90 (2H, m, 2×

CH=CH₂), 5.92–5.97 (1H, m, U1-H-1'), 6.21–6.23 (1H, m, U2-H-1'), 7.56–7.72 (2H, m, 2×H-6), 11.41–11.47 (2H, m, 2×NH); ³¹P NMR (DMSO-d₆) δ – 1.12; HiRes MALDI FT-MS *m*/*z* (M+Na) found/calcd 934.3442/934.3462.

4.1.17. Preparation of (5-allyl-2'-deoxy-3'-O-tert-butyldimethylsilyluridin-5'-yl) (5-allyl-2'-deoxy-5'-O-tertbutyldimethylsilyluridin-3'-yl) 2-cyanoethylphosphate (23). Compound 7 (0.156 g, 0.268 mmol) and compound 5 (0.111 g, 0.290 mmol) were coevaporated twice with anhydrous CH₃CN and dissolved in anhydrous CH₃CN (3 mL). A 0.45 M solution of 1H-tetrazole in CH₃CN (3.0 mL, 1.35 mmol) was added and the reaction mixture was stirred for 30 min. The reaction was quenched by the addition of CH₃OH (0.6 mL) and the mixture was concentrated under reduced pressure. The residue (crude phosphite, $R_{\rm f}$ 0.23 (75% ethyl acetate in petrol ether)) was dissolved in anhydrous CH₃CN (3 mL) and a 3 M solution of t-BuOOH in toluene (0.45 mL, 1.35 mmol) was added. The reaction mixture was stirred for 1.5 h and added another portion of the 3 M solution of t-BuOOH in toluene (0.23 mL, 0.69 mmol). The reaction mixture was stirred for 30 min and the reaction was quenched by the addition of CH₃OH (0.5 mL). The mixture was concentrated under reduced pressure, and the residue was dissolved in CH₂Cl₂ (50 mL), washed with a saturated aqueous solution of NaHCO₃ (25 mL), dried (Na₂SO₄) and concentrated under reduced pressure. The residue was purified by dry column chromatography (50-0% petrol ether in ethyl acetate) to give the product as a white foam and an epimeric mixture (0.142 g, 60%); (Found C, 53.01; H, 7.32; N, 7.42%; $C_{39}H_{62}N_5O_{12}PSi_2$ requires C, 53.23; H, 7.10; N, 7.96%); R_f 0.36 (75% ethyl acetate in petrolether); ¹H NMR (CDCl₃) δ 0.10-0.11 (12H, m, Si(CH₃)₂), 0.89-0.91 (18H, m, C(CH₃)₃), 2.08–2.68 (4H, m, T1-H-2', T2-H-2'), 2.75– 2.80 (2H, m, CH₂CN), 3.00-3.08 (4H, T1-5-CH₂, T2-5-CH₂), 3.80-4.03 (4H, m, T2-H-5', T2-H-4', T1-H-4'), 4.20-4.30 (5H, m, T1-H-5', T2-H-3', CH₂OP), 4.41-4.45 (1H, m, T1-H-3'), 5.04–5.17 (4H, m, $2 \times CH = CH_2$), 5.85–5.89 (2H, m, $2 \times CH = CH_2$), 6.10–6.17 (1H, m, T1-H-1'), 6.27–6.32 (1H, m, T2-H-1'), 7.20-7.22 (1H, m, H-6), 7.39-7.41 (1H, m, H-6), 9.00–9.13 (2H, m, 2×NH); ³¹P NMR (CDCl₃) δ -1.58, -1.44; HiRes MALDI FT-MS m/z (M+Na) found/ calcd 902.3581/902.3563.

4.1.18. Preparation of allyl (5-allyl-2'-deoxy-3'-O-tertbutyldimethylsilyluridin-5'-yl) (5'-O-tert-butyldimethylsilylthymidin-3'-yl) phosphate (24). Compound 5 (0.188 g, 0.49 mmol) and compound **1** (0.520 g, 0.96 mmol) were coevaporated twice with anhydrous CH₃CN (10 mL) and dissolved in anhydrous CH₃CN (25 mL). A 0.45 M solution of 1H-tetrazole in CH₃CN (5.3 mL, 2.38 mmol) was added over 5 min and the reaction mixture was stirred for 1.5 h at room temperature. A 3 M solution of t-BuOOH in toluene (0.82 mL, 2.46 mmol) was added, and the mixture was stirred for another 1.5 h. The reaction was quenched by the addition of CH₃OH (0.25 mL), and the mixture was concentrated under reduced pressure. The residue was dissolved in ethyl acetate (50 mL) and washed with a saturated aqueous solution of NaHCO3 $(2 \times 20 \text{ mL})$ and brine (20 mL). The aqueous phase was extracted with ethyl acetate (30 mL) and the combined organic phases were dried (Na₂SO₄) and concentrated under reduced pressure. The residue was purified by column chromatography (20–70% ethyl acetate in petrol ether) to give the product as a white foam and an epimeric mixture $(0.266 \text{ g}, 64\%); \text{mp} = 74-76 \,^{\circ}\text{C}; R_{f} \, 0.34 \, (75\% \text{ ethyl acetate})$ in petrol ether); ¹H NMR (CDCl₃) δ 0.01–0.12 (12H, m, Si(CH₃)₂), 0.83–0.92 (18H, m, C(CH₃)₃), 1.90–1.94 (3H, m, CH₃), 2.08–2.19 (1H, m, T1-H-2[']), 2.21–2.29 (2H, m, T2-H-2'), 2.50-2.58 (1H, m, T1-H-2"), 3.08-3.11 (2H, m, T2-5-CH₂), 3.85–3.92 (T1-H-5'), 4.01–4.03 (1H, m, T2-H-4'), 4.20-4.29 (3H, m, T2-H-5', T1-H-4'), 4.41-4.44 (1H, m, T2-H-3'), 4.55–4.60 (2H, m, CH₂OP), 5.00–5.04 (1H, T1-H-3'), 5.09–5.17 (2H, m, T2-CH=CH₂), 5.28–5.41 (2H, m, T1-CH=CH₂), 5.84-6.00 (2H, m, 2×CH=CH₂), 6.17-6.25 (1H, m, T2-H-1'), 6.32-6.37 (1H, m, T1-H-1'), 7.24-7.28 (1H, m, T2-H-6), 7.46-7.48 (1H, m, T1-H-6), 8.76-8.87 (2H, m, $2 \times \text{NH}$; ³¹P NMR (CDCl₃) $\delta - 0.61$; HiRes MALDI FT-MS m/z (M+Na) found/calcd 863.3454/863.3454.

4.1.19. Preparation of cyclic 5'-0.3'-0-bis-TBDMS protected dUpU containing a 5-to-5 (*E*/*Z*)-2-butenelinker (25). Compound 23 (26 mg, 0.030 mmol) was dissolved in anhydrous CH₂Cl₂ (3 mL). Grubbs second-generation catalyst \mathbf{X}^2 (1.4 mg, 1.6 µmol) was added and the reaction mixture was stirred at reflux for 22 h. The mixture was concentrated under reduced pressure, and the residue was purified by column chromatography (2–8% CH₃OH in CH₂Cl₂) to give the product as a white powder and a mixture of diastereomers (6 mg, 23%) as well as starting material 23 (7 mg, 27%); R_f 0.28 (10% CH₃OH in CH₂Cl₂); ³¹P NMR (CDCl₃) δ –2.01 (1), –0.90 (0.13), –0.56 (0.61), 0.02 (0.17); HiRes MALDI FT-MS *m*/*z* (M+Na) found/calcd 874.3199/874.3250.

4.1.20. Preparation of cyclic 5'-0,3'-0-bis-TBDMS protected dTpU containing a phosphate-to-5 (E/Z)-2butenelinker (26). Compound 24 (0.052 g, 0.062 mmol) was dissolved in anhydrous CH₂Cl₂ (6 mL). Grubbs secondgeneration catalyst \mathbf{X}^2 (2.8 mg, 3.2 µmol, 5.2 mol%) was added and the reaction mixture was stirred at reflux for 2 h. The mixture was concentrated under reduced pressure and the residue was purified by column chromatography (10-0%) petrol ether in ethyl acetate) to give the product as a white foam and an epimeric mixture (29 mg, 58%); $R_{\rm f}$ 0.29 (ethyl acetate); ¹H NMR (CDCl₃) δ 0.10–0.13 (12H, m, Si(CH₃)₂), 0.89–0.94 (18H, m, C(CH₃)₃), 1.90–1.94 (3H, m, T1-CH₃), 2.06-2.30 (3H, m, T2-H-2', T1-H-2'), 2.50-2.60 (1H, m, T1-H-2"), 3.10–3.14 (2H, m, T2-5-CH₂), 3.80–3.96 (2H, m, T1-H-5'), 3.98-4.49 (5H, m, T1-H-4', T2-H-5', T2-H-4', T2-H-3'), 4.51-4.90 (2H, m, CH2OP), 5.01-5.05 (1H, m, T1-H-3'), 5.84–5.89 (1H, m, CH=CHCH₂OP), 5.91–5.98 (1H, m, CH=CHCH₂OP), 6.36–6.42 (2H, m, T1-H-1['], T2-H-1'), 7.37-7.51 (2H, m, T1-H-6, T2-H-6), 9.02-9.20 (2H, m, NH); ³¹P NMR (CDCl₃) δ -0.65, 2.01. HiRes MALDI FT – MS m/z (M + Na) found/calcd 835.3148/835.3141; IR (KBr) ν cm⁻¹: 3441, 3065, 2954, 2930, 2857, 1694, 1471, 1276, 1100, 1006, 987, 837, 780.

4.1.21. Analytical formation of Tp-5-(4-amino-2-butenyl)-2'-deoxyuridine (27). Compound 26 (2 mg) was dissolved in 32% aqueous NH₃ (1 mL) for 24 h at room temperature. The reaction mixture was concentrated under reduced pressure to give the product as a white foam; ¹H NMR (DMSO-d₆) δ 1.78 (3H, s, CH₃), 1.91–2.18 (3H, m, T1-H-2', T2-H-2'), 2.24–2.33 (1H, m, T1-H-2"), 2.92–3.09 (2H, m, T2-5-CH₂), 3.40–3.43 (2H, m, CH₂N), 3.51–3.59 (2H, m, T1-H-5'), 3.75–4.31 (5H, m, T2-H-5', T1-H-4', T2-H-4', T2-H-4', T2-H-3'), 4.60–4.70 (1H, m, T1-H-3'), 5.20–5.32 (2H, m, 2×OH), 5.69–5.78 (1H, m, CH=CHCH₂N), 5.89–5.97 (1H, m, CH=CHCH₂N), 6.14–6.18 (1H, m, T1-H-1'), 6.25 (1H, t, J=7.4 Hz, T2-H-1'), 7.68 (1H, s, T2-H-6), 7.70 (1H, s, T1-H-6); ³¹P NMR (DMSO-d₆) δ – 0.86; MALDI-MS *m*/*z*: 601.27 (MH⁺), 623.19 (MNa⁺).

4.1.22. Preparation of cyclic 5'-0,3'-0-bis-TBDMS protected dTpU containing a phosphate-to-5 butanelinker (28). Compound 24 (0.255 g, 0.303 mmol) was dissolved in anhydrous CH_2Cl_2 (30 mL) and Grubbs second-generation catalyst \mathbf{X}^2 (14.0 mg, 16 μ mol, 5.4 mol%) was added. The reaction mixture was stirred at reflux for 3.5 h and then bubbled with hydrogen for 5 min. The mixture was placed in an autoclave at 1000 psi hydrogen at 50 °C overnight. The mixture was concentrated under reduced pressure and the residue was purified by column chromatography (20-0% petrol ether in ethyl acetate) to give the product as a white foam and an epimeric mixture (0.156 g, 63.0%); $R_{\rm f}$ 0.29 (1% CH₃OH in ethyl acetate); ¹H NMR $(CDCl_3) \delta 0.10-0.13 (12H, m, Si(CH_3)_2), 0.89-0.93 (18H, m$ C(CH₃)₃), 1.55–1.85 (4H, m, CH₂CH₂CH₂CH₂OP), 1.92– 1.94 (3H, m, CH₃), 2.10–2.58 (6H, m, T1-H-2', T2-H-2', T2-5-CH₂), 3.85–4.01 (4H, m, T1-H-5', CH₂OP), 4.09–4.44 (3H, m, T2-H-4', T2-H3', T1-H-4'), 4.45-4.60 (2H, m, T2-H-5'), 4.95-5.05 (1H, m, T1-H-3'), 6.29-6.40 (2H, m, T2-H-1', T1-H-1'), 7.39-7.52 (2H, m, T1-H-6, T2-H-6), 9.13-9.24 (2H, m, $2 \times \text{NH}$; ³¹P NMR (CDCl₃) δ -0.81, 1.29; HiRes MALDI FT-MS *m*/*z* (M+Na) found/calcd 837.3336/837.3298.

4.1.23. Preparation of cyclic dTpU containing a phosphate-to-5 butanelinker (29). Compound 28 (0.145 g, 0.178 mmol) was dissolved in a 90% aqueous solution of trifluoroacetic acid (5 mL) and stirred for 3 h. The mixture was concentrated under reduced pressure and coevaporated with 99.9% CH_3CH_2OH (3×5 mL) and with CH_3OH (5 mL) to give the product as a white powder and an epimeric mixture (0.104 g, 100%); ¹H NMR (CD₃OD) δ 1.56-1.75 (4H, m, CH₂CH₂CH₂CH₂OP), 1.77-1.80 (3H, s, CH₃), 2.08–2.50 (6H, m, T1-H-2', T2-H-2', T2-5-CH₂), 3.69–3.73 (2H, m, T1-H-5'), 3.90–4.42 (7H, m, T1-H-4', T2-H-5', T2-H-4', T2-H-3', POCH₂), 4.99-5.02 (1H, m, T1-H-3'), 6.18–6.28 (2H, m, T1-H-1', T2-H-1'), 7.40 (¹/₂H, s, T2-H-6), 7.50 (1/2H, s, T2-H-6), 7.69-7.70 (1H, m, T1-H-6); ³¹P NMR (CD₃OD) δ -1.05, 0.01 (¹H and ³¹P NMR data from DMSO- d_6 has been published in Ref. 9); HiRes MALDI FT-MS m/z (M+Na) found/calcd 609.1584/609, 1568; IR (KBr) ν cm⁻¹: 3435, 1690, 1472, 1277, 1022, 784.

4.1.24. Analytical formation of Tp-5-(4-aminobutanyl)-2'-deoxyuridine (30). Compound 29 (2 mg) was dissolved in 32% aqueous NH₃ (1 mL) for 5 days at 55 °C. The reaction mixture was concentrated under reduced pressure to give the product as a white foam; ¹H NMR (DMSO-d₆) δ 1.51–1.56 (4H, m, CH₂CH₂CH₂CH₂N), 1.78 (3H, s, CH₃), 2.03–2.10 (2H, m, T2-H-2'), 2.11–2.19 (2H, m, T2-5-CH₂), 2.23-2.28 (2H, m, T1-H-2'), 2.81–2.84 (2H, m, CH₂N), 3.58–3.62 (2H, m, T1-H-5'), 3.81–3.88 (2H, m, T2-H-5'), 3.94–3.96 (2H, m, T1-H-4', T2-H-4'), 4.30–4.32 (1H, m, T2-H-3'), 4.63–4.67 (1H, m, T1-H-3'), 5.31 (1H, br, OH), 6.16 (1H, t, J=6.7 Hz, T1-H-1[']), 6.26 (1H, t, J=7.0 Hz, T2-H-1[']), 7.70 (1H, s, T1-H-6), 7.73 (1H, s, T2-H-6), 8.2– 9.2 (3H, br, NH₃); ³¹P NMR (DMSO-d₆) δ –0.82; MALDI-MS *m*/*z*: 603 (M⁺), 625 (MNa⁺); IR (KBr) ν cm⁻¹: 3400, 2255, 2128, 1684, 1469, 1277, 1203, 1026, 825, 763.

Acknowledgements

The Danish National Research Foundation and The Danish Natural Science Research Council are thanked for financial support.

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Tetrahedron 60 (2004) 10967-10972

Electrochemical reductive cleavage of carbon-halogen bonds in 5-bromo-1,3-dichloro-2-iodobenzene

M. Arun Prasad and M. V. Sangaranarayanan*

Department of Chemistry, Indian Institute of Technology Madras, Chennai 600036, India

Received 28 June 2004; revised 23 August 2004; accepted 9 September 2004

Available online 8 October 2004

Abstract—The electrochemical reduction of carbon–halogen bonds in 5-bromo-1,3-dichloro-2-iodobenzene follows quadratic activation– driving force relationship except in one of the carbon–chlorine bonds. The variation of the transfer coefficient with the electrode potential has been estimated using the voltammetric data coupled with the convolution analysis. The standard potentials pertaining to the reduction of carbon–halogen bonds are evaluated using the Marcus theory of outer sphere electron transfer.

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1. Introduction

Electrochemical single electron transfer reactions constitute a frontier area of research¹ and the kinetics of these reactions can be analyzed using the Marcus–Hush theory of outer sphere electron transfer assuming the validity of the Born–Oppenheimer approximation. Electron transfer to an organic molecule (RX) is often accompanied by bond cleavage leading to a radical (R[•]) and an anion (X⁻) occurring either in a stepwise manner (reaction 1) or a single elementary step (reaction 2).

$$\mathbf{RX} + \mathbf{e}^{-} \rightleftharpoons \mathbf{RX}^{-} \left(E^{0}_{\mathbf{RX}/\mathbf{RX}^{-}} \right) \tag{1a}$$

$$\mathbf{R}\mathbf{X}^{\cdot-} \to \mathbf{R}^{\cdot} + \mathbf{X}^{-} \tag{1b}$$

$$RX + e^- \to R^+ + X^- (E^0_{RX/R^+ + X^-})$$
 (2)

Application of general electrochemical techniques yields the transfer coefficient and the forward electron transfer rate constant for the above reactions, but not the information about the standard potential (E^0) vis-à-vis the standard rate constant; since the dissociative electron transfer reactions are completely irreversible, E^0 values can not be obtained directly (for example, by cyclic voltammetry). However, the kinetic analysis of homogeneous redox catalysis of the electrochemical reduction leads to the determination of $E^{0.2}$ An elegant method for the estimation of standard potentials of irreversible systems obeying a quadratic activation– driving force relationship was demonstrated in the dissociative reduction of perbenzoates.³ The above methodology, making use of linear variation of transfer coefficient (α) with the electrode potential (*E*), was adopted in the electrochemical reduction of several organic^{4–6} and biologically relevant molecules.⁷ However, studies in this direction on the stepwise reductive cleavage reactions involving rapid decomposition of the anion radicals (especially aromatic compounds) are limited.

In this communication, we report the standard potentials of the irreversible reduction of aromatic carbon-halogen bonds in 5-bromo-1,3-dichloro-2-iodobenzene using the quadratic activation-driving force relationship. Convolution potential sweep voltammetry is the main tool for investigating the reaction kinetics, since, in contrast to homogeneous redox catalysis, the convolution approach allows one to obtain extensive data on the logarithmic electron transfer rate constant (ln $k_{\rm ET}$) versus electrode potential (*E*) variation. It has to be emphasized here that the present approach fails for systems involving linear variation of ln $k_{\rm ET}$ with *E* (Butler–Volmer kinetics)—an example of which also is demonstrated in the present study.

2. Experimental

The voltammetric studies were carried out in a single compartment electrochemical cell thermostated at 298 K, using the Bio Analytical Systems (BAS) 100A Electrochemical workstation. The working electrode was a glassy

Keywords: 5-Bromo-1,3-dichloro-2-iodobenzene; Marcus theory; Stepwise mechanism; Convolution analysis; Transfer coefficient; Outer-sphere electron transfer.

^{*} Corresponding author. Tel.: +91 22578269; fax: +91 22570545; e-mail: mvs@chem.iitm.ac.in

^{0040–4020/\$ -} see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.020

carbon disc (BAS) of 3 mm diameter while platinum foil (2 cm^2) served as the counter electrode. The working electrode was polished with the alumina slurry (BAS) and ultrasonically rinsed prior to use. The electrochemical pretreatment was carried out in the background solution using several cycles at 0.05–1 V s⁻¹ in a wide potential range. Tetra *n*-butyl ammonium bromide (*n*Bu₄NBr) (Fluka) was the supporting electrolyte and used as received. N,N'-dimethyl formamide (DMF) was initially distilled from anhydrous copper sulfate, then the distillate was again distilled from calcium hydride under reduced pressure and stored over 4 Å molecular sieves. Silver/silver ion (1 mM) electrode (BAS) was used as the quasi-reference electrode which was subsequently calibrated with the ferrocene/ ferrocenium couple under identical conditions of solvent and supporting electrolyte. The background subtracted voltammograms were analyzed by the convolution approach, the experimental and computational details of which have been described earlier.⁸ NMR spectra were recorded on Bruker Avance 400 spectrometer. UV-Vis spectrum was obtained with the Ocean Optics UV/Vis spectrometer.

5-Bromo-1,3-dichloro-2-iodobenzene was synthesized by the following procedure: 2,6-dichloroaniline was brominated by passing the vapours of bromine into a solution of 2,6-dichloroaniline in hydrochloric acid. The solid 4-bromo-2,6-dichloroaniline was filtered and purified by column chromatography (silica gel). 4-Bromo-2,6-dichloroaniline was then diazotized in hydrochloric acid (6 M) using aqueous sodium nitrite (6 M) and the resulting solution was slowly added to aqueous potassium iodide (5 M). When no gas was evolved, the crude product was filtered, washed with aqueous sodium hydroxide, subsequently with sodium metabisulphite and finally with water. The residue was purified by column chromatography (silica gel) using hexane as the eluent to give 5-bromo-1, 3-dichloro-2-iodobenzene as white solid: Mp 67.8-68.7 °C (literature:⁹ 67.5–68.2 °C); ¹H NMR (400 MHz, CDCl₃, TMS as internal standard) δ 7.48 (2H, s); ¹³C NMR (100 MHz, CDCl₃) δ 102.41, 122.36, 129.89, 141.33; UV-Vis (CHCl₃) broad band centered at 300 nm. The compound was crystallized in hexane (colorless crystals) for the electrochemical studies and the crystal structure confirms the identity of the compound^{\dagger}.

3. Results and discussion

Halobenzenes (RX) undergo irreversible electron transfers at the electrode surface and are capable of hosting transitorily the incoming electron in their π^* orbitals leading to the radical anions (RX^{•-}) (reaction 1a). The radical anions readily undergo decomposition with a first order rate constant greater than 10^4 s^{-1} to neutral radicals (R[•]) and halide ions (X⁻) (reaction 1b).¹⁰ The dissociation

of the radical anion may be viewed as an intramolecular dissociative single-electron transfer from the π^* orbital to the σ^* orbital of the carbon-halogen bond.¹¹ Interestingly, the neutral radicals are easier to reduce than the parent halobenzenes and immediately undergo a second electron transfer to form R⁻. However, the characteristic features of the reduction wave of halobenzene are solely governed by the kinetics of the first electron transfer. R⁻ abstracts a proton either from the solvent or the supporting electrolyte to give the hydrocarbon RH and it was observed that several halobenzenes and other aromatic halides upon electrolysis yielded 100% of RH.¹⁰ A recent investigation, involving in situ electrochemical-NMR spectroscopy,¹² has revealed that the aryl anion abstracts a proton preferably from the solvent rather than the supporting electrolyte. It is worth mentioning here that the hydrocarbon RH is reducible and, in fact, a second wave is observed in some cases before the background discharge.¹⁰ However, in most cases, the reduction wave of RH is suppressed by the background discharge current of the supporting electrolyte.

Figure 1 shows the cyclic voltammogram pertaining to the reduction of 5-bromo-1,3-dichloro-2-iodobenzene at the glassy carbon electrode in DMF containing 0.1 M nBu₄NBr as the supporting electrolyte. The reduction waves a, b, c and d represent, respectively, the two-electron reduction of carbon-iodine (C-I), carbon-bromine (C-Br) and two carbon-chlorine (C-Cl (1) and C-Cl (2)) bonds. This assignment follows from the fact that the carbon-halogen bonds are susceptible to reduction in the order of: C-I>C-Br>C-Cl>C-F. Each wave represents the hydrogenolysis of a carbon-halogen bond finally leading to the formation of benzene and hence an overall consumption of eight electrons in a single voltammetric cycle. The voltammogram B shows the reduction waves of 1,3-dichlorobenzene which corresponds to the peaks c and d of the voltammogram A. The π^* level of 5-bromo-1,3-dichloro-2iodobenzene being comparatively lower than the monosubstituted benzene is proved by the fact that the reduction potential of carbon-iodine bond (wave a) is ca. 336 mV more positive than that of iodobenzene. A similar behaviour also arises for the carbon-bromine bond, the reduction potential of which is ca. 486 mV more positive than that of bromobenzene. However, the peak potential of wave d corresponds to that of chlorobenzene.¹⁰ All the waves remain irreversible even at a scan rate of 2000 mV s⁻¹, indicating that the life time of the radical anion is less than 10^{-4} s. The peak currents of the waves a, b, c and d are proportional to the square root of the sweep rate. The transfer coefficient of the reduction of carbon-halogen bond can be calculated from the peak width measurements (Eq. 3) and the values are listed for various carbon-halogen bonds in Table 1.

$$\alpha = \frac{1.856RT}{F} \frac{1}{(E_{\rm P/2} - E_{\rm P})}$$
(3)

The α values close to or greater than 0.5 are expected for stepwise mechanism. However, this is not an absolute criterion³ and our systematic study has revealed that the reduction of carbon–iodine bond, for which the α value is less than 0.5, indeed follows a stepwise mechanism.¹³

[†] Crystallographic data have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication numbers CCDC 231991. Copies of the data can be obtained, free of charge, on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (Fax: +44(0)-1223-336033 or e-mail: deposit@ccdc.cam.ac.uk).



Figure 1. Cyclic voltammograms of (A) 5-bromo-1,3-dichloro-2-iodobenzene and (B) 1,3 dichlorobenzene in DMF/0.1 M nBu_4NBr at glassy carbon electrode. Scan rate: 200 mV s⁻¹; temperature: 298 K.

3.1. Convolution analysis

In order to analyze the thermodynamic and kinetic behaviour of the electron transfer, it is essential to deduce the electron transfer rate constant as a function of potential. In this respect, convolution voltammetry is a powerful electrochemical tool since it employs all the data points of the voltammetric wave rather than the peak characteristics alone.³ The convolution current (*I*) is related to the actual current (*i*) through the convolution integral¹⁴ (Eq. 4).

$$I = \frac{1}{\sqrt{\pi}} \int_0^t \frac{i(u)}{(t-u)^{1/2}} du$$
(4)

The plot between *I* versus *E* is sigmoidal in shape with the plateau being reached when the applied potential is sufficiently negative. Under this condition, *I* reaches its limiting value I_L defined as in Eq. 5

$$I_{\rm L} = nFAD^{1/2}C_{\rm b} \tag{5}$$

where D is the diffusion coefficient and C_b , the bulk concentration. Figure 2 shows the convolution potential sweep voltammogram of the reduction of 5-bromo-1,3-

dichloro-2-iodobenzene at a scan rate of 200 mV s⁻¹. In the case of closely spaced waves $I_{\rm L}$ was obtained at a potential at which the minimum occurs in the plot of derivative of the convolution current at the plateau region. The logarithmic analysis of the convolution current in conjunction with the voltammetric current yields the heterogeneous electron transfer rate constant¹⁴ as in Eq. 6.

$$\ln k_{\rm ET} = \ln D^{1/2} - \ln \frac{I_{\rm L} - I(t)}{i(t)}$$
(6)

Figure 3 depicts the variation of $\ln k_{\rm ET}$ with *E* at various scan rates for the reduction of C–I, C–Br, C–Cl (1) and C–Cl (2) bonds. In the cases of C–I, C–Br and C–Cl (2), the variations are parabolic, obeying the quadratic activation (ΔG^*) -driving force (ΔG^0) relationship (Eq. 7).

$$\Delta G^* = \frac{(\Delta G^0)^2}{16\Delta G_0^*} + \frac{\Delta G^0}{2} + \Delta G_0^* \tag{7}$$

However, in the reduction of C–Cl (1) bond, the variation of ln k_{ET} with *E* is linear. In this case the transfer coefficient is constant, conforming to Butler–Volmer kinetics (α obtained from the slope of the plot (Eq. 9) equals that obtained from

 Table 1. Electrochemical reduction behaviour of carbon-halogen bonds in 5-bromo-1,3-dichloro-2-iodobenzene

Carbon-halogen bond	Transfer coefficient from peak width measurements	Transfer coefficient from the Eq. 9	$E^0_{\text{RX/RX}^-}$ in mV versus SCE	$E^0_{\text{RX/RX}^-}$ for halobenzenes in mV versus SCE from Ref. 17
C–I	0.331	_	-1401	-2240 (iodobenzene)
C–Br	0.451	_	-2124	-2440 (bromobenzene)
C-Cl (1)	0.659	0.667	_	
C-Cl (2)	0.604	—	-2856	-2780 (chlorobenzene)

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Figure 2. Convolution potential sweep voltammogram of background subtracted voltammetric curve for the reduction of 5-bromo-1,3-dichloro-2-iodobenzene in DMF containing 0.1 M nBu_4NBr at the glassy carbon electrode. Scan rate: 200 mV s⁻¹; temperature: 298 K.



Figure 3. Potential dependence of logarithmic heterogeneous electron transfer rate constant for the reduction of (a) carbon–iodine bond (b) carbon–bromine bond (c) carbon–chlorine (1) bond and d) carbon–chlorine (2) bond at various scan rates.

the peak width measurements (Table 1)). It may be argued that a parabola with a small curvature would also fit the experimental $\ln k_{\rm ET}$ versus *E* plot for the reduction of C–Cl (1) bond. However, this possibility can be excluded, since the $\ln k_{\rm ET}$ versus *E* plot is too linear to be accounted for within the purview of a stepwise mechanism. Furthermore, α is too large to be accounted for by the theory within the framework of a concerted pathway.

3.2. Standard potential

The rate of change of activation energy (ΔG^*) with respect to the driving force (ΔG^0), yields the transfer coefficient as Eq. 8

$$\alpha = \frac{\partial \Delta G^*}{\partial \Delta G^0} = 0.5 + \frac{\Delta G^0}{8\Delta G^*_0} \tag{8}$$

Experimentally α is estimated from the derivative of $\ln k_{\text{ET}}$ versus *E* plot using Eq. 9

$$\alpha = -\frac{RT}{F} \frac{\mathrm{d}\ln k_{\mathrm{ET}}}{\mathrm{d}E} \tag{9}$$

wherein the symbols *R*, *T* and *F* assume the usual significance. Figure 4 shows the variation of α with *E* at various scan rates for the reduction of C–I, C–Br and C–Cl (2) bonds. Since the variation of ln k_{ET} with *E* is parabolic, α varies linearly with *E*. As implied by Eq. 8, most theoretical

models for outer sphere or dissociative electron transfers predict that α should be 0.5 at zero driving force^{15,16} $(\Delta G^0 = F(E - E^0) = 0)$. From the linear α versus *E* variation, $E_{\text{RX/RX}^-}^0$ of the reduction of respective carbon-halogen bonds can be estimated as the potential at which α becomes 0.5.⁴ Table 1 shows $E_{\text{RX/RX}^-}^0$ values for the reduction of C–I, C–Br and C–Cl (2) bonds obtained using the above methodology. Table 1 also provides $E_{\text{RX/RX}^-}^0$ values for the reduction of iodobenzene, bromobenzene and chlorobenzene for comparison.^{10,17} The standard potentials of the reduction of bromobenzene and chlorobenzene were determined through the kinetic analysis of homogeneous redox catalysis of the electrochemical reduction by Saveant et al.¹⁰ and that of the reduction of iodobenzene was obtained in an approximate way using the standard free energy of anion radical cleavage.¹⁷

The standard potentials of the reduction of C–I and C–Br are 839 and 316 mV more positive than those for the reduction of iodobenzene and bromobenzene, respectively (Table 1). This is consistent with the fact that the energy of the π^* orbital of the ring increases with the elimination of each halogen, viz. lowest at the first wave and highest at the fourth wave, which is reflected in the standard potentials of the reduction of respective carbon–halogen bonds. Even though α is substantially lower in the reduction of C–I, the energy of the π^* orbital of the ring is low enough to trap the unpaired electron before it



Figure 4. Variation of apparent transfer coefficient with electrode potential at various scan rates for the reduction of (a) carbon–iodine bond (b) carbon– bromine bond and (c) carbon–chlorine (2) bond.

dissociatively reduces the C–I bond in a successive step.¹³ A difference between the $E_{\rm RX/RX^{--}}^0$ for the reduction of C–Cl (2) and that for the reduction of chlorobenzene may be attributed to the different supporting electrolyte (*n*Bu₄NI) and working electrode (mercury) employed in the earlier study.¹⁰ Further, in our estimation of standard potentials, double layer corrections have not been applied; however it has been demonstrated¹⁸ that the standard potential calculations carried out incorporating the double layer effects amounts to a maximum difference of only 0.06–0.07 V. This fact is particularly significant, since good experimental data are obtained from the glassy carbon electrode—the double layer properties of which are unknown.

4. Summary

The electrochemical reduction of 5-bromo-1,3-dichloro-2iodobenzene results in four irreversible voltammetric waves consuming eight electrons in a single cycle. The reduction of C–I, C–Br and C–Cl (2) bonds lead to parabolic ln $K_{\rm ET}$ versus *E* plots obeying quadratic activation–driving force relationship. The analysis employing the Marcus theory of outer sphere electron transfer in conjunction with the convolution approach yields standard potentials of the reduction of carbon– halogen bonds in 5-bromo-1,3-dichloro-2-iodobenzene except in one of the carbon–chlorine bonds wherein the reduction follows Butler–Volmer kinetics.

Acknowledgements

M.A.P. thanks the CSIR, India for the award of a Senior Research Fellowship.

Supplementary data

Supplementary data associated with this article can be

found, in the online version, at doi:10.1016/j.tet.2004.09. 020

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Tetrahedron

Tetrahedron 60 (2004) 10973-10981

Photoinduced proton exchange between molecular switches

Silvia Giordani, Mabel A. Cejas and Françisco M. Raymo*

Center for Supramolecular Science, Department of Chemistry, University of Miami, 1301 Memorial Drive, FL 33146-0431, USA

Received 5 July 2004; revised 30 August 2004; accepted 8 September 2004

Available online 7 October 2004

Abstract—The identification of strategies to establish communication between independent molecules is an essential requirement for the development of operating principles to manipulate information at the molecular level. In this context, we have devised a strategy to exchange signals between pairs of complementary molecular switches. It is based on the photoinduced ring closing of a merocyanine to produce a spiropyran with the concomitant release of a proton. The liberated proton is captured by either one of two pyridine derivatives with the formation of their conjugate acids. This transformation induces a significant increase in chemical shift for the resonances of the pyridyl protons and, in one instance, also a pronounced color change. The overall process is fully reversible and the pair of communicating molecules reverts to the original state in the dark. Relying on this mechanism, an optical input is transduced into a detectable spectroscopic output after the controlled intermolecular exchange of protons. A simple analysis of the signal transduction operated by the communicating molecular switches reveals that a binary digit is passed unaltered from the input to the output even although the nature of the signal carrying the information changes at each step. Furthermore, the different nature of input and output implies that the state of the ensemble of molecules can be probed non-destructively at any point in time. The timescales of the switching steps, however, are seriously limited by the slow reaction kinetics. The photoinduced transformation occurs within minutes, but the thermal reaction reverts the switch state only after several hours. © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

The identification of experimental strategies to reproduce AND, NOT and OR operations with fluorescent compounds¹ has stimulated the development of chemical systems for digital processing.² The operating principles of these molecular logic gates are based on the interplay between chemical, electrical or optical stimulations and absorbance, luminescence or redox outputs. Generally, electron, energy or proton transfer processes and supramolecular events are invoked to transduce the input stimulations into detectable output changes. Relying on these mechanisms and on collections of relatively simple organic molecules in solution, diverse logic operations have been reproduced successfully.^{3–24} Some of them can even replicate the signal transduction of complex digital circuits composed of several AND, NOT and OR gates.²⁵ The chemical counterparts of these circuits, however, are single molecular switches.²⁶ They combine the functions of multiple logic gates into individual molecular skeletons. This approach to digital processing with molecules is extremely ingenious and fascinating but, at the present stage of development, lacks the modularity of conventional

0040–4020/\$ - see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.09.065

digital circuits. It is not yet possible to connect the molecular equivalents of AND, NOT and OR gates and build multimolecular logic circuits with desired configurations. This severe limitation will be overcome only after the identification of viable strategies to establish communication between independent molecular switches.

In search for efficient protocols to exchange signals between molecular switches, we have identified a mechanism for intermolecular communication based on photon transfer.^{17b,d} Our approach is based on the ability of one of the two states of a photochromic switch²⁷ to absorb the light emitted by a compatible fluorophore. Under these conditions, the optical inputs addressing the photochromic component modulate the emission intensity of the fluorescence component. Relying on these operating principles, we have reproduced the functions of complex logic circuits with multimolecular ensembles of fluorescent and photochromic compounds. In alternative to this mechanism, we have devised a strategy to exchange signals between molecular switches in the form of protons.^{17c,f} We have demonstrated that a photoacid can transfer a proton to either a chromogenic or an electroactive base upon illumination. The photoinduced protonation of the base triggers the release of an optical or electrical output. Overall, optical inputs modulate color or electrical current after the chemical communication established between pairs of compatible molecular switches. We have demonstrated that these processes can be exploited to

Keywords: Logic gates; Molecular switches; Photochromism; Proton transfer; Spiropyrans.

^{*} Corresponding author. Tel.: +1 305 284 2639; fax: +1 305 284 4571; e-mail: fraymo@miami.edu



Figure 1. The reversible interconversion of the spiropyran SP and the protonated merocyanine MEH.

reproduce the memory effects characteristic of sequential logic circuits. In this article, we report a nuclear magnetic resonance (NMR) spectroscopic investigation of the photo-induced exchange of protons between a photochromic spiropyran²⁸ and either one of two pyridine-based switches. In addition, we assess the timescales of these reversible processes by visible absorption spectroscopy and offer an alternative analysis of the signal transduction associated with these chemical systems.

2. Results and discussion

2.1. Design

Our design is based on the established properties of the spiropyran SP (Fig. 1).^{17a,g} In acetonitrile, this compound switches to the protonated merocyanine MEH upon addition of acid. The process is accompanied by the appearance of a yellow-green color, since MEH has an absorption band centered at 401 nm. The irradiation of MEH with light of wavelengths falling in its visible absorption band induces the release of a proton and the formation of the colorless SP. If a species able to capture the released proton is present in the same solution (Fig. 2) then 'chemical' communication, in the form of proton transfer, is established under illumination. Indeed, the photoacid MEH can send a signal to the base after the application of an optical input. The protonated form of the base can then release a detectable output, for example, a color change. Overall, the optical input addressing MEH is transduced into an output signal as a result of intermolecular communication established between two complementary molecular switches.



Figure 2. Transduction of an input stimulation into a detectable output after the transfer of a proton from one molecular switch to another.

2.2. Influence of acidification on the ¹H NMR spectra of two pyridyl switches

The ¹H NMR spectrum (a in Fig. 3) of the azopyridine AZ shows five sets of signals between 6.5 and 9.0 ppm. In addition, a singlet for the 6 equivalent methyl protons is observed at 3.12 ppm. Irradiation at a frequency corresponding to this singlet results in nuclear Overhauser

enhancements (NOE) of +6 and -2% for the resonances at 6.88 and 7.90 ppm associated with the two pairs of phenylene protons (Fig. S1). The absolute values the NOEs suggest that the protons resonating at 6.88 ppm are closer to the methyl groups than those appearing at 7.90 ppm. Thus, we can assign the doublet at 6.88 ppm to the pair of protons H^6 and that at 7.90 ppm to the pair of protons H^3 . The same experiment reveals NOEs of -1 and -4% for the two doublets at 7.67 and 8.66 ppm. These two doublets integrate for one proton each and, one the basis of the NOE values, can be assigned to the pyridyl protons H^4 and H^1 , respectively. A correlated spectroscopy (COSY) experiment shows cross peaks between the doublet for H¹ and the set of signals at 7.39 ppm (Fig. S2), which can be assigned to the pyridyl protons H⁵. Similarly, cross peaks between the doublet for H⁴ and the set of signals at 7.90 ppm indicate that the resonances of the pyridyl proton H^2 overlap those of the phenylene protons H^3 .

The addition of increasing amounts of CF₃CO₂H to a CD₃CN solution of **AZ** alters significantly the chemical shifts of the pyridyl protons (b–e in Fig. 3). The largest change is observed for the doublet of the proton H⁴, which shifts by ca. +0.41 ppm after the addition of 6 equiv of CF₃CO₂H.²⁹ These observations suggest that the acidification results in the protonation of the pyridyl nitrogen atom with the formation of the azopyridinium **AZH**. Consistently, the change in chemical shift for the phenylene (H³ and H⁶) and methyl protons is negligible, indicating that the appended dimethylamino group remains in a deprotonated form under these conditions.

The ¹H NMR spectrum (a in Fig. 4) of the 4,4'pyridylpyridinium cation BI shows singlets for the two methylene protons H^{12} and for the five phenyl protons H^{11} at 5.74 and 7.44 ppm, respectively. Irradiation at a frequency corresponding to the singlet for the methylene protons results in a NOE of +14% for each of the two doublets at 7.72 and 8.28 ppm (Fig. S3). The equal NOE values for both resonances suggest that the distance between the methylene protons H¹² and those associated with the two doublets are very similar. Thus, these signals must correspond to the pairs of pyridinium and pyridyl protons in the β -positions relative to the nitrogen atoms. Consistently, no cross peaks between these two doublets can be observed in the COSY (Fig. S4). Comparison of their chemical shifts with those of the 4,4'-bipyridine and 1,1'-dibenzyl-4,4'-bipyridinium protons suggests that the doublet at 7.72 ppm corresponds to the pair of pyridyl protons H^{10} and that at 8.28 ppm can be assigned to the pair of pyridinium protons H⁹. The remaining multiplet at 8.81 ppm integrates for four protons and must be associated with the two pairs of



Figure 3. Partial ¹H NMR spectra of AZ (5×10^{-3} M, CD₃CN, 25 °C) before (a) and after the addition of 1 equiv (b), 2 equiv (c), 4 equiv (d) and 6 equiv (e) of CF₃CO₂H.

4,4'-pyridylpyridinium protons H^7 and H^8 . In fact, the same NOE experiment revealed a change of 16% for this set of signals as a result of the close proximity of the pair of protons H^7 to the irradiated methylene protons H^{12} .

The addition of increasing amounts of CF₃CO₂H to a CD₃CN solution of **BI** induces a significant change in the chemical shift for the pair of pyridyl protons H^{10} (b–d in Fig. 4). Their doublet moves from 7.72 to 8.28 ppm after the addition of 10 equiv of CF₃CO₂H. These observations suggest that the acidification results in the protonation of the pyridyl nitrogen atom with the formation of the 4,4'-bipyridinium dication **BIH**.²⁹

2.3. Influence of light on the ¹H NMR spectra of two pyridyl switches in the presence of a photoacid

The pyridyl nitrogen atoms of **AZ** and **BI** capture protons when exposed to acid (Figs. 3 and 4). Thus, either one of these two bases can be employed in combination with the photoacid **MEH** to implement the operating principles illustrated in Figure 2. Under the influence of an optical input, **MEH** can transfer a proton to the pyridyl nitrogen atom of either **AZ** or **BI**. The formation of the conjugate acids **AZH** and **BIH** produces a detectable output in the form of a large change in chemical shift for the pyridyl protons H^4 and H^{10} , respectively. The concomitant transformation of **MEH** into **SP** can be followed by ¹H NMR spectroscopy, since the spectra of **SP** and **MEH** (a and b in Fig. 5) differ dramatically.^{17g}

The addition of CF₃CO₂H to an acetonitrile solution of **SP** is followed by the gradual formation of **MEH**, if the solution is maintained in the dark. The process is relatively slow and has a second-order rate constant of only $(54\pm2)\times$ 10^{-5} M⁻¹ s⁻¹.^{17g} For example, ca. 11 d are required for the quantitative conversion of **SP** into **MEH** in the presence of 10 equiv of CF₃CO₂H (a and b in Fig. 5). By contrast, the interconversion of **AZ** and **AZH** in the presence of acid is relatively fast and averaged signals can only be observed in the ¹H NMR spectra (b–e in Fig. 3) at ambient temperature. Thus, the ¹H NMR spectrum (a in Fig. 6), recorded



Figure 4. Partial ¹H NMR spectra (5×10⁻³ M, CD₃CN, 25 °C) of BI before (a) and after the addition of 2 equiv (b), 5 equiv (c) and 10 equiv (d) of CF₃CO₂H.

immediately after mixing equimolar amounts of SP, AZ and CF_3CO_2H in CD_3CN , reveals averaged resonances for AZ and AZH, the characteristic signals of SP but not those of MEH. In particular, a comparison with the ¹H NMR spectrum of AZ (a in Fig. 3) shows a shift of ca. +0.22 ppm for the doublet associated with pyridyl proton H^4 . This chemical shift change corresponds to a ratio between AZ and AZH of ca. 57:43.³⁰ Upon storage in the dark, SP switches gradually to MEH, reducing the concentration of acid available in solution. As a result, AZH is converted into AZ and the chemical shift of the pyridyl proton H^4 decreases. A stationary state is reached in ca. 4 d. The ¹H NMR spectrum (b in Fig. 6), recorded at this point, shows the characteristic resonances of SP and MEH and a chemical shift of 7.79 ppm for the pyridyl proton H^4 . This value corresponds to a ratio between AZ and AZH of ca. 76:24. Upon exposure of the solution to ambient light, MEH switches to SP releasing a proton. This transformation induces the protonation of AZ and the formation of AZH. Consistently, the chemical shift of the pyridyl proton H^4 increases to a stationary value of 7.87 ppm after ca. 8 h. This value corresponds to a ratio between AZ and AZH of ca. 61:39.

The reversible interconversion of **BI** and **BIH** in the presence of acid is also relatively fast. As a result, averaged signals are observed in the ¹H NMR spectra (b–d in Fig. 4) at ambient temperature. Thus, the ¹H NMR spectrum (a in Fig. 7), recorded immediately after mixing equimolar amounts of **SP**, **BI** and CF₃CO₂H, reveals a chemical shift of 8.02 ppm for the pyridyl protons H¹⁰. This value is 0.30 ppm higher than that of the same protons in the ¹H NMR spectrum of **BI** (a in Fig. 4). This significant change corresponds to a ratio between **BI** and **BIH** of ca. 53:47.³⁰ In addition to the averaged signals of **BI** and **BIH**, the characteristic resonances of **SP** can also be observed in the



Figure 5. Partial ¹H NMR spectra (5×10^{-3} M, CD₃CN, 25 °C) of SP before (a) and after (b) the addition of CF₃CO₂H (10 equiv) and storage in the dark for 11 d.

initial ¹H NMR spectrum (a in Fig. 7). Upon storage in the dark, **SP** switches gradually to **MEH** reducing the concentration of acid available in solution and inducing the transformation of **BIH** into **BI**. A stationary state is reached after ca. 6 d. Under these conditions, the resonances of both **SP** and **MEH** are clearly visible in the ¹H NMR spectrum (b in Fig. 7). Furthermore, the chemical shift for the pyridyl protons H¹⁰ is now 7.82 ppm. This value corresponds to a ratio between **BI** and **BIH** of ca. 84:16. Upon exposure of the solution to ambient light, **MEH** switches to **SP** releasing a proton, which induces the transformation of **BI** back to **BIH** and a decrease in the

chemical shift of the pyridyl protons H^{10} . A stationary state is reached after ca. 2 h. The ¹H NMR spectrum recorded at this point shows a chemical shift of 7.98 ppm for the probe protons H^{10} , indicating a ratio between **BI** and **BIH** of ca. 60:40.

2.4. Influence of light on the visible absorption spectra of two pyridyl switches in the presence of a photoacid

The visible absorption spectrum (a in Fig. 8) of AZ shows a band centered at 423 nm in acetonitrile.^{17c} This band fades upon addition of increasing amounts of CF_3CO_2H .



Figure 6. Partial ¹H NMR spectra (1.5×10^{-2} M, CD₃CN, 25 °C) recorded immediately after mixing equimolar amounts of **SP**, **AZ** and CF₃CO₂H (a) and after the subsequent storage in the dark for 4 d (b).



Figure 7. Partial ¹H NMR spectra (1.5×10^{-2} M, CD₃CN, 25 °C) recorded immediately after mixing equimolar solution of **SP**, **BI** and CF₃CO₂H (a) and after the subsequent storage in the dark for 6 d (b).



Figure 8. (a) Absorption spectra recorded before and after the addition of increasing amounts of CF₃CO₂H (0.5–13 equiv) to **AZ** (1×10^{-4} M, MeCN, 25 °C). (b) Evolution of the absorption spectrum from the photostationary state to the thermal equilibrium of a solution prepared by mixing equimolar amounts of **SP**, **AZ** and CF₃CO₂H (1×10^{-4} M, MeCN, 25 °C). (c) Time dependence of the absorbance at 554 nm during storage in the dark and the subsequent visible irradiation.

Concomitantly, a new band corresponding to AZH grows at 554 nm. As a result, the transformation of AZ into AZH is accompanied by a change in color from orange to purple. In the presence of SP and MEH, the interconversion of AZ and AZH can be followed by monitoring the absorbance of either one of these two visible bands. Indeed, the evolution of the absorption spectrum (b in Fig. 8) from the photostationary state to the thermal equilibrium of a solution prepared by mixing equimolar amounts of SP, AZ and CF₃CO₂H shows an absorbance increase at 423 nm with a concomitant absorbance decrease at 554 nm. These changes indicate that AZH switches gradually to AZ in the dark and are a consequence of the thermal interconversion of SP into MEH. The time dependence (c in Fig. 8) of the absorbance at 554 nm reveals the transition from the photostationary state to the thermal equilibrium to require ca. 24 h. This absorbance decay corresponds to an increase in the ratio between AZ and AZH from ca. 57:43 to 70:30.³¹ The photostationary absorbance value is fully restored after visible irradiation for ca. 15 min (c in Fig. 8). The photoinduced absorbance enhancement indicates that AZ switches back to AZH, as the photoacid MEH reverts to SP releasing a proton.

The base **BI** and its conjugate acid **BIH** do not absorb in the visible region. In the presence of **SP**, however, their interconversion can be monitored by probing the absorbance for the characteristic visible band of **MEH**. Indeed, the evolution of the absorption spectrum (a in Fig. 9) from the photostationary state to the thermal equilibrium of a solution prepared by mixing equimolar amounts of **SP**, **BI** and CF₃CO₂H shows an absorbance increase at 401 nm. Under these conditions, **SP** switches gradually to **MEH** and encourages the concomitant transformation of **BIH** into **BI**. The process requires ca. 24 h to reach thermal equilibrium (b in Fig. 9). Under visible irradiation, however, **MEH** reverts to **SP** releasing its proton and promoting the reconversion of **BI** into **BIH**. The photostationary state is



Figure 9. (a) Evolution of the absorption spectrum from the photostationary state to the thermal equilibrium of a solution prepared by mixing equimolar amounts of **SP**, **BI** and CF₃CO₂H (1×10^{-4} M, MeCN, 25 °C). (b) Time dependence of the absorbance at 401 nm during storage in the dark and the subsequent visible irradiation.

restored in ca. 15 min (b in Fig. 9). The ratio between **SP** and **MEH** at the photostationary state and at the thermal equilibrium can be estimated to be 96:4 and 64:36.³²

2.5. Analysis of the signal transduction operated by the two pairs of communicating switches

In the presence of CF₃CO₂H, **SP** switches to **MEH** (Fig. 1) consuming some of the H⁺ available in solution. Upon visible irradiation, MEH reverts to SP releasing back the captured H⁺. Thus, the concentration of H⁺ (chemical output) switches to a high value as the light (optical input) is turned on (a in Fig. 10). It returns to a low value when the light is turned off. Under a positive logic convention, we can assign a binary 0 to the off and low states of input (I) and output (**O**) and a binary 1 to their on and high states. A glance at the corresponding truth table (c in Fig. 10) reveals that this particular molecular switch transfers unaltered the binary value encoded in the optical input to the chemical output. It is important to stress, however, that the timescales of the switching steps are seriously limited by the relatively slow reaction kinetics. In particular, the thermal transformation of SP into MEH has a small second-order rate constant^{17g} and requires several hours to occur. Similarly,

а		Ь		С		
Light	H⁺	H+	δ	I	0	
Off	Low	Low	Low	0	0	
On	High	High	High	1	1	

Figure 10. Signal transductions operated by SP/MEH (a) and either AZ/AZH or BI/BIH (b) and equivalent truth table (c).

the photoinduced interconversion of **MEH** into **SP** has a modest quantum yield and occurs within several minutes.

The concentration of H^+ controls the interconversion of AZ and BI into the corresponding conjugate acids (Figs. 3 and 4). Both transformations induce significant changes in the chemical shifts (δ) of probe protons. Thus, the chemical shift (spectroscopic output) switches to a high value when the concentration of H^+ (chemical input) is high (b in Fig. 10). It reverts to a low value when the concentration of H^+ is low. Once more, we can assign a binary 0 to the low values of input (I) and output (O) and a binary 1 to their high values, under a positive logic convention. Again, the corresponding truth table (c in Fig. 10) shows that a binary digit is passed unaltered from the chemical input to the spectroscopic output.³³

When the molecular switch **SP/MEH** is operated together with either **AZ/AZH** or **BI/BIH**, the concentration of H^+ becomes a common signal to both systems. Indeed, the binary digit encoded in the optical input addressing **SP/ MEH** (a in Fig. 10) is communicated chemically to either **AZ/AZH** or **BI/BIH** and passed to the final spectroscopic output (b in Fig. 10). Thus, a binary value travels unaltered through a sequence of two communicating molecular switches, while the nature of the signal carrying the information is converted from light, to H^+ and finally to chemical shift.

It is worth stressing that the communication between **SP**/**MEH** and **AZ/AZH** can also be monitored by following the absorbance changes at 554 nm. At this particular wavelength, neither **SP** nor **MEH** absorb. Thus, the state of the photochromic component **SP/MEH** can be probed optically, but non-destructively, through its chromogenic partner **AZ/AZH**. In fact, the need to read the state of a photochromic switch without altering it has been one of the major limitations in the development of optical memories based on this class of compounds.²⁷ It follows that our protocol for signal transduction might also contribute to the identification of viable solutions to this challenging problem.

3. Conclusions

We have identified a mechanism to establish communication between pairs of molecular switches. It is based on the reversible interconversion of a spiropyran and a protonated merocyanine in the presence of acid and under the influence of visible light. In the dark, the spiropyran switches to the merocyanine consuming H⁺. Under illumination, the process is fully reversed with the liberation of H⁺. The capture and release of H⁺ can be exploited to regulate the interconversion of either one of two pyridyl switches. Their protonation produces the corresponding conjugate acids and induces a significant enhancement in the chemical shift of the pyridyl protons. Thus, an optical input can modulate a chemical shift output on the basis of intermolecular proton exchange. Furthermore, these processes can be monitored by visible absorption spectroscopy, following the absorbance changes associated with the bands of the protonated form of one of the two communicating switches. These analyses indicate that the transition from the photostationary state to the thermal equilibrium requires approximately one day in acetonitrile at submillimolar concentrations. The opposite and photoinduced transformation, instead, is considerably faster and the photostationary state is restored within minutes. The timescales of both processes are certainly not compatible with any practical computing scheme. These results, however, demonstrate that relatively simple reactions can be coupled, with a judicious choice of compatible molecular components, to transduce incoming optical inputs in well-defined spectroscopic outputs. Methods to reduce the switching times by several orders of magnitude now need to be identified. Under these conditions, our strategy might be employed to exchange effectively signals between collections of molecules operating in solution.

4. Experimental procedures

4.1. General methods

Chemicals were purchased from commercial sources. Acetonitrile (MeCN) was distilled over calcium hydride (CaH₂). The spiropyran **SP** and the 4,4'-pyridylpyridinum cation **BI** were prepared according to literature procedures.^{17f,g 1}H NMR spectra were recorded with a Bruker Avance 400 (400 MHz) using quartz tubes (diameter = 5 mm). Absorption spectra were recorded with a Varian Cary 100 Bio using quartz cells (path length = 5 mm). The cells were irradiated with a Cole-Parmer Fiber Optic Illuminator 9745-00 coupled to a band pass filter (360–650 nm).

Acknowledgements

We thank the National Science Foundation (CAREER Award CHE-0237578) for financial support and the University of Miami for a Robert E. Maytag Fellowship to S.G.

Supplementary data

Difference nuclear Overhauser enhancement (NOE) and ${}^{1}\text{H}{-}^{1}\text{H}$ correlated spectroscopy (COSY) spectra of AZ and **BI**. Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tet.2004.09. 065

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- 29. Analyses of the chemical shift changes with the concentration of CF_3CO_2H indicate the limiting values ($\Delta\delta_L$) to be 0.51 and 0.65 ppm for AZ and BI, respectively.
- 30. The **AZ/AZH** and **BI/BIH** ratios were determined from the molar fractions (χ) of **AZH** and **BIH**, respectively. The χ values are equal to the ratios between the observed ($\Delta \delta_{O}$) and limiting ($\Delta \delta_{L}$) chemical shift changes of the probe protons. The values of $\Delta \delta_{L}$ are quoted in Ref. 29.
- 31. The ratios were calculated from the absorbance values at 554 nm, considering molar extinction coefficients for AZ and AZH of ca. 600 and 46,000 M^{-1} cm⁻¹, respectively, at this wavelength. The molar extinction coefficient of AZH was determined by non-linear curve fitting of the absorbance profile at 554 nm (a in Fig. 8) of a solution of AZ treated with increasing amounts of CF₃CO₂H.
- 32. The ratios were calculated from the absorbance values at 401 nm, considering molar extinction coefficients for **SP** and **MEH** of ca. 200 and 32,000 M^{-1} cm⁻¹, respectively, at this wavelength (Ref. 17g).
- 33. The spectroscopic output (chemical shift) chosen to illustrate the signal transduction mechanism requires a rather cumbersome detection protocol (¹H NMR spectroscopy). However, these operating principles are not limited to chemical shift measurements and can be easily extended to more practical electrical or optical detection methods. In fact, the signal transduction associated with the combination of SP/MEH and AZ/AZH results in an optical output, which can be monitored with simple transmission absorption measurements.



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Tetrahedron

Tetrahedron 60 (2004) 10983-10992

Stepwise and one-pot cross-coupling–heteroannulation approaches toward 2-substituted C5-, C6-, and C7-nitroindoles^{\star}

Li-Ping Sun,^b Xiang-Hong Huang^b and Wei-Min Dai^{a,b,*}

^aLaboratory of Asymmetric Catalysis and Synthesis, Department of Chemistry, Zhejiang University, Hangzhou 310027, China ^bCombinatorial Chemistry Laboratory, The Biotechnology Research Institute and Department of Chemistry, The Hong Kong University of Science and Technology, Clear Water Bay, Kowloon, Hong Kong SAR, China

Received 22 June 2004; revised 25 August 2004; accepted 1 September 2004

Available online 30 September 2004

Abstract—A general and efficient synthesis of 2-substituted C5-, C6-, and C7-nitroindoles has been established. Starting from commercially available 2-amino nitrophenols, C5-, C6-, and C7-nitroindoles were synthesized via the stepwise Pd-catalyzed cross-coupling of nitro 2-trifloxyanilides with 1-alkynes followed by the *t*-BuOK-mediated heteroannulation. A Pd-catalyzed one-pot coupling–heteroannulation procedure was carried out by using nitro 2-trifluoroacetamidoaryl triflates. © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

Nitroindoles are known to be biologically active¹ and serve as the useful precursors to a variety of nitrogen-substituted indole derivatives, which exhibit diverse regulatory activities on bio-macromolecules. Selected examples are given in Figure 1, including SDZ-216525 (a 5-HT₇ selective agonist),² Delavirdine (a non-nucleoside reverse



Figure 1. Selected bioactive indoles possessing a nitrogen-based substituent.

* Part 4 of Chemistry of Aminophenols. Some of the results were communicated in a preliminary report, see Ref. 20j.

Keywords: Cross-coupling; Heteroannulation; Nitroindoles; Aryl triflates; 2-Aminophenols.

* Corresponding author. Tel.: +86 571 87953128; fax: +86 571 87953128/+852 23581594; e-mail addresses: chdai@zju.edu.cn, chdai@ust.hk



Scheme 1. Synthesis of 2-alkynylanilines 4-6.

transcriptase inhibitor approved for HIV therapy),³ Zafirlukast (an LTD₄ antagonist),⁴ LY334370 (a selective 5-HT_{1F} receptor agonist),⁵ and E7070 (an anticancer agent thought to affect the progression of the cell cycle in the G1 phase with inhibition of expression of cyclin E and phosphorylation of cdk2).⁶ Direct nitration of indoles is a straightforward method for synthesis of nitroindoles,^{1,7} but it suffers from non-selectivity in some cases.¹ A number of methodologies⁸ have been used to form nitroindoles. These cover the Fischer indole synthesis,⁹ the Bergman indole synthesis,¹⁰ and various nucleophilic/electrophilic cyclization approaches by using the nitro-containing substrates.¹¹ However, low yields and/or formation of mixtures of nitroindoles were reported.^{9e,f,11c-e}

The metal-catalyzed cross-coupling reactions have emerged as the most powerful methods for carbon-carbon bond formation in recent years and they have been used for heterocycle synthesis.¹² Among the metal-catalyzed indole syntheses,⁸ the cross-coupling and heteroannulation procedures enjoy wide applications. 2-Haloanilines and derivatives are the common substrates, which are transformed into 2-alkynylanilines via Pd(0)-Cu(I)-catalyzed Sonogashira cross-coupling with 1-alkynes¹³ followed by metal-catalyzed, $^{13-19}$ or base- $^{20-22}$ and iodonium(I)mediated²³ heteroannulation. Alternatively, 2-alkynylanilines undergo aminopalladation-reductive elimination with aryl/vinyl iodides and triflates to form highly functionalized indoles.^{24,25} Moreover, 2-iodoanilines react with internal alkynes via Pd-catalyzed annulation to afford 2,3-disubstituted indoles (Larock indole synthesis).14,26 There are two reports on synthesis of C5- and C7-nitroindoles through the cross-coupling and heteroannulation procedures.^{17f,20h,k} In both cases, 2-iodo nitroanilines were used as the starting materials, which were synthesized from nitroanilines by using bis(pyridine)iodonium(I) tetrafluoroborate (IPy₂BF₄) as the iodinating agent.^{17f} The nitro-containing 2-alkynylanilines cyclized to nitroindoles via CuI- and *t*-BuOK-mediated heteroannulation, respectively.^{17f,20h,k} We report here on a general and efficient synthesis of 2-substituted C5-, C6-, and C7-nitroindoles starting from commercially available 2-amino nitrophenols.^{20j}

2. Results and discussion

2-Iodoanilines could be prepared from 4-substituted anilines by using IPy_2BF_4 as the iodinating agent.^{17f} However, the C4 substituent is necessary for directing the iodination at C2.²⁷ Alternatively, the oxygen-substituted 2-iodoanilines could be prepared from protected aminophenols by directed *ortho*-lithiation followed by trapping with I_2 .^{20g} We used the nitro 2-trifloxyanilides 3 to substitute 2-iodoanilines in the metal-catalyzed indole synthesis. Before our work, 20i, j a 2-trifloxyanilide was used for the Pd-catalyzed crosscoupling with alkynylstannanes to provide 2-alkynylanilides.^{15g} The Pd(0)–Cu(I)-catalyzed Sonogashira crosscoupling of 2-trifloxyanilides with 1-alkynes was virtually unknown. We found a remarkable additive effect on the cross-coupling of 2-trifloxyanilides with 1-alkynes and high yields of 2-alkynylanilides were obtained in the presence of 1.5 equiv of n-Bu₄NI.²⁰ⁱ As shown in Scheme 1, three nitro 2-trifloxyanilides 3a-c were readily prepared from 2-amino-5-nitrophenol (1a), 2-amino-4-nitrophenol (1b), and 2-amino-3-nitrophenol (1c) via selective N-acylation²⁸ followed by formation of triflates from **2a–c**. The butyryl anilides were selected for their relatively good solubility in common organic solvents compared with the acetamides. Cross-coupling of 3a-c with 1-alkynes took place at room temperature in CH₃CN-Et₃N (5:1) with 1.5 equiv of n-Bu₄NI as the additive to give the 2-alkynyl nitroanilides 4-6 in excellent yields. The results are summarized in Table 1.

Table 1. Cross-coupling of aryl triflates 3 with 1-alkynes^a

Entry	Aryl triflate	R	Yield (%)
1	3a : 5-NO ₂	Ph	4a : 95
2	3a : 5-NO ₂	<i>n</i> -Pr	4b : 95
3	3b : 4-NO ₂	Ph	5a : 96
4	3b : 4-NO ₂	<i>n</i> -Pr	5b : 90
5	3c : 3-NO ₂	Ph	6a : 90
6	3c : 3-NO ₂	<i>n</i> -Pr	6b : 91

 $^{\rm a}$ Carried out with 10 mol% Pd(PPh_3)_4, 30 mol% CuI, and 150 mol% $n{\rm -}\,{\rm Bu_4NI}$.





Dirity	1 (lu olindole)		11010 (70)
1	5-NO ₂	Ph	7a : 85
2	5-NO ₂	<i>n</i> -Pr	7b : 84
3	6-NO ₂	Ph	8a : 84
4	$6-NO_2$	<i>n</i> -Pr	8b : 86
5	$7-NO_2$	Ph	9a : 76
6	$7-NO_2$	<i>n</i> -Pr	9b : 72

^a 1.2 equiv of *t*-BuOK were used.

Heteroannulation of 2-alkynyl nitroanilides **4–6** was carried out in NMP with 1.2 equiv of *t*-BuOK at 60–70 °C for 7 h to furnish 2-substituted C5-, C6-, and C7-nitroindoles **7–9** in 72–86% yields (Table 2). In Knochel's nitroindole synthesis, a unprotected 2-alkynylaniline was used.^{20h} We found that the *N*-acyl group, the position of nitro group, and the nature of R in **4–6** did not affect the efficiency of the base-mediated heteroannulation.²⁹ Nitroindoles **7–9** are all crystalline compounds; the C6-nitroindole **8b** was analyzed by X-ray crystallography, showing a flat nitroindole skeleton. 30

Next, we examined a one-pot cross-coupling and heteroannulation procedure toward nitroindoles 7-9 as shown in Scheme 2. A Cu(I)-catalyzed one-pot synthesis of indoles from 2-iodotrifluoroacetanilide and 1-alkynes was recently reported.^{17h} In our nitroindole synthesis, the trifluoroacetamidoaryl triflates 11a-c were prepared in good yields by selective N-acylation of 1a-c followed by triflate formation from 10a-c and PhNTf₂. The one-pot cross-coupling and heteroannulation of 10a-c with various 1-alkynes was investigated and the results are given in Table 3. We observed solvent effect on the one-pot reactions and used three solvent combinations, A (DMF-Et₃N=5:1), B (CH₃CN-Et₃N=5:1), and C (DMF-TMG=5:1). For formation of C5- and C6-nitroindoles 7 and 8 (entries 1-10, Table 3), solvents A and B could be used and the product yields are comparable to those given in Table 2. However, the one-pot reaction times were significantly prolonged than the stepwise protocol. Under the one-pot reaction conditions, some functional groups such as OH, CN, and Cl remained intact and protection of OH was not required. Formation of C7-nitroindoles 9 by the one-pot procedure was somewhat problematic. After screening on catalyst precursor, solvent, and base, three C7-nitroindoles 9a-c could be obtained albeit in lower yields (entries 11-13,



Scheme 2. One-pot synthesis of nitroindoles 7–9.

Table 3.	One-pot synt	hesis of	nitroindoles	via Pd	l-catalyzed	coupling a	nd	heteroannul	ation	2
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Entry	Nitroindoles	R	<i>t</i> (h)	Yield (%)
1	5-NO ₂	Ph	41	7a: 85
2	5-NO ₂	<i>n</i> -Pr	24	7b : 90
3 ^b	5-NO ₂	(CH ₂) ₂ OH	42	7c : 68
4	5-NO ₂	(CH ₂) ₃ CN	25	7d : 75
5 ^b	5-NO ₂	(CH ₂) ₃ Cl	24	7e : 45
6 ^b	6-NO ₂	Ph	17	8a : 88
7	6-NO ₂	<i>n</i> -Pr	21	8b : 84
8	6-NO ₂	(CH ₂) ₂ OH	19	8c : 69
9	6-NO ₂	(CH ₂) ₃ CN	8	8d : 73
10 ^b	6-NO ₂	(CH ₂) ₃ Cl	20	8e : 66
11 ^{c-e}	7-NO ₂	Ph	3.5	9a : 52
12 ^{c,d}	7-NO ₂	<i>n</i> -Pr	7.5	9b : 39
13 ^d	7-NO ₂	(CH ₂) ₂ OH	12	9c : 48

^a Carried out in solvent A (DMF-Et₃N=5:1) with 10 mol% Pd(PPh₃)₄, 30 mol% CuI, and 150 mol% *n*-Bu₄NI.

^b Carried out in solvent B (CH₃CN-Et₃N=5:1).

^c Carried out in solvent C (DMF-TMG=5:1).

^d Pd(PPh₃)₂Cl₂ was used to replace Pd(PPh₃)₄.

^e Carried out at 100 °C. TMG=1,1,3,3-tetramethylguanidine.
Table 3). Use of $Pd(PPh_3)_2Cl_2$ as the catalyst precursor seems essential along with a stronger base, TMG. Higher reaction temperature (100 °C) was also applied for the formation of **9a**. These results suggest that the *ortho* nitro group in **11c** may form hydrogen bond with the amido moiety, thus interfering the indole ring closure reaction. This type of intramolecular hydrogen bond would be destroyed when a strong base such as *t*-BuOK was used for the heteroannulation. Therefore, no difference was recognized among the stepwise synthesis of C5-, C6-, and C7-nitroindoles.

3. Conclusion

In summary, we have established general and efficient cross-coupling-heteroannulation procedures toward 2-substituted C5-, C6-, and C7-nitroindoles starting from the commercially available 2-amino nitrophenols. Both stepwise and one-pot protocols were examined and the former approach seems much more reliable and tolerant to the position of nitro group on the benzene ring. Nitroindoles can be converted, by hydrogenation over Pd/C, into aminoindoles,^{20j} which are useful precursors to a variety of indole derivatives possessing nitrogen-based ring substituents.

4. Experimental

¹H and ¹³C NMR spectra were recorded in CDCl₃ or acetone- d_6 (300 MHz for ¹H and 75 MHz for ¹³C, respectively) with CHCl₃ or acetone as the internal reference. IR spectra were taken on a FT-IR spectrophotometer. Mass spectra (MS) were measured by the +CI method. Elemental analyses were performed by Zhejiang University and Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences. All reactions were carried out under a nitrogen atmosphere and monitored by thinlayer chromatography on 0.25-mm E. Merck silica gel plates (60 F-254) using UV light, or 7% ethanolic phosphomolybdic acid and heating as the visualizing methods. E. Merck silica gel (60, particle size 0.040-0.063 mm) was used for flash column chromatography. Yields refer to chromatographically and spectroscopically (¹H NMR) homogeneous materials. Reagents were obtained commercially and used as received. Room temperature is around 20 °C.

4.1. Representative procedure for the acylation of 2-amino nitrophenols 1a–c

4.1.1. *N*-(2'-Hydroxy-5'-nitrophenyl)butyramide (2b). To a solution of 2-amino-4-nitrophenol 1b (462.0 mg, 3.00 mmol) in dry THF (25 mL) cooled in an ice-water bath was added pyridine (0.30 mL, 3.80 mmol) and butyryl chloride (0.34 mL, 3.30 mmol) through a syringe, respectively. The resultant mixture was stirred for 60 h at refluxing temperature under a nitrogen atmosphere. The reaction was quenched by water and the resultant mixture was extracted with EtOAc (30×3 mL). The combined organic layer was washed with brine, dried over anhydrous MgSO₄, and evaporated under reduced pressure. The residue was purified by flash column chromatography (silica gel, 25%)

EtOAc in hexane) to give **2b** (650.0 mg, 97%) as a pale yellow crystalline solid; mp 191–192 °C (CH₂Cl₂–hexane); R_f =0.53 (33% EtOAc in hexane); IR (film) 3406, 1646, 1529, 1498, 1343, 1289 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 10.85 (br s, 1H), 9.24 (br s, 1H), 8.99 (d, J=2.8 Hz, 1H), 8.06 (dd, J=8.9, 2.8 Hz, 1H), 7.20 (d, J=9.0 Hz, 1H), 2.68 (t, J=7.4 Hz, 2H), 1.88 (sextet, J=7.4 Hz, 2H), 1.13 (t, J=7.4 Hz, 3H); ¹³C NMR (75 MHz, acetone- d_6) δ 172.8, 152.8, 140.2, 126.9, 120.2, 116.3, 115.8, 37.9, 18.4, 12.7; MS (+CI) m/z 225 (M+H⁺, 100). Anal. Calcd for C₁₀H₁₂N₂O₄: C, 53.57; H, 5.39; N, 12.49. Found: C, 53.81; H, 5.40; N, 12.31%.

4.1.2. *N*-(2'-Hydroxy-4'-nitrophenyl)butyramide (2a). Prepared in 86% yield from 2-amino-5-nitrophenol **1a**. *Compound* **2a**. A colorless crystalline solid; mp 175–176 °C (CH₂Cl₂–hexane); $R_{\rm f}$ =0.51 (50% EtOAc in hexane); IR (film) 3409, 1664, 1504, 1421, 1341 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 9.22 (br s, 1H), 8.36 (d, *J*=8.8 Hz, 1H), 7.94–7.87 (m, 2H), 2.69 (t, *J*=7.4 Hz, 2H), 1.87 (sextet, *J*=7.4 Hz, 2H), 1.12 (t, *J*=7.4 Hz, 3H); ¹³C NMR (75 MHz, acetone- d_6) δ 172.5, 146.3, 143.2, 133.2, 119.6, 115.4, 110.1, 38.1, 18.3, 12.7; MS (+CI) *m/z* 225 (M+H⁺, 100). Anal. Calcd for C₁₀H₁₂N₂O₄: C, 53.57; H, 5.39; N, 12.49. Found: C, 53.62; H, 5.48; N, 12.38%.

4.1.3. *N*-(2'-Hydroxy-6'-nitrophenyl)butyramide (2c). Prepared in 60% yield from 2-amino-3-nitrophenol 1c using NaH to replace pyridine as the base (room temperature, 24 h). *Compound* 2c. A yellow crystalline solid; mp 119–120 °C (CH₂Cl₂–hexane); $R_{\rm f}$ =0.34 (25% EtOAc in hexane); IR (film) 3395, 3141 (br), 1663, 1540, 1511, 1367 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 10.09 (br s, 1H), 9.32 (s, 1H), 7.71 (dd, *J*=8.4, 1.5 Hz, 1H), 7.36 (dd, *J*=8.2, 1.5 Hz, 1H), 7.24 (t, *J*=8.3 Hz, 1H), 2.56 (t, *J*=7.4 Hz, 2H), 1.82 (sextet, *J*=7.4 Hz, 2H), 1.04 (t, *J*=7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 175.0, 151.2, 141.2, 127.1, 126.1, 122.2, 117.9, 39.4, 19.0, 13.5; MS (+CI) *m*/z 225 (M+H⁺, 100). Anal. Calcd for C₁₀H₁₂N₂O₄: C, 53.57; H, 5.39; N, 12.49. Found: C, 53.72; H, 5.45; N, 12.42%.

4.2. Representative procedure for the synthesis of nitroaryl triflates using NaH as the base

4.2.1. N-[5'-Nitro-2'-(((trifluoromethane)sulfonyl)oxy)phenyl]butyramide (3b). To a suspension of NaH (60%, 8.0 mg, 2.03 mmol) in dry MeCN (15 mL) cooled in an icewater bath under a nitrogen atmosphere was added a solution of **2b** (363.0 mg, 1.62 mmol) in dry MeCN (30 mL) followed by stirring at the same temperature for 20 min. Tf₂O (0.30 mL, 1.78 mmol) was then added dropwise, and the resultant mixture was stirred for 6 h at -5-0 °C. The reaction was quenched by water and the resultant mixture was extracted with EtOAc (30×2 mL). The combined organic layer was washed with 5% aqueous HCl, saturated aqueous NaHCO₃ and brine, dried over anhydrous MgSO₄, and evaporated under reduced pressure. The residue was purified by flash column chromatography (silica gel, 25%) EtOAc in hexane) to give 3b (461.0 mg, 80%) as a white crystalline solid; mp 73–74 °C (CH₂Cl₂–hexane); $R_{\rm f}$ =0.51 (25% EtOAc in hexane); IR (film) 3271 (br), 2969, 1681, 1542, 1430, 1217 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ

9.30 (d, J=2.7 Hz, 1H), 8.04 (dd, J=9.0, 2.7 Hz, 1H), 7.55–7.47 (br s, 1H), 7.48 (d, J=9.0 Hz, 1H), 2.45 (t, J=7.4 Hz, 2H), 1.79 (sextet, J=7.4 Hz, 2H), 1.03 (t, J=7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.3, 148.6, 141.9, 131.6, 122.1, 119.6, 118.5, 118.3 (q, $J_{C-F}=$ 318.4 Hz), 39.4, 18.5, 13.6; MS (+CI) *m/z* 357 (M+H⁺, 100). Anal. Calcd for C₁₁H₁₁F₃N₂O₆S: C, 37.08; H, 3.11; N, 7.86. Found: C, 37.45; H, 3.13; N, 7.77%.

4.2.2. N-[4'-Nitro-2'-(((trifluoromethane)sulfonyl)oxy)phenyl]butyramide (3a). Prepared from 2a in 87% yield. Compound 3a. A colorless crystalline solid; mp 88–89 °C (EtOAc); $R_f = 0.45$ (25% EtOAc in hexane); IR (film) 3274 (br), 2972, 1690, 1516, 1435, 1350, 1217 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{acetone-}d_6) \delta 9.68 \text{ (br s, 1H)}, 8.68-8.48 \text{ (m, 3H)},$ 2.70 (t, J = 7.4 Hz, 2H), 1.88 (sextet, J = 7.4 Hz, 2H), 1.12 (t, J=7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.5, 142.7, 138.8, 136.8, 123.9, 123.6, 118.3 (q, $J_{C-F}=$ 318.4 Hz), 117.4, 38.0, 17.9, 12.7; MS (+CI) m/z 357 $(M+H^+, 100)$. Anal. Calcd for $C_{11}H_{11}F_3N_2O_6S$: C, 37.08; H, 3.11; N, 7.86. Found: C, 37.40; H, 3.21; N, 7.66%.

4.3. Procedure for the synthesis of nitroaryl triflate using Et₃N as the base

4.3.1. N-[6'-Nitro-2'-(((trifluoromethane)sulfonyl)oxy)phenyl]butyramide (3c). To a solution of 2c (395.0 mg, 1.76 mmol) and Et₃N (0.30 mL, 2.20 mmol) in dry CH₂Cl₂ (20 mL) cooled in an ice-water bath under a nitrogen atmosphere was added Tf₂O (0.33 mL, 1.93 mmol) dropwise. The resultant mixture was stirred at the same temperature for 7 h. After removal of CH₂Cl₂ under reduced pressure, the residue was dissolved in 25 mL EtOAc and then washed with 5% aqueous HCl, saturated aqueous NaHCO₃, and brine. The organic layer was dried over anhydrous MgSO₄ and evaporated under reduced pressure. The residue was purified by flash column chromatography (silica gel, 25% EtOAc in hexane) to give 3c (590.0 mg, 94%) as a white crystalline sold; mp 119-119.5 °C (CH₂Cl₂-hexane); $R_f = 0.40$ (25% EtOAc in hexane); IR (film) 3248 (br), 2973, 1676, 1541, 1518, 1425, 1209 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.18 (br s, 1H), 8.10 (dd, J=8.4, 1.4 Hz, 1H), 7.62 (dd, J=8.4, 1.4 Hz, 1H), 7.48 (t, J=8.4 Hz, 1H), 2.46 (t, J=7.4 Hz, 2H), 1.77 (sextet)J=7.4 Hz, 2H), 1.02 (t, J=7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.2, 145.1, 145.0, 127.5, 126.9, 126.1, 124.9, 118.4 (q, J_{C-F} =318.5 Hz), 38.5, 18.4, 13.6; MS (+CI) m/z 357 (M+H⁺, 100). Anal. Calcd for C₁₁H₁₁F₃N₂O₆S: C, 37.08; H, 3.11; N, 7.86. Found: C, 37.16; H, 3.15; N, 8.06%.

4.4. Representative procedure for the cross-coupling of triflates with 1-alkynes

4.4.1. *N*-[(5'-Nitro-2'-phenylethynyl)phenyl]butyramide (5a). To a suspension of triflate 3b (142.0 mg, 0.40 mmol), $Pd(PPh_3)_4$ (46.0 mg, 4.0×10^{-2} mmol), CuI (23.0 mg, 0.12 mmol), and *n*-Bu₄NI (222.0 mg, 0.60 mmol) in degassed dry MeCN (5 mL) was added Et_3N (1.0 mL) and phenylacetylene (90 µL, 0.80 mmol), respectively, through a syringe under a nitrogen atmosphere. The resultant mixture was stirred at room temperature for 1 h. The reaction was quenched by saturated aqueous NH₄Cl and the resultant mixture was extracted with EtOAc (20×2 mL). The combined organic layer was washed with brine, dried over anhydrous MgSO₄, and concentrated under reduced pressure. The residue was purified by flash column

chromatography (silica gel, 30% EtOAc in hexane) to give 5a (126.0 mg, 96%) as a yellow crystalline solid; mp 168–169 °C (CH₂Cl₂-hexane); $R_{\rm f}$ =0.44 (25% EtOAc in hexane); IR (film) 3290, 2960, 2214, 1665, 1534, 1341 cm⁻ ¹H NMR (300 MHz, CDCl₃) δ 9.36 (d, J = 2.2 Hz, 1H), 8.09 (br s, 1H), 7.92 (dd, J=8.5, 2.2 Hz, 1H), 7.62 (d, J=8.5 Hz, 1H), 7.58–7.55 (m, 2H), 7.47–7.40 (m, 3H), 2.47 (t, J=7.4 Hz, 2H), 1.82 (sextet, J=7.4 Hz, 2H), 1.05 (t, J=7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.3, 147.9, 139.5, 132.0, 131.7 (×2), 129.9, 128.8 (×2), 121.2, 118.0, 117.8, 114.3, 100.8, 82.8, 39.8, 18.8, 13.7; MS (+CI) m/z $309 (M + H^+, 100)$. Anal. Calcd for $C_{18}H_{16}N_2O_3$: C, 70.12; H, 5.23; N, 9.09. Found: C, 69.75; H, 6.00; N, 9.20%.

4.4.2. *N*-[(4'-Nitro-2'-phenylethynyl)phenyl]butyramide (4a). Prepared from the triflate 3a and 1-phenylacetylene in 95% yield. Compound 4a. A colorless crystalline solid; mp 149–150 °C (CH₂Cl₂-hexane); $R_f = 0.54$ (25% EtOAc in hexane); IR (film) 3306, 2959, 2212, 1677, 1574, 1506 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.68 (d, J = 9.2 Hz, 1H), 8.38 (d, J = 2.6 Hz, 1H), 8.25 (br s, 1H), 8.20 (dd, J = 8.6, 2.6 Hz,1H), 7.58–7.55 (m, 2H), 7.47–7.40 (m, 3H), 2.49 (t, J =7.4 Hz, 2H), 1.82 (sextet, J=7.4 Hz, 2H), 1.05 (t, J=7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.4, 143.9, 142.6, 131.6 (×2), 129.7, 128.8 (×2), 127.1, 125.2, 121.2, 118.7, 112.3, 98.5, 82.02, 40.0, 18.8, 13.7; MS (+CI) m/z $309 (M + H^+, 100)$. Anal. Calcd for $C_{18}H_{16}N_2O_3$: C, 70.12; H, 5.23; N, 9.09. Found: C, 70.34; H, 5.63; N, 8.87%.

4.4.3. *N*-[(4'-Nitro-2'-(pentyn-1"-yl))phenyl]butyramide (4b). Prepared from the triflate 3a and 1-pentyne in 95% yield. Compound 4b. A white crystalline solid; mp 87-88 °C (CH₂Cl₂-hexane); $R_f = 0.59$ (25% EtOAc in hexane); IR (film) 3335, 2965, 2228, 1678, 1503, 1345 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 8.59 \text{ (d}, J = 9.2 \text{ Hz}, 1\text{H}), 8.23-8.15 \text{ (br}$ s, 1H), 8.19 (d, J=2.6 Hz, 1H), 8.09 (dd, J=9.2, 2.6 Hz, 1H), 2.51 (t, J=7.0 Hz, 2H), 2.42 (t, J=7.4 Hz, 2H), 1.81-1.65 (m, 4H), 1.08 (t, J=7.4 Hz, 3H), 1.02 (t, J=7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.4, 144.0, 142.3, 126.9, 124.4, 118.2, 112.9, 100.3, 74.2, 39.9, 21.9, 21.4, 18.7, 13.6, 13.5; MS (+CI) m/z 275 (M+H⁺, 100). Anal. Calcd for C₁₅H₁₈N₂O₃: C, 65.68; H, 6.61; N, 10.21. Found: C, 66.03; H, 6.73; N, 10.05%.

4.4.4. *N*-[(5'-Nitro-2'-(pentyn-1"-yl))phenyl]butyramide (5b). Prepared from the triflate 3b and 1-pentyne in 90% yield. Compound 5b. A white crystalline solid; mp 91-92 °C (CH₂Cl₂-hexane); $R_f = 0.38$ (25% EtOAc in hexane); IR (film) 3290, 2962, 2223, 1668, 1529, 1344 cm⁻¹; ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 9.31 \text{ (d, } J=2.2 \text{ Hz}, 1\text{H}), 8.04 \text{ (br s,}$ 1H), 7.85 (dd, J = 8.5, 2.2 Hz, 1H), 7.47 (d, J = 8.5 Hz, 1H), 2.55 (t, J=7.0 Hz, 2H), 2.42 (t, J=7.4 Hz, 2H), 1.82–1.67 (m, 4H), 1.11 (t, J=7.4 Hz, 3H), 1.04 (t, J=7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.2, 147.4, 139.5, 131.9, 118.5, 117.8, 113.9, 102.9, 75.0, 39.8, 21.9, 21.6, 18.8, 13.7, 13.6; MS (+CI) m/z 275 (M+H⁺, 100). Anal. Calcd for C₁₅H₁₈N₂O₃: C, 65.68; H, 6.61; N, 10.21. Found: C, 65.48; H, 6.79; N, 9.86%.

4.4.5. *N*-[(6'-Nitro-2'-phenylethynyl)phenyl]butyramide (**6a**). Prepared from the triflate **3c** and 1-phenylacetylene in 90% yield. *Compound* **6a**. A white crystalline solid; mp 167–168 °C (CH₂Cl₂-hexane); $R_{\rm f}$ =0.31 (25% EtOAc in hexane); IR (film) 3275, 2923, 2215, 1668, 1509 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.13 (br s, 1H), 7.89 (dd, *J*=8.3, 1.4 Hz, 1H), 7.76 (dd, *J*=8.3, 1.4 Hz, 1H), 7.53–7.48 (m, 2H), 7.42–7.35 (m, 3H), 7.29 (t, *J*=8.1 Hz, 1H), 2.44 (t, *J*= 7.4 Hz, 2H), 1.78 (sextet, *J*=7.4 Hz, 2H), 1.01 (t, *J*= 7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 170.9, 144.3, 136.7, 131.6 (×2), 131.4, 129.3, 128.6 (×2), 125.3, 124.9, 121.8, 120.9, 97.2, 83.7, 38.9, 18.8, 13.7; MS (+CI) *m/z* 309 (M+H⁺, 100). Anal. Calcd for C₁₈H₁₆N₂O₃: C, 70.12; H, 5.23; N, 9.09. Found: C, 70.31; H, 5.91; N, 8.82%.

4.4.6. *N*-[(6'-Nitro-2'-(pentyn-1"-yl))phenyl]butyramide (6b). Prepared from the triflate 3c and 1-pentyne in 91% yield. *Compound* 6b. A white crystalline solid; mp 144– 145 °C (CH₂Cl₂–hexane); R_f =0.36 (25% EtOAc in hexane); IR (film) 3272, 2961, 2225, 1670, 1511, 1536 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.95 (br s, 1H), 7.79 (dd, *J*= 8.0, 1.4 Hz, 1H), 7.60 (dd, *J*=8.0, 1.4 Hz, 1H), 7.20 (t, *J*= 8.0 Hz, 1H), 2.44 (t, *J*=7.0 Hz, 2H), 2.39 (t, *J*=7.4 Hz, 2H), 1.83–1.56 (m, 4H), 1.05 (t, *J*=7.3 Hz, 3H), 1.01 (t, *J*= 7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 170.8, 144.3, 136.4, 131.0, 124.9, 124.2, 121.0, 99.2, 75.3, 38.7, 21.9, 21.4, 18.7, 13.6, 13.5; MS (+CI) *m*/*z* 275 (M+H⁺, 100). Anal. Calcd for C₁₅H₁₈N₂O₃: C, 65.68; H, 6.61; N, 10.21. Found: C, 64.19; H, 6.05; N, 10.50%.

4.5. Representative procedure for the synthesis of trifluoroacetanilides 10a–c

4.5.1. *N*-(2'-Hydroxy-5'-nitrophenyl)trifluoroacetamide (10b). To a solution of 2-amino-4-nitrophenol 1b (3.124 g, 20 mmol) in dry THF (83 mL) and pyridine (2.4 mL, 29.95 mmol) cooled in an ice-water bath was added dropwise a solution of trifluoroacetic anhydride (3.1 mL, 22 mmol) in dry THF (7 mL). The resultant mixture was stirred for 16 h at room temperature under a nitrogen atmosphere. The reaction was quenched by adding water (10 mL) and brine (10 mL). The organic layer was separated and the aqueous layer was extracted with EtOAc $(20 \times 2 \text{ mL})$. The combined organic layer was washed with 5% HCl (15 mL), saturated aqueous NaHCO₃ (20 mL), and brine (20 mL), dried over anhydrous Na₂SO₄, and evaporated under reduced pressure. The crude product was purified by flash column chromatography (silica gel, 50% EtOAc in hexane) to give 10b (5.000 g, 99%) as a yellow crystalline solid; mp 188–189 °C (EtOAc–hexane); $R_f = 0.30$ (50%) EtOAc in hexane); IR (KBr): 3386, 3188 (br), 1696 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 11.20–10.30 (br s, 1H), 9.90–9.55 (br s, 1H), 9.02 (d, J = 2.7 Hz, 1H), 8.22 (dd, J =9.1, 2.8 Hz, 1H), 7.34 (d, J=9.0 Hz, 1H); ¹³C NMR (75 MHz, acetone-d₆) δ 153.7, 146.6, 140.0, 123.4, 122.4, 118.0, 115.4 (q, J_{C-F} =287.1 Hz), 114.9; MS (+CI) m/z 251 $(M+H^+,100)$. Anal. Calcd for $C_8H_5F_3N_2O_4$: C, 38.41; H, 2.01; N, 11.20. Found: C, 38.32; H, 1.88; N, 11.00%.

4.5.2. *N*-(2'-Hydroxy-4'-nitrophenyl)trifluoroacetamide (10a). Prepared in 99% yield from 2-amino-5-nitrophenol 1a after reaction at room temperature for 4 h. *Compound* 10a. As a yellow crystalline solid; mp 164–165 °C (EtOAc–

hexane); $R_{\rm f}$ =0.32 (25% EtOAc in hexane); IR (KBr) 3374, 3222 (br), 1697 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 10.60–9.35 (br s, 2H), 8.40 (d, J=8.7 Hz, 1H), 8.02–7.90 (m, 2H); ¹³C NMR (75 MHz, acetone- d_6) δ 154.7 (q, $J_{\rm C-F}$ =45.3 Hz), 147.7, 145.1, 129.7, 121.5, 115.6 (q, $J_{\rm C-F}$ =271.7 Hz), 115.2, 109.7; MS (+CI) m/z 251 (M+H⁺, 100). Anal. Calcd for C₈H₅F₃N₂O₄: C, 38.41; H, 2.01; N, 11.20. Found: C, 38.15; H, 1.94; N, 11.39%.

4.5.3. *N*-(2'-Hydroxy-6'-nitrophenyl)trifluoroacetamide (10c). Prepared in 86% yield from 2-amino-3-nitrophenol 1c after reaction at room temperature for 9 h. *Compound* 10c. As a yellow crystalline solid; mp 136–137 °C (EtOAc– hexane); $R_{\rm f}$ =0.41 (50% EtOAc in hexane); IR (KBr) 3310 (br), 1729 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 11.04 (br s, 1H), 8.05–7.75 (br s, 1H), 7.85 (dd, *J*=7.8, 2.1 Hz, 1H), 7.47–7.38 (m, 2H); ¹³C NMR (CDCl₃) δ 153.4 (q, *J*_{C-F}= 43.5 Hz), 146.4, 146.0, 128.5, 120.7, 115.3, 115.5 (q, *J*_{C-F}= 286.8 Hz), 115.0; MS (+CI) *m*/*z* 251 (M+H⁺, 100). Anal. Calcd for C₈H₅F₃N₂O₄: C, 38.41; H, 2.01; N, 11.20. Found: C, 38.41; H, 1.85; N, 10.80%.

4.6. Representative procedure for the synthesis of nitroaryl triflates 11a-c

4.6.1. *N*-[5'-Nitro-2'-(((trifluoromethane)sulfonyl)oxy)phenyl]trifluoroacetamide (11b). To a suspension of NaH (0.54 g, 13.5 mmol) in dry THF (80 mL) cooled in an ice-water bath was added dropwise a solution of **10b** (2.25 g, 9.0 mmol) and PhNTf₂ (4.82 g, 13.5 mmol) in dry THF (20 mL). The resultant mixture was stirred for 19 h at room temperature under a nitrogen atmosphere. The reaction was quenched by water (25 mL) and brine (25 mL). The organic layer was washed with saturated aqueous NaHCO₃ (45 mL \times 5) and brine (35 mL), dried over anhydrous Na₂SO₄, evaporated under reduced pressure. The crude product was purified by flash column chromatography (silica gel, 25% EtOAc in hexane) to give 11b (3.20 g, 93%) as a yellow crystalline solid; mp 88-89 °C (EtOAc-hexane); $R_{\rm f}$ =0.54 (25% EtOAc in hexane); IR (KBr) 3292 (br), 1717 cm^{-1} ; ¹H NMR (300 MHz, CDCl₃) δ 9.17 (d, J=2.7 Hz, 1H), 8.28 (br s, 1H), 8.24 (dd, J=9.3, 3.0 Hz, 1H), 7.62 (d, J=9.0 Hz, 1H); ¹³C NMR $(75 \text{ MHz}, \text{CDCl}_3) \delta 155.9 (q, J_{C-F} = 39.4 \text{ Hz}), 148.0, 143.8,$ 129.4, 123.7, 123.2, 120.1, 119.2 (q, $J_{C-F}=320.9$ Hz), 115.8 (q, J_{C-F} =288.4 Hz); MS (+CI) *m*/*z* 383 (M+H⁺, 100). Anal. Calcd for C₉H₄F₆N₂O₆S: C, 28.28; H, 1.05; N, 7.33. Found: C, 28.39; H, 0.89; N, 6.65%.

4.6.2. *N*-[4'-Nitro-2'-(((trifluoromethane)sulfonyl)oxy)phenyl]trifluoroacetamide (11a). Prepared from 10a in 84% yield after reaction at room temperature for 4 h. *Compound* 11a. As a yellow crystalline solid; mp 66–67 °C (EtOAc–hexane); R_f =0.38 (25% EtOAc in hexane); IR (KBr) 3284, 1725 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 8.62 (d, *J*=9.3 Hz, 1H), 8.46–8.32 (br s, 1H), 8.38 (dd, *J*= 9.0, 2.7 Hz, 1H), 8.31 (d, *J*=2.4 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 155.8 (q, *J*_{C-F}=45.1 Hz), 145.4, 138.3, 134.1, 125.4, 123.5, 119.1 (q, *J*=320.2 Hz), 118.8, 117.2 (q, *J*=288.1 Hz); MS (+CI) *m*/*z* 383 (M+H⁺, 100). Anal. Calcd for C₉H₄F₆N₂O₆S: C, 28.28; H, 1.05; N, 7.33. Found: C, 28.53; H, 0.98; N, 6.86%. **4.6.3.** *N*-[6'-Nitro-2'-(((trifluoromethane)sulfonyl)oxy)phenyl]trifluoroacetamide (11c). Prepared from 10c in 88% yield after reaction at room temperature for 24 h. *Compound* **11c**. As a yellow crystalline solid; mp 108.5– 109.5 °C; R_f =0.37 (25% EtOAc in hexane); IR (KBr) 3255, 1732 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.10 (br s, 1H), 8.22 (dd, *J*=8.2, 1.3 Hz, 1H), 7.75 (dd, *J*=8.4, 1.5 Hz, 1H), 7.66 (t, *J*=8.4 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 156.1 (q, J_{C-F} =43.2 Hz), 145.6, 145.3, 129.8, 129.1, 126.1, 123.6, 119.1 (q, J_{C-F} =320.6 Hz), 115.9 (q, J_{C-F} = 288.2 Hz); MS (+CI) *m*/*z* 383 (M+H⁺, 100). Anal. Calcd for C₉H₄F₆N₂O₆S: C, 28.28; H, 1.05; N, 7.33. Found: C, 28.34; H, 0.96; N, 7.25%.

4.7. Representative procedure for the *t*-BuOK-promoted heteroannualtion toward nitroindoles

4.7.1. 6-Nitro-2-phenylindole (8a). A mixture of 5a (60.0 mg, 0.19 mmol), t-BuOK (25.0 mg, 0.22 mmol) in dry NMP (2.0 mL) was heated at 60-70 °C for 7 h under a nitrogen atmosphere. After cooling to room temperature, water (2 mL) and EtOAc (50 mL) were added to the reaction mixture, respectively. The separated aqueous layer was extracted with EtOAc (20×3 mL). The combined organic layer was washed with brine, dried over anhydrous MgSO₄, concentrated under reduced pressure. The residue was purified by flash column chromatography (silica gel, 30% EtOAc in hexane) to give 8a (37.0 mg, 84%) as a yellow crystalline solid; mp 211–212 °C (CH₂Cl₂–hexane); $R_{\rm f} = 0.61$ (33% EtOAc in hexane); IR (KBr) 3322, 2923, 1298 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 11.61 (br s, 1H), 8.51 (d, J = 2.0 Hz, 1H), 8.12–8.06 (m, 3H), 7.88 (d, J=8.8 Hz, 1H), 7.70–7.63 (m, 2H), 7.61–7.54 (m, 1H), 7.25 (dd, J=2.0, 0.8 Hz, 1H); ¹³C NMR (75 MHz, acetone- d_6) δ 144.0, 142.5, 135.5, 133.9, 131.0, 128.9 (×2), 128.6, 125.5 (×2), 119.9, 114.7, 107.5, 99.7; MS (+CI) *m*/*z* 239 (M+ H^+ , 100). Anal. Calcd for $C_{14}H_{10}N_2O_2$: C, 70.58; H, 4.23; N, 11.76. Found: C, 70.40; H, 4.11; N, 11.64%.

Other results are listed in Table 2.

4.8. Representative procedure for the Pd-catalyzed onepot cross-coupling and heteroannulation toward nitroindoles

4.8.1. 6-Nitro-2-propylindole (8b). A mixture of 11b (115.0 mg, 0.3 mmol), n-Bu₄NI (169.6 mg, 0.45 mmol), Pd(PPh₃)₄ (11.1 mg, 0.03 mmol), CuI (17.14 mg, 0.09 mmol), and 1-pentyne (40.8 mg, 60 µL, 0.6 mmol) in degassed DMF (5 mL) containing Et₃N (1 mL) was heated at ca. 80 °C for 21 h. After cooling to room temperature, the reaction mixture was diluted with EtOAc (20 mL) and washed with saturated aqueous NH₄Cl (10 mL) and brine (15 mL). The organic layer was dried over anhydrous MgSO₄, and evaporated under reduced pressure. The crude product was purified by flash column chromatography (silica gel, MeOH–CH₂Cl₂–hexane = 1:40:30) to give **8b** (51.7 mg, 84%, entry 7, Table 3) as a yellow crystalline solid;³⁰ mp 94–95 °C (CH₂Cl₂–hexane); $R_{\rm f}$ =0.54 (33%) EtOAc in hexane); IR (film) 3367, 2962, 1302 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 10.98 (br s, 1H), 8.42 (d, J= 2.0 Hz, 1H), 8.04 (dd, J=8.8, 2.2 Hz, 1H), 7.73 (d, J=8.8 Hz, 1H), 6.55 (d, J=0.5 Hz, 1H), 2.98 (t, J=7.5 Hz,

2H), 1.94 (sextet, J=7.5 Hz, 2H), 1.13 (t, J=7.5 Hz, 3H); ¹³C NMR (75 MHz, acetone- d_6) δ 141.5, 119.3, 118.6, 114.8, 114.0, 107.3, 106.7, 99.8, 29.9, 21.7, 12.8; MS (+CI) m/z 205 (M+H⁺, 100). Anal. Calcd for C₁₁H₁₂N₂O₂: C, 64.69; H, 5.92; N, 13.72. Found: C, 64.74; H, 6.08; N, 14.13%.

Other results are summarized in Table 3.

4.8.2. 5-Nitro-2-phenylindole (7a). A yellow crystalline solid; mp 190–191 °C (CH₂Cl₂–hexane); $R_{\rm f}$ =0.37 (25% EtOAc in hexane); IR (film) 3344, 1329 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 11.52 (br s, 1H), 8.69 (d, J= 2.2 Hz, 1H), 8.26 (dd, J=8.9, 2.2 Hz, 1H), 8.05–8.00 (m, 2H), 7.76–7.60 (m, 3H), 7.56–7.50 (m, 1H), 7.28 (s, 1H); ¹³C NMR (75 MHz, acetone- d_6) δ 141.3, 139.8, 130.9, 128.6 (×2), 128.2, 128.1, 128.0, 125.0 (×2), 116.6, 116.5, 110.8, 100.4; MS (+CI) m/z 239 (M+H⁺, 100). Anal. Calcd for C₁₄H₁₀N₂O₂: C, 70.58; H, 4.23; N, 11.76. Found: C, 70.32; H, 4.11; N, 11.73%.

4.8.3. 5-Nitro-2-propylindole (7b). A yellow crystalline solid; mp 125–126 °C (CH₂Cl₂–hexane); $R_{\rm f}$ =0.34 (50% EtOAc in hexane); IR (KBr) 3325, 1314 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 10.86 (br s, 1H), 8.57 (d, J= 2.1 Hz, 1H), 8.11 (dd, J=9.0, 2.1 Hz, 1H), 7.58 (d, J= 9.0 Hz, 1H), 6.56 (d, J=0.7 Hz, 1H), 2.92 (t, J=7.4 Hz, 2H), 1.90 (sextet, J=7.5 Hz, 2H), 1.12 (t, J=7.5 Hz, 3H); ¹³C NMR (75 MHz, acetone- d_6) δ 143.6, 140.6, 138.9, 127.6, 115.4, 115.2, 109.8, 100.4, 28.1, 21.4, 12.5; MS (+ CI) m/z 205 (M+H⁺, 100). Anal. Calcd for C₁₁H₁₂N₂O₂: C, 64.69; H, 5.92; N, 13.72. Found: C, 64.74; H, 5.85; N, 13.46%.

4.8.4. 5-Nitro-2-(2'-hydroxyethyl)indole (7c). A crystalline solid; mp 114–115 °C (EtOAc–hexane); R_f =0.29 (67% EtOAc in hexane); IR (KBr) 3469, 3185 (br), 1512, 1471, 1337 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 10.85 (br s, 1H), 8.60 (d, J=2.2 Hz, 1H), 8.11 (dd, J=9.0, 2.3 Hz, 1H), 7.75 (d, J=8.9 Hz, 1H), 6.67 (d, J=0.9 Hz, 1H), 4.22 (t, J=5.3 Hz, 1H), 4.07 (q, J=6.3 Hz, 2H), 3.18 (t, J=6.2 Hz, 2H); ¹³C NMR (75 MHz, acetone- d_6) δ 142.0, 141.1, 139.3, 127.9, 115.8, 115.6, 110.5, 101.5, 60.8, 31.4; MS (+CI) *m/z* 207 (M+H⁺, 100). Anal. Calcd for C₁₀H₁₀N₂O₃: C, 58.25; H, 4.89; N, 13.59. Found: C, 58.25; H, 5.00; N, 13.75%.

4.8.5. 5-Nitro-2-(3'-cyanopropyl)indole (7d). A crystalline solid; mp 125.5–126 °C (EtOAc–hexane); $R_{\rm f}$ =0.41 (50% EtOAc in hexane); IR (KBr) 3321, 2250, 1471, 1334 cm⁻¹; ¹H MNR (300 MHz, acetone- d_6): δ 10.9 (br s, 1H), 8.59 (d, J=2.1 Hz, 1H), 8.11 (dd, J=8.7, 2.4 Hz, 1H), 8.60 (d, J= 8.7 Hz, 1H), 6.70 (s, 1H), 3.16 (t, J=7.5 Hz, 2H), 2.72 (t, J=7.2 Hz, 2H), 2.33–2.22 (m, 2H); ¹³C NMR (75 MHz, acetone- d_6) δ 142.0, 141.5, 138.5, 128.1, 119.2, 116.1, 116.0, 110.6, 101.5, 26.7, 24.7, 15.8; MS (+CI) *m/z* 230 (M+H⁺, 100). Anal. Calcd for C₁₂H₁₁N₃O₂: C, 62.87; H, 4.84; N, 18.33. Found: C, 62.75; H, 4.89; N, 18.37%.

4.8.6. 5-Nitro-2-(3'-chloropropyl)indole (7e). A crystalline solid; mp 107–107.5 °C (EtOAc–hexane); $R_{\rm f}$ =0.34 (25% EtOAc in hexane); IR (KBr): 3336, 1323 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 10.86 (br s, 1H), 8.59 (d, J= 1.8 Hz, 1H), 8.11 (dd, J=8.7, 2.1 Hz, 1H), 7.61 (d, J= 8.7 Hz, 1H), 6.68 (s, 1H), 3.84 (t, J=6.3 Hz, 2H), 3.17 (t, J=7.8 Hz, 2H), 2.44–2.35 (m, 2H); ¹³C NMR (75 MHz, acetone- d_6) δ 142.2, 140.8, 139.2, 127.7, 115.7, 115.6, 110.2, 101.0, 43.2, 30.8, 24.2; MS (+CI) m/z 239 (M+H⁺, 100). Anal. Calcd for C₁₁H₁₁ClN₂O₂: C, 55.36; H, 4.65; N, 11.74. Found: C, 55.28; H, 4.71; N, 11.76%.

4.8.7. 6-Nitro-2-(2'-hydroxyethyl)indole (8c). A crystalline solid; mp 141–142 °C (EtOAc–hexane); R_f =0.28 (50% EtOAc in hexane); IR (KBr) 3490, 3275 (br), 1501, 1316, 1051 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 10.90 (br s, 1H), 8.47 (d, J=2.1 Hz, 1H), 8.03 (dd, J=8.4, 2.1 Hz, 1H), 7.73 (d, J=9.0 Hz, 1H), 6.60 (d, J=2.4 Hz, 1H), 4.25 (br s, 1H), 4.08 (br t, J=6.0 Hz, 2H), 3.21 (t, J=6.3 Hz, 2H); ¹³C NMR (75 MHz, acetone- d_6) δ 145.3, 142.1, 134.2, 133.7, 118.8, 114.1, 107.2, 100.7, 60.7, 31.6; MS (+CI) *m/z* 207 (M+H⁺, 100). Anal. Calcd for C₁₀H₁₀N₂O₃: C, 58.25; H, 4.89; N, 13.59. Found: C, 58.29; H, 4.77; N, 13.37%.

4.8.8. 6-Nitro-2-(3'-cyanopropyl)indole (8d). A crystalline solid; mp 117–118 °C (EtOAc–hexane); $R_{\rm f}$ =0.47 (50% EtOAc in hexane); IR (KBr) 3326, 2254, 1499, 1308 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 10.99 (br s, 1H), 8.41 (d, J=1.8 Hz, 1H), 8.18 (dd, J=8.7, 2.4 Hz, 1H), 7.75 (d, J= 8.7 Hz, 1H), 6.63 (d, J=3.0 Hz, 1H), 3.19 (t, J=7.8 Hz, 2H), 2.72 (t, J=8.4 Hz, 2H), 2.35–2.27 (m, 2H); ¹³C NMR (75 MHz, acetone- d_6) δ 145.9, 142.8, 134.6, 133.6, 131.6, 119.1, 114.3, 107.1, 100.6, 26.9, 24.7, 15.9; MS (+CI) *m/z* 230 (M+H⁺, 100). Anal. Calcd for C₁₂H₁₁N₃O₂: C, 62.87; H, 4.84; N, 18.33. Found: C, 62.72; H, 4.85; N, 18.46%.

4.8.9. 6-Nitro-2-(3'-chloropropyl)indole (8e). A white crystalline solid; mp 98–99 °C (EtOAc–hexane); R_f =0.49 (25% EtOAc in hexane); IR (KBr) 3329, 1505, 1337, 1298 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 10.95 (br s, 1H), 8.42 (d, *J*=1.2 Hz, 1H), 8.04 (dd, *J*=8.7, 2.1 Hz, 1H), 7.74 (d, *J*=8.7 Hz, 1H), 6.60 (d, *J*=3.0 Hz, 1H), 3.83 (t, *J*=6.6 Hz, 2H), 3.19 (t, *J*=7.8 Hz, 2H), 2.45–2.35 (m, 2H); ¹³C NMR (75 MHz, acetone- d_6) δ 145.7, 142.0, 134.6, 133.7, 118.1, 114.3, 107.1, 100.4, 44.0, 31.6, 25.2; MS (+ CI) *m/z* 239 (M+H⁺, 100). Anal. Calcd for C₁₁H₁₁ClN₂O₂: C, 55.36; H, 4.65; N, 11.74. Found: C, 55.28; H, 4.51; N, 11.72%.

4.8.10. 7-Nitro-2-phenylindole (9a). A white crystalline solid; mp 142–143 °C (CH₂Cl₂–hexane); $R_{\rm f}$ =0.74 (25% EtOAc in hexane); IR (film) 3149, 2921, 1338, 1292 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 11.09 (br s, 1H), 8.25 (dd, J=8.1, 0.9 Hz, 1H), 8.18 (dd, J=8.1, 0.9 Hz, 1H), 8.18 (dd, J=8.1, 0.9 Hz, 1H), 8.13–8.09 (m, 2H), 7.67–7.55 (m, 3H), 7.40 (t, J=7.9 Hz, 1H), 7.25 (d, J=1.6 Hz, 1H); ¹³C NMR (75 MHz, acetone- d_6) δ 140.8, 132.9, 130.8, 129.8, 128.9, 128.6 (×2), 128.2, 128.0, 125.8 (×2), 119.1, 118.3, 100.4; MS (+CI) *m*/z 239 (M+H⁺, 44), 153 (100). Anal. Calcd for C₁₄H₁₀N₂O₂: C, 70.58; H, 4.23; N, 11.76. Found: C, 70.36; H, 4.25; N, 12.00%.

4.8.11. 7-Nitro-2-propylindole (9b). A yellow crystalline solid; mp 79–80 °C (CH₂Cl₂–hexane); $R_{\rm f}$ =0.71 (25% EtOAc in hexane); IR (KBr) 3409, 2956, 1512, 1339 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 9.69 (br s, 1H), 8.05 (d, *J*= 8.1 Hz, 1H), 7.80 (d, *J*=7.6 Hz, 1H), 7.13 (t, *J*=7.9 Hz, 1H), 6.38 (s, 1H), 2.81 (t, *J*=7.4 Hz, 2H), 1.80 (sextet, *J*=

7.4 Hz, 2H), 1.04 (t, J=7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 142.7, 132.6, 129.4, 128.4, 127.6, 118.9, 117.9, 100.7, 30.0, 22.2, 13.8; MS (+CI) *m*/*z* 205 (M+H⁺, 100). Anal. Calcd for C₁₁H₁₂N₂O₂: C, 64.69; H, 5.92; N, 13.72. Found: C, 64.90; H, 6.11; N, 13.75%.

4.8.12. 7-Nitro-2-(2'-hydroxyethyl)indole (9c). A yellow crystalline solid; mp 117–118 °C (EtOAc–hexane); R_f = 0.47 (50% EtOAc in hexane); IR (KBr): 3394, 3291 (br), 2923, 1514, 1338 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 11.11 (br s, 1H), 8.19 (d, J=7.8 Hz, 1H), 8.09 (d, J= 7.8 Hz, 1H), 7.34 (t, J=7.8 Hz, 1H), 6.67 (d, J=0.9 Hz, 1H), 4.29 (br s, 1H), 4.11 (t, J=6.2 Hz, 2H), 3.26 (t, J= 6.4 Hz, 2H); ¹³C NMR (75 MHz, acetone- d_6) δ 141.7, 132.8, 132.2, 129.1, 127.4, 118.5, 117.2, 101.0, 61.0, 31.0; MS (+CI) m/z 207 (M+H⁺, 100). Anal. Calcd for C₁₀H₁₀N₂O₃: C, 58.25; H, 4.89; N, 13.59. Found: C, 58.39; H, 4.89; N, 13.70%.

Acknowledgements

This work is supported in part by the Innovation and Technology Fund (ITS/119/00) from the Innovation and Technology Commission of the Hong Kong Special Administration Region, China, the Department of Chemistry, HKUST, and a research grant provided by Zhejiang University. Elemental analyses performed by Zhejiang University are acknowledged. W. -M. Dai is the recipient of Cheung Kong Scholars Award of The Ministry of Education of China.

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Tetrahedron

Tetrahedron 60 (2004) 10993-10998

Asymmetric epoxidation of *cis*-1-propenylphosphonic acid (CPPA) catalyzed by chiral tungsten(VI) and molybdenum(VI) complexes

Xin-Yan Wang, Hong-Chang Shi,* Chuan Sun and Zhi-Guo Zhang

Department of Chemistry, Tsinghua University, Beijing 100084, China

Received 19 January 2004; revised 25 August 2004; accepted 30 August 2004

Available online 5 October 2004

Abstract—In the presence of 5.0 mol% chiral tungsten(VI) and molybdenum(VI) complexes, the catalytic asymmetric epoxidation of *cis*-1-propenylphosphonic acid (CPPA) with 30% aqueous H_2O_2 affording (1*R*,2*S*)-(-)-(1, 2)-epoxypropyl phosphonic acid (fosfomycin) was first described. The enantioselectivities of the tungsten and molybdenum catalysts depend strongly on the ligands, reaction temperature and solvent. In CH₂Cl₂ at 0 °C for 72 h, complex MoO₂[(+)-campy]₂ catalyzed the asymmetric epoxidation in a 100% conversion of CPPA with the highest 80% ee. The mechanism of the present epoxidation could be described as direct oxygen transfer occurred on the interface of the biphasic H₂O-nonprotic system.

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1. Introduction

(1R,2S)-(-)-(1,2)-Epoxypropyl phosphonic acid (fosfomycin) is a clinically important drug with wide-spectrum antibiotic activity. It was isolated originally from fermentation broth of *Streptomyces fradiae*¹ and prepared mainly by epoxidation of *cis*-1-propenylphosphonic acid (CPPA) (1)^{1,2} followed by an optical resolution of the racemic epoxide with chiral amines.^{1,3} Although a few of asymmetric synthetic methods were reported for the preparation of fosfomycin,⁴ no catalytic asymmetric epoxidation of CPPA has been reported. It is well known that the tungsten and molybdenum derivatives efficiently catalyzed the epoxidation of electron-deficient olefin.⁵ Therefore, we were encouraged to explore the asymmetric epoxidation of CPPA catalyzed by chiral salen tungsten(VI) and salen



Scheme 1. Synthesis of chiral salen tungsten(VI) and molybdenum(VI) complexes.

Keywords: cis-1-Propenylphosphonic acid; Chiral tungsten and molybdenum complexes; Catalytic asymmetric epoxidation; Enantioselectivity. * Corresponding author. Tel.: +86 10 627 83910; fax: +86 10 627 85569; e-mail: shihc@mail.tsinghua.edu.cn



Scheme 2. Synthesis of chiral tungsten(VI) and molybdenum(VI) 2'-pyridinyl alcoholate complexes.

molybdenum(VI) complexes. In this paper, we have reported the catalytic activities and selectivity of several chiral tungsten(VI) and molybdenum(VI) complexes in the asymmetric epoxidation of CPPA.

2. Results and discussion

2.1. Preparation of chiral tungsten(VI) and molybdenum(VI) complexes

Although several salen tungsten(VI) complexes have been prepared,⁶ none of those is chiral complex. We first synthesized chiral salen tungsten(VI) and molybdenum(VI) complexes by the reactions of WCl₂O₂, Mo(acac)₂ with the ligand shown in Scheme 1.⁷

Chiral tungsten and molybdenum 2'-pyridinyl alcoholate complexes have been also used to catalyze the epoxidation



Scheme 3. Catalytic asymmetric epoxidation of CPPA.

Table 1. Asymmetric epoxidation of CPPA at 25 °C^a

of olefins.⁸ We prepared $WO_2[(+)$ -campy]₂, $MoO_2[(+)$ -campy]₂, $WO_2[(-)$ -fenpy]₂ and $MoO_2[(-)$ -fenpy]₂ complexes according to the reported methods^{8e,f} as shown in Scheme 2.

The ligand (+)-campy stands for (1R,2R,4R)-1,7,7-trimethyl-2-(2'-pyridinyl)bicyclo-[2.2.1] heptan-2-ol; the (-)-fenpy stands for (1R,2R,4S)-1,3,3-trimethyl-2-(2'-pyridinyl)bicyclo-[2.2.1] heptan-2-ol.

2.2. Asymmetric epoxidation of *cis*-1-propenyl-phosphonic acid

We studied the asymmetric epoxidation of CPPA with 30% aqueous H_2O_2 as oxidant in the presence of catalytic amount of chiral tungsten(VI) and molybdenum(VI) complexes, as shown in Scheme 3.

To a solution of CPPA (0.2 mmol) in ethanol (10 mL) was added racemic α -phenylethylamine (0.2 mmol), then catalyst (0.01 mmol) and H₂O₂ (1.0 mmol, 105 μ L of 30% aqueous). The resulting mixture was stirred at 25 °C and the reaction was monitored by ¹H NMR. After 24 h, the conversion of CPPA reached 100%, with the ee value of 45%.¹⁰ (Table 1, entry 1). The same reaction in CH₂Cl₂ gave a higher ee value of 69% (Table 1, entry 2). Although salen molybdenum catalyst showed low catalytic activity, it exhibited a similar enantioselectivity as salen tungsten did (Table 1, entries 3 and 4).

Entry	Catalyst	Solvent	Conv. of CPPA (%) ^b	ee (%) of $(1R, 2S)$ -epoxide
1	Salen W	C ₂ H ₅ OH	100	45
2		CH_2Cl_2	100	69
3	Salen Mo	C ₂ H ₅ OH	20	46
4		CH_2Cl_2	30	69
5	$WO_2[(+)-campy]$	C ₂ H ₅ OH	100	62
6		CH ₂ Cl ₂	100	74
7	$MoO_2[(+)-campy]_2$	C ₂ H ₅ OH	100	63
8		CH ₂ Cl ₂	100	74
9	$WO_2[(-)-fenpy]_2$	C ₂ H ₅ OH	100	52
10		CH ₂ Cl ₂	100	71
11	$MoO_2[(-)-fenpy]_2$	C ₂ H ₅ OH	100	59
12		CH ₂ Cl ₂	100	72

^a Reactions were carried out at 25 °C for 24 h using 0.2 mmol of CPPA, 0.2 mmol of α-phenylethylamine, 1.0 mmol H₂O₂ (30% aqueous) and 0.01 mmol of catalyst in 10 mL of solvent.

^b Determined by ¹H NMR.

^c Determined by ¹H NMR with Eu(tfc)₃.



Scheme 4. Preparation of the dimethyl ester of the epoxide.

The optical purity of 2 was determined from the corresponding dimethyl ester by ¹H NMR analysis with the optically active shift reagent Eu(tfc)₃. Its chemical name is tri[3-(trifluoromethylhydroxymethylene)-(+)-camphorato]europium(III). The preparation of the dimethyl ester is shown in Scheme 4 and in Section 4.

As shown in Table 1, the enantioselectivities of tungsten and molybdenum catalysts were more dependent on the ligands than on the metals. Tungsten and molybdenum catalysts bearing the same ligand show similar enantioselectivities in either C₂H₅OH or CH₂Cl₂ solvents. For instance, salen tungsten and salen molybdenum (entries 1-4); WO₂[(+)campy]₂ and MoO₂[(+)-campy]₂ (entries 5–8); WO₂[(-)fenpy]₂ and $MoO_2[(-)-fenpy]_2$ (entries 9–12) catalyzed the epoxidation of CPPA resulting in almost same ee value. In addition, CH₂Cl₂ is a better solvent for the present catalytic epoxidation. When (+)-campy ligand was employed in CH₂Cl₂ at 25 °C, the highest ee value (74%) was achieved (entries 6 and 8).

Table 2 shows reaction temperature impact on the enantioselectivities in CH₂Cl₂. For all used catalysts, at low temperature (0 °C), catalysts showed the higher ee values than at 50 °C or at 25 °C (Table 1). In the case of $MoO_2[(+)-campy]_2$, the optical yield of the catalytic epoxidation was up to 80% ee.

At 0 °C, we have studied the catalytic asymmetric epoxida-

	Table	2.	Asv	ymmetric	epoxidation	of	CPPA	at 0	and 50	$^{\circ}C^{a}$
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tion in different solvent. It was found that CHCl₃ and toluene were also suitable solvents. In CHCl₃ or toluene, $WO_2[(+)-campy]_2$ catalyzed the reaction in a 100% conversion of CPPA with ee values of 78 or 74%, respectively. Similarly, under the same reaction conditions, $MoO_2[(-)-campy]_2$ gave the similar results [100% conversion, 79% ee in CHCl₃ and 76% ee in toluene].

2.3. Proposed mechanism: a direct oxygen transfer on biphase interface

Mimoun type diperoxo complexes, such as $MoO(O_2)_2$ - (OPR_3) , R = NMe₂ is often used to study on the mechanism of olefin epoxidation catalyzed by tungsten(VI) and molybdenum(VI) complexes. An important characteristic is that every molecule of the complexes possesses two metal-dioxygen rings. Mimoun-type diperoxocomplexes9 were obtained from the reaction of MoO₃ with H₂O₂ in the presence of ligand, such as HMPA. MoO₃ has three oxo oxygen atoms, among them two have been perhydrolyzed into the two metal-dioxygen ring in the peroxocomplex (Scheme 5). A great number of experiments indicate the last oxo oxygen atom cannot be converted by H₂O₂ into a metaldioxygen ring.

However, the molecules of above $WO_2[(-)-fenpy]_2$ and $MoO_2[(-)-fenpy]_2$ or salen tungsten(VI) and molybdenum(VI) complexes have only two oxo oxygen atoms (see Schemes 1 and 2) because an oxo oxygen atom has been

Entry	Catalyst	Temperature (°C)	Conv. of CPPA $(\%)^{b}$	ee (%) of (1 R ,2 S)-epoxide ^c
1	Salen W	0	100	74
2		50	100	59
3	Salen Mo	0	24	75
4		50	58	62
5	$WO_2[(+)-campy]$	0	100	78
6		50	100	67
7	$MoO_2[(+)-campy]_2$	0	100	80
8		50	100	68
9	$WO_2[(-)-fenpy]_2$	0	100	76
10		50	100	63
11	$MoO_2[(-)-fenpy]_2$	0	100	77
12	/ 100-	50	100	64

^a Reactions were carried out at 0 °C for 72 h, at 50 °C for 1 h by using 0.2 mmol of CPPA, 0.2 mmol of α-phenylethylamine, 1.0 mmol H₂O₂ (30% aqueous) and 0.01 mmol of catalyst in 10 mL CH₂Cl₂.

^b Determined by ¹H NMR. ^c Determined by ¹H NMR using Eu(tfu)₃.

$$O = M_0 = O + HMPA + 2H_2O_2 \longrightarrow O = M_0O - H_2O + HMPA + 2H_2O_2 \longrightarrow O = M_0O - H_2O + H_2O +$$

R= NMe₂



Scheme 6. Proposed mechanism for the epoxidation of CPPA in H₂O–nonprotic solvent.

replaced by two oxygen atoms in the ligands. Therefore, they can be only perhydrolyzed into a monoperoxocomplexes with one metal-dioxygen ring.

In recent years, quantum chemical study on epoxidation mechanism has made great progress.¹⁰ The results revealed that a nucleophilic attack of π -electron of the olefin toward the σ^* orbital of the peroxo bond maybe involved. This strongly supported the direct oxygen transfer mechanism suggested by Sharpless.¹¹ Therefore, we consider that the mechanism of the epoxidation of CPPA should be a direct oxygen transfer from the metal-dioxygen ring in the monoperoxocomplexes.

In the epoxidation, 30% aqueous H_2O_2 was used as oxidant. When CH₂Cl₂, CHCl₃ and C₆H₅CH₃ were used as solvents, the epoxidation was carried out in a biphasic system. H₂O₂ is a molecule with minor polarity and can get into nonprotic solvent such as CH₂Cl₂. Therefore, the chiral tungsten or molybdenum complexes in organic phase can be perhydrolyzed into a monoperoxocomplex (see TS in Scheme 6). On the other hand, both methyl group and double bond in CPPA are lipophilic groups, which gets into the organic phase readily. Meanwhile, -PO₃H⁻ group easily dissolve in H₂O because it is a very strong hydrophilic group. Therefore, the oxygen transfer should occur on the interface of H₂O–nonprotic solvent as shown in Scheme 6. Moreover, the epoxide produced by the epoxidation dissolves easily in water because the three-membered oxygen-containing ring has relatively strong hydrophilic property.

3. Conclusion

CPPA is an α , β -unsaturated acid and an electron-defect olefin. We have developed chiral tungsten(VI) and chiral molybdenum(VI) complexes to catalyze asymmetric epoxidation of CPPA. The ee values depend on the nature of ligands, reaction temperatures and solvents. At 0 °C in CH₂Cl₂, MoO₂[(+)-campy]₂ catalyzed the epoxidation reaction to give the product with the highest ee value of 80%.

4. Experimental

4.1. General

All melting points were determined on a Yanaco melting

point apparatus and are uncorrected. IR spectra were recorded on a Nicolet FT-IR 5DX spectrometer as KBr pellets. The NMR spectra were recorded on Bruker AC 300 MHz or AC 500 MHz spectrometer in CDCl₃ with TMS as the internal reference. The elemental analyses were performed on a Perkin–Elmer 240C instrument. Optical rotations were determined on a Perkin–Elmer 343 polarimeter.

4.1.1. Preparation of chiral salen tungsten complexes. Under argon atmosphere, salen ligand (50 mg, 0.1 mmol) was dissolved in toluene (5 mL), followed by the addition of WO_2Cl_2 (26 mg, 0.1 mmol) and pyridine (0.014 mL, 0.178 mmol). The resulting mixture was refluxed for 48 h and then cooled to room temperature and washed several times with water. The organic layer was separated and condensed to give a yellow oil. The crude product was purified by flash chromatography (eluent: CHCl₃/hexane 2:1) to give a yellow solid in 65% yield.

*WO*₂ (*salen*). Mp 161−162 °C. $[\alpha]_D^{25} - 32.7^\circ$ (*c* 0.2, CHCl₃). Anal. Calcd for C₃₈H₄₂N₂O₄W: C, 58.92; H, 5.46; N, 3.62. Found: C, 59.02; H, 5.40; N, 3.47%. IR (KBr, cm⁻¹): ν (W=O) 880, 928. ¹H NMR 7.93 (s, 1H), 7.84 (s, 1H), 7.39– 7.46 (m, 5H), 7.36 (s, 1H), 7.33 (s, 1H), 7.22–7.27 (m, 5H), 6.85 (s, 1H), 6.83 (s, 1H), 6.36 (d, *J*=4.1 Hz, 1H), 4.75 (d, *J*=5.4 Hz, 1H), 2.28 (s, 3H), 2.18 (s, 3H), 1.50 (s, 9H), 1.21 (s, 9H). ¹³C NMR 166.9, 165.7, 162.7, 157.9, 141.0, 140.1, 138.5, 133.7, 132.5, 129.3 (2C), 129.2 (2C), 128.3 (2C), 128.0 (2C), 122.4, 121.5, 118.1, 80.2, 79.3, 74.7, 35.3, 35.1, 34.7, 32.3 (2C), 29.7 (2C), 29.3 (2C), 20.6, 20.5. ESI-MS: *m/z* 775 (M⁺ + H).

4.1.2. Preparation of chiral salen molybdenum complexes. Under argon atmosphere, salen ligand (50 mg, 0.1 mmol) was dissolved in anhydrous alcohol (5 mL), followed by adding $MoO_2(acac)_2$ (26 mg, 0.1 mmol). The resulting mixture was refluxed for 0.5 h and then was cooled to room temperature. Precipitate was observed when water was added to the reaction mixture. After filtration, the solid was purified by flash chromatography (eluent: CHCl₃/ hexane 1:1) to give red solid in 86% yield.

*MoO*₂ (*salen*). Mp 193–195 °C. $[\alpha]_D^{25}$ –40.6° (*c* 0.2, CHCl₃). Anal. Calcd for C₃₈H₄₂N₂O₄Mo: C, 66.47; H, 6.16; N, 4.08. Found: C, 66.63; H, 6.25; N, 3.84%. IR (KBr,

cm⁻¹): ν (Mo=O) 881, 916. ¹H NMR 7.97 (s, 1H), 7.84 (s, 1H), 7.47–7.53 (m, 5H), 7.41 (s, 1H), 7.38 (s, 1H), 7.25–7.27 (m, 5H), 6.84 (s, 1H), 6.80 (s, 1H), 6.12 (d, J=4.7 Hz, 1H), 4.70 (d, J=5.5 Hz, 1H), 2.27 (s, 3H), 2.16 (s, 3H), 1.48 (s, 9H), 1.21 (s, 9H). ¹³C NMR 167.6, 165.7, 161.7, 155.9, 140.0, 139.3, 138.5, 132.7, 131.4, 127.8 (2C), 127.5 (2C), 126.3 (2C), 125.9 (2C), 120.4, 119.3, 116.4, 79.4, 78.5, 73.8, 34.3, 34.0, 33.4, 31.7 (2C), 28.8 (2C), 28.2 (2C), 19.3, 19.1. ESI-MS: m/z 689 (M⁺ + H).

4.1.3. Asymmetric epoxidation of *cis*-1-propenylphosphonic acid. *cis*-1-Propenylphosphonic acid (25 mg, 0.2 mmol) dissolved in C₂H₅OH or CH₂Cl₂ (10 mL) was added racemic α -phenylethylamine (25 μ L, 0.2 mmol) to neutralize the acid. Then catalyst (0.01 mmol) was added to the mixture. After the addition of H₂O₂ (0.105 mL, 1.0 mmol, 30% aqueous), the reaction mixture was stirred for up to 72 h at 0 °C, 24 h at 25 °C or 1 h at 50 °C, respectively. The solvent was removed followed by the addition of water in order to dissolve the epoxide. The mixture was filtered and the filtrate was dried to give the product **2**.

Compound **2**. Mp 83–85 °C. $[\alpha]_{25}^{25}$ – 38.4° (*c* 0.2, CHCl₃). Anal. Calcd for C₁₁H₁₈NO₄P: C, 50.96; H, 7.00; N, 5.40. Found: C, 51.05; H, 7.12; N, 5.29%. IR (KBr, cm⁻¹): ν 3168, 2975, 2944, 2848, 2750, 1630, 1537, 1157, 1053, 914, 848, 702, 565. ¹H NMR 7.21–7.16 (d, *J*=9.6 Hz, 1H), 4.30– 4.23 (m, 1H), 3.09–3.04 (m, 1H), 2.72–2.63 (dd, *J*₁= 3.3 Hz, *J*₂=14.7 Hz, 1H), 1.38–1.36 (d, *J*=4.3 Hz, 3H), 0.92–0.87 (d, *J*=5.5 Hz, 1H). ¹³C NMR 137.9, 129.3 (2C), 129.2, 126.6 (2C), 54.4, 51.7, 51.0, 19.4, 13.4. ESI-MS: *m/z* 260 (M⁺ + H).

4.1.4. Synthesis of the dimethyl ester of fosfomycin. The crude product **2** (25.6 mg, 0.1 mmol) was dissolved in water followed by the addition of KOH (11.2 mg, 0.2 mmol). After stirring for 10 min, CH_2Cl_2 was added. The mixture was separated and the water layer was evaporated to obtained white solid. The solid was then dissolved in CH_3OH (5 mL). Upon the addition of methyl sulphoacid (19.5 mg, 0.2 mmol), much precipitate was produced. Under tempestuously stir, diazomethane in ether was added to the mixture until the yellow color was not disappeared. The precipitate was filtered off and the filtrate was concentrated to give a yellow oil. The crude dimethyl ester was then purified by flash chromatography. (CHCl₃/ C_2H_5OH 35:1) to obtain colorless oil.

Compound **3**. Anal. Calcd for $C_5H_{11}O_4P$: C, 36.15; H, 6.67. Found: C, 36.27; H, 6.53%. IR (KBr, cm⁻¹): ν 3475, 2960, 1413, 1248, 1034, 834, 786, 562. ¹H NMR 4.00–3.98 (d, J = 5.5 Hz, 3H), 3.94–3.92 (d, J = 5.4 Hz, 3H), 3.49–3.36 (m, 1H), 3.17–3.15 (d, J = 6.8 Hz, 1H), 3.04–3.01 (d, J = 6.8 Hz, 1H), 1.72–1.70 (d, J = 8.3 Hz, 3H). ¹³C NMR 53.2, 52.7, 50.6, 47.8, 13.9. ESI-MS: m/z 167 (M⁺ + H).

4.1.5. Determination of the ee value of the products. The pure dimethyl ester of fosfomycin **3** (10 mg, 0.06 mmol) was dissolved in CDCl₃ (0.5 mL) followed by adding optically active shift reagent $Eu(tfc)_3$ (5.38 mg, 0.006 mmol). The chemical shifts of CH₃O- in levo and

dextral dimethyl ester of fosfomycin were different in ¹H NMR and the ee value was determined by calibration.

Acknowledgements

We are grateful to the Institute of Medical Science, Tsinghua University, for financial support of this research and to Professor Ruimao Hua, Institute of Physical Chemistry, Tsinghua University, for the helpful discussion on kinetics and mechanism. We thank Professor Yuping Feng, Organic Phosphorus National Laboratory, for ¹H NMR support.

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Tetrahedron

Tetrahedron 60 (2004) 10999-11010

Efficient electrocatalytic intramolecular anion radical cyclobutanation reactions

Greg A. N. Felton and Nathan L. Bauld*

Department of Chemistry and Biochemistry, The University of Texas at Austin, Austin, TX 78712, USA

Received 3 June 2004; revised 18 August 2004; accepted 26 August 2004

Available online 2 October 2004

Abstract—Electrochemically initiated, intramolecular anion radical cyclobutanations of bis(enones) and related substrates are presented. The formation of novel anion radical Diels–Alder adducts in minor amounts is also observed. Total yields of pericyclic products, which include both *cis*- and *trans*-cyclobutanes and a single Diels–Alder adduct, are generally high (51–88%), with electrocatalytic factors in the range of 1.5–5. Mechanistically, strong evidence for the intervention of distonic anion radical intermediates as precursors of both types of pericyclic products is presented. The scope and limitations of these reactions are rather extensively explored and defined, and in particular the tendency, in some cases, for electrogenerated base-catalyzed reactions to compete with these anion radical pericyclic reactions. © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

Cyclobutanation reactions of the cation radicals of alkenes with neutral alkenes have by now become rather commonplace and are characterized by impressively high cycloaddition rates and low activation barriers, especially in comparison to the corresponding thermal reactions.^{1,2} There have been some recent indications that this extensive body of cation radical cyclobutanation chemistry may have a close counterpart in the domain of anion radical chemistry. Specifically, the reduction of phenyl vinyl sulfone under electrochemical conditions (mercury pool cathode) has been reported to yield trans-1,2-bis(phenylsulfonyl)cyclobutane.³ Subsequently, the cyclodimerizations of a variety vinylpyridines and vinylquinolines under similar conditions have also been established.⁴ Still more recently, a few intramolecular anion radical cyclobutanations of tethered bis(enones) have been described from these laboratories.^{5,6} These anion radical reactions are of special interest because they represent rare examples of intramolecular anion radical cycloaddition, rather than the more common electrohydrocyclization/dimerization (EHC or EHD).^{7,8}

The environmentally benign nature of these electrochemical conversions, inherent in the simplified workup and the consumption of electricity as the sole reagent (in catalytic amounts), further add to their experimental appeal and

0040–4020/\$ - see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.08.088

potential utility. The present paper reports the development of substantially more efficient conditions than those previously reported for carrying out these electrocatalytic intramolecular cycloaddition reactions in high yields, and further extends the scope and defines the limitations of these reactions. In particular, the use of tetraalkylammonium tetrafluoroborates as electrolytes in acetonitrile solution is developed as a particularly efficient method for these cyclobutanations. From a mechanistic viewpoint, experiments are described which strongly support the formulation of the cycloaddition reaction as proceeding in a stepwise fashion, via a distonic anion radical intermediate. Other new mechanistic and theoretical aspects of the reaction are clarified, including the requirement of at least one aroyl group, and the preference for two aroyl moieties.

2. Results and discussion

2.1. Electrolysis of 1a

The cyclobutanations of substrates **1a** and **1g** (Scheme 1) have previously been reported from these laboratories. Using lithium perchlorate (0.1 M) as the electrolyte, the reaction of **1a** was found to afford a total yield of pericyclic products (**2**, **3**) of just 45%, and the ratio of *trans-2/cis-2/3* was found to be 2.4:2.0:1. The total yield of pericyclic products obtained from **1g** was even smaller (35%).⁵ Consequently, various electrolytes (and other procedural changes) were investigated in the present work in order to determine whether synthetically useful procedures might be developed for carrying out these novel reactions.

Keywords: Anion radical; Cyclobutanation; Catalytic; Electrochemical reduction; Pericyclic; Electrogenerated base.

^{*} Corresponding author. Tel.: +1-512-471-3017; fax: +1-512-471-8696; e-mail addresses: bauld@mail.utexas.edu; greg_felton@hotmail.com



Scheme 1. Pericyclic products in the electrocatalytic, intramolecular anion radical cyclobutanation reactions of various bis(enones).

2.1.1. The anion radical mechanism. The anion radical chain cycloaddition mechanism proposed for these reactions is rather novel and is summarized in Scheme 2. According to this mechanism, reduction of the substrate at the cathode leads to a substrate anion radical, which then cyclizes to a distonic anion radical intermediate. This intermediate cyclizes to the anion radical of the cyclobutane product. The anion radical moiety presumably resides upon one of the benzoyl groups. Finally, exergonic electron transfer (ET) from the product anion radical to a molecule of the neutral substrate occurs, setting up the chain process and affording the neutral cyclobutane product. The distonic anion radical intermediate can also cyclize to a Diels–Alder adduct anion radical, where the anion radical moiety again resides upon a benzoyl moiety.

2.1.2. Tetraalkylammonium tetrafluoroborate electrolytes. The use of tetraalkylammonium tetrafluoroborates as electrolytes was explored on the assumption that ion pairing to the tetraalkylammonium cation should be much looser than with the lithium ion, resulting in a possible increase in reactivity of the anion radical intermediates. In fact, in the case of **1a**, a dramatic increase in yield to 88% was observed under these conditions. As will be noted in Table 1, the yield of the *trans* cyclobutane product (*trans-2*) is elevated to 59%. Since the *cis* isomer (obtained in 17% yield) is readily isomerized to the *trans* isomer under acidic or basic conditions, this reaction sequence makes the *trans* isomer available in an overall 71% yield. The novel Diels–Alder product (**3**), obtained in 13% yield, is also of inherent interest in that it represents the first documented instance of an anion radical Diels–Alder reaction. Since complete conversion of **1a** to products is accomplished after a maximum⁹ of 21% of the theoretical charge had flowed, the reaction is mildly electrocatalytic, with a catalytic factor of 4.7 (representing a 0.21 F mol⁻¹ process).

2.2. Electrolysis of 1b

2.2.1. Effect of electrogenerated bases. The extension of this chemistry to the ethereal substrate 1b was investigated next. Under conditions similar to the earlier work (using lithium perchlorate as the electrolyte)⁵ a total yield of 43%of pericyclic products was isolated, with *trans-2b* being the major product. Under both the present and former conditions the catalytic factor was <3. When tetrabutylammonium tetrafluoroborate was used as the electrolyte, pericyclics were isolated in 53% yield. Interestingly, this reaction proceeded somewhat more efficiently in terms of the catalytic factor (10.6) than any of the other reactions, but the yield of pericyclic products was diminished in comparison to the corresponding reduction of substrate 1a when using the same electrolyte because of a competing reaction of the substrate (Scheme 3). The formation of 4b implies a base catalyzed deprotonation of 1b, followed by a Michael type addition of the conjugate base via the alpha carbon of the extended enolate to the beta carbon of the enone moiety. Formation of this product in the reaction using tetrabutylammonium tetrafluoroborate as the electrolyte, and not with any of the other electrolytes, is in



Scheme 2. Proposed mechanism for the anion radical chain cyclobutanation reaction of 1,7-dibenzoyl-1,6-heptadiene.

Substrate	Procedure ^a	Yield of <i>trans</i> -2	Yield of cis-2	Yield of 3	Total pericyclic yield	Other products	Catalytic factor
1a	А	59%	17%	13%	88%		4.7
1b	А	39	11	3	53	4b; 11%	10.6
1b	В	21	20	2.0	43	_	1.6
1b	С	21	39	28	88		1.8
1c	А	52	11	12	75	5c ; 5%	5.1
1d	B+C	7	33	16	56	6d; 6%	1.3
1d	D	3	0	0	3	5d ; 17% ^b	7.3
1e	А	29	14	8	51	5e; 17%	5.2
1f	С	7	21	25	53	5f; 13%, 7f; 7%	1.9
1f	А	12	0	0	12	5f ; 17%	1.4
1g	С	0	9	0	9	7g ; 30% ^c	<1
1g	D	19 ^d	0	0	19	_	1.8
1i	В	32 ^e	0	0	32	_	<1
1j	D	0	0	0	0	5j ; 42%	4.5

Table 1. Yields and catalytic factors for the electrocatalytic intramolecular anion radical pericyclic reactions of various bis(enone) substrates (1)

^a The electrolytes used in the respective procedures were: A=0.1 M tetrabutylammonium tetrafluoroborate; B=0.1 M lithium perchlorate; C=0.1 M magnesium perchlorate; D=0.1 M tetraethylammonium tetrafluoroborate.

^b Also obtained 35% of an unidentified polymer.

^c Two isomers.

 d Both trans isomers were observed: 12 and 7% ($\sim\!2{:}1$ ratio), X-ray of major isomer provided.

^e Both *trans* isomers were observed: 28 and 4% (7:1 ratio).

agreement with the postulate that the intermediate anionic species are less tightly ion paired under these conditions, and are therefore substantially more reactive (basic). Since a product analogous to 4b was not observed in the electrochemical reaction of 1a, it is apparent that the substitution of oxygen for carbon tends to acidify the adjacent C-H bond. A similar competition has been reported in the EHC of butenolides.¹⁰ In the case of substrate 1b, a product corresponding to the isomerization of the double bond, into conjugation with the carbonyl group, was not observed. However, in several other instances products corresponding to such a structure, 5,¹¹ were obtained instead of 4. The nature of the electrogenerated base (EGB)^{12,13} species responsible for the deprotonation is unknown, but likely candidates might be a substrate or product anion radical or a dianionic intermediate produced by further reduction of the proposed distonic anion radical intermediate.

2.2.2. Use of $Mg(ClO_4)_2$ electrolyte. Although the use of tetraalkylammonium tetrafluoroborates as the electrolyte appears to be the more general method of choice for carrying out most of these anion radical cycloadditions with

optimal efficiency, in the specific case of **1b** the competing reaction described above tended to lower the yields of the pericyclic products. In view of that, it appeared worthwhile to investigate the effect of using a more strongly ion-pairing counterion than even the lithium ion, that is, the magnesium ion. When the electrochemical reaction was carried out in the presence of magnesium perchlorate as the electrolyte, an 88% yield of pericyclic products was obtained. The increase in the *cis/trans* ratio in the cyclobutane product may also reflect the stronger ion pairing between the magnesium ion and the anion radical moiety in the transition state for the final cyclization step (vide infra).

2.3. Electrolysis of 1c

It had been found in the previously reported work⁵ that the introduction of electron donating substituents, such as 4-methoxy, onto the aryl ring of **1a** sharply inhibits cyclobutanation. It is presumed that this is the result, at least in part, of the inability of the distonic anion radical intermediate to cyclize to the product cyclobutane anion radical, which would require the anion radical moiety to



Scheme 3. The electrogenerated base (EGB) catalyzed products (4 and 5) obtained in the electrochemical reduction of substrates 1, in the presence of tetraalkylammonium tetrafluoroborate as the electrolyte.



Scheme 4. Formation of an aldol-type side product (6), via reduction of the distonic anion radical to an enolate dianion.

reside in a higher energy SOMO (that of a 4-methoxybenzoyl moiety) than in the case of the unsubstituted substrate. Consequently, the facility of bis(enone) anion radical cyclobutanation of substrates that would provide a lower energy product anion radical SOMO was investigated. Substrate 1c, which has electron withdrawing 4-chloro substituents on both of its benzoyl groups, was found to undergo smooth anion radical cycloaddition under the tetrabutylammonium tetrafluoroborate electrolyte conditions, affording a 75% yield of total pericyclics, of which 52% was the trans cyclobutane. It is of special interest that the chlorine substituent is retained in the product, even though the 4-chlorobenzoyl anion radical moiety was potentially susceptible to chloride ion loss. Presumably, electron transfer from the product anion radical to substrate is sufficiently rapid as to suppress the potential loss of chloride ion.

2.4. Electrolysis of 1d

The introduction of an even more strongly electron withdrawing meta chloro substituent into the substrate, in addition to the *para* chloro substituent, as in substrate 1d would be expected to accelerate the rate of the second cyclization step even further by providing a lower energy SOMO for the product anion radical. However, the considerably increased stabilization of the substrate anion radical, and the expected shift in electron density in the SOMO from the alkene moiety to the now rather strongly electron-deficient aroyl ring could also be expected to have an adverse effect upon the rate of the first cyclization step. In accord with the latter idea, when 1d is electrolytically reduced in the presence of the tetraethylammonium tetrafluoroborate electrolyte, the yield of pericyclic products falls precipitously to 3%, the main products consisting of a 17% yield of a base-catalyzed cyclization/isomerization product (5d) and an uncharacterized polymer (35%). However, in the presence of a mixed lithium perchlorate/ magnesium perchlorate electrolyte, the desired pericyclic reactions are observed to occur with moderate efficiency, but the yields of the pericyclic products are lower than in the case of 1c, and an aldol-type product (6d) is also formed. This latter product is considered to result from reduction of the intermediate distonic anion radical to a dianion, which subsequently is protonated and undergoes cyclization (Scheme 4).

2.5. Electrolysis of 1e

The replacement of the phenyl group with a β -naphthyl or 4-biphenylyl group could also provide a more suitable venue for the stabilization of the anion radical of the product than in the case of 1a, but again the lowering of the SOMO energy and the shifting of the SOMO density toward the aroyl moiety could retard the first cyclization step. The cyclobutanation of substrate 1e (which has β -naphthyl substituents), using tetrabutylammonium tetrafluoroborate as the electrolyte, was found to proceed, albeit in moderate yield (51%), suggesting that the predominant effect of the increased delocalization provided by the naphthyl group is the lowering of the SOMO energy and the shift in density away from the alkene linkage. However, the retardation of the cyclization is evidently much less than in the case of the dichloro substrate 1d. As in this latter case, the basecatalyzed cyclization/isomerization product 5e was also formed (17%; Table 2). It is worth noting that when the reaction is not run to completion ($\sim 80\%$ complete), the yield of pericyclic products is relatively unchanged, but the amount of the base catalyzed product is greatly reduced. This may well be indicative of a build up of electrogenerated bases within the solution, accelerating the cyclization to 5 in the latter stages of the reaction. This build up of base could also provide an explanation for the increase in the trans/cis ratio observed in the more complete reaction (vide infra).

2.6. Electrolysis of 1f

Substrate **1f**, which contains 4-biphenylyl moieties, responds in a manner rather similar to that of **1d**. Limited *trans-***2f** formation is observed, along with greater formation of the base-catalyzed cyclization/isomerization product **5f** (tetraalkylammonium electrolyte). However the use of $Mg(ClO_4)_2$ afforded a moderate pericyclic yield of 53%. Substrate **1f** has limited solubility in acetonitrile, so that these reductions were carried out in 1:1 THF/acetonitrile solutions.

Table 2. The effect of the extent of reaction upon the yields of anion radical and base-catalyzed products

Substrate ^a	Yield of <i>trans</i> -2	Yield of cis-2	Yield of 3	Total pericyclic yield	Other products	Catalytic factor
1e	29	14	8	51	5e ; 17%	5.2
1e ^b	17	21	10	48	5e ; 6%	5.4

^a The electrolyte was 0.1 M tetrabutylammonium tetrafluoroborate.

^b Reaction run to 79% completion (based upon recovered 1e), yields and catalytic factor are corrected for 1e recovery.

2.7. SOMO requirements for cyclization

2.7.1. Electrolysis of 1g and 1h. In contrast to the aroyl groups considered above, acetyl groups provide a much less extensively delocalized, higher energy SOMO for the product cyclobutane or Diels–Alder anion radical. The potential anion radical pericyclic chemistry of substrate **1h**, which has two acetyl substitutents, was therefore not expected to be as efficient as when aroyl groups are present. In accord with this supposition, no pericyclic products at all could be detected in the electroreduction of this substrate.

However, since the anion radical of the pericyclic products only requires (and perhaps can only utilize) a single aroyl moiety, it was considered likely that pericyclic chemistry might occur with unsymmetrical substrate 1g, which has one benzoyl and one acetyl substituent. The previously reported results, using lithium perchlorate as an electrolyte, have already confirmed this conjecture, but further experiments were carried out in the present work in connection with magnesium perchlorate and tetraethylammonium tetrafluoroborate as the electrolytes. The observed yields in both cases are quite modest. However, an additional feature of interest emerges when tetraethylammonium tetrafluoroborate is used as the electrolyte, namely, that both possible trans-2g isomers are formed, in a 2:1 ratio, with the major cyclobutane having the benzoyl group syn to the cyclopentane ring.

2.7.2. Electrolysis of 1i. The reduction of **1i**, which has one benzoyl and one carboethoxy substituent, provides a still further example of the relative inefficiency of pericyclic chemistry when only a single benzoyl group is present. When electrolyzed in the presence of lithium perchlorate, **1i** provides a 32% yield of two isomers of *trans*-**2i**. The major isomer appears, on the basis of NMR comparisons, to be structurally analogous to the major *trans*-**2g** isomer obtained from **1g**. This reduction is unusual in that it is not catalytic. An intriguing possible interpretation of this data is that while benzoyl reduction step is sharply retarded by the ineffectiveness of the ester function at delocalizing and stabilizing the SOMO in the transition state. This could require cyclization to occur via the rarer reduction of the

unsaturated ester function. This higher energy anion radical could then rapidly cyclize to the reactive benzoyl ene function. The key factor here is that catalysis would necessarily involve electron transfer from a product anion radical to a substrate molecule, and this chemical electron transfer undoubtedly is highly selective for formation of the more stable, and apparently unreactive, anion radical corresponding to the benzoyl enone function. Additionally, this chemical electron transfer is likely to be irreversible in nature, halting catalysis. The chemically reduced substrate may prove reactive in non-cyclobutanation mechanisms, which may account for the observed low yield.

2.7.3. Diminished cyclization rates. Since the desired pericylic reaction products provide the required low energy SOMO associated with an aroyl function (in this case, benzoyl), and the starting substrate provides a readily reducible aroyl enone function, it appears likely that the lower efficiency of the desired pericyclic chemistry in the substrates which contain only a single aroyl function must arise from diminished cyclization rates in either one or both of the cyclization steps. It therefore seems reasonable to propose that, in the transition states for both cyclization steps, the SOMO is at least partially delocalized over both enone moieties, as indicated in Scheme 5. The delocalization is presumably greatest, and the SOMO energy the lowest, when both keto functions are of the aroyl type.

2.8. Evidence for the distonic anion radical intermediate

In most cases the electrolyte systems (lithium/magnesium perchlorate or tetraalkylammonium tetrafluoroborate) lead to the formation of a mixture of the *cis* and *trans* cyclobutane isomers. At the earliest point of the electrochemical reactions at which these products could be detected, the *trans* cyclobutane was always formed in modest excess over the *cis* isomer. As the reaction progressed further toward completion, the *trans/cis* ratio progressively increased. The values given in Table 1 correspond to reactions run essentially to completion. This progressive change in the *trans/cis* ratio suggested the possibility that a portion of the *cis* isomer in the course of the reaction. Although a base-catalyzed isomerization was



Scheme 5. Proposed delocalization of the SOMO over both carbonyl groups in the transition states for both cyclization steps.



Scheme 6. Mechanism of formation of 7, via protonation of distonic intermediate.

formally a possibility, it also appeared possible that anion radicals of the *cis*-cyclobutane were being reformed during the reaction and subsequently reverting to the distonic anion radical, which could once again cyclize to give either cyclobutane isomer or the Diels-Alder adduct. A distinction between these two mechanistic possibilities is therefore possible, based upon the predicted formation of small amounts of the Diels-Alder adduct in the latter mechanism. To test this possibility, isomers of **2b** were isolated and used in electrolysis reactions with lithium perchlorate as the electrolyte. A quantity of cis-2b was reduced in isolation, leading to the formation of *trans*-2b (32%), 3b (7%), an aldol product **6b** (11%), a dihydrocyclopentane product **7b** (20%; see Scheme 6), with a further 18% of unreacted cis-**2b**. The attempted reduction of *trans*-**2b**, as expected, failed to lead to any reaction. A degree of reactivity was seen under extreme conditions (very negative potentials, large amounts of charge). Although some trans-2b was still returned, no other known products were obtained. This clearly indicates that, while the *trans* isomer is stable toward continued reduction at normal potentials, the cis isomer readily reverts to the proposed distonic intermediate. This not only allows for the formation of the *trans* and Diels-Alder products, but also of **7b**, which represents protonation after the first cyclization, effectively 'trapping' the distonic anion. In further accord with the postulate of cis-trans isomerization via regeneration of the distonic anion radical intermediate is the previously discussed absence of any base-catalyzed products (e.g., 4 and 5) in any perchlorate reaction system. Since the distonic anion radical intermediate is evidently involved in the reversal of the cycloaddition, an application of the law of microscopic reversibility strongly suggests that the forward reaction also involves the same distonic intermediate.

2.9. Trapping of the distonic anion radical intermediate/inhibition of EGB pathway

It was thought that addition of a slight excess of a weak acid would not only inhibit EGB pathways but would also trap (protonate) the proposed distonic anion radical. Indeed this was realized with a 1.6 M excess of acetic acid placed in the solution from the beginning of electrolysis. The naphthoyl substrate **1e** was chosen as it leads to modest base-catalyzed product formation (17% of **5e**). The expected trapping is clearly the dominant mechanism, represented by both **6e** and **7e** formation (Table 3). The expected reduction in basecatalysis (product **5e**) is also seen, down to just 3%. Also, by limiting the excess of acetic acid, we were still able to obtain small amounts of our primary cyclobutanation product (*trans*-**2e**), although formation of *cis*-**2e** and **3e** were reduced to levels below detection. The proposed mechanism for the formation of **7** is given in Scheme 6.

2.10. Mediated electrolysis

The possibility of developing a mediated electrochemical reduction method was also probed. The strategy adopted was to provide a mediator which is (in the ideal case, selectively) reduced at a less negative potential than the substrate, and which forms a relatively long-lived anion radical capable of mildly endergonic electron transfer to the substrate molecules.^{14–16} This strategy should provide for a low substrate anion radical concentration, which could minimize anion radical to anion radical coupling as well as over-reduction of the substrate (dianion formation). Initial attempts utilized **1b** as the substrate and magnesium or lithium perchlorate as the electrolyte. Benzil (diphenyl diketone), dypnone (*E*-3-phenyl-2-butenoylbenzene), and benzophenone were investigated as mediators, but all three

Table 3. Effect of a weak acid in solution during electrolysis of 1e

Additive	% Yield of trans-2e	% Yield of cis-2e	% Yield of 3e	% Yield of 5e	% Yield of 6e	% Yield of 7e	% Yield of products
None	29	14	8	17	0	0	68
1.6 M excess of acetic acid	6	0	0	3	14	38	61

Table 4. Anion radical versus base-catalyzed reactions in the mediated electrolysis of 1c

Mediator	% Yield of <i>trans</i> -2c	% Yield of cis-2c	% Yield of 3c	% Yield of 5c
None	52	11	12	5
Benzophenone	42	11	7	13
Benzil	0	0	0	28 ^a

^a Along with 42% unidentified polymer.

proved to be ineffective. Mediator reduction did occur at less negative potentials (as evidenced by a temporary solution color change) than that of the substrate, but products did not form until the potential was reduced to the usual value for reduction of the substrate.

Further studies were directed toward the use of tetraethylammonium tetrafluoroborate as the electrolyte, this time using 1c as the substrate. Although these conditions failed to provide an increase in yield of pericyclic products, they did provide important insights into the interplay between anion radical cyclization and electrogenerated base catalysis (Table 4). Benzophenone reduction was found to occur at nearly the same potential as for 1c, when benzil reduction occurred at a much less negative potential. In the case of benzil, this occurred at a potential that did not lead to substrate reduction, so no pericyclic products were formed. On the other hand, in the absence of a mediator, base-catalyzed products (such as 5c) were formed in a low vield. When benzophenone was employed as the mediator, both types of products were formed, indicating that both the substrate and mediator are being reduced at the cathode. Apparently, anion radical cyclobutanation occurs only when the substrate is reduced, and reduction of the mediator leads essentially only to the base-catalyzed reaction. These observations indicate that the desired mildly endergonic electron transfer is too slow to compete with the basecatalyzed reactions.

2.11. Efficient base-catalyzed reactions

The observations noted above immediately suggested the possibility of employing an excess of benzil to engender a highly catalytic method for selectively and efficiently forming the base-catalyzed product. This possibility was realized in the case of the electrochemical reduction of 1b in the presence of a relatively large excess of benzil, which gave only 4b, in 68% yield. The catalytic factor here was rather modest at 2.8, although low concentrations of starting material and a large excess of benzil made judging the end of the reaction difficult. Comparison of this result with that obtained for 1c (with benzil) suggests that the use of mediators to engender these base-catalyzed cyclizations may prove problematic, due to competition with polymerization. The efficiency seen with formation of 4b may be engendered by the greater acidity of the protons (alpha to the bridging oxygen) in 1b.

3. Conclusions

The present paper reports the development of conditions for carrying out electrochemically initiated, intramolecular anion radical cyclobutanations of bis(enones) and related substrates in high yields and with substantially less than the theoretical consumption of electricity (i.e., electrocatalytically). The solvent/electrolyte combination acetonitrile/ tetraalkylammonium tetrafluoroborate is found to be an especially effective one for producing high yields and large catalytic factors. The formation of novel anion radical Diels-Alder adducts in minor amounts is also verified. The scope and limitations of these reactions are rather extensively explored and defined. In particular, the reactions have been found to have an absolute requirement for at least one aroyl ketone moiety and a significant preference for both ketonic moieties to be of the aroyl type. Theoretical rationales for these requirements and preferences are presented. Strongly electron-withdrawing substituents (upon the aroyl moiety) tend to decrease reaction efficiency by diminishing the rate of the first cyclization step, such that a competition between anion radical mediated and electrogenerated base-catalyzed reactions is observed. Evidence for a stepwise (as opposed to concerted) cycloaddition mechanism involving a distonic anion radical intermediate is presented, and the distonic anion intermediate has been trapped.

4. Experimental

4.1. General electrolysis procedure

A typical experiment utilizes 100 mg of a given bis(enone) substrate, which equates to 0.329 mmol for substrate **1a**. The substrate is dissolved in 22 mL of electrolyte solution, giving a typical substrate concentration of 0.0150 M, and added to the working electrode (WE) compartment of the electrolysis cell. The electrolyte solution is 0.100 M (unless stated) in either alkylammonium or perchlorate salt (detailed below) in dry acetonitrile (unless stated). The acetonitrile is distilled fresh for each electrolysis from a reservoir containing phosphorus pentoxide. Electrolyte solution (6 mL) is added to the counter electrode (CE) compartment.

Electrolysis of the substrate was carried out at specific voltages (detailed below) with stirring under positive nitrogen flow at room temperature. The voltage is commonly increased through the course of an electrolysis to help maintain the current (tracked by coulometer). This increase is detailed in the description of each electrolysis. Electrolysis voltages were versus a 'pseudo-standard' silver wire (encased in porous vycor glass) reference electrode (RE).¹⁷ The RE used is seen to have a calibration to SCE of approximately +0.1 V, when in 0.1 M Et₄NBF₄ acetonitrile solution. The CE and WE consisted of reticulated vitreous carbon (25 mm×5 mm), their corresponding compartments separated by a course frit. The RE was placed within 0.5 cm of the WE. The reaction was stopped when thin-layer chromatography (TLC) indicated that the starting

material had been consumed. The reactant solution (WE compartment only) then underwent an aqueous workup with sequential benzene washings (alkyl ammonium salt) or dichloromethane (perchlorate salt). The organic phase was retained and dried with Na₂SO₄. The benzene/dichloromethane was removed by rotary evaporation, with the crude solution being purified by preparative TLC (1 mm thick, elution with ethyl acetate/petroleum ether mixture, 1:9 ratio, unless stated). Bands were identified, collected by scraping, and extracted with dichloromethane. Filtering removed the silica, with a rotary evaporator again employed to remove the solvent, yielding the desired products.

Notes. In cases when starting material is recovered, the yields of products and the catalytic factors are corrected accordingly. Characterization of products is in most cases completed by comparison to the products obtained from **1b** electrolysis, and to results previously published (as referenced). The products from **1b** are more fully characterized both by 500 MHz NMR (1 H/ 13 C) and HRMS. Their ready crystallization also allowed for X-ray structural determination (provided previously⁶). All novel products are also characterized by HRMS or, in two cases, by X-ray crystallography.

4.2. Analysis

Room temperature ¹H NMR spectra were recorded on a Varian Unity + 300 as solutions in CDCl₃. ¹³C NMR and COSY spectra were recorded on a Varian Unity Inova 500 spectrometer. Chemical shifts (δ) are relative to tetra-methylsilane, and coupling constants (*J*) are given in Hertz (Hz). Splitting patterns are designated as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; dd, doublet of doublets; br, broad. X-ray diffraction analyses were conducted using a Nonius Kappa CCD diffractometer. Low-resolution mass spectra (LRMS) were recorded on a Finnigan MAT TSQ-70 mass spectrometer, with high-resolution mass spectra (HRMS) recorded on a VGZAB-2E mass spectrometer.

4.3. Equipment/reagents

A potentiostatic controller, the Electrosynthesis Company (ESC) model 415, was used to control the applied potential. The charge used was tracked by a digital coulometer, ESC model 640. The applied potential was confirmed using a digital multimeter (Wavetek DM7), operating as a potentiometer. The electrode material was Duocel 80 PPI reticulated vitreous carbon. The substrates were kindly produced within the Krische group or by Dr. Jingkui Yang.^{5,6} Reagent purity was assayed by NMR/LRMS. Electrolytes were used as purchased, from Alfa Aesar (98% purity).

4.4. Electrolysis of *E,E*-1,7-dibenzoyl-1,6-heptadiene (1a)

Electrolysis of 111 mg (0.0166 M) of **1a** with Bu_4NBF_4 electrolyte, at -2.0 V versus RE (first 3.0 C at -1.5 V). The reaction appeared complete after 7.5 C, or 21.3% (of required charge) had passed through the cell. PTLC purification of the 105 mg of recovered crude yielded *cis*-**2a** (19 mg, 17%), *trans*-**2a** (65 mg, 59%), and **3a** (14 mg,

13%) for a total of 98 mg of pericyclic products (88%). No starting material was recovered, and no other products were identified.

4.4.1. Compound *cis*-2a^{5,6}. ¹H NMR (300 MHz, CDCl₃): 1.65 (2H, m), 1.84 (2H, m), 2.02 (2H, m), 3.20 (2H, br.s), 3.85 (2H, d, *J*=4.2 Hz), 7.35 (4H, br.t, *J*=7.2 Hz,), 7.44 (2H, br.t, *J*=7.2 Hz), 7.75 (2H, br.d, *J*=8.1 Hz).

4.4.2. Compound *trans*-2a^{5,6}. ¹H NMR (300 MHz, CDCl₃): 1.42 (2H, m), 1.52 (1H, m), 1.85 (3H, br.m), 3.06 (1H, q, *J*=6.9 Hz), 3.24 (1H, m), 4.28 (1H, dd, *J*=6.6, 7.5 Hz), 4.57 (1H, dd, *J*=7.8, 10.5 Hz), 7.46 (4H, m), 7.55 (2H, m), 7.95 (2H, br.d, *J*=6.9 Hz), 8.02 (2H, br.d, *J*=7.2 Hz).

4.4.3. Compound 3a^{5,6}. ¹H NMR (300 MHz, CDCl₃): 1.60 (2H, m), 1.84 (2H, m), 2.01 (2H, m), 2.69 (2H, m), 4.89 (1H, d, *J*=6.6 Hz), 5.59 (1H, d, *J*=3.0 Hz), 7.26 (3H, m), 7.48 (4H, m), 8.08 (2H, d, *J*=7.5 Hz).

4.5. Electrolysis of *E*,*E*-1,7-dibenzoyl-4-oxa-1,6-heptadiene (1b)

Electrolysis of 100 mg (0.0149 M) of **1b** with Mg(ClO₄)₂ electrolyte, at increasing voltages versus RE. The first 3.0 C at -2.0 V, then 1.8 C at -3.0 V, followed by 1.3 C at -3.5 V, and 6.0 C at -4.0 V. The reaction appeared complete after 12.1 C, or 38.4% (of the required charge) had passed through the cell. PTLC purification (1:4 ratio of EA/PET) of the 156 mg of recovered crude yielded *cis*-**2b** (26 mg, 39%), *trans*-**2b** (14 mg, 21%), and **3b** (19 mg, 28%), giving a total of 59 mg of pericyclic products (88%). While no other products were identified, 33 mg of starting material was recovered. The above percent of required charge is based upon the 100 mg of starting material being used up; this becomes 54.8% based upon the 67 mg of starting material used up (not recovered).

Electrolysis of 205 mg (0.0305 M) of **1b** with LiClO₄ electrolyte, at -1.3 V for 40.0 C (62.7% of required charge). PTLC separation of the 170 mg of crude yielded *cis*-**2b** (41 mg, 20%), *trans*-**2b** (42.2 mg, 21%), **3b** (4 mg, 2%), along with 2.8 mg of unreacted **1b**.

Electrolysis of 105 mg (0.0156 M) of **1b** with Bu_4NBF_4 electrolyte, at -1.6 V for 2.1 C and -2.0 V for 0.9 C. This corresponds to 9.4% of required charge. PTLC separation of the 108 mg of crude yielded *cis*-**2b** (12 mg, 11%), *trans*-**2b** (41 mg, 39%), **3b** (3 mg, 3%), and **4b** (11 mg, 11%).

Electrolysis of 28 mg (0.0042 M) of **1b** with Et_4NBF_4 electrolyte, and 157 mg (0.340 M) of benzil, at -1.4 V for 3.2 C. This corresponds to 36.2% of required charge, or a catalytic factor of 2.76. PTLC separation proved problematic due to the large benzil excess, however a yield of 19.1 mg (68%) of **4b** was obtained by NMR integration of several mixed **4b**/benzil PTLC bands. No other products or unreacted starting material were observed.

Electrolysis of 66 mg (0.0098 M) of *cis*-**2b** with LiClO₄ electrolyte, at -2.5 V for 27.5 C, -3.0 V for 4.5 C, and then at -3.5 V for a further 30.0 C. PTLC separation of the crude yielded *trans*-**2b** (17.3 mg, 32%), **3b** (4 mg, 7%), an

aldol product **6b** (5.7 mg, 11%), a dihydro product **7b** (11 mg, 20%), with a further 12 mg (18%) of unreacted *cis*-**2b**.

Electrolysis of 52 mg (0.0077 M) of *trans*-**2b** with LiClO₄ electrolyte, at -3.0 V for 11 C, -3.5 V for 130 C, and -4.0 V for 20 C. No product spots were observed until after ~ 40 C, indicating that no reaction was occurring, hence the continued flow of charge. PTLC of the 68 mg of crude yielded 8 mg of impure *trans*-**2b**, along with 19 mg of unidentified product (single benzoyl moiety). None of the characterized products (such as *cis*-**2b**) were observed.

4.5.1. Compound *cis*-**2b.**⁶ ¹H NMR (500 MHz, CDCl₃): 3.42 (2H, m), 3.63 (2H, d.m, J=10.2 Hz), 4.14 (4H, m), 7.34 (4H, br.t, J=7.6 Hz), 7.45 (2H, br.t, J=7.3 Hz), 7.72 (4H, br.d, J=8.2 Hz); ¹³C NMR (500 MHz in CDCl₃): 39.72, 47.70, 73.46, 127.81, 128.59, 132.8, 136.05, 198.08; NMR COSY (500 MHz), and X-ray crystallography confirm structure; ⁶ HRMS (CI+): calcd; 307.133420. Found; 307.132575.

4.5.2. Compound *trans*-2b.⁶ ¹H NMR (500 MHz, CDCl₃): 3.36 (3H, br.m), 3.46 (1H, dd, J=4.6, 9.6 Hz), 3.68 (1H, d, J=9.8 Hz), 4.06 (1H, d, J=9.6 Hz), 4.41 (1H, m), 4.53 (1H, m), 7.45 (4H, m), 7.54 (2H, br.m), 7.90 (2H, m), and 8.01 (2H, m); ¹³C NMR (500 MHz in CDCl₃): 39.71, 40.74, 42.99, 43.01, 69.27, 72.55, 128.29, 128.71, 128.76, 128.91, 133.41, 133.43, 135.27, 135.61, 196.70, 199.70; NMR COSY (500 MHz), and X-ray crystallography confirms the structure; ⁶ HRMS (CI+): calcd; 307.133420. Found; 307.133048.

4.5.3. Compound 3b.⁶ ¹H NMR (500 MHz, CDCl₃): 2.99 (1H, m), 3.06 (1H, m), 3.58 (1H, m), 3.75 (1H, dd, J=4.4, 9.6 Hz), 4.17 (2H, m), 4.98 (1H, d, J=8.6 Hz), 5.58 (1H, d, J=4.2 Hz), 7.26 (2H, m), 7.48 (5H, m), 7.61 (1H, m), 8.08 (2H, m); ¹³C NMR (500 MHz in CDCl₃): 35.54, 36.94, 70.30, 74.03, 76.30, 98.89, 124.65, 128.24, 128.44, 128.62, 129.51, 133.70, 134.44, 135.41, 151.80, 196.01; NMR COSY (500 MHz) and X-ray crystallography confirms the structure; ⁶ HRMS (CI+): calcd; 307.133420. Found; 307.132957.

4.5.4. Compound 4b. ¹H NMR (300 MHz, CDCl₃): 3.08 (1H, m), 3.17 (1H, d, J=5.7 Hz), 3.20 (1H, d, J=7.5 Hz), 3.85 (1H, m), 3.98 (1H, dd, J=3.3, 10.8 Hz), 4.23 (1H, dd, J=2.7, 10.8 Hz), 4.72 (1H, td, J<1.5, 6.0 Hz), 6.52 (1H, dd, J=1.8, 6.0 Hz), 7.48 (4H, br.t, J=7.5 Hz), 7.58 (2H, m), 8.00 (4H, m). X-ray crystallography confirms structure (CCDC 240331).

4.5.5. Compound 6b.⁶ ¹H NMR (300 MHz, CDCl₃): 1.98 (1H, ddd, J=1.5, 8.1, 12.9 Hz), 2.45 (1H, dd, J=8.1, 12.9 Hz), 3.22 (2H, m), 3.64 (2H, dd, J=6.3, 9.0 Hz), 3.85 (2H, t, J=9.9 Hz), 4.12 (1H, d, J=9.6 Hz), 5.35 (1H, d, J=2.1 Hz), 7.14 (1H, m), 7.24 (2H, m), 7.40 (2H, m), 7.52 (2H, m), 7.59 (1H, m), 7.80 (2H, dd, J=1.5, 8.7 Hz).

4.5.6. Compound *cis*-**7b.** ¹H NMR (300 MHz, CDCl₃): 3.04 (4H, br.m), 3.16 (2H, dd, J=3.0, 14.1 Hz), 3.60 (2H, dd, J=4.2, 8.7 Hz), 4.12 (2H, m), 7.47 (4H, br.t, J=

7.8 Hz), 7.58 (2H, br.t, *J*=7.8 Hz), 7.94 (4H, m); HRMS (CI+): calcd; 309.149070. Found; 309.149110.

4.6. Electrolysis of *E*,*E***-1**,**7**-bis(4-chlorobenzoyl)-1,6-heptadiene (1c)

Electrolysis of 99 mg (0.012 M) of **1c** with Bu_4NBF_4 electrolyte, at -1.2 V for 2.1 C, -1.6 V for 2.2 C, and -1.8 V for 0.8 C. The reaction appeared complete after 5.0 C, or 19.5% (of required charge) had passed through the cell. PTLC purification of the 124 mg of recovered crude yielded *cis*-**2c** (11 mg, 11%), *trans*-**2c** (51.5 mg, 52%), and **3c** (11.5 mg, 12%), for a total yield of 74 mg of pericyclic products (75%). No starting material was recovered. However, a further 5 mg (5%) of **5c** was recovered.

Electrolysis of 55 mg (0.0067 M) of **1c** with 32 mg (0.0080 M) of benzophenone and Et_4NBF_4 electrolyte, at -1.8 V for 1.5 C (10.5% of required charge). **1c**/benzophenone=1:1.19. PTLC separation yielded *cis*-**2c** (6 mg, 11%), *trans*-**2c** (23 mg, 42%), and **3c** (4 mg, 7%), for a total yield of 33 mg of pericyclic products (60%). A further 7 mg (13%) of **5c** was recovered.

Electrolysis of 99 mg (0.012 M) of **1c** with 165 mg (0.036 M) of benzil and Et_4NBF_4 electrolyte, at -1.3 V for 2.5 C (9.7% of required charge). **1c**/benzil=1:1.2.96. PTLC separation yielded 28 mg (28%) of **5c** along with 42 mg (42%) of an unidentified polymer.

4.6.1. Compound *cis*-**2c.**⁶ ¹H NMR (300 MHz, CDCl₃) 1.68 (2H, m), 1.82 (2H, dd, J=9.9, 2.4 Hz), 2.00 (2H, m), 3.16 (2H, m), 3.77 (2H, d, J=3.9 Hz), 7.32 (4H, d, J=8.4 Hz), 7.66 (4H, d, J=9.0 Hz); HRMS (CI+): calcd; 373.076210. Found 373.076164.

4.6.2. Compound trans-2c.⁶ ¹H NMR (300 MHz, CDCl₃): 1.38 (2H, m), 1.58 (1H, m), 1.82 (3H, m), 3.04 (1H, dd, J =6.9, 13.2 Hz), 3.22 (1H, m), 4.20 (1H, dd, J = 0.9, 7.5 Hz), 4.49 (1H, dd, J = 2.4, 10.2 Hz), 7.44 (4H, dd, J = 2.7, 8.4 Hz), 7.88 (2H, br. d, J = 8.4 Hz), 7.95 (2H, br.d, J =9.0 Hz):); HRMS (CI+): calcd; 373.076210. Found; 373.075821.

4.6.3. Compound 3c.⁶ ¹H NMR (300 MHz, CDCl₃): 1.41 (1H, m), 1.59 (2H, m), 1.78 (1H, m), 2.01 (2H, m), 2.68 (2H, m), 4.80 (1H, d, J=6.9 Hz), 5.52 (1H, d, J<1.5 Hz), 7.40 (2H, m), 7.47 (4H, m), 8.01 (2H, d, J=8.4 Hz); HRMS (CI+): calcd; 373.076210. Found; 373.076898.

4.6.4. Compound 5c. ¹H NMR (300 MHz, CDCl₃): 1.64 (1H, m), 1.72 (3H, m), 2.30 (2H, br.m), 2.79 (1H, dd, J = 10.2, 14.7 Hz), 3.34 (1H, dd, J = 3.6, 14.7 Hz), 3.43 (1H, br.s), 6.61 (1H, td, J = 3.9, 1.2 Hz), 7.43 (3H, m), 7.51 (1H, m), 7.62 (2H, m), 7.99 (2H, m); HRMS (CI+): calcd; 373.076210. Found; 373.076307.

4.7. Electrolysis of *E*,*E*-1,7-bis(3,4-dichlorobenzoyl)-1,6-heptadiene (1d)

Electrolysis of 102 mg (0.0105 M) of **1d** with LiClO₄ and Mg(ClO₄)₂ electrolyte (both 0.10 M), at -4.0 V (first 1.5 C at -3.0 V). The reaction appeared complete after 17.0 C, or

76.3% (of required charge) had passed through the cell. PTLC purification of the 87 mg of recovered crude yielded *cis*-2d (34 mg, 33%), *trans*-2d (7 mg, 7%), and 3d (16 mg, 16%), for a total yield of 57 mg of pericyclic products (56%). A further 6 mg (6%) of a product was isolated and identified as 6d.

Electrolysis of 100 mg (0.0103 M) of 1d with Et₄NBF₄ electrolyte, at -3.2 V for 3.0 C (first 0.75 C at -2.5 V, then 0.75 C at -3.0 V). This corresponds to 13.7% of required charge. PTLC purification of the 150 mg of crude yielded 3 mg (3%) of *trans*-2d, 17 mg (17%) of 5d, and 35 mg (35%) of an unidentified polymer.

4.7.1. Compound *cis*-**2d.**⁶ ¹H NMR (300 MHz, CDCl₃): 1.74 (2H, m), 1.85 (2H, dd, J = 11.7, 3.3 Hz), 2.05 (2H, m), 3.19 (2H, d, J = 2.4 Hz), 3.77 (2H, d, J = 3.6 Hz), 7.46 (2H, d, J = 8.4 Hz), 7.57 (2H, dd, J = 8.7, 2.1 Hz), 7.81 (2H, d, J = 2.1 Hz); HRMS (CI+): calcd; 440.998266. Found; 440.997859.

4.7.2. Compound *trans*-2d. ¹H NMR (300 MHz, CDCl₃): 1.38 (2H, m), 1.55 (1H, m), 1.83 (3H, m), 3.04 (1H, m), 3.24 (1H, m), 4.15 (1H, m), 4.47 (1H, dd, J=2.1, 10.2 Hz), 7.55 (1H, d, J=3.3 Hz), 7.57 (1H, d, J=3.0 Hz), 7.76 (1H, br.d), 7.81 (1H, br.d), 8.02 (1H, d, J=2.1 Hz) 8.08 (1H, d, J=1.8 Hz); HRMS (CI+): calcd; 440.998266. Found; 440.997697.

4.7.3. Compound 3d. ¹H NMR (300 MHz, CDCl₃): 1.60 (2H, m), 1.85 (2H, m), 2.07 (2H, m), 2.83 (2H, m), 5.09 (1H, d, J = 6.9 Hz), 5.73 (1H, d, J < 3 Hz), 7.39 (2H, m), 7.62 (3H, m), 7.74 (3H, m), 7.95 (4H, m), 8.15 (1H, m), 8.74 (1H, br.s); HRMS (CI+): calcd; 440.998266. Found; 440.997720.

4.7.4. Compound 5d. ¹H NMR (300 MHz, CDCl₃): 1.73 (4H, m), 2.31 (2H, br.m), 2.84 (1H, dd, J=9.6, 15.0 Hz), 3.66 (1H, dd, J=3.6, 15.0 Hz), 3.40 (1H, br.s), 6.66 (1H, t, J=3.3 Hz), 7.51 (2H, m), 7.56 (1H, d, J=8.4 Hz), 7.74 (1H, d, J=1.8 Hz), 7.90 (1H, dd, J=2.1, 8.4 Hz), 8.12 (1H, d, J=2.1 Hz); HRMS (CI+): calcd; 440.998266. Found; 440.998110.

4.7.5. Compound 6d. ¹H NMR (300 MHz, CDCl₃): 1.71 (3H, m), 1.98 (1H, m), 2.17 (1H, m), 2.34 (1H, m), 2.96 (3H, m), 3.17 (1H, br.d, J = 17.4 Hz), 3.65 (1H, d, J = 9.0 Hz), 5.24 (1H, d, J < 1.5 Hz), 7.31 (1H, m), 7.54 (2H, m), 7.76 (1H, br.d), 7.81 (1H, d, J = 2.1 Hz), 8.00 (1H, m); HRMS (CI+): calcd; 443.013916. Found; 443.012405.

4.8. Electrolysis of *E*,*E***-1**,7**-di-1**-naphthoyl-1,6-heptadiene (1e)

Electrolysis of 97 mg (0.0109 M) of **1e** with Bu_4NBF_4 electrolyte, at -1.8 V (first 3.0 C at -1.5 V). The reaction appeared complete after 4.5 C, or 19.4% (of required charge) had passed through the cell. PTLC purification of the 123 mg of recovered crude yielded *cis*-**2e** (14 mg, 14%), *trans*-**2e** (28 mg, 29%), and **3e** (8 mg, 8%), for a total yield of 50 mg of pericyclic products (51%). No starting material was recovered. A further 16 mg (17%) of **5e** was recovered.

Electrolysis of 104 mg (0.0117 M) of **1e**, with Et_4NBF_4 electrolyte and 25 mg of acetic acid, giving a 1.6:1 excess of acetic acid. Initially at -2.5 V for 15.0 C and then at -3.0 V for 32.0 C. PTLC purification of the crude yielded *trans-2e* (6.0 mg, 6%), **5e** (2.8 mg, 3%), **6e** (14.1 mg, 14%), and **7e** (39.2 mg, 35%).

Electrolysis of 98 mg (0.0110 M) of **1e** with Bu_4NBF_4 electrolyte, at -1.5 V. The reaction was stopped after 3.4 C, or 14.5% (of required charge, corrected to 18.4%) had passed through the cell. PTLC purification of the 122 mg of recovered crude yielded *cis*-**2e** (16.6 mg, 21%), *trans*-**2e** (13 mg, 17%), and **3e** (8 mg, 10%), for a total yield of 37.6 mg of pericyclic products (48%). 20.4 mg (21%) of starting material was recovered, along with 5 mg (6%) of **5e**. Reaction is 79% complete based upon recovered **1e**.

4.8.1. Compound *cis*-**2e.**⁶ ¹H NMR (300 MHz, CDCl₃) 1.74 (2H, m), 1.93 (2H, dd, J = 14.1, 5.4 Hz), 2.15 (2H, m), 3.29 (2H, d, J = 2.4 Hz), 4.08 (2H, d, J = 4.2 Hz), 7.45 (4H, m), 7.74 (4H, m); 7.81 (4H, m), 8.22 (2H, s); HRMS (CI+): calcd; 405.185455. Found 405.185760.

4.8.2. Compound *trans*-2e.⁶ ¹H NMR (300 MHz, CDCl₃): 1.45 (1H, m), 1.62 (1H, m), 1.85 (2H, m), 1.99 (2H, m), 3.16 (1H, m), 3.35 (1H, m), 4.50 (1H, m), 4.79 (1H, dd, J=2.4, 10.2 Hz), 7.58 (4H, m), 7.89 (4H, m), 8.03 (4H, br.m), 8.48 (1H, br.s), 8.60 (1H, br.s); HRMS (CI+): calcd; 405.185455. Found; 405.186095.

4.8.3. Compound 3e.⁶ ¹H NMR (300 MHz, CDCl₃): 1.60 (2H, m), 1.85 (2H, m), 2.07 (2H, m), 2.83 (2H, m), 5.09 (1H, d, J=6.9 Hz), 5.73 (1H, d, J<3 Hz), 7.39 (2H, m), 7.62 (3H, m), 7.74 (3H, m), 7.95 (4H, m), 8.15 (1H, m), 8.74 (1H, br.s); HRMS (CI+): calcd; 405.185455. Found; 405.186005.

4.8.4. Compound 5e.¹¹ ¹H NMR (300 MHz, CDCl₃): 1.71 (1H, m), 1.87 (3H, m), 2.35 (2H, br.m), 3.00 (1H, dd, J = 11.4, 15.6 Hz), 3.66 (1H, dd, J = 3.6, 15.6 Hz), 3.77 (1H, br.s), 6.75 (1H, br.t), 7.60 (4H, m), 7.92 (5H, m), 7.99 (1H, m), 8.05 (1H, m), 8.14 (1H, m), 8.22 (1H, s), 8.71 (1H, s).

4.8.5. Compound 6e.⁶ ¹H NMR (300 MHz, CDCl₃). Partial only: 1.60 (m), 1.91 (m), 3.20 (2H, m), 5.74 (1H, d, J = 1.8 Hz), 7.41 (2H, m), 7.59 (6H, br.m), 7.76 (2H, m), 7.87 (2H, m), 8.00 (2H, m).

4.8.6. Compound *trans*-7e. ¹H NMR (300 MHz, CDCl₃): 1.35 (1H, m), 1.67 (2H, m), 1.88 (1H, m), 2.04 (2H, m), 2.31 (1H, m), 2.78 (1H, m), 2.95 (1H, dd, J=8.1, 15.6 Hz), 3.10 (1H, dd, J=8.1, 16.2 Hz), 3.27 (1H, dd, J=5.7, 15.3 Hz), 3.38 (1H, dd, J=4.8, 16.2 Hz), 7.56 (4H, m), 7.88 (4H, m), 7.95 (2H, br.d, J=7.8 Hz), 8.02 (2H, m), 8.47 (2H, br.d, J= 6.0 Hz); HRMS (CI+): calcd; 407.201105. Found; 407.202010.

4.9. Electrolysis of *E*,*E*-1,7-bis(4-phenylbenzoyl)-1,6-heptadiene (1f)

Electrolysis of 76 mg (0.0076 M) of **1f** with Bu_4NBF_4 electrolyte (in a 1:1 THF/acetonitrile solution), at -3.5 V (first 1.8 C at -2.5 V, then 1.1 C at -3.0 V). The reaction

appeared complete after 11.7 C, or 69.6% (of required charge) had passed through the cell. PTLC purification of the 107 mg of recovered crude yielded *trans-2f* (9 mg, 12%). Also recovered was 13 mg (17%) of 5f.

Electrolysis of 55 mg (0.0055 M) of **1f** with Mg(ClO₄)₂ (0.1 M in 1:1 THF/acetonitrile) at -2.0 V for 5.0 C (corrected for recovered **1f** to 53.7% of required charge), PTLC purification of the 61 mg of crude yielded *cis*-**2f** (9 mg, 21%), *trans*-**2f** (3.1 mg, 7%), and **3f** (11 mg, 25%), for a total yield of 23.1 mg of pericyclic products (53%). 11 mg of unreacted **1f** was recovered, allowing the corrected yields given above. Two additional products were isolated; 5.8 mg (13%) of **6f**; and 3.2 mg (7%) of **7f**.

4.9.1. Compound *cis*-**2f.**⁶ ¹H NMR (300 MHz, CDCl₃): 1.84 (4H, m), 2.06 (2H, m), 3.27 (2H, m), 3.92 (2H, m), 7.37 (6H, m), 7.55 (8H, m), 7.84 (4H, d, *J*=8.7 Hz).

4.9.2. Compound *trans-***2f.** ¹H NMR (300 MHz, CDCl₃): 1.47 (2H, m), 1.66 (2H, m), 1.91 (2H, m), 3.13 (1H, m), 3.30 (1H, m), 4.33 (1H, m), 4.61 (1H, m), 7.45 (6H, m), 7.65 (2H, m), 7.70 (2H, d, *J*=8.4 Hz), 8.04 (2H, d, *J*=9.0 Hz), 8.11 (2H, d, *J*=8.7 Hz); HRMS (CI+): calcd; 457.216755. Found; 457.215380.

4.9.3. Compound 3f. ¹H NMR (300 MHz, CDCl₃): 1.56 (3H, m), 1.78 (1H, m), 2.05 (2H, m), 2.75 (2H, m), 4.93 (1H, m), 5.61 (1H, m), 7.40–7.72 (12H, m), 8.19 (2H, m): calcd; 457.216755. Found; 457.218026.

4.9.4. Compound 5f. ¹H NMR (300 MHz, CDCl₃): 1.67 (1H, m), 1.78 (3H, m), 2.31 (2H, br.m), 2.85 (1H, dd, J= 10.5, 14.7 Hz), 3.47 (1H, dd, J=6.6, 13.8 Hz), 3.54 (1H, br.s), 6.70 (1H, m), 7.43 (6H, m), 7.66 (6H, m), 7.79 (2H, d, J=8.4 Hz), 8.03 (2H, d, J=8.7 Hz), 8.16 (2H, d, J= 8.7 Hz); HRMS (CI+): calcd; 457.216755. Found; 457.216312.

4.9.5. Compound 6f.⁶ ¹H NMR (300 MHz, CDCl₃). Partial only: 3.94 (1H, d, *J*=8.7 Hz), 5.63 (1H, d, *J*~1.5 Hz).

4.9.6. Compound *cis*-**7f.** ¹H NMR (300 MHz, CDCl₃): 1.34 (2H, m), 1.67 (2H, m), 2.02 (2H, m), 2.24 (2H, m), 2.98 (2H, dd, *J*=7.2, 17.4 Hz), 3.26 (2H, dd, *J*=4.5, 16.2 Hz), 7.44 (6H, m), 7.65 (8H, m), 8.03 (4H, d, *J*=8.7 Hz); HRMS (CI+): calcd; 459.232406. Found; 459.231734.

4.10. Electrolysis of *E*,*E*-1-acetyl-7-benzoyl-1,6-heptadiene (1g)

Electrolysis of 98 mg (0.018 M) of **1g** with Mg(ClO₄)₂ electrolyte, at -4.5 V for 150 C (initial 4.0 C at -4.0 V, last 16 C at -5.0 V), corresponding to 459% of required charge. PTLC separation yielded *cis*-**2g** (7 mg, 9%), and **7g** (14 mg, 17%), along with 11 mg (13%) of an unidentified product, tentatively described as an isomer of **7g** (LRMS (CI+): 245, 227). 16 mg of unreacted **1g** was also recovered.

Electrolysis of 73 mg (0.014 M) of 1g with Et₄NBF₄ electrolyte, at -2.0 V for 16.0 C (initial 2.2 C at -1.6 V), corresponding to 55.0% of required charge.

PTLC separation yielded two isomers of *trans*-2g, 9 mg (12%) of the isomer where the benzoyl group is *syn* to the cyclopentane ring, and 5 mg (7%) with an *anti* benzoyl group.

4.10.1. Compound *cis*-**2g.**⁶ ¹H NMR (300 MHz, CDCl₃): 1.59 (2H, m), 1.68 (4H, br.m), 2.03 (3H, s), 3.00 (2H, m), 3.11 (1H, m), 3.79 (1H, dd, J=4.5, 9.9 Hz), 7.45 (2H, br.t, J=7.2 Hz), 7.52 (1H, br.t, J=7.2 Hz), 7.83 (2H, br.d, J= 8.1 Hz); HRMS (CI+): calcd; 243.138505. Found; 243.138743.

4.10.2. Compound *trans-2g* (benzoyl group *syn* to the cyclopentane ring). ¹H NMR (300 MHz, CDCl₃): 1.36 (2H, m), 1.74 (4H, m), 2.13 (3H, s), 2.93 (1H, m), 3.13 (1H, m), 3.49 (1H, t, J=8.1 Hz), 4.32 (1H, dd, J=2.1, 10.5 Hz), 7.47 (2H, br.t, J=7.2 Hz), 7.57 (1H, br.t, J=6.9 Hz), 7.93 (1H, br.d, J=7.2 Hz). X-ray crystallography confirms structure (CCDC240332).

4.10.3. Compound *trans*-2g.⁶ (benzoyl group *anti* to the cyclopentane ring) ¹H NMR (300 MHz, CDCl₃): 1.58 (2H, m), 1.80 (4H, m), 2.11 (3H, s), 2.94 (1H, m), 3.07 (1H, m), 3.38 (1H, m), 3.86 (1H, dd, *J*=2.4, 10.5 Hz), 3.99 (1H, t, *J*=6.3 Hz), 7.45 (2H, br.t, *J*=7.2 Hz), 7.55 (1H, br.t, *J*=7.2 Hz), 7.95 (1H, br.d, *J*=7.2 Hz).

4.10.4. Compound 7g. ¹H NMR (300 MHz, CDCl₃). 1.23 (2H, m), 1.61 (2H, m), 1.98 (4H, br.m), 2.13 (3H, s), 2.40 (1H, dd, *J*=7.8, 16.8 Hz), 2.67 (1H, dd, *J*=4.2, 16.8 Hz), 2.91 (1H, dd, *J*=7.8, 16.5 Hz), 3.14 (1H, dd, *J*=4.2, 16.8 Hz), 7.46 (2H, br.t, *J*=7.5 Hz), 7.56 (1H, br.t, *J*=7.2 Hz), 7.95 (1H, br.d, *J*=7.2 Hz); HRMS (CI+): calcd; 245.154155. Found; 245.154390.

4.11. Electrolysis of *E*,*E*-1,7-diacetyl-1,6-heptadiene (1h)

Electrolysis of 97 mg (0.025 M) of **1h** with Et_4NBF_4 electrolyte, at -2.5 V for 10.0 C (initial 4.0 C at -2.0 V). ¹H NMR (300 MHz, CDCl₃) of the 121 mg of recovered crude showed only starting material present (plus electrolyte).

4.12. Electrolysis of *E*,*E*-7-ethoxy-1-benzoyl-1,6-heptadiene (1i)

Electrolysis of 117 mg (0.020 M) of **1i** with LiClO₄ electrolyte (0.3 M), at -2.0 V for 10.0 C, -2.2 V for 20.0 C, and -3.0 V for 10.0 C (136% of required charge). PTLC separation yielded 23 mg (28%) of one isomer of *trans*-**2i**. Thought to be the same isomer as the major isomer seen in the **1g** electrolysis (based upon NMR comparison). A further 3 mg (4%) of the alternate *trans*-**2i** isomer was also obtained. 34 mg of starting material **1i** was recovered.

Electrolysis of 136 mg (0.023 M) of **1i** with Et_4NBF_4 electrolyte, at -2.0 V for 10.5 C. Yielding only unidentified polymers

4.12.1. Compound *trans-2i* (benzoyl group *syn* to the cyclopentane ring). ¹H NMR (300 MHz, CDCl₃): 0.98 (3H, t, *J*=7.2 Hz), 1.63 (2H, m), 1.72 (2H, m), 1.91 (2H, m), 3.04 (2H, m), 3.22 (1H, m), 3.69 (1H, dd, *J*=5.1, 9.9 Hz), 3.87

(2H, q, *J*=7.2 Hz), 7.43 (2H, br.t, *J*=8.1 Hz), 7.53 (1H, br.t, *J*=6.9 Hz), 7.84 (1H, br.d, *J*=7.2 Hz). HRMS (CI+): calcd; 273.149070. Found; 273.148638.

4.12.2. Compound *trans-2i* (benzoyl group *anti* to the cyclopentane ring). ¹H NMR (300 MHz, CDCl₃): 0.98 (3H, t, *J*=7.2 Hz), 1.69 (4H, m), 2.01 (2H, m), 2.99 (1H, m), 3.21 (1H, m), 3.38 (1H, m), 4.14 (2H, q, *J*=7.2 Hz), 4.34 (1H, dd, *J*=2.7, 10.5 Hz), 7.47 (2H, br.t, *J*=6.3 Hz), 7.56 (1H, br.t, *J*=6.9 Hz), 7.93 (1H, br.d, *J*=6.6 Hz). HRMS (CI+): calcd; 273.149070. Found; 273.148198.

4.13. Electrolysis of *E*,*E*-1,7-dicarbethoxy-1,6-heptadiene (1j)

Electrolysis of 112 mg (0.021 M) of 1j with Et₄NBF₄ electrolyte, at -2.5 V for 10.0 C, corresponding to 22.2% of required charge. PTLC purification of the 112 mg of recovered crude yielded 47 mg (42%) of 5j. No starting material was recovered.

Electrolysis of 96 mg (0.018 M) of 1j with LiClO₄ electrolyte, at -3.5 V for 4.5 C, -3.5 V for 7.9 C, and -4.0 V for 55 C. ¹H NMR (300 MHz, CDCl₃) of the recovered crude showed only starting material present.

4.13.1. Compound 5j. ¹H NMR (500 MHz, CDCl₃): 1.26 (3H, t, J=7.5 Hz), 1.29 (3H, t, J=7.0 Hz), 1.64 (4H, m), 2.18 (2H, br.m), 2.27 (1H, dd, J=10.5, 15.0 Hz), 2.64 (1H, dd, J=3.5, 15.0 Hz), 3.09 (1H, br.s), 4.16 (4H, m), 7.03 (1H, t, J=4.0 Hz). ¹³C NMR (500 MHz in CDCl₃): 14.21 (2C), 17.11, 25.77, 26.52, 29.92, 38.40, 60.20, 60.26, 132.79, 140.99, 166.86, 172.53; NMR COSY (500 MHz) (consistent with structure). HRMS (CI+): calcd; 241.143984. Found; 241.142945.

4.14. X-ray data

Crystallographic data (excluding structure factors) for **4b** and the primary *trans*-**2g** isomer, have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication numbers CCDC 240331 and CCDC 240332, respectively. Copies of the data can be obtained, free of charge, via www.ccdc.cam.ac.uk/data_request/cif or on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (fax: +44(0)-1223-336033 or e-mail: deposit@ccdc.cam.ac.uk).

Acknowledgements

The authors acknowledge the assistance of Michael J. Krische, Jingkui Yang, and thank the Robert A. Welch Foundation (F-149) for support of this research.

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Tetrahedron

Tetrahedron 60 (2004) 11011-11027

Nitroalkylation and nitroalkenylation reactions of γ -lactone enolates. A facile ring switch from polysubstituted γ -lactones to polysubstituted γ -lactams

C. Forzato,^a P. Nitti,^{a,*} G. Pitacco,^a E. Valentin,^{a,*} S. Morganti,^b E. Rizzato,^b D. Spinelli,^b C. Dell'Erba,^c G. Petrillo^c and C. Tavani^c

^aDipartimento di Scienze Chimiche, Università di Trieste, via L. Giorgieri 1, I-34127 Trieste, Italy

^bDipartimento di Chimica Organica 'A. Mangini', Università di Bologna, via San Donato 15, I-40127 Bologna, Italy ^cDipartimento di Chimica e Chimica Industriale, Università di Genova, Via Dodecaneso 31, I-16146 Genova, Italy

Received 11 June 2004; revised 5 July 2004; accepted 28 July 2004

Available online 7 October 2004

Abstract—Michael addition of lithium enolates of γ -butyrolactone 1 and α -methyl- γ -butyrolactone 1' to (E)-1-nitropropene 2, (E)- β nitrostyrene 3 and (E)-2-nitro-1-phenylpropene 4 is described. Reactions of the lithium enolate of 1' with 2 and 4 occurred with high diasteroselectivity (80 and 92% d.e., respectively). Reactions of the zinc enolate of 1' with two β -nitroenamines and two methylthiosubstituted 1-amino-2-nitro-1,3-dienes were also examined. Catalytic reduction of the nitroalkylated and nitroalkenylated products allowed the achievement of functionalized γ -lactams and/or cyclic hydroxamic acids.

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1. Introduction

Polyfunctionalized γ -lactones are frequently encountered in the literature,¹ owing to the fact that many natural products, in particular those belonging to the sesquiterpene series, contain the γ -lactone ring in their structure. Many examples of α -functionalization and α, α -difunctionalization of γ lactone rings are present in the literature³ and among them a new class of anticonvulsant drugs can be mentioned.⁴ α -Nitroalkenylation reactions were extensively studied,⁵ with the aim of preparing compounds possessing quaternary stereocentres,⁶ whereas no examples of nitroalkylation reactions of γ -lactones can be found. On the contrary, Enders and co-workers⁷ obtained excellent results in the diastereo- and enantio-selective Michael additions of enolates of γ -lactams to aliphatic and aromatic nitroolefins. Seebach and coworkers⁸ investigated the nitroalkylation reactions of lithium enolates of other five-membered ring heterocycles, such as chiral non-racemic 2-t-butyl-1,3dioxolan-4-ones, 2-t-butylimidazolidin-4-ones and 2-tbutyloxazolidin-5-ones, to verify the 1,3 asymmetric induction on the reaction products. In all cases examined the diastereoselectivity was high, depending however on

whether the position α to the carbonyl group was substituted or not.

In this paper we describe the nitroalkylation reactions of the lithium enolates of γ -butyrolactone 1 and α -methyl- γ butyrolactone $\mathbf{1}'$ with a few conjugated nitroolefins, such as (E)-1-nitropropene 2, (E)- β -nitrostyrene 3 and (E)-2-nitro-1-phenylpropene 4. Nitroalkenylation reactions have also been carried out, by reacting the zinc enolate of compound 1' with the β -nitroenamines **5**⁹ and **6**¹⁰ as well as with the nitrodienes **7**, ^{11,12} and **7**'¹² (Fig. 1).

2. Results and discussion

2.1. Nitroalkylation reactions

The butanolides 1 and 1' were enolized with lithium diisopropylamide in THF at -78 °C to the corresponding lithium enolates 8 and 8', which reacted with the nitroolefins 2, 3, and 4, to provide the corresponding nitronate salts 9–14 (Scheme 1).

No attempt to isolate the lithium nitronate intermediates 9-14 was undertaken. Treatment of the crude reaction mixtures with a weak acid afforded the corresponding nitroalkylated lactones 15-20. The nitroalkylated lactones 15, 16, 17 and 18, for which $R^2 = H$, were mixtures of syn/

Keywords: Substituted nitroalkenes; Substituted nitroalkadienes; y-Lactams; Cyclic hydroxamic acids.

^{*} Corresponding authors. Tel.: +39-040-5583917; fax: +39-040-5583903; e-mail: valentin@dsch.univ.trieste.it

^{0040-4020/\$ -} see front matter © 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.07.084



Figure 1.

anti diastereomers, **a** and **b** respectively, owing to the presence of two adjacent chiral centres, while **19** was a mixture of four diastereomers and **20** was a mixture of three diastereomers out of the four possible ones. In order to evaluate the diastereoselectivity of the reaction, the *syn* and *anti* descriptors were also used for **19** and **20**, to indicate the relative configurations of the two stereocentres of the newly formed C–C bond.

Since for the nitroalkylated γ -lactones 15–20 the syn/anti stereochemical assignments were not straightforward, a stereochemical correlation was desirable: therefore, such lactones were converted into the corresponding trans and cis lactam derivatives 21-26 to take advantage of the more rigid polysubstituted lactams to determine the configurations of the stereocentres. On the other hand, γ -lactams themselves are attractive targets because they possess a variety of biological activities¹³ and have been used to produce γ -aminobutyric acid (GABA) analogues by hydrolysis.¹⁴ Thus, reduction of the nitro group of compounds 15–20 with hydrogen on Raney Ni and subsequent cyclization¹⁵ of the corresponding aminoalkyl lactone intermediates, which were not isolated, afforded the corresponding lactam derivatives 21–26 (Scheme 1). In this manner, the stereochemical assignments were made on the lactam derivatives 21–26 either by a comparison of their ¹³C NMR spectra or by means of NOE measurements. When this latter method was unsatisfactory, the nitroalkylated γ -lactones 15–19 were transformed into the corresponding cyclic hydroxamic

acids **27–31**,¹⁶ using 10% Pd/C as the catalyst. The latter compounds proved better substrates for NOE measurements, which supported the stereochemical assignments previously made on the lactam derivatives. All resonances of each compound were identified by means of 2D correlated experiments.

2.2. Reactions of enolates 8 and 8' with (E)-1-nitropropene 2

The reaction of lithium enolate 8 with (E)-1-nitropropene 2 led, after acidification with aqueous satd. NH₄Cl, to the formation of two diastereomeric (nitroalkyl)lactones 15a and 15b in 2:3 ratio, which were separated by flash chromatography (Scheme 2). They were assigned the syn and anti configuration, respectively, after transformation into the corresponding lactams 21a and 21b whose geometries were demonstrated to be trans and cis, respectively. In fact the ¹³C NMR spectrum of **21b** showed an upfield shift for the methyl group with respect to the same resonance in **21a**, thus demonstrating its *cis* relationship with the hydroxyethyl chain (Table 1). The same *trans/cis* assignment was made for the corresponding cyclic hydroxamic acids 27a and 27b that were obtained in admixture with the corresponding lactams 21a and 21b when the reduction was performed using Pd on carbon as the catalyst.

Similarly, the reaction between the enolate 8' and 2 afforded the corresponding nitroalkylated products **16a** and **16b** in





Scheme 2.

1:9 ratio, as determined by HRGC analysis of the crude reaction mixture. However, since in this case the two diastereomers were not separable by flash chromatography, the subsequent reductions of the nitro group were performed on the mixture. When Raney Ni was used as the catalyst, a 1:9 mixture of the corresponding lactams **22a** and **22b** was isolated. They were assigned the $(3R^*, 4R^*)$ and $(3R^*, 4S^*)$ configurations, respectively, by comparison of their ¹³C NMR spectra (Table 1): an upfield shift was always observed for those carbon atoms which suffered from steric effects.¹⁷ Thus, the methyl group at C-3 resonated at 15.1 ppm in **22a** and at 20.5 ppm in **22b**, as a consequence of a less constrained situation in the latter diastereomer.

When the reduction of 16a,b was performed using Pd on carbon, two cyclic hydroxamic acids, namely 28a and 28b, were obtained in 1:9 ratio (Scheme 2). The relevant configurations were assigned on the basis of NOE measurements performed on the major product 28b, which was separated in pure form by fractional crystallization. Irradiation of its methyl singlet at 1.18 ppm enhanced the signal of H-4 (5%), while irradiation of the α -methylene hydrogens of the hydroxyethyl chain at 1.74 and 1.50 ppm produced enhancement of the methyl doublet at 1.04 ppm (2%). The NOE enhancement values were low, however this assignment was supported by a comparison of the ¹³C NMR spectra of the two isomers, whose significant digits are reported in Table 1. Again an upfield shift was observed for those carbon atoms which suffered from steric effects. Thus the methyl group at C-3 is shifted to higher field in 28a, being cis to the methyl group at C-4. The same trend was observed for the α -methylene hydrogens of the hydroxyethyl chain in 28b. As a consequence, the configuration of the chiral centres in compounds 22a and 28a is $(3R^*, 4R^* \text{ or }$ like¹⁸) and that of **22b** and **28b** is $(3R^*, 4S^* \text{ or unlike}^{18})$. The same configurations can be assigned to the corresponding parent lactones 16a and 16b, whose relative configurations are therefore *syn* and *anti* respectively.

2.3. Reactions of enolates 8 and 8' with (E)- β -nitrostyrene 3

The reactions between the nitroalkene 3 and the enolates 8

			γ -Lactams						Cycl	ic hydroxamic ac	ids		
Entry	Me at C-3	Me at C-4	CH ₂ CH ₂ OH	C-3	C-4	C-5	Entry	Me at C-3	Me at C-4	CH ₂ CH ₂ OH	C-3	C-4	C-5
21a		17.3	32.4	49.7	37.1	48.7	27a		17.6	33.0	47.8	31.5	55.2
21b	I	14.6	28.5	45.5	33.5	48.6	27b		14.9	28.8	43.4	28.4	55.5
22a	15.1	11.4	38.2	45.9	39.7	47.0	28a	16.0	11.5	38.5	44.7	33.8	53.6
22b	20.5	12.6	34.0	44.6	42.0	46.6	28b	21.3	12.0	35.0	43.6	37.3	53.6
23a			32.4	49.0	48.4	48.9	29a			34.1	46.4	43.5	56.3
23b			29.9	45.3	44.3	48.0	29b			30.5	42.6	40.0	55.8
24a	17.4		38.5	47.5	51.3	44.5	30a	19.5		40.1	47.1	45.0	52.5
24b	21.4		35.7	46.5	53.6	44.4	30b	23.3		37.0	46.5	45.0	52.4



Scheme 3.

and 8' were poorly diastereoselective, as the former reaction afforded a ca. 1:1 mixture of diastereomers 17a and 17b (Scheme 3), and the latter reaction gave a ca. 2:1 mixture of 18a and 18b (Scheme 4). In both cases the diastereomers could not be completely separated: anyway, for an easier stereochemical assignment, different mixtures of 17a,b or 18a,b, enriched in either diastereomer, were converted into the corresponding lactams (23a,b or 24a,b) or cyclic hydroxamic acids (29a,b or 30a,b).

The *trans* and *cis* stereochemical assignments for the γ -lactams **23a** and **23b** and for the cyclic hydroxamic acids **29a** and **29b** were based on ¹³C NMR spectra (Table 1). The carbon resonances for the *cis* diastereomers **23b** and **29b** were shifted upfield with respect to those for the *trans* diastereomers **23a** and **29a**. Furthermore, NOE measurements carried out on **29a** and **29b** confirmed the previous assignments. Irradiation of the H-3 signal at 2.71 ppm in **29a** produced enhancement (4%) of the *ortho* hydrogens of the phenyl ring, while irradiation of the H-3 signal at 2.95 ppm in **29b** enhanced the H-4 signal at 3.64 ppm (5%). Therefore, since the configuration of **23a** and **29a** is $(3R^*, 4S^*)$ and that of **23b** and **29b** is $(3R^*, 4R^*)$, the relative configurations of lactones **17a** and **17b**, from which they are derived, are *syn* and *anti*, respectively.

In a similar manner, the NOE experiments performed on the diastereomeric cyclic hydroxamic acids **30a** and **30b**, allowed the correct *syn* and *anti* attributions to the parent lactones **18a** and **18b** to be made. Thus, the configuration of the cyclic hydroxamic acid **30a**, derived from **18a**, was proved to be $(3R^*, 4R^*)$ and that of **30b**, derived from **18b**, was $(3R^*, 4S^*)$. In fact, upon irradiation of the methyl group at 1.30 ppm in **30b**, the H-4 signal was enhanced (10%). On the other hand, irradiation of the M-4 signal at 3.63 ppm in **30a** caused enhancement of the α -methylene protons of the chain (5%), while irradiation of the methyl group at 0.74 ppm enhanced the aromatic *o*-hydrogens (6%). All these assignments were also supported by a comparison of the ¹³C NMR spectra of each pair of diastereomers, as shown in Table 1.

It should be noted that in spite of the fact that the geometries of the products are the same as those observed for the products of the reaction between 8 or 8' with 1-nitropropene, the descriptors are different, owing to a different priority of the groups.

2.4. Reactions of enolates 8 and 8' with (E)-2-nitro-1-phenylpropene 4

In the reaction between the enolate **8** and (E)-2-nitro-1phenylpropene **4** four diastereomers, namely **19a**, **19b**, **19c**, and **19d** were formed, in 53:22:14:11 relative ratio (Scheme 5), as determined by HRGC analysis of the crude reaction mixture. Their separation on chromatographic column allowed the isolation of all stereoisomers as pure compounds with the exception of **19c** which was isolated in admixture with **19a**. Treatment under basic conditions (DBU in chloroform) of the pure diastereomer **19a** afforded a 1:1 mixture of **19a** and **19b**, thus demonstrating that they differed in the configuration of the nitromethine carbon atom. The same was proved for compounds **19c** and **19d**.

As above, the stereochemical assignments for **19a** and **19b** were made on the corresponding lactam **25a** and on the cyclic hydroxamic acid **31b**, respectively. Reduction of **19a** with hydrogen on Raney Ni afforded lactam **25a** as a single





Scheme 5.

diastereomer, while reduction of 19b using 10% Pd/C as the catalyst afforded a mixture of lactam 25b and cyclic hydroxamic acid **31b** in the 1:3 ratio. The stereochemistry of 25a was established by means of NOE measurements. In fact, irradiation of the H-4 signal enhanced the signals of H-3 (4%) and H-5 (5%). Compound **31b** was less soluble than the corresponding lactam 25b in ethyl acetate and therefore it could be separated from the mixture in pure state. The NOE experiment carried out on 31b revealed that the hydroxyalkyl chain was cis to the phenyl group which in turn was trans to the methyl group. In fact, irradiation of the methyl group enhanced the signal of H-4 (6%) as well as that of H-3 (3%), while irradiation of H-4 enhanced the signals of H-3 (8%) and that of the methyl group (6%). Its configuration was therefore $(3R^*, 4R^*, 5R^*)$ and, accordingly, the relative configuration of 19b was assigned as anti, anti. Since the stereochemical relationship between 19a and 19b was known from the previous equilibration reaction, the configuration of 19a must be anti, syn.

The reaction between the enolate 8' and the nitroolefin 4 was somewhat more complicated by the fact that acidification of the nitronate salt intermediate 14 (Scheme 1) required strictly controlled conditions and the use of a weak acid (see Section 4) in order to obtain the nitroalkylated lactones 20. When glacial acetic acid in THF was used, a 61:35:4 mixture of 20a, 20b and 20c was obtained (Scheme 6), which were isolated by flash chromatography. Equilibration of **20a**, carried out under basic conditions with DBU at room temperature, partially converted it into **20b**, thus demonstrating that **20a** and **20b** differed in the configuration of the nitromethine carbon atom.

Reduction of a 1:1 mixture of lactones 20a and 20b using Raney Ni as catalyst furnished a 1:1 mixture of lactams 26a and 26b. The stereochemistry of lactam 26a was assigned by means of NOE measurements: irradiation of the α -methylene hydrogens of the hydroxyethyl chain at 1.90 ppm produced enhancement of the signal of H-4 (5%) as well as that of H-5 (8%) indicating that the hydroxyalkyl chain was *trans* to the phenyl group, which in turn was *cis* to the methyl group at C-5. Its configuration was therefore $(3R^*, 4R^*, 5R^*)$ and that of **26b** was $(3R^*, 4R^*, 5S^*)$. Accordingly, the relative configuration of 20a was assigned as syn, syn and that of 20b as syn, anti, both deriving from the same type of attack of the enolate onto the nitroolefin. The diastereoselective excess of the reaction, with reference to the syn configuration around the newly formed C-C bond, was 92%.

2.5. Products of the Nef reaction

The nitronate salt intermediates 13 (R=H) and 14 (R=Me) (Scheme 1) were treated with 3 N HCl,¹⁹ with the aim of obtaining the corresponding Nef products.²⁰ Thus, a 75:25 diastereomeric mixture of 13 furnished 32a (isolated in19%)



yield) and **32b** in a ca. 3:2 ratio, in admixture with the oxime **34** (isolated in 15% yield), the latter most likely resulting from the autoxidation-reduction of the not detected nitronic acid intermediate 36^{21} Acidic equilibration of the 3:2 mixture of 32a and 32b changed its composition to 3:1. The thermodynamically more stable 32a was tentatively assigned the $(3R^*, 1'R^*)$ configuration, by a comparison of the values of the ${}^{3}J$ coupling constants between H-3 and the benzylic proton in the two isomers: 7.7 and 5.1 Hz for 32a and **32b**, respectively.²² In the ¹³C NMR spectra, the only significant difference between the two diastereomers was the resonance value of C-3, 42.8 ppm for 32b and 41.3 ppm for 32a, suggesting a slightly more crowded situation for the latter compound.

From the nitronic salt intermediates 14 (96:4 diastereomeric ratio) a single Nef product 33 (isolated in 17% yield) and a single oxime **35** (isolated in 10% yield) were obtained by the same acid treatment as above.

Remarkably, when acidification of the crude reaction mixture was performed with a saturated solution of ammonium chloride, followed by separation of the organic phase, the subsequent treatment of the mother liquors with 3 N HCl afforded the nitronic acid 37 (isolated in 10%) yield). This latter compound was separated as a white solid, stable at -20 °C. It must be underlined that reduction of the nitronic acid 37 with Raney Ni afforded the same lactams 26a and 26b, in the ratio of 7:3, as previously obtained from 20a and 20b, thus demonstrating their stereochemistry. The relative configuration reported in Figure 2 for the products of the Nef reaction are correlated with the diastereomeric values of the nitronate lithium salt formation.

2.6. Mechanism of the nitroalkylation reactions

The results relating to the geometry of the products are summarized in Table 2 (the syn and anti descriptors are also used for 19 and 20, to indicate the relative configurations of the two stereocentres of the newly formed C–C bond). The diastereoselectivity observed is generally low except for

Table 2. Diastereomeric ratios and yields for the nitroalkylated lactones 15 - 20

Entry	Product	syn/anti	Yield (%)
1	15	40/60	70
2	16	10/90	60
3	17	45/55	62
4	18	65/35	80
5	19	25/75	64
6	20	96/4	45



32a.b

compounds 16, for which the *anti* diastereomer largely prevailed, and for compound 20, for which the syn diastereomer was formed almost exclusively.

Formation of the syn and anti diastereomers would involve a different topological approach of the donor and acceptor.²³ Thus the Re^*, Re^* (like) approach of the enolate to the nitroolefin would lead to the syn products, while the Re*, Si* (unlike) one would give the anti products. It appears that the unlike approach was slightly preferred over the like one when the lactone enolate was unsubstituted (R = H) (entries 1, 3, 5). When R was methyl (entries 2, 4, 6) the reactions showed the opposite diastereoselection, with the exception of the reaction of 1-nitropropene. The following simplified model transition states A and B (Fig. 3), in which the pyramidalization of the reacting carbon atoms is ignored as are the role of the solvent and the state of aggregation of the lithium enolates,²⁴ would account for the different selectivity observed. When R and R^2 are hydrogens no remarkable differences in the transition states exist, and the resulting products syn and anti 15 and syn and anti 17 are formed in almost equal amount. Substitution of a hydrogen for a methyl group ($R^2 = Me$) slightly disfavours the like approach of the reactants and in fact compound anti-19 was formed with 50% d.e. When the lactone bears a methyl group at the α -position, the steric situation seems to be dominated by the presence of the phenyl group $(R^1 = Ph)$, which disfavours the unlike approach and consequently favours the formation of the syn products (30% d.e. for 18 and 92% d.e. for 20).

The prevalent formation of compound 16 in anti configuration from 1-nitropropene is not in accordance with the results found for the Michael addition of cyclohexanone lithium enolate to the same nitroolefin²⁵ which always afforded the syn-product under several reaction conditions. Probably this is due to the fact that in our case the presence of the heterocyclic oxygen atom allows the unlike or endo orientation of the nitroolefin.

2.7. Nitroolefination and nitrodienylation reactions

The reactivity of lithium enolates of γ - and δ -lactones bearing no substituent at α -position with a few β nitroenamines has already been reported²⁶ to lead to the corresponding Nef products²⁰ and not to the desired nitroolefinated lactones. On the contrary, when the lactone bears a substituent at the α -position, nitroolefination of the lithium enolate by β -nitroenamines proceeds diastereo- and enantio-selectively, as proved by Severin and coworkers.^{5a,27} An exchange of the counter ion from lithium to zinc had the





Figure 3.

effect of increasing the reactivity and the enantioselectivity of the reactions when a chiral non-racemic β -nitroenamine derived from (*S*)-prolinol was used.²⁸ In particular, when the zinc enolate²⁹ of lactone **1**['] reacted with (*E*)-1-(1-morpholinyl)-2-nitroethene and (*E*)-1-(1-morpholinyl)-2-nitropropene^{5c} the corresponding addition–elimination products in (*E*) configuration were obtained.

Herein an analogous reaction has been carried out on the lithium enolate **8**' with the nitroenamines (*E*)-1-(1-morpho-linyl)-2-nitro-2-phenylethene **5** and (*E*)-1-(1-pyrrolidinyl)-2-nitro-2-[2-(methylthio)phenyl]ethene **6** (Scheme 7). The corresponding products, **38** and **39**, were obtained as 85:15 and 65:35 *E/Z* mixtures, respectively, as determined by ¹H NMR analysis of the crude reaction mixtures. In fact, when the vinyl proton was *cis* to the nitro group, it resonated at lower field (7.75 ppm for **38** and 7.81 ppm for **39**) than in the case it was *trans* to it (6.43 ppm for **38** and 7.73 ppm for **39**). ^{12,30}

The E/Z mixtures **38** and **39** were reduced under different conditions. When Pd on carbon was used as the catalyst, lactones **38** afforded a 4:1 mixture of *cis* and *trans* diastereomers of lactam derivatives **40a** and **40b**. Differently from the cases previously reported for the nitroalkylated γ -lactones, no traces of the corresponding cyclic hydroxamic acid derivatives were detected. The geometry of the lactams **40a** and **40b** was established by NOE measurements. Irradiation of the methyl group signal at 1.33 ppm in 40a caused enhancement of the H-5 signal at 4.75 ppm (6%). On the contrary, under the same conditions, the nitroalkenylated lactones 39 furnished the corresponding oxime 41. Using Raney Ni as the catalyst, both mixtures 38 and 39 afforded the same diastereomeric mixture of lactams 40a and 40b, as a result of the concomitant hydrogenolysis of the methylthio group in 39. Reduction with sodium borohydride was not satisfactory, even using the reagent supported on Amberlyst[®] A 26, which is known to reduce regioselectively the carbon–carbon double bond of the nitrovinyl moiety to the corresponding nitroalkane.³¹

The same conditions as above were used for the nitroalkenvlation reaction of the enolate 8' with the dienes 7 and 7' to afford the corresponding addition–elimination products 42 and 43, isolated from the respective reaction mixtures in 52 and 70% yield, the former as the (*E*,*Z*) diastereomer, the latter as the (*E*) isomer (Scheme 8).

Interestingly, in deuteriated chloroform, compound 42 slowly equilibrated into a 4:1 mixture of (E,Z) and (Z,Z)-diastereomers. This equilibration did not occur in the parent nitrodienamine 7'. In the major isomer the H-1 vinyl proton resonated at 7.24 ppm, while in the minor isomer it absorbed at 6.29 ppm, values which are consistent with the *cis* and *trans* relationship of the same proton with the nitro group. DIFNOE measurements supported the Z geometry for the





Scheme 8.

C(3)–C(4) double bond, as it was originally in the reagent. On the contrary, an analogous equilibration between the (E) and (Z) forms was not observed for compound **43**, under the same conditions.

Reductions with hydrogen and metal, as a catalyst, were unsatisfactory as furnished complex mixtures of products not identified as yet. Treatment of compound **43** with polymer-supported borohydride resulted in the reduction of the sole C(1)–C(2) double bond,³¹ affording a 3:2 **44a**,**b** diastereomeric mixture which was assigned the structure indicated in Scheme 8. These compounds however were not stable in CDCl₃ solution and were converted into the fully conjugated system **45** in (*E*) configuration. In this manner a conjugated ketene *S*,*S*-acetal was obtained whose reactivity as a precursor of an acyl anion³² will be further investigated.

3. Conclusions

Differently from the cases of lithium enolates and enamines from cycloalkanones, for which the Michael addition of nitroolefins proceeded with high diastereoselectivity,^{24,33} in general the lactone lithium enolates 1 and 1' showed low to moderate diastereoselectivity, with the exception of the case in which both reactants were substituted by bulky groups. The stereochemical assignments were made either on the lactam derivatives or on the cyclic hydroxamic acids formed by reduction of the nitro group under different reaction conditions. The reduction with Raney Ni however needed milder conditions than those reported in the literature for other heterocyclic nitroalkylated compounds.^{8b,15}

The nitroolefination reaction proceeded smoothly and quantitatively on the zinc enolates of $\mathbf{1}'$ and afforded new α,β -unsaturated nitroderivatives in *E* and *Z* configurations, the percentage of the latter increasing with the size of the substituent on the carbon bearing the nitro group.

4. Experimental

4.1. General

FT-IR spectrometer. ¹H NMR and ¹³C NMR spectra were run on a Jeol EX-400 spectrometer (400 MHz for proton, 100.5 MHz for carbon) and on a Jeol EX-270 (270 MHz for proton, 67.9 MHz for carbon), using deuteriochloroform as a solvent and tetramethylsilane as the internal standard. Coupling constants are given in Hz. Mass spectra were recorded on a VG 7070 (70 eV) spectrometer and on an ion trap instrument Finnigan GCQ (70 eV). GLC analyses were run on a Carlo Erba GC 8000 instrument, the capillary columns being OV 1701 (25 m×0.32 mm) (carrier gas He, 40 KPa, split 1:50). TLC's were performed on Polygram[®] Sil G/UV₂₅₄ silica gel pre-coated plastic sheets (eluant: light petroleum-ethyl acetate). CHN analyses were run on a Carlo Erba 1106 Elemental Analyser. Flash chromatography was run on silica gel 230-400 mesh ASTM (Kieselgel 60, Merck). Light petroleum refers to the fraction with bp 40-70 °C and ether to diethyl ether. y-Butyrolactone 1, α -methyl- γ -butyrolactone 1' and (E)-2-nitrostyrene **3** were purchased from Aldrich. (E)-1-Nitropropene **2**,³⁴ (*E*)-2-nitro-1-phenylpropene **4**,³⁵ (*E*)-1-(2-methylthio-phenyl)-1-nitro-2-pyrrolidinoethene **6**,¹⁰ (1*E*,3*Z*)-4methylthio-2-nitro-1-pyrrolidino-1,3-butadiene 7¹¹ and 4,4bis(methylthio)-2-nitro-1-pyrrolidino-1,3-butadiene 7^{12} were prepared according to the literature.

4.1.1. (*E*)-2-Morpholinyl-1-nitro-1-phenylethene 5.⁹ The synthesis was accomplished in accordance with a literature procedure,³⁶ using phenylnitromethane³⁷ as the nitroaliphatic component. Thus phenylnitromethane (0.09 mol, 12.3 ml), triethyl orthoformate (0.1 mol, 17 ml), morpholine (0.09 mol, 7.8 ml) and *p*-toluenesulfonic acid (0.09 g, 0.5 mmol) were heated under reflux for 1 h. Then the solvent was evaporated, the residue diluted with dichloromethane and purified through an alumina column (30 \times 2.5 cm) using dichloromethane as eluting agent. Although the crude reaction mixture contained both diastereomers, the nitroenamine 5 was isolated as pure E-isomer and crystallized from ethanol. 21% Yield, mp 150–152 °C, [lit. for a 1:1 mixture of E- and Z-isomers, mp 125 °C]; IR (cm⁻ nujol): 3056 (=CH), 1626, 1592, 1573, 785, 772, 725, 696 (Ph), 1488, 1377 (NO₂); ¹H NMR (δ , ppm): 8.41 (1H, s, H-C=C), 7.42 (3H, m, Ph), 7.26 (2H, m, Ph) 3.59 (4H, s, CH₂–O), 3.14 (4H, s, CH₂–N).

4.2. General procedure for the Michael addition of nitroalkenes to lactone enolates

To a solution of lithium diisopropylamide (3.5 mmol, 2.3 ml of a 1.5 M solution in THF) in THF (16 ml), a solution of the γ -lactone (2.9 mmol) in 2.5 ml of THF was slowly added, at -78 °C. The mixture was stirred at -78 °C for 1 h and the appropriate nitroalkene (3.5 mmol) dissolved in 2.5 ml of THF was added dropwise. The mixture was stirred at -78 °C for 2 h, the temperature was allowed to raise to -40 °C and the reaction mixture was quenched by addition of a saturated aqueous solution of NH₄Cl. The aqueous phase was extracted three times with CH₂Cl₂. The combined organic phases were dried over anhydrous Na₂SO₄. After removal of the solvent, the residue was analysed by HRGC and purified by flash chromatography (light petroleum/ethyl acetate).

4.2.1. Reaction of the lactone 1 with (*E*)-1-nitropropene **2.** syn- and anti-4,5-Dihydro-3-(1-methyl-2-nitroethyl)-**2(3H)-furanone 15a and 15b.** The isomers **15a** and **15b** (70% yield) were only partially separable by column chromatography, yellow oil, IR (cm⁻¹, neat): 1760 (OC=O), 1550, 1380 (NO₂); MS (*m*/*z*): 126 (7), 86 (56), 83 (10), 82 (16), 81 (10), 69 (14), 68 (16), 67 (34), 55 (100). Anal. Calcd for $C_7H_{11}NO_4$: C, 48.55; H, 6.40; N, 8.09. Found: C, 48.4; H, 6.20; N, 7.89.

Compound **15a.** ¹H NMR (δ , ppm): 4.73 (1H, dd, J_1 = 6.6 Hz, J_2 =12.8 Hz, CHNO₂), 4.48 (1H, dd, J_1 =7.1 Hz, J_2 =12.8 Hz, CHNO₂), 4.39 (1H, dt, J_1 =2.4 Hz, J_2 = J_3 = 9.0 Hz, H-5), 4.24 (1H, dt, J_1 =6.9 Hz, J_2 = J_3 =9.0 Hz, H-5), 2.74 (2H, m, H-3 and CHCH₃), 2.42 (1H, m, H-4), 2.14 (1H, m, H-4), 1.15 (3H, d, J=6.6 Hz, CH₃); ¹³C NMR (δ , ppm): 176.7 (s), 78.5 (t, CH₂NO₂), 66.5 (t, C-5), 41.2 (d, C-3), 32.4 (d, CHCH₃), 26.2 (t, C-4), 13.3 (q, CH₃).

Compound **15b.** ¹H NMR (δ , ppm): 4.71 (1H, dd, J_1 = 6.2 Hz, J_2 =12.4 Hz, CHNO₂), 4.51 (1H, dd, J_1 =7.1 Hz, J_2 =12.4 Hz, CHNO₂), 4.38 (1H, dt, J_1 =1.8 Hz, J_2 = J_3 = 9.0 Hz, H-5), 4.21 (1H, dt, J_1 =6.7 Hz, J_2 = J_3 =9.0 Hz, H-5), 2.79 (1H, sept, J=6.9 Hz, CHCH₃), 2.69 (1H, ddd, J_1 =11.3 Hz, J_2 =8.4 Hz, J_3 =6.9 Hz, H-3), 2.36 (1H, m, H-4), 2.10 (1H, m, H-4), 1.08 (3H, d, J=6.9 Hz, CH₃); ¹³C NMR (δ , ppm): 177.1 (s), 78.9 (t, CH₂NO₂), 66.3 (t, C-5), 41.2 (d, C-3), 32.5 (d, CHCH₃), 25.3 (t, C-4), 14.5 (q, CH₃).

4.2.2. Reaction of the lactone 1' with (*E*)-1-nitropropene **2.** *syn-* and *anti-*4,5-Dihydro-3-methyl-3-(1-methyl-2nitroethyl)-2(3*H*)-furanone 16a and 16b. The isomers 16a and 16b (60% yield) were obtained as a 1:9 inseparable mixture, yellow oil, IR (cm⁻¹, neat): 1764 (OC=O), 1550, 1380 (NO₂); MS (*m*/*z*): 140 (3), 100 (35), 97 (8), 96 (18), 83 (10), 82 (16), 81 (34), 71 (10), 69 (29), 67 (24), 57 (20), 55 (100). Anal. Calcd for C₈H₁₃NO₄: C, 51.33; H, 7.00; N, 7.48. Found: C, 51.4; H, 6.92; N, 7.19.

For clarity sake the NMR values are given separately for each isomer.

Compound **16a**. ¹H NMR (δ , ppm): 4.93 (1H, dd, J_1 = 4.0 Hz, J_2 = 12.4 Hz, CHNO₂), 4.19 (1H, dd, J_1 = 9.9 Hz,

 J_2 =12.4 Hz, CHNO₂), 1.22 (3H, s, CH₃), 1.05 (3H, d, J= 7.0 Hz, CH₃); ¹³C NMR (δ , ppm): 180.0 (s), 77.6 (t, CH₂NO₂), 64.8 (t, C-5), 44.5 (s, C-3), 36.9 (d, CHCH₃), 32.8 (t, C-4), 18.7 (q, CH₃ at C-3), 13.5 (q, CH₃ of the chain).

Compound **16b.** ¹H NMR (δ , ppm): 4.53 (1H, dd, J_1 = 4.0 Hz, J_2 =12.1 Hz, CHNO₂), 4.33 (1H, dd, J_1 =10.4 Hz, J_2 =12.1 Hz, CHNO₂), 4.26 (2H, m, 2H-5), 2.63 (1H, m, CHCH₃), 2.22 (1H, dt, J_1 =13.2 Hz, J_2 = J_3 =8.8 Hz, H-4), 1.98 (1H, ddd, J_1 =4.0 Hz, J_2 =7.3 Hz, J_3 =13.2 Hz, H-4), 1.31 (3H, s, CH₃), 1.10 (3H, d, J=7.0 Hz, CH₃); ¹³C NMR (δ , ppm): 179.5 (s), 78.2 (t, CH₂NO₂), 65.0 (t, C-5), 44.6 (s, C-3), 37.6 (d, CHCH₃), 32.5 (t, C-4), 21.5 (q, CH₃ at C-3), 13.1 (q, CH₃ of the chain).

4.2.3. Reaction of lactone 1 with (*E*)-2-nitrostyrene 3. syn and *anti*-4,5-Dihydro-3-(2-nitro-1-phenylethyl)-2(3*H*)-furanone 17a and 17b. The isomers 17a and 17b (62% yield) were obtained as a 45:55 inseparable mixture, white solid, mp 65–69 °C, IR (cm⁻¹, nujol): 1762 (OC=O), 1603 (Ph), 1552, 1378 (NO₂); ¹³C NMR (δ , ppm): 176.9 (s), 176.8 (s), 136.7 (s), 135.4 (s), 129.2 (d), 129.1 (d), 128.5 (d), 128.4 (d), 128.3 (d), 127.7 (d), and for 17a: 77.7 (t, CH₂NO₂), 66.3 (t, C-5), 44.3 (d, CHPh), 41.3 (d, C-3), 27.6 (t, C-4), and for 17b: 76.6 (t, CH₂NO₂), 66.7 (t, C-5), 43.8 (d, CHPh), 41.7 (d, C-3), 26.1 (t, C-4). MS (*m*/*z*): 235 (M⁺, 2), 217 (8), 189 (21), 188 (66), 160 (23), 145 (51), 143 (22), 131 (18), 130 (100), 128 (19), 118 (34), 116 (34), 106 (26), 105 (90), 103 (33), 91 (85), 86 (13), 78 (18), 77 (30). Anal. Calcd for C₁₂H₁₃NO₄: C, 61.27; H, 5.57; N, 5.95. Found: C, 61.3; H, 5.50; N, 6.04.

For clarity sake the NMR values are given separately for each isomer.

Compound **17a.** ¹H NMR (δ , ppm): 7.30 (5H, m, Ph), 5.44 (1H, dd, J_1 =5.5 Hz, J_2 =13.2 Hz, CHNO₂), 4.80 (1H, dd, J_1 =9.9 Hz, J_2 =13.2 Hz, CHNO₂), 4.25 (1H, dt, J_1 = 2.8 Hz, J_2 = J_3 =8.8 Hz, H-5), 4.12 (1H, m, H-5), 3.80 (1H, m, CHPh), 2.92 (1H, q, J=9.8 Hz, H-3), 1.98 (2H, m, 2H-4); ¹H NMR (δ , ppm, CDCl₃+drops of C₆D₆): 7.20 (5H, m, Ph), 5.33 (1H, dd, J_1 =5.3 Hz, J_2 =12.9 Hz, CHNO₂), 4.67 (1H, dd, J_1 =9.9 Hz, J_2 =12.9 Hz, CHNO₂), 4.06 (1H, dt, J_1 =2.5 Hz, J_2 = J_3 =8.8 Hz, H-5), 3.93 (1H, m, H-5), 3.63 (1H, m, CHPh), 2.69 (1H, dt, J_1 =8.8 Hz, J_2 = J_3 =10.2 Hz, H-3), 1.84 (1H, m, H-4), 1.71 (1H, m, H-4).

Compound **17b.** ¹H NMR (δ , ppm): 7.30 (5H, m, Ph), 5.20 (1H, dd, $J_1 = 7.3$ Hz, $J_2 = 13.5$ Hz, CHNO₂), 5.05 (1H, dd, $J_1 = 8.4$ Hz, $J_2 = 13.5$ Hz, CHNO₂), 4.12 (1H, m, H-5), 3.80 (2H, m, CHPh, H-5), 3.06 (1H, dt, $J_1 = 4.6$ Hz, $J_2 = J_3 = 9.4$ Hz, H-3), 2.38 (1H, m, H-4), 1.98 (1H, m, H-4); ¹H NMR (δ , ppm, CDCl₃+drops of C₆D₆): 7.20 (5H, m, Ph), 5.07 (1H, dd, $J_1 = 6.9$ Hz, $J_2 = 13.4$ Hz, CHNO₂), 4.93 (1H, dd, $J_1 = 8.0$ Hz, $J_2 = 13.4$ Hz, CHNO₂), 3.93 (1H, m, H-5), 3.71 (1H, dt, $J_1 = 4.5$ Hz, $J_2 = J_3 = 8.0$ Hz, CHPh), 3.63 (1H, m, H-5), 2.85 (1H, dt, $J_1 = 4.5$ Hz, $J_2 = J_3 = 9.3$ Hz, H-3), 2.10 (1H, m, H-4), 1.84 (1H, m, H-4).

4.2.4. Reaction of lactone 1' with (*E*)-2-nitrostyrene 3. syn- and anti-4,5-Dihydro-3-methyl-3-(2-nitro-1-phenyl-ethyl)-2(3*H*)-furanone 18a and 18b. The isomers 18a and

18b (80% yield), obtained in the ratio of 65:35 were not separable by flash chromatography, white solid, mp 99–101 °C; IR (cm⁻¹, nujol): 1756 (OC=O), 1602 (Ph), 1548, 1339 (NO₂); MS (*m*/*z*): 250 (MH⁺, 1), 204 (4), 203 (13), 143 (16), 129 (13),128 (10), 116 (14), 114 (13), 106 (26), 105 (100), 104 (18), 101 (85), 100 (10), 98 (10), 92 (40), 85 (15), 83 (11), 78 (13), 77 (26), 71 (23), 69 (20), 57 (40), 56 (12), 55 (34). Anal. Calcd for $C_{13}H_{15}NO_4$: C, 62.64; H, 6.07; N, 5.62. Found: C, 62.5; H, 6.17; N, 5.81.

For clarity sake the NMR values are given separately for each isomer.

Compound **18a**. ¹H NMR (δ , ppm): 7.28 (5H, m, Ph), 5.28 (1H, dd, J_1 =4.0 Hz, J_2 =13.4 Hz, CHNO₂), 4.93 (1H, dd, J_1 =11.5 Hz, J_2 =13.4 Hz, CHNO₂), 4.17 (1H, m, H-5), 4.08 (1H, m, H-5), 3.82 (1H, dd, J_1 =4.0 Hz, J_2 =11.5 Hz, CHPh), 2.28 (1H, m, H-4), 1.72 (1H, ddd, J_1 =5.5 Hz, J_2 = 7.3 Hz, J_3 =13.2 Hz, H-4), 1.29 (3H, s, CH₃); ¹³C NMR (δ , ppm): 179.8 (s), 135.0 (s), 128.8 (d), 128.5 (d), 128.3 (d), 75.2 (t, CH₂–NO₂), 64.8 (t, C-5), 47.3 (d, CHPh), 44.6 (s, C-3), 34.0 (t, C-4), 19.6 (q, CH₃).

Compound **18b.** ¹H NMR (δ , ppm): 7.28 (5H, m, Ph), 5.09 (1H, dd, J_1 =11.2 Hz, J_2 =13.2 Hz, CHNO₂), 4.89 (1H, dd, J_1 =4.2 Hz, J_2 =13.2 Hz, CHNO₂), 4.08 (1H, m, H-5), 3.70 (1H, dd, J_1 =4.2 Hz, J_2 =11.2 Hz, CHPh), 3.59 (1H, dt, J_1 = J_2 =9.0 Hz, J_3 =4.8 Hz, H-5), 2.28 (1H, m, H-4), 1.98 (1H, ddd, J_1 =4.8 Hz, J_2 =8.1 Hz, J_3 =12.8 Hz, H-4), 1.36 (3H, s, CH₃); ¹³C NMR (δ , ppm): 179.9 (s), 135.8 (s) 129.0 (d), 128.4 (d), 128.3 (d), 76.5 (t, CH₂–NO₂), 65.5 (t, C-5), 50.2 (d, CHPh), 45.5 (s, C-3), 33.3 (t, C-4), 23.4 (q, CH₃).

4.2.5. Reaction of the lactone 1 with (*E*)-2-nitro-1phenylpropene **4. 4,5-Dihydro-3-(2-nitro-1-phenylpro**pyl)-2(3*H*)-furanone 19a,b,c,d. Four isomers 19a, 19b, 19c and 19d in 53:22:14:11 ratio were identified in the ¹H NMR spectrum of the crude reaction mixture. They were separated by flash chromatography.

Compound 19a. 30% Yield, white solid, mp 125-126 °C, from ether; IR (cm⁻¹, nujol): 1756, (OC=O), 1546, 1377 (NO₂); ¹H NMR (δ, ppm): 7.28 (5H, m, Ph), 5.72 (1H, dq, $J_1 = 11.0 \text{ Hz}, J_2 = J_3 = J_4 = 6.6 \text{ Hz}, \text{ CHNO}_2), 4.09 (1H, q)$ J = 8.2 Hz, H-5), 3.64 (1H, dt, $J_1 = 4.8$ Hz, $J_2 = J_3 = 8.2$ Hz, H-5), 3.45 (1H, dd, $J_1 = 4.8$ Hz, $J_2 = 11.0$ Hz, CHPh), 3.12 (1H, dt, J_1 =4.8 Hz, J_2 = J_3 =8.2 Hz, H-3), 2.38 (1H, m, H-4), 1.92 (1H, dq, $J_1 = 13.0$ Hz, $J_2 = J_3 = J_4 = 8.2$ Hz, H-4), 1.82 (3H, d, J = 6.6 Hz, CH₃), ¹³C NMR (δ , ppm): 177.0 (s), 135.6 (s), 128.9 (d), 128.5 (d), 128.4 (d), 85.4 (d, CHNO₂), 66.8 (t, C-5), 50.0 (d, CHPh), 39.9 (d, C-3), 26.1 (t, C-4), 18.6 (q, CH₃); MS (*m*/*z*): 250 (MH⁺, 4), 203 (26), 202 (31), 144 (25), 143 (63), 142 (38), 131 (29), 129 (21), 118 (100), 116 (27), 106 (19), 103 (31), 102 (29), 91 (57), 79 (13), 77 (19). Anal. Calcd for C13H15NO4: C, 62.64; H, 6.07; N, 5.62. Found: C, 62.5; H, 5.87; N, 5.28.

Compound **19b.** 17% Yield; white solid, mp 95–97 °C, IR (cm⁻¹, nujol): 1762, (OC=O), 1600 (Ph), 1550, 1388 (NO₂); ¹H NMR (δ , ppm): 7.36 (3H, m, Ph), 7.26 (2H, m, Ph), 5.89 (1H, dq, J_1 =11.1 Hz, J_2 = J_3 = J_4 =6.7 Hz, CHNO₂), 4.06 (1H, q, J=8.5 Hz, H-5), 3.70 (1H, dt, J_1 = 4.3 Hz, J_2 = J_3 =8.5 Hz, H-5), 3.44 (1H, dd, J_1 =4.2 Hz,

 J_2 =11.1 Hz, CHPh), 2.88 (1H, dt, J_1 =4.2 Hz, J_2 = J_3 = 8.5 Hz, H-3), 2.46 (1H, m, H-4), 1.91 (1H, dq, J_1 =12.8 Hz, J_2 = J_3 = J_4 =8.5 Hz, H-4), 1.33 (3H, d, J=6.7 Hz, CH₃); ¹³C NMR (δ , ppm): 176.8 (s), 135.0 (s), 129.4 (d), 129.3 (d), 128.6 (d), 83.5 (d, CHNO₂), 66.6 (t, C-5), 50.6 (d, CHPh), 40.5 (d, C-3), 26.5 (t, H-4), 19.5 (q, CH₃); MS (m/z): 249 (M⁺, 3), 203 (26), 202 (36), 143 (14), 131 (37), 129 (27), 118 (38), 117 (100), 115 (29), 105 (15), 91 (67), 84 (13), 77 (14). Anal. Calcd for C₁₃H₁₅NO₄: C, 62.64; H, 6.07; N, 5.62. Found: C, 62.9; H, 6.23; N, 5.33.

Compound **19c** (in admixture with **19a**). ¹H NMR (δ , ppm): 7.21 (5H, m, Ph), 5.68 (1H, dq, J_1 =5.5 Hz, J_2 = J_3 = J_4 = 6.6 Hz, CHNO₂), 4.13 (2H, m, 2H-5), 3.40 (2H, m, CHPh and H-3), 2.12 (1H, m, H-4), 1.85 (1H, m, H-4), 1.54 (3H, d, J=6.6 Hz, CH₃); ¹³C NMR (δ , ppm): 177.5 (s), 134.6 (s), 128.7 (d), 128.6 (d), 128.2 (d), 82.5 (d, CHNO₂), 66.2 (t, C-5), 50.1 (d, CHPh), 39.7 (d, C-3), 27.2 (t, H-4), 17.5 (q, CH₃).

Compound **19d.** Oil, 6% yield; IR (cm⁻¹, neat): 1766, 1712 (OC=O), 1602 (Ph), 1550, 1390 (NO₂); ¹H NMR (δ , ppm): 7.34 (3H, m, Ph), 7.16 (2H, m, Ph), 5.31 (1H, dq, J_1 = 9.1 Hz, J_2 = J_3 = J_4 =6.6 Hz, CHNO₂), 4.13 (2H, m, 2H-5), 3.87 (1H, dd, J_1 =6.2 Hz, J_2 =9.1 Hz, CHPh), 2.84 (1H, dt, J_1 =6.2 Hz, J_2 = J_3 =9.6 Hz, H-3), 2.34 (1H, m, H-4), 2.09 (1H, m, H-4), 1.45 (3H, d, J=6.6 Hz, CH₃); MS,(m/z): 203 (15), 202 (34), 176 (19), 148 (28), 143 (15), 131 (46), 130 (47), 129 (36), 118 (35), 117 (100), 116 (20), 115 (42), 105 (18), 104 (14), 103 (10), 91 (95), 77 (22).

4.2.6. Reaction of the lactone 1' with (E)-2-nitro-1phenylpropene 4. 4,5-Dihydro-3-methyl-3-(2-nitro-1phenylpropyl)-2(3H)-furanone 20a,b,c. The isomers 20a, 20b and 20c (61%, 35% and 4% relative ratio) were obtained acidifying the reaction mixture with 0.43 ml (7 mmol) of glacial acetic acid in 1.5 ml of THF, at -78 °C. The temperature was allowed to raise to -40 °C and, after 15 min, 25 ml of water was added. The aqueous phase was extracted three times with CH₂Cl₂. The combined organic phases were dried over anhydrous Na₂SO₄. After removal of the solvent, the crude reaction mixture was purified by flash chromatography (light petroleum/ethyl acetate) to give the mixture of lactones **20a,b,c** (45% yield) which could not be separated. White solid, mp 89-92 °C, IR $(cm^{-1}, nujol)$: 1755 (OC=O), 1602 (Ph), 1545, 1334 (NO₂); MS (*m*/*z*): 263 (M⁺, 0.8), 221 (10), 206 (44), 145 (14), 131 (18), 129 (13), 119 (100), 118 (39), 116 (21), 106 (42), 102 (13), 101 (75), 99 (10), 91 (48), 77 (20), 69 (14), 57 (36), 56 (16), 55 (43). Anal. Calcd for C₁₄H₁₇NO₄: C, 63.87; H, 6.51; N, 5.32. Found: C, 64.0; H, 6.74; N, 5.31.

For clarity sake the NMR values are given separately for each isomer.

Compound **20a**. ¹H NMR (δ , ppm): 7.31 (3H, m, Ph), 7.23 (2H, m, Ph), 5.31 (1H, quintet, J_1 =6.6 Hz, CHNO₂), 4.18 (1H, dt, J_1 =8.0 Hz, J_2 =9.2 Hz, H-5), 4.02 (1H, dd, J_1 = 4.8 Hz, J_2 =8.0 Hz, J_3 =9.2 Hz, H-5), 3.44 (1H, d, J= 6.6 Hz, CHPh), 2.44 (1H, dt, J_1 =8.0 Hz, J_2 =13.2 Hz, H-4), 2.08 (1H, ddd, J_1 =4.8 Hz, J_2 =8.0 Hz, J_3 =13.2 Hz, H-4), 1.59 (3H, d, J=6.6 Hz, CH₃), 1.50 (3H, s, CH₃); ¹³C NMR (δ , ppm): 180.2 (s), 135.6 (s), 130.0 (d), 128.7 (d),

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128.1 (d), 83.2 (d, CHNO₂), 64.8 (t, C-5), 54.3 (d, CHPh), 45.7 (s, C-3), 33.3 (t, C-4), 22.2 (q, CH₃), 19.9 (q, CH₃).

Compound **20b.** ¹H NMR (δ , ppm): 7.33 (5H, m, Ph), 5.22 (1H, dq, J_1 =9.5 Hz, J_2 = J_3 =6.6 Hz, CHNO₂), 4.18 (1H, m, H-5), 4.10 (1H, ddd, J_1 =4.8 Hz, J_2 =8.4 Hz, J_3 = 9.5 Hz, H-5), 3.62 (1H, d, J=9.5 Hz, CHPh), 2.47 (1H, m, H-4), 2.05 (1H, m, H-4), 1.34 (3H, d, J=6.6 Hz, CH₃), 1.26 (3H, s, CH₃); ¹³C NMR (δ , ppm): 179.4 (s), 135.3 (s), 129.8 (d), 128.6 (d), 128.2 (d), 83.6 (d, CHNO₂), 65.1 (t, H-5), 53.4 (d, CHPh), 45.7 (s, C-3), 33.4 (t, H-4), 22.2 (q, CH₃), 19.9 (q, CH₃).

Compound **20c.** Only a few signals were identified: ¹H NMR (δ , ppm): 5.45 (1H, dq, J_1 =6.6 Hz, J_2 =9.5 Hz, CHNO₂), 3.89 (1H, dt, J_1 =3.9 Hz, J_2 =9.2 Hz, H-5), 3.61 (1H, d, J=9.5 Hz, CHPh), 2.63 (1H, m, H-4), 1.93 (1H, m, H-4), 1.29 (3H, d, J=6.6 Hz, CH₃); ¹³C NMR (δ , ppm): 129.1 (d), 82.5 (d, CHNO₂), 65.7 (t, C-5), 54.8 (d, CHPh), 32.6 (s).

4.3. Acidification of the lithium nitronates 13 and 14 with 3 N HCl

The crude reaction mixture containing the lithium nitronate was acidified with 20 ml of 3 N HCl, the mixture was stirred overnight and extracted three times with CH₂Cl₂. The combined organic phases were dried over anhydrous Na₂SO₄. After removal of the solvent, the residue was purified by flash chromatography (light petroleum/ethyl acetate). In the case of nitronate **13**, from the complex mixture containing the Nef products **32a** and **32b** in 3:2 ratio and the oxime **34**, only **32a** and **34** could be isolated as pure compounds, by flash chromatography. In the case of nitronate **14** the Nef product **33** and the oxime **35** could be isolated by flash chromatography (see text).

4.3.1. 4,5-Dihydro-3-(2-oxo-1-phenylpropyl)-2(3H)-furanone 32a. 19% Yield, white solid, mp 87–88 °C from ether; IR (cm⁻¹, nujol): 1753 (OC=O), 1714 (C=O); ¹H NMR (δ , ppm): 7.33 (3H, m, Ph), 7.21 (2H, m, Ph), 4.15 (2H, m, 2H-5), 4.08 (1H, d, *J*=7.7 Hz, CHPh), 3.56 (1H, ddd, *J*₁= 7.7 Hz, *J*₂=8.7 Hz, *J*₃=12.1 Hz, H-3), 2.20 (3H, s, CH₃), 2.12 (1H, m, H-4), 1.82 (1H, m, H-4); ¹³C NMR (δ , ppm): 206.1 (s, C=O), 177.9 (s, COO), 134.7 (s), 129.0 (d), 128.8 (d), 128.0 (d), 66.9 (t, C-5), 58.3 (d, CHPh), 41.3 (d, C-3), 29.4 (q, CH₃), 26.4 (t, C-4); MS (*m*/*z*): 218 (M⁺⁺, 1), 176 (53), 149 (76), 148 (32), 132 (47), 131 (100), 118 (18), 117 (19), 116 (28), 105 (31), 92 (74), 77 (18). Anal. Calcd for C₁₃H₁₄O₃: C, 71.54; H, 6.47. Found: C, 71.3; H, 6.39.

4.3.2. 4,5-Dihydro-3-(2-oxo-1-phenylpropyl)-2(3H)-fura-none 32b. The lactone **19d** was submitted to the Nef reaction conditions according to the literature,¹⁹ furnishing a 3:1 mixture of **32a** and **32b**.

Compound **32b**. ¹H NMR (δ , ppm): 7.36 (5H, m, Ph), 4.48 (1H, dt, $J_1 = 2.3$ Hz, $J_2 = J_3 = 9.0$ Hz, H-5), 4.28 (1H, d, J = 5.1 Hz, CHPh), 4.16 (1H, m, H-5), 2.89 (1H, ddd, $J_1 = 5.1$ Hz, $J_2 = 9.5$ Hz, $J_3 = 11.0$ Hz, H-3), 2.56 (1H, quintet, J = 10.4 Hz, H-4), 2.21 (1H, m, H-4), 2.08 (3H, s, CH₃); ¹³C NMR (δ , ppm): 206.3 (s, C=O), 177.8 (s, COO), 136.3 (s),

129.3 (d), 128.7 (d), 127.9 (d), 66.6 (t, C-5), 58.2 (d, CHPh), 42.8 (d, C-3), 29.1 (q, CH₃), 25.1 (t, C-4).

4.3.3. 4,5-Dihydro-3-methyl-3-(2-oxo-1-phenylpropyl)-2(*3H*)-furanone **33.** 17% Yield; white solid, mp 67– 70 °C; IR (cm⁻¹, nujol): 1772 (OC=O), 1701(C=O), 1580 (Ph); ¹H NMR (δ , ppm): 7.33 (3H, m, Ph), 7.28 (2H, m, Ph), 4.27 (1H, s, CHPh), 4.06 (1H, q, J_1 =8.6 Hz, H-5), 3.58 (1H, dt, J_1 =5.1 Hz, J_2 =8.6 Hz, H-5), 2.63 (1H, m, H-4), 2.23 (1H, m, H-4), 2.12 (3H, s, CH₃–CO), 1.38 (3H, s, CH₃); ¹³C NMR (δ , ppm): 205.6 (s, C=O), 180.6 (s, COO), 133.4 (s), 130.0 (d), 128.9 (d), 128.2 (d), 65.6 (t, C-5), 62.7 (d, CHPh), 46.1 (s, C-3), 31.1 (q, CH₃–CO), 30.9 (t, C-4), 23.3 (q, CH₃); MS (*m*/*z*): [232 (M⁺⁺) at 20 eV], 190 (25), 175 (15), 162 (21), 144 (44), 129 (25), 117 (22), 115 (20), 106 (52), 105 (65), 91 (100), 77 (41), 65 (10), 57 (10). Anal. Calcd for C₁₄H₁₆O₃: C, 72.39; H, 6.94. Found: C, 72.5; H, 6.80.

4.3.4. 4,5-Dihydro-3-(2-hydroxyimino-1-phenylpropyl)-2(3H)-furanone 34. 15% Yield; white solid, mp 127-129 °C from light petroleum/ether; IR (cm⁻¹, nujol): 3329 (OH), 1730 (OC=O), 1655 (C=N), 1600 (Ph); ¹H NMR (δ , ppm): 8.00 (1H, bs, OH), 7.27 (5H, m, Ph), 4.40 (1H, dt, $J_1 = 3.0 \text{ Hz}, J_2 = J_3 = 8.8 \text{ Hz}, \text{ H-5}$, 4.16 (1H, q, J = 8.8 Hz, Hz) H-5), 4.10 (1H, d, J=5.9 Hz, CHPh), 2.99 (1H, dt, $J_1=$ 5.9 Hz, $J_2 = J_3 = 8.8$ Hz, H-3), 2.66 (1H, dq, $J_1 = 12.0$ Hz, $J_2 = J_3 = J_4 = 8.8$ Hz, H-4), 1.99 (1H, m, H-4), 1.75 (3H, s, CH₃); ¹³C NMR (δ , ppm): 178.3 (s, COO), 155.8 (s, C=N), 138.5 (s), 128.8 (d), 128.4 (d), 127.4 (d), 66.7 (t, C-5), 50.4 (d, CHPh), 43.2 (d, C-3), 24.7 (t, C-4), 13.8 (q, CH₃); MS (m/z): 233 (M⁺⁺, 15), 216 (79), 185 (20), 175 (22), 174 (40), 173 (32), 172 (18), 170 (26), 157 (26), 156 (17), 149 (41), 148 (49), 147 (20), 132 (32), 130 (49), 128 (22), 121 (26), 118 (30), 116 (65), 92 (100), 89 (29), 81 (39),77 (65). Anal. Calcd for C₁₃H₁₅NO₃: C, 66.94; H, 6.48; N, 6.00. Found: C, 67.2; H, 6.58; N, 6.02.

4.3.5. 4,5-Dihydro-3-methyl-3-(2-hydroxyimino-1-phenylpropyl)-2(3H)-furanone 35. 10% Yield, white solid, mp 117–119 °C. IR (cm⁻¹, nujol): 3271 (OH), 1758 (C=O), 1670 (C=N); ¹H NMR (δ , ppm): 7.80 (1H, bs, OH), 7.30 (5H, m, Ph), 4.03 (1H, dt, $J_1 = 7.3$ Hz, $J_2 =$ 8.8 Hz, H-5), 3.89 (1H, s, CHPh), 3.56 (1H, dt, $J_1 = 5.1$ Hz, $J_2 = 8.8$ Hz, H-5), 2.65 (1H, ddd, $J_1 = 7.3$ Hz, $J_2 = 8.8$ Hz, $J_3 = 13.6$ Hz, H-4), 2.17 (1H, m, H-4), 1.79 (3H, s, CH₃C=N-OH), 1.45 (3H, s, CH₃); ¹³C NMR (δ, ppm): 181.4 (s, COO), 156.6 (s, C=N), 136.1 (s), 130.1 (d), 128.5 (d), 127.8 (d), 65.4 (t, C-5), 56.5 (d, CHPh), 46.7 (s, C-3), 31.7 (t, C-4), 24.5 (q, CH₃), 15.5 (q, CH₃); MS (m/z): 247 $(M^{+}, 16), 202 (10), 186 (18), 149 (43), 148 (100), 131 (25),$ 130 (42), 129 (26), 117 (19), 116 (15), 115 (33), 106 (25), 105 (17), 100 (13), 95 (17), 91 (42), 77 (25), 69 (10), 55(13). Anal. Calcd for C14H17NO3: C, 68.00; H, 6.93; N, 5.66. Found: C, 67.9; H, 7.00; N, 5.56.

4.3.6. 1-(Tetrahydro-3-methyl-2-oxofuryl)-1-phenylpropan-2-ylideneazinic acid 37. The reaction between the lithium enolate of 1' and (*E*)-2-nitro-1-phenylpropene **4**, carried out in accordance with the general procedure, was quenched by addition of a saturated aqueous solution of NH₄Cl. The aqueous phase was extracted with CH₂Cl₂ three times in order to separate the nitroalkylated lactones. From
the mother liquors, acidified to pH 2 with 3 N HCl, the nitronic acid **37** precipitated as a white solid which was washed with ether. 10% Yield, mp 100–102 °C; IR (cm⁻¹, nujol): 2668 (OH); 1754 (COO), 1658 (C=N); ¹H NMR (DMSO-d₆, δ , ppm) 7.20 (5H, m, Ph), 4.70 (1H, s, CHPh), 4.18 (1H, q, *J*=8.4 Hz, H-5), 3.93 (1H, dt, *J*₁=2.7 Hz, *J*₂= *J*₃=8.4 Hz, H-5), 2.43 (1H, m, H-4), 2.12 (1H, m, H-4), 2.03 (3H, s, CH₃), 1.23 (3H, s, CH₃); ¹³C NMR (DMSO-d₆, δ , ppm): 180.9 (s), 137.9 (s), 129.3 (d), 129.0 (d), 127.7 (d), 121.2 (s), 65.4 (t), 50.8 (d), 46.4 (t), 31.2 (s), 23.0 (q), 16.6 (q); MS (*m*/*z*): 262 (M-H^{¬+}, 0.3), 247 (0.4), 216 (1.3), 190 (30), 175 (18), 162 (13), 145 (27), 144 (58), 130 (35), 118 (27), 116 (28), 106 (44), 100 (10), 99 (5), 92 (100), 77 (47), 55 (7), 51 (17). Anal. Calcd for C₁₄H₁₇NO₄: C, 63.87; H, 6.51; N, 5.32. Found: C, 63.7; H, 6.71; N, 5.26.

4.4. General procedure for the reduction with Raney Ni

The nitro compound (0.85 mmol) was dissolved in 1:1 EtOH/ethyl acetate (20 ml) and one teaspoon of Raney Ni (Aldrich) was added. The apparatus was evacuated and flushed with H_2 . The mixture was stirred at room temperature under H_2 atmosphere for 16 h, then filtered on Celite and the solvent was evaporated.

4.4.1. (3*R**,4*R**)- and (3*R**,4*S**)-3-(2-Hydroxyethyl)-4methyl-2-pyrrolidinone 21a and 21b. Reduction of the nitroalkylated lactone 15a gave the *trans* isomer 21a: 80% yield, white solid, mp 78–80 °C, IR (cm⁻¹, nujol): 3165 (OH and NH), 1672 (NHC=O); ¹H NMR (δ , ppm): 7.03 (1H, bs, NH), 4.9 (1H, bs, OH), 3.84 (1H, m, CHOH), 3.72 (1H, m, CHOH), 3.49 (1H, t, *J*=9.0 Hz, H-5), 2.99 (1H, t, *J*=9.0 Hz, H-5), 2.18 (1H, m), 2.09 (1H, m), 1.78 (2H, m, CH₂CH₂OH), 1.15 (3H, d, *J*=6.9 Hz, CH₃); ¹³C NMR (δ , ppm): 181.6 (s), 62.2 (t, CH₂OH), 49.7 (d, C-3), 48.7 (t, C-5), 37.1 (d, C-4), 32.4 (t, CH₂CH₂OH), 17.3 (q, CH₃); MS (*m*/*z*): 144 (MH⁺, 41), 126 (6), 113 (9), 112 (10), 99 (22), 98 (33), 96 (17), 85 (18), 84 (100), 67 (9). Anal. Calcd for C₇H₁₃NO₂: C, 58.72; H, 9.15; N, 9.78. Found: C, 58.5; H, 8.90; N, 9.59.

Reduction of nitroalkylated lactone **15b** gave the *cis* isomer **21b**: 80% yield, white solid, mp 62–65 °C, IR (cm⁻¹, nujol): 3165 (OH and NH), 1672 (NHC=O); ¹H NMR (δ , ppm): 6.77 (1H, bs, NH), 4.4 (1H, bs, OH), 3.82 (1H, m, CHOH), 3.70 (1H, m, CHOH), 3.48 (1H, dd, J_1 =6.0 Hz, J_2 =9.5 Hz, H-5), 2.94 (1H, bd, J=9.5 Hz, H-5), 2.50 (2H, m, H-3 and H-4), 1.70 (2H, m, CH₂CH₂OH), 0.97 (3H, d, J=6.9 Hz, CH₃); ¹³C NMR (δ , ppm): 180.3 (s), 62.2 (t, CH₂OH), 48.6 (t, C-5), 45.5 (d, C-3), 33.5 (d, C-4), 28.5 (t, CH₂CH₂OH), 14.6 (q, CH₃); MS (*m*/*z*): 144 (MH⁺, 49), 126 (14), 112 (22), 99 (27), 98 (69), 96 (23), 85 (23), 84 (100), 67 (22). Anal. Calcd for C₇H₁₃NO₂: C, 58.72; H, 9.15; N, 9.78. Found: C, 58. 5; H, 9.00; N, 9.58.

4.4.2. (*3R**,*4R**)- and (*3R**,*4S**)-3-(2-Hydroxyethyl)-3,4dimethyl-2-pyrrolidinone 22a and 22b. The two isomers were obtained as a 1:9 inseparable mixture, 80% yield, white solid, mp 88 °C (from light petroleum/ethyl acetate); IR (cm⁻¹, nujol): 3330 (OH and NH), 1680 (NHC=O); MS (*m*/*z*): 158 (M+H^{¬+}, 13), 113 (40), 112 (54), 98 (100), 84 (17), 67 (11), 55 (22). Anal. Calcd for C₈H₁₅NO₂: C, 61.12; H, 9.62; N, 8.91. Found: C, 61.3; H, 9.41; N, 9.14. For clarity sake the NMR values of the two isomers are given separately.

Compound **22a** in admixture with **22b**. ¹H NMR (δ , ppm) (only few signals were identified): 6.97 (1H, bs, NH), 2.25 (1H, m, H-4), 1.70 (1H, ddd, J_1 =5.8 Hz, J_2 =10.2 Hz, J_3 = 15.7 Hz, CHCH₂OH), 1.42 (1H, m, CHCH₂OH), 0.94 (3H, s, CH₃); ¹³C NMR (δ , ppm): 184.5 (s), 58.7 (t, CH₂OH), 47.0 (t, C-5), 45.9 (s, C-3), 39.7 (d, C-4), 38.2 (t, CH₂CH₂OH), 15.1 (q, CH₃ at C-3), 11.4 (q, CH₃ at C-4).

Compound **22b.** ¹H NMR (δ , ppm). 6.92 (1H, bs, NH), 3.82 (1H, m, CHOH), 3.79 (1H, bs, OH), 3.63 (1H, m, CHOH), 3.35 (1H, dd, J_1 =8.2 Hz, J_2 =9.0 Hz, H-5), 2.92 (1H, dd, J_1 =8.2 Hz, J_2 =9.7 Hz, H-5), 2.13 (1H, m, H-4), 1.73 (1H, ddd, J_1 =5.3 Hz, J_2 =9.0 Hz, J_3 =14.1 Hz, CHCH₂OH), 1.35 (1H, dt, J_1 =4.7 Hz, J_2 =14.1 Hz, CHCH₂OH), 1.13 (3H, s, CH₃), 0.96 (3H, d, J=7.3 Hz, CH₃); ¹³C NMR (δ , ppm): 184.0 (s), 58.9 (t, CH₂OH), 46.6 (t, C-5), 44.6 (s, C-3), 42.0 (d, C-4), 34.0 (t, CH₂CH₂OH), 20.5 (q, CH₃ at C-3), 12.6 (q, CH₃ at C-4).

4.4.3. (3*R**,4*S**)- and (3*R**,4*R**)-3-(2-Hydroxyethyl)-4phenyl-2-pyrrolidinone 23a and 23b. Treatment of the crude reaction mixture with light petroleum/ethyl acetate gave a 3:2 mixture of 23a and 23b as a white solid (70% yield), mp 90–93 °C, IR (cm⁻¹, nujol): 3366, 3262 (OH and NH), 1693 (NHC=O), 1638 (Ph); MS (*m*/*z*): 206 (MH⁺, 16), 160 (100), 159 (90), 147 (13), 118 (13), 117 (43), 115 (33), 104 (16), 91 (13), 84 (14), 78 (11). Anal. Calcd for $C_{12}H_{15}NO_2$: C, 70.22; H, 7.37; N, 6.82. Found: C, 70.0; H, 7.11; N, 6.64.

For clarity sake the NMR values of the isomeric mixture are given separately for each isomer.

Compound **23a**. ¹H NMR (δ , ppm): 7.64 (1H, bs, NH), 7.28 (5H, m, Ph), 4.5 (1H, bs, OH), 3.74–3.50 (3H, m, CH₂OH, H-5), 3.41 (1H, t, *J*=9.5 Hz, H-5), 3.27 (1H, q, *J*=9.5 Hz, H-4), 2.70 (1H, dt, *J*₁=*J*₂=9.5 Hz, *J*₃=4.0 Hz, H-3), 1.85 (1H, m, CHCH₂OH), 1.74 (1H, m, CHCH₂OH); ¹³C NMR (δ , ppm): 180.4 (s), 140.0 (s), 129.0 (2d), 128.7 (2d), 127.6 (d), 61.6 (t, CH₂OH), 49.0 (d, C-3), 48.9 (t, C-5), 48.4 (d, C-4), 32.4 (t, CH₂CH₂OH).

Compound **23b.** ¹H NMR (δ , ppm): 7.67 (1H, bs, NH), 7.28 (5H, m, Ph), 4.5 (1H, bs, OH), 3.79 (1H, dd, J_1 =7.1 Hz, J_2 =9.7 Hz, H-5), 3.74–3.50 (4H, m, CH₂OH, H-5, H-4), 2.92 (1H, dt, J_1 = J_2 =8.7 Hz, J_3 =5.0 Hz, H-3), 1.48 (1H, m, CHCH₂OH), 1.30 (1H, m, CHCH₂OH); ¹³C NMR (δ , ppm): 181.2 (s), 140.4 (s), 127.8 (2d), 127.5 (2d), 127.3 (d), 61.4 (t, CH₂OH), 48.0 (t, C-5), 45.3 (d, C-3), 44. 3 (d, C-4), 29.9 (t, CH₂CH₂OH).

4.4.4. (3*R**,4*R**)- and (3*R**,4*S**)-3-(2-Hydroxyethyl)-3methyl-4-phenyl-2-pyrrolidinone 24a and 24b. The 65:35 crude mixture of 24a and 24b (70% overall yield), obtained from the reduction, was purified by flash chromatography. The two isomeric lactams could be separated only partially. Semisolid material, IR (cm⁻¹, neat): 3260 (OH and NH), 1685 (NHC=O); MS (*m*/*z*): 220 (MH⁺, 20), 204 (27), 175 (100), 174 (69), 160 (68), 158 (28), 131 (14), 129 (18), 128 (15), 117 (11), 115 (19), 104 (21), 98 (54), 91 (16), 78 (11). Anal. Calcd for C₁₃H₁₇NO₂: C, 71.21; H, 7.81; N, 6.39. Found: C, 71.0; H, 7.88; N, 6.25.

Compound **24a**. ¹H NMR (δ , ppm). 7.33 (3H, m, *m*-, *p*-ArH), 7.23 (2H, m, *o*-ArH), 6.31 (1H, bs, NH), 4.53 (1H, dd, J_1 =8.4 Hz, J_2 =2.6 Hz, OH), 3.85–3.60 (4H, m, CH₂OH, 2H-5), 3.48 (1H, dd, J_1 =10.1 Hz, J_2 =7.8 Hz, H-4), 1.90 (1H, m, CHCH₂OH), 1.72 (1H, m, CHCH₂OH), 0.93 (3H, s, CH₃); ¹³C NMR (δ , ppm): 183.4 (s), 136.4 (s), 129.1 (2d), 128.5 (2d), 127.7 (d), 58.6 (t, CH₂OH), 51.3 (d, C-4), 47.5 (s, C-3), 44.5 (t, C-5), 38.5 (t, CH₂CH₂OH), 17.4 (q, CH₃).

Compound **24b.** ¹H NMR (δ , ppm). 7.33 (3H, m, *m*-, *p*-ArH), 7.21 (2H, m, *o*-ArH), 6.34 (1H, bs, NH), 3.85–3.60 (4H, m, CH₂OH, H-5, H-4), 3.62 (1H, dd, J_1 =2.1 Hz, J_2 = 10.1 Hz, OH), 3.36 (1H, t, J=7.7 Hz, H-5), 1.66 (1H, m, CHCH₂OH), 1.37 (3H, s, CH₃), 0.98 (1H, m, CHCH₂OH); ¹³C NMR (δ , ppm): 183.6 (s), 137.8 (s), 128.7 (2d), 128.4 (2d), 127.6 (d), 58.7 (t, CH₂OH), 53.6 (d, C-4), 46.5 (s, C-3), 44.4 (t, C-5), 35.7 (t, CH₂CH₂OH), 21.4 (q, CH₃).

4.4.5. (3R*,4R*,5S*)-3-(2-Hydroxyethyl)-5-methyl-4phenyl-2-pyrrolidinone 25a. The crude reaction mixture obtained from the reduction of 19a was treated with light petroleum/ethyl acetate to afford 25a (70% yield) as a white solid, mp 123–125 °C; IR (cm⁻¹, nujol): 3260 (OH and NH), 1690 (NHC=O), 1600 (Ph); ¹H NMR (CD₃OD, δ , ppm): 7.27 (3H, m, *m*-, *p*-ArH), 7.12 (2H, bd, *o*-ArH), 4.09 (1H, dq, $J_1 = 6.6$ Hz, $J_2 = 5.8$ Hz, H-5), 3.54 (1H, dd, $J_1 =$ 7.7 Hz, J₂=5.8 Hz, H-4), 3.45 (2H, m, CH₂OH), 3.03 (1H, ddd, J₁=8.9 Hz, J₂=7.7 Hz, J₃=5.8 Hz, H-3), 1.86 (1H, m, CHCH₂OH), 1.30 (1H, m, CHCH₂OH), 0.83 (3H, d, J= 6.6 Hz, CH₃); ¹³C NMR (δ, ppm): 183.7 (s), 139.3 (s), 132.5 (2d), 131.3 (2d), 130.4 (d), 65.1 (t, CH₂OH), 56.0 (d), 54.2 (d), 51.2 (d, C-3), 32.4 (t, CH₂CH₂OH), 19.5 (q, CH₃); MS (*m/z*): 220 (MH⁺, 14), 175 (100), 174 (19), 131 (15), 118 (27), 117 (65), 115 (33), 91 (10). Anal. Calcd for C13H17NO2: C, 71.21; H, 7.81; N, 6.39. Found: C, 69.9; H, 7.41; N, 6.14.

4.4.6. ($3R^*$, $4R^*$, $5R^*$)- and ($3R^*$, $4R^*$, $5S^*$)-3,5-Dimethyl-3-(2-hydroxyethyl)-4-phenyl-2-pyrrolidinone 26a and 26b. A 1:1 mixture of nitroalkylated lactones 20a and 20b was reduced under the above mentioned conditions to give a 1:1 mixture of the corresponding lactams 26a and 26b (70% overall yield). The same compounds were obtained from the reduction with Raney Ni of the nitronic acid 37, although in a 7:3 molar ratio. The two isomers could not be separated. White solid, mp 94–100 °C, IR (cm⁻¹, CHCl₃): 3336 (OH and NH), 1680 (NHC=O), 1602 (Ph); MS (*m*/*z*): 234 (MH⁺, 14), 218 (49), 203 (16), 190 (27), 189 (100), 188 (82), 174 (62), 162 (28), 161 (14), 160 (15), 132 (28), 131 (87), 129 (42), 128 (20), 118 (28), 117 (55), 116 (22), 115 (37), 112 (35), 91 (46). Anal. Calcd for C₁₄H₁₉NO₂: C, 72.07; H, 8.21; N, 6.00. Found: C, 71.9; H, 8.11; N, 5.89.

For clarity sake the NMR values of the isomeric mixture are given separately for each isomer.

Compound **26a**. ¹H NMR (CD₃OD, δ , ppm). 7.25 (3H, m, *m*-, *p*-ArH), 7.12 (2H, bd, *o*-ArH), 4.25 (1H, dq, J_1 =6.2 Hz, J_2 =6.9 Hz, H-5), 3.8–3.6 (2H, m, CH₂OH), 3.38 (1H, d,

 J_1 =6.2 Hz, H-4), 1.90 (2H, m, CH_2CH_2OH), 0.91 (3H, d, J=6.9 Hz, CH₃), 0.81 (3H, s, CH₃); ¹³C NMR (δ , ppm): 183.0 (s), 136.9 (s), 130.1 (2d), 128.1 (2d), 127.7 (d), 59.2 (t, CH₂OH), 57.4 (d, C-4), 50.6 (d, C-5), 47.7 (s, C-3), 40.5 (t, CH₂CH₂OH), 18.7 (q, CH₃ at C-3), 17.1 (q, CH₃ at C-5).

Compound **26b.** ¹H NMR (CD₃OD, δ , ppm). 7.25 (3H, m, *m*-, *p*-ArH), 7.12 (2H, bd, *o*-ArH), 4.12 (1H, dq, J_1 =6.2 Hz, J_2 =9.9 Hz, H-5), 3.8–3.6 (2H, m, CH₂OH), 3.07 (1H, d, J_1 =9.9 Hz, H-4), 1.70 (2H, m, CH₂CH₂OH), 1.15 (3H, d, J=6.2 Hz, CH₃), 0.85 (3H, s, CH₃); ¹³C NMR (δ , ppm): 181.9 (s), 134.8 (s), 129.4 (2d), 128.5 (2d), 127.9 (d), 60.7 (d, C-4), 58.3 (t, CH₂OH), 51.7 (d, C-5), 48.8 (s, C-3), 38.4 (t, CH₂CH₂OH), 19.7 (q, CH₃ at C-3), 19.6 (q, CH₃ at C-5).

4.5. Reduction of the nitro group with 10% Pd on carbon

The appropriate nitroalkylated γ -lactone (0.8 mmol) was dissolved in 6 ml of MeOH and 10% Pd on activated carbon (54 mg) was added. The mixture was stirred at room temperature under H₂ for 4 h. The mixture was filtered on Celite and the solvent was evaporated.

4.5.1. (*3R**,*4R**)- and (*3R**,*4S**)-1-Hydroxy-3-(2-hydroxyethyl)-4-methyl-2-pyrrolidinone 27a and 27b. The two isomers were obtained in admixture with the corresponding lactams 21a and 21b from the corresponding parent lactones 15a and 15b. For clarity sake the NMR values of the isomeric mixture are given separately for each isomer.

Compound **27a.** ¹H NMR (δ , ppm): 9.0 (1H, vbs, OH), 4.6 (1H, bs, OH), 3.75 (2H, m, CH₂OH), 3.72 (1H, m, H-5), 3.22 (1H, bd, *J*=9.1 Hz, H-5), 2.14 (2H, m, H-3 and H-4), 1.80 (2H, m, CH₂CH₂OH), 1.14 (3H, d, *J*=6.2 Hz, CH₃); ¹³C NMR (δ , ppm): 172.4 (s), 61.5 (t, CH₂OH), 55.2 (t, C-5), 47.8 (d, C-3), 33.0 (t, CH₂CH₂OH), 31.5 (d, C-4), 17.6 (q, CH₃).

Compound **27b.** ¹H NMR (δ , ppm): 9.0 (1H, vbs, OH), 4.6 (1H, bs, OH), 3.75 (2H, m, CH₂OH), 3.73 (1H, m, H-5), 3.21 (1H, dd, J_1 =9.5 Hz, J_2 =11.0 Hz, H-5), 2.66 (1H, m, H-3), 2.52 (1H, m, H-4), 1.70 (2H, m, CH₂CH₂OH), 1.04 (3H, d, J=7.0 Hz, CH₃); ¹³C NMR (δ , ppm): 172.0 (s), 61.5 (t, CH₂OH), 55.5 (t, C-5), 43.4 (d, C-3), 28.8 (t, CH₂CH₂-OH), 28.4 (d, C-4), 14.9 (q, CH₃).

4.5.2. (3*R**,4*R**) and (3*R**,4*S**)-1-Hydroxy-3-(2-hydroxyethyl)-3,4-dimethyl-2-pyrrolidinone 28a and 28b. The two isomers were obtained from the corresponding nitroalkylated lactones 16a and 16b. The crude reaction mixture was purified by flash chromatography (eluant: ethyl acetate, 70% yield) and the isomer 28b crystallized on standing at room temperature.

Compound **28a** (in admixture with **28b**). Oil, only a few signals were identified; ¹H NMR (δ , ppm): 3.36 (1H, m), 2.95 (1H, m), 2.23 (1H, m, H-4), 0.94 (3H, s, CH₃), 0.92 (3H, d, *J*=7.3 Hz, CH₃); ¹³C NMR (δ , ppm): 58.5 (t, CH₂OH), 53.6 (t, C-5), 44.7 (s, C-3), 38.5 (t, CH₂CH₂OH), 33.8 (d, C-4), 16.0 (q, CH₃ at C-3), 11.5 (q, CH₃ at C-4).

Compound **28b**. White solid, mp 90–92 °C, IR (cm⁻¹, neat): 3350 (OH), 1680 (NC=O); ¹H NMR (δ , ppm): 10.0 (1H,

vbs, NOH), 5.00 (1H, vbs, OH), 3.82 (1H, m, CHOH), 3.66 (2H, m, H-5 and CHOH), 3.29 (1H, t, J=9.0 Hz, H-5), 2.18 (1H, sextet, J=7.3 Hz, H-4), 1.74 (1H, m, CHCH₂OH), 1.50 (1H, dt, J_1 = J_2 =5.1 Hz, J_3 =13.9 Hz, CHCH₂OH), 1.18 (3H, s, CH₃), 1.04 (3H, d, J=7.3 Hz, CH₃); ¹³C NMR (δ , ppm): 174.0 (s), 58.3 (t, CH₂OH), 53.6 (t, C-5), 43.6 (s, C-3), 37.3 (d, C-4), 35.0 (t, CH₂CH₂OH), 21.3 (q, CH₃ at C-3), 12.0 (q, CH₃ at C-4). MS (m/z): 156 (M – OH^{¬+}, 12), 129 (18), 128 (34), 127 (11), 114 (100), 113 (33), 100 (14), 99 (68), 83 (21), 82 (39), 81 (13), 70 (22), 69 (66), 67 (65), 55 (80). Anal. Calcd for C₈H₁₅NO₃: C, 55.47; H, 8.73; N, 8.09. Found: C, 55.4; H, 8.29; N, 7.89.

4.5.3. (3*R**,4*S**)- and (3*R**,4*R**)-1-Hydroxy-3-(2-hydroxyethyl)-4-phenyl-2-pyrrolidinone 29a and 29b. Treatment of the crude reaction mixture, obtained by reduction of 17a and 17b, with ethyl acetate led to the crystallization of the isomer 29a (6% yield). The isomer 29b was recovered in 1:1 admixture with 29a (89% yield).

Compound **29a.** White solid, mp 171–173 °C; IR (cm⁻¹, nujol): 3210 (OH), 1685 (NC=O); ¹H NMR (CD₃OD, δ , ppm): 7.35 (5H, m, Ph), 3.85 (1H, t, J=8.6 Hz, H-5), 3.66–3.48 (3H, m, CH₂OH and H-5), 3.25 (1H, q, J=8.5 Hz, H-4), 2.71 (1H, m, H-3), 1.94 (1H, sextet, J=6.9 Hz, CHCH₂OH), 1.73 (1H, sextet, J=6.9 Hz, CHCH₂OH); ¹³C NMR (CD₃OD, δ , ppm): 173.3 (s), 141.2 (s), 129.9 (d), 128.6 (d), 128.4 (d), 60.6 (t, CH₂OH), 56.3 (t, C-5), 46.4 (d, C-3), 43.5 (d, C-4), 34.1 (t, CH₂CH₂OH); MS (*m*/*z*): 175 (19), 174 (13), 161 (93), 160 (43), 148 (16), 131 (25), 130 (16), 129 (16), 128 (11), 118 (26), 117 (100), 116 (33), 115 (48), 104 (67), 91 (48), 84 (30), 78 (17), 77 (25). Anal. Calcd for C₁₂H₁₅NO₃: C, 65.14; H, 6.83; N, 6.33. Found: C, 65.4; H, 6.71; N, 6.16.

Compound **29b.** ¹H NMR (CD₃OD, δ , ppm): 7.35 (5H, m, Ph), 3.99 (1H, t, J=8.2 Hz, H-5), 3.64 (2H, m, H-5 and H-4), 3.47 (2H, m, CH₂OH), 2.95 (1H, q, J=7.7 Hz, H-3), 1.59 (1H, sextet, J=7.2 Hz, CHCH₂OH), 1.25 (1H, m, CHCH₂OH); ¹³C NMR (CD₃OD, δ , ppm): 172.9 (s), 141.6 (s), 129.6 (d), 128.2 (d), 128.1 (d), 60.6 (t, CH₂OH), 55.8 (t, C-5), 42.6 (d, C-3), 40.0 (d, C-4), 30.5 (t, CH₂CH₂OH).

4.5.4. (*3R**,*4R**)- and (*3R**,*4S**)-1-Hydroxy-3-(2-hydroxyethyl)-3-methyl-4-phenyl-2-pyrrolidinone 30a and 30b. Treatment of the crude reaction mixture, obtained by reduction of 18a and 18b, with ethyl acetate led to the crystallization of the isomer 30a, while 30b was obtained by flash chromatography of the mother liquors (eluant: ethyl acetate).

Compound **30a**. 57% Yield, white solid, mp 175–178 °C; IR (cm⁻¹, nujol): 3200 (OH), 1681 (NC=O); ¹H NMR (CD₃OD, δ , ppm): 7.31 (5H, m, Ph), 3.90 (1H, dd, J_1 = 8.4 Hz, J_2 = 9.1 Hz, H-5), 3.80 (1H, dd, J_1 = 7.8 Hz, J_2 = 9.0 Hz, H-5), 3.77–3.66 (2H, m, CH₂OH) 3.63 (1H, t, J= 8.1 Hz, H-4), 1.80 (2H, m, CH₂CH₂OH), 0.74 (3H, s, CH₃); ¹³C NMR (CD₃OD, δ , ppm): 175.3 (s), 138.7 (s), 129.9 (d), 129.5 (d), 128.4 (d), 59.3 (t, CH₂OH), 52.5 (t, C-5), 47.1 (s, C-3), 45.0 (d, C-4), 40.1 (t, CH₂CH₂OH), 19.5 (q, CH₃); MS (m/z): 218 (M – OH^{¬+}, 1), 190 (18), 189 (18), 175 (100), 174 (37), 160 (23), 131 (43), 129 (23), 117 (28), 116 (19), 115 (27), 104 (77), 98 (57), 91 (62), 78 (15), 77 (18). Anal.

Calcd for C₁₃H₁₇NO₃: C, 66.36; H, 7.28; N, 5.95. Found: C, 66.5; H, 7.31; N, 5.81.

Compound **30b.** 12% Yield; oil, IR (cm⁻¹, film): 3200 (OH), 1681 (NC=O); ¹H NMR (CD₃OD, δ , ppm): 7.31 (5H, m, Ph), 3.92 (1H, dd, J_1 =7.9 Hz, J_2 =9.3 Hz, H-5), 3.81 (1H, dd, J_1 =7.7 Hz, J_2 =9.2 Hz, H-5), 3.55 (1H, m, CHOH), 3.35 (2H, m, CHOH and H-4), 1.57 (1H, ddd, J_1 = 6.4 Hz, J_2 =9.5 Hz, J_3 =13.9 Hz, CHCH₂OH), 1.30 (3H, s, CH₃), 1.15 (1H, ddd, J_1 =5.2 Hz, J_2 =9.4 Hz, J_3 =14.1 Hz, CHCH₂OH); ¹³C NMR (CD₃OD, δ , ppm): 175.2 (s), 138.7 (s), 129.7 (d), 129.5 (d), 128.6 (d), 58.9 (t, CH₂OH), 52.4 (t, C-5), 46.5 (s, C-3), 45.0 (d, C-4), 37.0 (t, CH₂CH₂OH), 23.3 (q, CH₃).

4.5.5. (3*R**,4*R**,5*R**)-1-Hydroxy-3-(2-hydroxyethyl)-5methyl-4-phenyl-2-pyrrolidinone 31b. Reduction of 19b afforded a 1:3 mixture of the lactam 25b and the cyclic hydroxamic acid 31b (80% yield), from which 31b was isolated by treatment with ethyl acetate.

Compound **25b** not separated: ¹H NMR (CDCl₃+CD₃OD, δ , ppm) (only a few signals were identified): 3.96 (1H, bq, J=5.7 Hz, H-5), 3.25 (1H, dd, $J_1=4.6$ Hz, $J_2=8.6$ Hz, H-4), 1.29 (3H, d, J=6.2 Hz, CH₃); ¹³C NMR (CDCl₃+ CD₃OD, δ , ppm): 180.1 (s), 138.8 (s), 128.6 (2d), 127.9 (2d), 127.1 (d), 61.0 (t, CH₂OH), 54.8 (d, C-5), 52.1 (d, C-4), 44.2 (d, C-3), 29.6 (t, CH₂CH₂OH), 20.6 (q, CH₃).

Compound 31b. White solid, mp 158 °C, 18% yield, IR (cm⁻¹, nujol): 3220 (OH), 1676 (NC=O); ¹H NMR (CDCl₃+CD₃OD, δ, ppm): 7.24 (3H, m, Ph), 7.14 (2H, m, Ph), 3.94 (1H, dq, J_1 =4.4 Hz, J_2 = J_3 = J_4 =6.2 Hz, H-5), 3.45 (2H, m, CH₂OH,), 3.20 (1H, dd, $J_1 = 4.4$ Hz, $J_2 =$ 8.8 Hz, H-4), 2.91 (1H, dt, $J_1 = 8.8$ Hz, $J_2 = J_3 = 6.2$ Hz, H-3), 1.47 (1H, m, CHCH₂OH), 1.33 (3H, d, *J*=6.2 Hz, CH₃), 1.24 (1H, m, CHCH₂OH); ¹³C NMR (CDCl₃+CD₃OD, δ , ppm): 171.3 (s), 138.3 (s), 128.7 (2d), 128.0 (2d), 127.3 (d), 60.8 (d, C-5), 60.7 (t, CH₂OH), 47.6 (d, C-4), 41.7 (d, C-3), 30.0 (t, CH₂CH₂OH), 17.1 (q, CH₃); MS (m/z): 217 (M- H_2O^{++} , 1), 216 (1), 176 (28), 175 (32), 148 (13), 133 (11), 132 (25), 131 (13), 119 (51), 118 (100), 117 (29), 116 (38), 92 (35), 77 (14); MS (m/z, 20 eV): 236 (MH⁺, 2), 235 (M⁺ 2). Anal. Calcd for C₁₃H₁₇NO₃: C, 66.36; H, 7.28; N, 5.95. Found: C, 66.5; H, 7.27; N, 5.90.

4.6. Reactions of α -methyl- γ -butyrolactone 1' with (*E*)-2-morpholinyl-1-nitro-1-phenylethene (5), (*E*)-1-(2methylthiophenyl)-2-pyrrolidinyl-1-nitroethene (6), (*Z*,*E*)-1-methylthio-4-pyrrolidinyl-2-nitrobutadiene (7) and (*E*)-1,1-dimethylthio-4-pyrrolidinyl-2nitrobutadiene (7'). General procedure for the Michael addition of nitroenamines to α -methyl- γ -butyrolactone zinc enolate 8'

To a solution of lithium diisopropylamide (1.5 M solution in THF) (2 mmol, 1.3 ml for the reaction with **5**, **7** and **7**'; 3.5 mmol, 2.3 ml for the reaction with **6**) in THF (1.3 ml), a solution of the lactone **1**' (0.162 g, 1.62 mmol) in 1.3 ml of THF was slowly added, at -78 °C. The mixture was stirred for 1 h at -78 °C. A solution of 1M ZnCl₂ (1.62 ml) was then added and the temperature was raised to -40 °C. After 1 h at this temperature, the solution was transferred to the

appropriate aminonitroalkene (0.81 mmol) dissolved in 5.2 ml of THF at -78 °C. The mixture was stirred at -78 °C for 2 h, the temperature was allowed to raise and the solution was kept overnight at room temperature. The reaction mixture was then quenched with 2 N HCl and extracted with dichloromethane. The organic phase was dried over anhydrous Na₂SO₄. After removal of the solvent, the residue was purified by flash chromatography (light petroleum/ethyl acetate 3:2).

4.6.1. (E)- and (Z)-4,5-Dihydro-3-methyl-3-(2-nitro-2phenylethenyl)-2(3H)-furanone 38. The crude reaction mixture (88% yield), whose composition was 85:15 in favour of the (E)-isomer, was treated with ether and ethanol at 0 °C. The major isomer (E)-38 was isolated as pure compound (30% yield), mp 90–92 °C; IR (cm⁻¹, nujol): 1778 (COO), 1672 (C=C), 1528 (NO₂). ¹H NMR (δ, ppm): 7.75 (1H, s, HC=C), 7.50 (3H, m, Ar-H), 7.32 (2H, m, Ar-H), 4.13 (2H, m, 2H-5), 2.10 (1H, dt, $J_1=9.3$ Hz, $J_2=$ 13.2 Hz, H-4), 1.60 (1H, ddd, $J_1 = 2.9$ Hz, $J_2 = 6.2$ Hz, $J_3 =$ 13.2 Hz, H-4), 1.41 (3H, s, CH₃); ¹³C NMR (δ, ppm): 177.8 (s, COO), 152.2 (s, C=CNO₂), 137.6 (d, CH=CNO₂), 130.7 (2d, o-Ar-H), 130.4 (2d, m-Ar-H), 129.1 (s), 128.6 (d, *p*-Ar–H), 65.1 (t, C-5), 43.2 (s, C-3), 34.6 (t, C-4), 23.5 (q, CH₃). MS (m/z): 247 (M⁺⁺, 0.01), 201 (72), 174 (15), 173 (100), 171 (14), 170 (13), 156 (13), 155 (10), 145 (23), 143 (20), 142 (43), 141 (38), 130 (12), 129 (65), 128 (67), 127 (25), 117 (17), 115 (49), 105 (58), 104 (18), 103 (52), 102 (31), 99 (10), 91 (40), 84 (32), 77 (73), 76 (28), 75 (15), 69 (17), 65 (10), 63 (17), 56 (11), 55 (29), 51 (66), 53 (13). Anal. Calcd for C₁₃H₁₃NO₄: C, 63.15; H, 5.30; N, 5.67. Found: C, 63.0; H, 5.19; N, 5.90.

The compound (Z)-38 was not isolated as pure isomer.

¹H NMR (δ , ppm): 7.43 (5H, m, Ar–H), 6.43 (1H, s, HC=C), 4.44 (1H, dt, $J_1 = 1.5$ Hz, $J_2 = J_3 = 9.2$ Hz, H-5), 4.32 (1H, ddd, $J_1 = 6.3$ Hz, $J_2 = 9.2$ Hz, $J_3 = 10.7$ Hz, H-5), 2.51 (1H, ddd, $J_1 = 9.2$ Hz, $J_2 = 10.7$ Hz, $J_3 = 12.8$ Hz, H-4), 2.35 (1H, ddd, $J_1 = 1.5$ Hz, $J_2 = 6.3$ Hz, $J_3 = 12.8$ Hz, H-4), 1.58 (3H, s, CH₃); ¹³C NMR (δ , ppm): 178.2 (s), 152.2 (s), 137.6 (d), 128.8 (d), 128.5 (d), 128.2 (d), 65.3 (t, C-5), 43.8 (s, C-3), 32.9 (t, C-4), 22.7 (q, CH₃); the singlet relative to C-1 of the phenyl ring was hidden under other signals.

4.6.2. (E)- and (Z)-4,5-Dihydro-3-methyl-3-[2-(2methylthiophenyl)-2-nitroethenyl]-2(3H)-furanone 39. The crude reaction mixture, whose composition was 67:33 in favour of the (E)-isomer, was purified on flash chromatography, 50% yield, yellow oil, IR (cm⁻¹, neat): 1775 (COO), 1520, (C=C-NO₂); ¹H NMR (δ, ppm): 7.81 (0.67H, s, C=CH), 7.73 (0.33H, s, C=CH), 7.47 (1H, m, Ar-H), 7.31 (1H, bd, J=7.7 Hz, Ar-H), 7.23 (2H, m, Ar-H), 4.23 (0.67H, dt, $J_1 = 9.0$ Hz, $J_2 = 2.6$ Hz, H-5), 4.09 (1.33H, m, H-5), 2.45 $(3H, s, SCH_3)$, 2.26 $(0.33H, dt, J_1 =$ 8.9 Hz, $J_2 = 13.1$ Hz, H-4), 2.16 (0.67H, dt, $J_1 = 9.1$ Hz, $J_2 = 12.8$ Hz, H-4), 1.78 (0.33H, ddd, $J_1 = 3.7$ Hz, $J_2 =$ 6.6 Hz, $J_3 = 13.1$ Hz, H-4), 1.48 (0.67H, ddd, $J_1 = 2.4$ Hz, $J_2 = 6.6$ Hz, $J_3 = 12.8$ Hz, H-4), 1.43 (0.90H, s, CH₃), 1.38 (2.1H, s, CH₃); ¹³C NMR (δ, ppm): (*E*)-**39**: 177.7 (s), 150.8 (s), 140.6 (s), 139.0 (d), 131.6 (d), 131.3 (d), 128.5 (s), 126.0 (d), 125.0 (d), 65.4 (t), 43.6 (s), 33.3 (t), 23.3 (q), 15.7 (q); (Z)-**39**: 178.0 (s), 150.2 (s), 140.1 (s), 139.2 (d), 132.2 (d),

131.3 (d), 127.9 (s), 126.1 (d), 125.0 (d), 65.4 (t), 43.3 (s), 34.6 (t), 22.9 (q), 15.8 (q); MS (m/z): 293 (M⁺, 28), 248 (12), 247 (49), 220 (16), 219 (100), 201 (12), 194 (36), 191 (12), 189 (17), 186 (14), 174 (15), 173 (36), 161 (14), 151 (41), 149 (25), 148 (13), 147 (28), 129 (14), 128 (12), 115 (18), 83 (11). Anal. Calcd for C₁₄H₁₅NO₄S: C, 57.32; H, 5.15; N, 4.77; S, 10.93. Found: C, 56.4; H, 5.19; N, 4.60.

4.6.3. *cis*- and *trans*-**3**-(**2-Hydroxyethyl**)-**3-methyl**-**5-phenyl-2-pyrrolidinone 40a and 40b.** The nitroalkenylated lactones **38** and **39** were reduced using Pd on carbon and Raney Nickel as a catalyst as reported in the general procedure.

A 2:3 mixture of lactams **40a** and **40b** were obtained in 70% yield. The two isomers were only partially separable by flash chromatography; yellow oil, IR (cm⁻¹, CHCl₃): 3420, 3153 (NH and OH), 1685 (NHC=O); MS (*m*/*z*): 220 (MH⁺, 33), 219 (M⁺⁺, 38), 202 (10), 191 (65), 190 (46), 175 (100), 174 (69), 173 (35), 172 (29), 160 (40), 158 (40), 147 (18), 146 (49), 145 (17), 144 (13), 143 (14), 132 (13), 131 (41), 130 (26), 129 (34), 128 (32), 120 (11), 117 (12), 115 (23), 106 (16), 104 (21), 91 (26), 84 (11), 77 (14). Anal. Calcd for C₁₃H₁₇NO₂: C, 71.21; H, 7.81; N, 6.39. Found: C, 70.5; H, 7.65; N, 6.30.

For sake of clarity the NMR values of the isomeric mixture are given separately for each isomer.

Compound **40a**. ¹H NMR (δ , ppm): 7.33 (5H, m, Ph), 6.23 (1H, bs, NH), 4.75 (1H, t, J=6.6 Hz, H-5), 4.09 (1H, bd, J=6.2 Hz, OH), 3.92 (1H, m, *CHOH*), 3.70 (1H, m, *CHOH*), 2.30 (1H, dd, J_1 =6.6 Hz, J_2 =12.6 Hz, H-4), 1.90 (2H, m, *CHCH*₂OH and H-4), 1.71 (1H, m, *CHCH*₂OH), 1.33 (3H, s, CH₃); ¹³C NMR (δ , ppm): 183.3 (s), 141.6 (s), 129.0 (2d), 128.0 (d), 125.7 (2d), 58.7 (t, CH₂OH), 55.8 (d, C-5), 46.6 (t, C-4), 44. 6 (s, C-3), 40.3 (t, *CH*₂CH₂OH), 21.8 (q).

Compound **40b.** ¹H NMR (δ , ppm): 7.38 (2H, m, Ph), 7.31 (3H, m, Ph), 5.94 (1H, bs, NH), 4.77 (1H, t, J=7.3 Hz, H-5), 3.94 (1H, m, CHOH), 3.75 (1H, m, CHOH), 2.97 (1H, bd, OH), 2.55 (1H, dd, J_1 =7.3 Hz, J_2 =13.0 Hz, H-4), 2.02 (1H, m, CHCH₂OH), 1.90 (1H, dd, J_1 =7.3 Hz, J_2 =13.0 Hz, H-4), 1.71 (1H, m, CHCH₂OH), 1.28 (3H, s, CH₃); ¹³C NMR (δ , ppm): 183.3 (s), 142.2 (s), 129.1 (2d), 128.1 (d), 125.8 (2d), 59.4 (t, CH₂OH), 55.0 (d, C-5), 46.8 (t, C-4), 43. 3 (s, C-3), 40.6 (t, CH₂CH₂OH), 23.5 (q).

4.6.4. 4,5-Dihydro-3-(2-(hydroxyimino)-2-(2-methylthiophenyl))ethyl-3-methyl-2(3H)-furanone 41. 51% Yield, yellow oil, IR (cm⁻¹) 3350 (OH), 1766 (COO), 1625 (C=N), 1587, 1560 (Ph), ¹H NMR (δ , ppm): 8.8 (1H, vbs, OH), 7.27 (4H, m, Ph), 4.25 (2H, m, 2H-5), 3.21 (2H, AB system, J=13.2 Hz, CH₂ of the chain), 2.44 (3H, s, SCH₃), 2.30 (1H, dt, J_1 = J_2 =7.6 Hz, J_3 =12.9 Hz, H-4), 1.90 (1H, ddd, J_1 =5.1 Hz, J_2 =7.3 Hz, J_3 =12.9 Hz, H-4), 1.17 (3H, s, CH₃); ¹³C NMR (δ , ppm): 181.0 (s), 156.7 (s), 137.7 (s), 135.7 (s), 129.4 (d), 129.1 (d), 126.6 (d), 124.9 (d), 65.3 (t, C-5), 42.4 (s, C-3), 34.6 (t), 34.3 (t), 22.9 (q, CH₃), 16.5 (q, CH₃); MS (m/z): 264 (M-CH₃⁺, 100), 262 (76), 247 (50), 232 (42), 229 (16), 203 (13), 188 (10), 164 (17), 152 (14).

Anal. Calcd for C₁₄H₁₇NO₃S: C, 60.19; H, 6.13; N, 5.01; S, 11.48. Found: C, 60.3; H, 6.25; N, 4.96.

4.6.5. (*E*,*Z*)- and (*Z*,*Z*)-4,5-Dihydro-3-methyl-3-(4-methylthio-2-nitrobutadienyl)-2(3*H*)-furanone 42. The product (*E*,*Z*)-42 was purified on column chromatography (52% yield). Oil, IR (cm⁻¹, CHCl₃): 1775 (COO), 1525 (C=C-NO₂); ¹H NMR (δ , ppm): 7.32 (1H, d, *J*=1.5 Hz, H-1 of the chain), 6.64 (1H, d, *J*=10.2 Hz, H-3 of the chain), 6.04 (1H, dd, *J*₁=10.2 Hz, *J*₂=1.5 Hz, H-4 of the chain), 4.32 (2H, m, 2H-5), 2.55 (1H, dt, *J*₁=8.5 Hz, *J*₂=13.1 Hz, H-4), 2.32 (1H, m, H-4), 2.36 (3H, s, SCH₃), 1.52 (3H, s, CH₃); ¹³C NMR (δ , ppm): (*E*,*Z*)-42: 177.7 (s), 148.5 (s), 141.3 (d), 141.2 (d), 113.1 (d), 65.4 (t, C-5), 43.4 (s, C-3), 33.3 (t, C-4), 23.3 (q, SCH₃), 15.7 (q, CH₃); Anal. Calcd for C₁₀H₁₃NO₄S: C, 49.37; H, 5.39; N, 5.76; S, 13.18. Found: C, 48.8; H, 5.49; N, 5.64. On standing in CDCl₃, the (*E*,*Z*)-isomer converted into its (*Z*,*Z*) isomer for an amount of 20%.

Compound (Z,Z)-42. ¹H NMR (δ , ppm) (a few signals were hidden under those of the (*E*)-isomer): 6.54 (1H, d, *J*= 11.0 Hz, H-3 of the chain), 6.29 (1H, s, H-1 of the chain), 6.09 (1H, d, *J*=11.0 Hz, H-4 of the chain), 4.44 (1H, dt, *J*₁=1.5 Hz, *J*₂=9.2 Hz, H-5), 2.41 (3H, s, SCH₃), 1.57 (3H, s, CH₃); ¹³C NMR (δ , ppm): 180.6 (s), 147.5 (s), 137.9 (d), 132.6 (d), 115.2 (d), 65.4 (t, C-5), 43.6 (s, C-3), 34.3 (t, C-4), 23.9 (q, SCH₃), 18.9 (q, CH₃).

4.6.6. (E)-4,5-Dihydro-3-methyl-3-(4,4-dimethylthio-2nitro)butadienyl-2(3H)-furanone 43. The product (70%) yield) was purified by flash chromatography (eluant: light petroleum:ethyl acetate, gradient), mp 82-84 °C; IR (cm⁻ film): 1770 (COO), 1666 (C=C), 1528, 1368 (NO₂); ¹H NMR (δ , ppm): 7.24 (1H, d, J = 1.8 Hz, H-1 of the chain), 5.96 (1H, d, J = 1.8 Hz, H-3 of the chain), 4.32 (2H, dd, $J_1 =$ 6.0 Hz, $J_2 = 7.7$ Hz, 2H-5), 2.48 (1H, dt, $J_1 = 7.7$ Hz, $J_2 =$ 12.8 Hz, H-4), 2.45 (3H, s, SCH₃), 2.35 (3H, s, SCH₃), 2.30 $(1H, dt, J_1 = 6.0 Hz, J_2 = 12.8 Hz, H-4), 1.51 (3H, s, CH_3);$ ¹³C NMR (δ, ppm): 177.7 (s), 149.8 (s), 147.9 (s), 137.0 (d, C-1 of the chain), 111.0 (d, C-3 of the chain), 65.3 (t, C-5), 43.2 (s, C-3), 35.9 (t, C-4), 22.6 (q, CH₃), 16.4 (q, SCH₃), 16.2 (q, SCH₃); MS (m/z): 289 (M⁺⁺, 3), 243 (100), 215 (48), 195 (11), 167 (22), 139 (11), 91 (11). Anal. Calcd for C₁₁H₁₅NO₄S₂: C, 45.66; H, 5.22; N, 4.84; S, 22.16. Found: C, 46.5; H, 5.35; N, 4.76.

4.6.7. 4,5-Dihydro-3-methyl-3-[(**4,4-dimethylthio-2-nitro)but-3-enyl]-2(3***H***)-furanone 44a,b.** To a solution of the nitrodiene **43** (0.04 g, 0.14 mmol) in MeOH (7 ml), 0.105 g of supported borohydride (2 mmol/g of Amberlyst A26, Aldrich product) was added under stirring and the reaction was monitored by TLC. After 15 min, the polymer was filtered off and washed with MeOH, the filtrated was evaporated and purified by flash chromatography. Compounds **44a** and **44b** (60 and 40%, respectively) were not separable by flash chromatography. For sake of clarity the NMR values of the isomeric mixture are given separately for each isomer.

Compound **44a**. ¹H NMR (δ , ppm): 5.52 (1H, d, J=8.8 Hz, H–C=C), 4.50 (1H, dt, J_1 = J_2 =8.8 Hz, J_3 =4.8 Hz, CH–NO₂), 4.26 (2H, m, 2H-5), 2.44 (1H, m), 2.35 (3H, s, SCH₃),

2.31 (3H, s, SCH₃), 2.03 (2H, m), 1.70 (1H, m), 1.29 (3H, s, CH₃).

Compound **44b**. ¹H NMR (δ , ppm): 5.57 (1H, d, J=8.4, H– C=C), 4.58 (1H, ddd, J_1 =3.0, J_2 =8.4, J_3 =10.6, CH– NO₂), 4.26 (2H, m, 2H-5), 2.44 (1H, m), 2.35 (3H, s, SCH₃), 2.30 (3H, s, SCH₃), 2.03 (2H, m), 1.70 (1H, m), 1.30 (3H, s, CH₃).

4.6.8. (*E*)-4,5-Dihydro-3-methyl-3-(4,4-dimethylthio)butadienyl-2(*3H*)-furanone 45. On standing in CDCl₃, 44a and 44b convert into the diene system 45. Oil. IR (cm⁻¹, CHCl₃): 1770 (COO), 1605, 1558 (C=C); ¹H NMR (δ , ppm): 6.78 (1H, dd, J_1 =15.7 Hz, J_2 =10.2 Hz, H-2 of the chain), 6.32 (1H, d, J=10.2 Hz, H-3 of the chain), 5.77 (1H, d, J=15.7 Hz, H-1 of the chain), 4.29 (2H, m, 2H-5), 2.40 (1H, m, H-4), 2.33 (3H, s, SCH₃), 2.32 (3H, s, SCH₃), 2.17 (1H, m, H-4), 1.40 (3H, s, CH₃); ¹³C NMR (δ , ppm): 168.0 (s), 133.6 (s), 133.2 (d), 129.4 (d), 126.9 (d), 65.2 (t), 45.3 (s), 35.8 (t), 23.3 (q), 17.4 (q), 16.7 (q); MS (m/z): 245 (MH⁺, 19), 244 (M⁺, 21), 231 (19), 230 (24), 229 (100), 213 (13), 212 (15), 201 (36), 185 (12), 183 (43), 155 (10), 138 (10), 137 (13), 123 (10), 91 (19), 77 (12). Anal. Calcd for C₁₁H₁₆O₂S₂: C, 54.06; H, 6.60; S, 26.24. Found: C, 55.0; H, 6.67.

Acknowledgements

The authors wish to thank the Universities of Trieste, Bologna, and Genova and M.I.U.R. (Rome) for financial support to this research (PRIN 2001-2003).

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Tetrahedron

Tetrahedron 60 (2004) 11029-11039

Kinetic versus thermodynamic access to imidazoisoindolones, benzimidazoisoindolones, and [1,4]diazepinoisoindolones: intramolecular nitrogen and π -aromatic trapping of *N*-acyliminium cation

Armelle Cul,^a Adam Daïch,^{a,*} Bernard Decroix,^a Gérard Sanz^b and Luc Van Hijfte^b

^aLaboratoire de chimie, URCOM, EA 3221, UFR des Sciences and Techniques de l'Université du Havre, B.P: 540, 25 rue Philippe Lebon, F-76058 Le Havre Cedex, France

^bJohnson & Johnson, Pharmaceutical Research & Development, Division of Janssen-Cilag, Campus de Maigremont, B.P: 615, F-27106 Val-de-Reuil Cedex, France

Received 24 May 2004; revised 28 June 2004; accepted 5 July 2004

Available online 5 October 2004

Abstract—Efficient assembly of substituted imidazo[2,1-*a*]isoindolones **I** is reported from suitable α , β -diamine **IV** (or corresponding β -nitroamine) and phthalic anhydride (1) in a three- or four-step sequence in good yields. The key step of this methodology is based on an intramolecular α -aza-amidoalkylation of the *N*-acyliminium species. Furthermore, when R₂ is an aromatic moiety a competing α -amidoalkylation took place and imidazo[2,1-*a*]isoindolones (or benzimidazo[2,1-*a*]isoindolones) **I** and/or isoindolo[1,4]benzodiazepines **III** were obtained under kinetic or thermodynamic control. The chemoselectivity of these transformations is also discussed. © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

The importance of the imidazo [2, 1-a] isoindolone skeleton is well recognized due to its significance as a subunit of a wide panel of synthetic pharmaceutical compounds. Some of these structures are patented and have been reported to possess a wide variety of biological activities as: psychostimulant, analgesic, anti-inflammatory, antifungal, antipyretic and hypertensive,¹ blood pressure lowering, spasmolytic, antitussive and tranquilizer properties,² and use in the treatment of rheumatism.³ Furthermore, they are useful intermediates in organic synthetic chemistry, especially in the elaboration of imidazo[2,1-*a*]isoindolol-based anorectics,^{1,4a,b} central nervous system (CNS) stimulants^{1,4c,d} and antidepressants,⁵ respectively. Finally, this class of compounds has also demonstrated to be highly effective plant growth regulating agents⁶ with effects on the plant budding process.⁷ More recently related benzimidazo[2,1-a]isoindolones have been reported and their biological activities evaluation show Batracylin comparable anti-tumor activities⁸ as well as their

ability to induce unscheduled DNA synthesis in rat hepatocytes.⁹

The traditional synthesis of imidazo[2,1-*a*]isoindolones involves the reaction between a 1,2-diamine and an aromatic or non aromatic keto acid, or equivalent, under azeotropic removal of water with¹⁰ or without¹¹ a catalytic amount of acid (i.e., *p*-toluenesulfonic acid). More recent methods include a reaction of a 1,2-diamine with phthalic anhydride (or dicarboxylic acid equivalent) followed by thermal cyclodehydration,^{9,12} a palladium catalyzed reaction via carbonylative cyclization between an α , β -diamine and 2-bromobenzaldehyde under controlled carbon monoxide pressure,¹³ an iminocyclization of *N*-azidoalkyl(or aryl) phthalimides^{14a} or the *N*-(*O*-aminoaryl)phthalimides^{14b} via intramolecular Aza–Wittig reaction, and finally a cationic cyclization involving *N*-acyliminium species.^{15,16}

2. Results and discussion

In our laboratory we are interested in the development of synthetic methodologies towards original aza-heterocyclic systems containing imidazole, benzimidazole and benzodiazepine moieties with promising pharmaceutical activities. In association with our recent reports dealing with

Keywords: Isoindole; Imidazole; Benzimidazole; [1,4]Diazepine; N-Acyliminium ion; α -Aza-amidoalkylation; Kinetic versus thermodynamic control.

^{*} Corresponding author. Tel.: +33 2 32 74 44 03; fax: +33 2 32 74 43 91; e-mail: adam.daich@univ-lehavre.fr

^{0040–4020/\$ -} see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2004.07.107



Scheme 1. Retrosynthetic scheme leading to imidazo(or benzimidazo)[2,1-a]isoindolones I and related isoindolo[1,4]benzodiazepines III.

intramolecular α -thio-amidoalkylation¹⁷ and α -oxo-amidoalkylation,¹⁸ we reasoned that a suitably substituted N-acyliminium precursor of type II (Scheme 1) could allow a facile approach to the tricyclic derivative (I) or tetracyclic (I and III) cores of the title targets. The cationic cyclization using a nitrogen atom as an internal nucleophile has been mentioned first during the synthesis of 9a-phenyl-5,6,6a,11tetrahydroisoindolo[2,1*a*]quinazoline-5,11-dione without isolation of the opened amide-hydroxylactam intermediate.^{16a} Some 10 years later it was used by Speckamp et al.^{16b} in the total synthesis of (\pm) -physostigmine $((\pm)$ -eserine), the principal alkaloid of Calabar bean. The essence of the few reports of this process concern its use in chiral version to access imidazo[2,1-a]isoindole-2,5-diones,¹⁵ imidazoisoquinolinones,^{16c} functionalized peptidomimetics,^{16d} and azabicyclo[4.4.0]alkane amino acids.¹⁹ To the best of our knowledge utilization of the present application in an intramolecular N-acyliminium mediated cyclization reaction, as depicted in Scheme 1, to form a central five- or sevenmembered ring as a cyclic N,N-acetal or [1,4]diazepine

structure, represents a novel illustration of this chemistry. In the intermediate **II**, cyclisation to the *N*-acyliminium cation may indeed be originated from intramolecular attack of either the nitrogen nucleophile or from the π -aromatic system, in case the latter is sufficiently activated.

As a starting point of our study, the α -hydroxylactam derivative **5a**, as *N*-acyliminium precursor, constituted a valuable target molecule. We expected to obtain it in a twostep sequence from phthalic anhydride (1) and *o*-nitroaniline (**3a**) by thermal amino-anhydride condensation in refluxing acetic acid for 24 h,²⁰ followed by selective sodium borohydride reduction of one of the carbonyl functions of the imide function under mild conditions (Scheme 2). In the case of borohydride reduction of **4a**, the process led to a mixture of compounds containing none of the desired amino-alcohol product **5a**. Instead, the isoindolobenzimidazole **8a** was identified as a minor product (13%), while the major compound proved to be 10*b*-methoxyisoindolobenzimidazole **8'a** (39% yield) which resulted from the methanol

Table 1. Yields of the intermediates 4a-c, 5a-c, 6a-c and the cyclized isoindolo[2,1-a]benzimidazole 8a,b,d and 8'a produced via Scheme 1

Products 4	Yield (%)	Products 5	Yield (%)	Products 6	Yield (%)	Products 8	Yield (%) ^a
4a 4b 4c 4d	89 91 91 91	5a 5b 5c 5d	b 65 	6a 6b 6c 6d	ь ь 85 89	8a 8b 8'a 8d	45–61 (82) ^c 63 (54) ^a 39 91

^a After recrystallization or chromatographic purification.

^b Product not isolated.

^c Isolated yields after reaction of diamines **7a**,**b** and 2-formylbenzoic acid (2).



Scheme 2. Reagents and conditions: (i) Phthalic anhydride (1), AcOH, reflux, 24 h; (ii) 1.5–3 equiv of NaBH₄, MeOH, -5 to 0 °C, 45 min to 1 h; (iii) Fe, 50% aq. AcOH, 60 °C, 4 h or H₂/Pd-C, AcOH–H₂O (v/v), 50 psi, 6 h; (iv) see Ref. 25a for product 3c and Ref. 25b for product 3d; (v) 1, toluene, reflux, NEt₃, Dean–Stark, 12 h; (vi) (a) acid hydrolysis during the work-up of the reduction of 5a,b with 10% HCl or 6 M H₂SO₄; (b) For 6c,d: TFA, CH₂Cl₂, rt, 4 h.

addition onto the imine function of **8a** under the protic acid activation conditions used during the reduction. Furthermore, in the presence of 2 equiv of additional nickel (II) chloride hexahydrate, the borohydride reduction reaction showed the same profile with however product **8a** obtained in larger quantity. Interestingly, treatment of **4a** with conjointly iron powder as a reductant and a 50% aqueous acetic acid as a proton source at 60 °C over 4 h,⁹ provided only 6-oxoisoindolo[2,1-*a*]benzimidazole (**8a**) in 44% yield after chromatography. The product **8a** was also obtained in 61% yield, without formation of the methoxy adduct **8'a**, from **4a** via catalytic hydrogenation at 50 psi over Pd–C 10% for 6 h in ethanol in the presence of acetic acid as a proton source. Interestingly, in this case no ethoxy adduct analogous to **8'a** could be observed (Table 1).

The ease of intramolecular α -aza-amidoalkylation by the amino-ketone cyclodehydration^{9,14} or the *N*-cyclization of the supposed amino-alcohol intermediate 6a, depending of the reduction sequence, caused us to investigate additional stabilization of these species. To this end, we considered the para-methyl substituted series based on the idea that an electron-donating methyl group would provide some increased stability of the nitro functions in 4b with respect to reducing agents. Interestingly, treatment of 4b using 3 equiv of NaBH₄ in methanol at 0 °C during 2 h (conditions ii outlined in Scheme 2), gave successfully the nitrohydroxylactam **5b** as a crystalline and stable product in 65% yield. This observation was in line with the fact that the reduction of the nitro-imide functionality into the nitroalcohol was already achieved by us on related products using sodium borohydride at lower temperature.²¹ Furthermore, reaction of 5b under conditions iii as outlined above (Scheme 2), gave directly 2-methyl-6-oxoisoindolo[2,1-a]



Scheme 3. Univocally 'one-pot' procedure access to 11-oxoisoindolo[2,1-*a*]-benzimidazole derivatives 8a and 8b.

benzimidaz-ole (**8b**) in comparable yields (63%) after chromatography and recrystallisation from dry ethanol.

The synthetic pathway leading to **8b** occurred through a cascade process by subsequent reduction of the nitro group of **5b** into the non isolated amino-alcohol **6b**, followed by loss of water, to form the *N*-acyliminium cation **A**, nucleophilic attack by the nitrogen atom (Scheme 2) and finally by spontaneous oxidation²²-elimination of the resulting unstable 10b,11-dihydro-6-oxoisoindolo[2,1-*a*]-benzimidazole (**B**). In the case of **8a**, there was no direct evidence for the nucleophilic attack of the nitrogen atom onto the *N*-acyliminium cation since the nitro-hydroxylactam **5a** was not isolated under the set of reduction conditions.

The structure of **8a,b** was equally confirmed chemically, as outlined in Scheme 3, by treatment of an equimolar amount of *o*-phenylenediamine (**7a**), or 2-amino-4-methyl-aniline (**7b**) and 2-formylbenzoic acid (**2**) under azeotropic removal of water according to the classical reported procedure.²³

It is interesting to note that this process in the case of the 4-methyl derivative **7b** proceeded in a highly regioselective manner. Scheme 4 demonstrates a plausible mechanism of the formation of the sole 2-methyl-6-oxoisoindolo[2,1-a]benzimidazole (8b). So, because the amine function at C_1 of **7b** is more nucleophilic than the one at C_2 , the aminoaldehyde condensation yielded the imine C which then cyclized to afford the imidazoline D. This latter, after an intramolecular cyclodehydration into B followed in an ultimate step by a spontaneous oxidation/elimination reaction afforded the corresponding imidazole derivative **8b**.²⁴ In contrast, the opposite profile was obtained with the o-phenylenediamine substrate 7e. In fact, the condensation of 2 with 7e took another course with a more reactive amine function at C₂, giving the imine E. This latter intermediate, after a sequential set of reactions as outlined for 8b, led to the product **8e** with the ester function at the C_8 position.²⁵

We decided next to explore another approach starting from o-phenylenediamine (**7a**) which was protected at one nitrogen atom with di-*t*-butyldicarbonate (Boc₂O) and acetic anhydride (Ac₂O) into carbamate $3c^{26a}$ and acetamide $3d^{26b}$ respectively (Scheme 2). The choice of these groups was based on two considerations: first, the NHBoc and NHAc groups were rarely engaged in the intramolecular cationic cyclization, and we expected that both groups,



Scheme 4. Plausible sequential mechanism leading to 2-methyl(or 3-methoxycarbonyl)-6-oxoisoindolo[2,1-a]benzimidazole 8a or 8b.

especially the *t*-Boc, could be removed easily during^{16d} or after cyclization to form the benzimidazole derivative 8a,b, respectively. Second, both the *t*-butyloxy carbamate and acetyl groups are electron-withdrawing groups which could render the amino group less nucleophilic leading to the amino-alcohols 6c,d as isolable intermediates. Thus, treatment of 3c,d with 1 equiv of phthalic anhydride (1) in toluene at reflux in the presence of a catalytic amount of triethylamine over 12 h gave the imides 4c and 4d in 85 and 91% yields, respectively. These imides were then converted regioselectively to the N-acyliminium ions precursors 6c,d by borohydride reduction as described above for 5b in 85 and 89% yield.

According to our previous reports showing that trifluoroacetic acid (TFA) and acetic acid (AcOH) are good catalysts for the intramolecular α -amidoalkylation and α -heteroamidoalkylation, treatment of hydroxy-lactam 6c with TFA in CH₂Cl₂ for 24 h or neat TFA for 4 h at room temperature afforded in both cases 8a in about 73% yield. This product, which was identical to the one obtained from the nitro-imide 4a (Scheme 2), resulted from an intramolecular cyclization of the *N*-acyliminium intermediate of type A with a nitrogen atom as nucleophile in parallel with Boc deprotection. We do have some evidence that the *t*-Boc deprotection occurs subsequently to the cyclization due to different deprotection kinetics, but this was not confirmed. Taking into account that TFA is a standard deprotecting agent for the t-Boc



Scheme 5. Sequential set leading to α -hydroxylactam precursors 11a-c. Reagents and conditions: (i) Toluene, cat. NEt3, reflux, Dean-Stark, 12 h; (ii) 3 equiv of NaBH₄, MeOH, -5 to 0 °C, aq. HCl in ethanol, 1 h.

group, we decided to use a weaker acid, such as AcOH, for the cyclization process, hoping to be able to maintain the t.Boc protective group. Thus, α -hydroxy-lactam **6c** upon treatment with neat AcOH for 8 h or in CH₂Cl₂ for 24 h at reflux gave again 8a in comparable yields of about 67%. In order to avoid the deprotection process, the aza-acylated α -hydroxylactam **6d** (R₂=Ac) was used as *N*-acyliminium ion precursor. In fact, the acetyl derivative 6d, after treatment with neat TFA at rt over 4 h gave 11-acyl-6oxoisoindolo[2,1-a]benzimidazo-le (8d) in 80% yield after recrystallization from ethanol.

Having established that the intramolecular a-aza-amidoalkylation route is effective for the preparation of the isoindolo[2,1-a]benzimidazole derivatives **8a–d**, we decided to elaborate another class of α -hydroxylactam precursors in which a nitrogen atom bear another nucleophile such as an aromatic or heteroaromatic system. Thus, as depicted in Scheme 5, the requisite α -hydroxy-lactams 11a-c were obtained in a two-step sequence using classical procedures. Imides 10a-c were readily prepared by amineanhydride condensation from commercial diamines 9a-c and phthalic anhydride (1) as indicated for 4c,d in 76, 85 and 86%, respectively. The reaction was accelerated by adding dry triethylamine in catalytic quantity. Regioselective reduction of imides 10a-c was accomplished with a large excess of sodium borohydride in methanol at -5 to 0 °C. In all cases, a regular addition of an ethanolic hydrogen chloride solution was necessary as already mentioned elsewhere for related structures, ^{17,18,21} and after 1 h of the reaction, α -hydroxylactams **11a**-c were isolated in, respectively, 80, 88 and 82% yield.

In the first set of cationic cyclizations in this series, substrate 11a was chosen as a model for the N-acyl-iminium ion precursor. So, the subjection of α -hydroxy-lactam 11a to weak AcOH (method A), weak TFA (method B) or catalytic p-toluenesulfonic acid (PTSA) (method C) in CH₂Cl₂ at room temperature for 12 h (Table 2, entry 1, 2 or 3) afforded only 11-phenyl-6-oxoisoindolo[2,1-a]benzimidazole (12a) in 65, 80 or 79% yield, respectively. This product resulted invariably from an intramolecular aza-cationic cyclization of the endocyclic N-acyliminium ion intermediate of type **H** (Scheme 6).

On the basis on our precedent work in this field and to avoid the intramolecular α -aza-amidoalkylation process to the

Table 2. Isoindolo[2,1-a]benzimidazole and corresponding isoindolo[1,4]dibenzodiazepine derivatives 12a,b and 13a,b produced via Scheme 6

	Reactant	Quantity ^a mmol	Conditions	Method	Product	Yield (%) ^b
1	11a	2.0	4 equiv of AcOH, CH ₂ Cl ₂ , rt, 12 h	Α	12a	65
2	11a	2.5	4 equiv of TFA, CH ₂ Cl ₂ , rt, 12 h	В	12a	80
3	11a	3.0	Catalytic PTSA, CH ₂ Cl ₂ , rt, 12 h	С	12a	79
4	11a	4.0	Neat AcOH, rt, 12 h	D	13a	69
5	11a	4.0	Neat TFA, rt, 12 h	E	13a	74
6	12a	3.0	Neat AcOH, reflux, 24 h	F	13a	83
7	12a	6.0	Neat TFA, reflux, 24 h	G	13a	81
8	12a	5.0	Catalytic PTSA, toluene, reflux, 24 h	Н	13a	90
9	11b	5.0	Catalytic PTSA, CH ₂ Cl ₂ , rt, 12 h	С	12b	82
10	11b	5.0	Neat TFA, rt, 12 h	E	13b ^c	93
11	12b	4.5	Neat TFA, reflux, 24 h	G	13b	89

^a The reaction was conducted on 2–6 mmol of reactant under stirring. For entries 6–8, the isoindolo[2,1-a]benzimidazole 12a was used as starting material. ^b Isolated yield after purification by recrystallization or chromatography on silica gel column.

^c A trace of the kinetic product **12b** were detected by ¹H NMR analysis and its quantity not exceed 5% of the products mixture.



Scheme 6. Reagents and conditions: (i) 2, toluene, cat. NEt₃, reflux, Dean-Stark, 12 h; (ii) See text and Table for others procedures.

detriment of others, different reaction conditions were considered. So, the intramolecular arylation leading to isoindolo[1,4]dibenzodiazepine product **13a** as a single product occurred when neat AcOH (method D, Table 2, entry 4, 69%) or neat TFA (method E, Table 2, entry 5, 74%) was used as a proton source. Furthermore, to check the reversibility of the aza-cyclization reaction, taking into account that cyclic *N*,*N*-aminal **12a** could generate the *N*-acyliminium ion under acidic influence according to previously observations,^{18c,27} we envisaged the treatment of acetal **12a** with neat AcOH (method F, Table 2, entry 6), neat TFA (method G, Table 2, entry 7) or catalytic amount of PTSA in toluene (method H, Table 2, entry 8) at reflux. Under these conditions all reactant **12a** disappeared (monitored by TLC) and the expected isoindolodibenzodiazepine product **13a** was obtained in good yield (81–90%).

The formation of these cycles 12a and 13a in acidic medium seems to proceed by invoking the kinetic vs thermodynamic control using the formal N-acyliminium ion H as intermediate (Schemes 6, 7). In fact, under mild conditions (methods A, B and C), the cationic intermediate H lead to isoindolo[2,1-a]benzimidazole derivative **12a** as the sole reaction product under kinetic process (fast reaction). In contrary, under stronger acidic conditions (neat acid at room temperature; methods D and E) and/or higher temperatures, the same intermediate provided in an alternate pathway isoindolo[1,4]dibenzodiazepine 13a under thermodynamic control (slow reaction). The difference in reactivity between the two pathways can easily be contributed to the fact that the formation of 13a requires higher activation energy due to the loss of aromaticity in the transition state. In addition, the compound 13a proves to be about 20 kcal/mol more stable than its corresponding imidazoline 12a according to AM1 calculations (86.29 kcal/mol vs 106.66 kcal/mol in the case of $R-R_1=Ph$), what clearly explains the complete conversion from 12a to 13a under more drastic conditions. These results confirm that the formation of the CH-N linkage of the kinetic product 12a is reversible depending on

the acidic activation. Under stronger conditions, cleavage of the CH–N bond in 12a led back to the *N*-acyliminium ion congener **H** which in turn led to the diazepine compound 13a as the thermodynamically more stable product.

To establish the generality and versatility of this process we studied the effect of varying the nucleophilicity of the nitrogen atom which is interacts with the *N*-acyliminium ion during the cyclization process. For this purpose, two kinds of *N*-acyliminium ion precursors were considered; **11b** and **11c** in which the nitrogen atom nucleophilicity is altered with respect to the one in the reactant **11a**, and which bear a benzene or pyridine ring as a competing π -nucleophile or aza-nucleophile, respectively.

So, treatment of hydroxylactam **11b** according to method C (Table 2, entry 9), gave exclusively 1-phenyl-5-oxoisoindolo[2,1-*a*]imidazole (**12b**) under the kinetic control in 82% yield (Schemes 6 and 7). Similarly, reaction under conditions of method E, **11b** led to the cyclized thermodynamic [1,4]benzodiazepine product **13b** in excellent yield (Table 2, entry 10, 93%). During this reaction, the 1-phenyl-5-oxoisoindolo[2,1-*a*]imidazole structure **12b** was detected as a minor product but its yield did not exceed 5% in all cases.²⁸ As for the diazepine derivative **13a**, the product **13b** was also isolated in 89% yield starting from the kinetic product **12b** under conditions outlined in Table 2 (method G, entry 11) in a one pot procedure involving *N*-acyliminium ion intermediate **H** (Schemes 6 and 7).

In contrast, the hydroxylactam **11c**, with a reduced nucleophilicity with respect to both the N and π -aromatic nucleophilic centers due to the nitro group, afforded in all attempts with differing acidic and/or temperature conditions, the imidazoline derivative **12c** in comparable yields (85%). These results suggest that the formation of the azepine type structures **13** and **14** requires sufficiently activated aromatic systems (Scheme 8).²⁹



Scheme 7. Kinetic vs thermodynamic scheme leading to imidazole and diazepine derivatives 12a,b and 13a,b. Reagents and conditions: (i) H₂, 10% Pd–C, AcOH–HCl, 50 °C, 6.5 h (see Ref. 30 for more details); (ii) 2, toluene, cat. NEt₃, reflux, Dean–Stark, 12 h.



Scheme 8. Possible structures which could be resulted form a pyridine attack on the *N*-acyliminium ion intermediate **H**.

As a direct method for structural confirmation of imidazole and benzimidazole derivatives, the condensation of *o*-formylbenzoic acid (2) with diamines **7a,b** and **12a–c** under azeotropic removal of water was used successfully, thus providing isoindoloimidazoles **12b** (82%) and **12c** (96%), and isoindolobenzimidazoles **8a** (69%), **8b** (54%), and **12a** (89%) identical to the compounds obtained by the *N*-acyliminium sequence. Furthermore, the structure elucidation of these products as well as all compounds intermediates was based on their spectroscopic data (IR, ¹H NMR and ¹³C NMR including NOE Difference and DEPT experiments) as well as their microanalyses.

The NMR data of the imidazole product **8a** has been previously reported in some details.^{9,14b} For related compounds **8'a**, **8b**, and **8e**, the ¹H NMR data have a same profile globally as for **8a** with however a side chain signal corresponding to a methoxy (δ =3.79 ppm), methyl (δ =2.47 ppm), and methoxycarbonyl (δ =3.95 ppm) groups, respectively, with an additional NH signal at δ =10.48 ppm for **8'a**. No signals for the proton in position C_{4b} of the isoindole system, appearing classically at about δ =6.0 ppm, was observed in theses cases.

In the imidazoisoindolones structures **12a**, **12b** and **12c**, the angular protons appear as singlets at $\delta = 7.24$, 6.12, and 6.61 ppm, respectively. These latter absorb downfield compared to the same protons of their hydroxylactams congeners **11a** ($\delta = 6.22$ ppm), **11b** ($\delta = 5.78$ ppm), and **11c** ($\delta = 5.83$ ppm), respectively. The same fact, was also observed for the diazepinoisoindolones in comparaison to their precursors, with however a little difference on the chemical shift values which are $\Delta \delta = +0.09$ ppm and $\Delta \delta = +0.05$ ppm in favour to **13a** and **13b** to the detriment of **12a** and **12b**, respectively. These observations are in agreement with the fact that C–N and C–C bonds, formed during the cyclization process, have not the same effect on the CH angular absorbance. These results are also in agreement with previous reports on analogous compounds.^{15–18}

Furthermore, the key feature in the ¹³C NMR spectra of 8'a and 8a-d or 8e, was the appearance of fourteen or fifteen signals, respectively, in the aromatic region. One of these

disappears in the corresponding DEPT program spectra as the consequence of the aza-amidoalkylation cyclization process. Interestingly, if the quaternary angular carbon of the C=N bond appears at δ =156.4 ppm (**8a**), δ =152.4 ppm (**8b**), and δ =155.0 ppm (**8e**) in comparable chemical shift values reported in the literature for related products,^{14b} the one of **8'a** appeared downfield with more significant deshielding at δ =166.7 ppm. This fact is due to the proximity of three heteroatoms which belong to amide, amine and etheroxide functionalities, respectively. Especially diagnostic was the differentiation between the formed five-membered ring products **8b** and **12a–c** and the cyclized seven-membered ring ones **13a,b**.

In fact, for the isoindolobenzimidazole **8b** the appearance of the C=N signal at δ =152.4 ppm constitutes the consequence of the intramolecular cyclization of **4b**. This value is similar to those obtained for related structures.¹⁴ Interestingly, it can be seen that the aza-cyclization process of **11a,b** into **12a,b** induces a weak variation in the carbon angular absorbance which is $\Delta\delta$ =+1.2 ppm and $\Delta\delta$ =+7.3 while the π -cyclization into **13a,b** of **11a,b** shifted dramatically the absorbance of the angular carbon to higher fields. In these case, an important deshielding of $\Delta\delta$ =+21.2 ppm and $\Delta\delta$ =+20.5 ppm was observed.

3. Conclusion

In summary, we have shown that *N*-acyliminium ion precursors **6a–d** could be generated in two pathways from nitro-hydroxylactams **5a,b** or corresponding amino-imides **4c,d** by regioselective reduction processes using iron/acetic acid or sodium borohydride/methanol, respectively. The *N*-acyliminium ion in turn furnished via an intramolecular α -aza-amidoalkylation with a nitrogen atom as nucleophile various and new 6-oxoisoindolo[2,1-*a*]benzimidazole products **8a–d** in good yields. In some cases, the amino-hydroxylactam intermediates **6** were not isolated but cyclized directly into product **8**.

Similar α -hydroxylactams **11a**,**b**, under the same process, produced efficiently under thermodynamic control the expected isoindolo[1,4]benzodiazepines 13a,b in good yields and excellent regiocontrol. These latter were also obtained starting from isoindoloimidazoles 12a,b, which turned out to be the kinetically formed products from cyclization of the nitrogen onto the iminium ion species produced by acid treatment of the α -hydroxylactams **11a**,**b**. The hydroxylactam **11c** only gave rise to the imidazoline derivative 12c, clearly demonstrating the influence of electronic factors in the π -aromatic attack. Finally, the structures of the isoindoloimidazole derivatives 8a,b and 12a,b as well as 12c was confirmed chemically by an alternative synthesis from o-formylbenzoic acid (2) and corresponding diamines 7a,b or 9a-c in an one pot procedure. For full exploitation of this route which provides a novel synthesis of imidazole, benzimidazole and diazepine derivatives and which is short, facile, general and more competitive, further work is currently underway to enlarge the scope of this application by accessing a wider variety of these structures.

4. Experimental

4.1. General

All melting points were measured on a Boetius micro hotstage and are uncorrected. The infrared spectra of solids (potassium bromide) and liquids (neat) were recorded on a Perkin Elmer FT-IR paragon 1000 spectrometer. The ¹H and ¹³C NMR spectra were recorded on a Bruker AC-200 (200 MHz) instrument in deuteriochloroform unless other indicated solvent and chemical shifts (δ) are expressed in ppm relative to TMS as internal standard. Ascending thin layer chromatography was performed on precoated plates of silica gel 60 F 254 (Merck) and the spots visualised using an ultraviolet lamp or iodine vapour. Mass spectral measurements were recorded on a AEI MS 902 S spectrophotometer. The elemental analyses were carried out by the microanalysis laboratory of INSA, F-76130 Mt. St. Aignan, France.

4.2. General procedure for synthesis of imides (4a,b)

A mixture of powdered phthalic anhydride (1, 1.48 g, 10 mmol) and *o*-nitroaniline (**3a**, 1.38 g, 10 mmol) or 4-methyl-2-nitroaniline (**3b**, 1.52 g, 10 mmol) in 100 mL of glacial acetic acid was heated at reflux for 24 h. After cooling, the precipitate formed was collected by filtration, washed with cyclohexane, diethyl ether and air dried. The resulting products were purified by recrystallization from ethanol to give imides **4a** and **4b** as yellow needles.

4.2.1. *N*-(*o*-Nitrophenyl)phthalimide (4a). This product was isolated as yellow solid in 89% yield; mp=190–195 °C (lit., ³¹ mp=202–203 °C); IR (KBr) v 3093, 1715, 1525 cm⁻¹; ¹H NMR (CDCl₃) δ 7.52 (d, 1H, 1H_{benzene}, *J*=7.8 Hz), 7.61 (t, 1H, 1H_{benzene}, *J*=7.8, 7.0 Hz), 7.74–7.83 (m, 3H, 1H_{benzene} and 2H_{phthalimide}), 7.93–7.98 (m, 2H, 2H_{phthalimide}), 8.18 (d, 1H, 1H_{benzene}, *J*=7.8 Hz); ¹³C NMR (CDCl₃) δ 124 (2CH), 125.6 (Cq), 125.9 (CH), 129.8 (CH), 131.0 (CH), 131.9 (Cq), 134.3 (CH), 134.9 (2CH), 145.8 (2Cq), 166.5 (2CO); Anal. Calcd for C₁₄H₈N₂O₄ (268.22): C, 62.69; H, 3.01; N, 10.44. Found: C, 62.48; H, 2.95; N, 10.22.

4.2.2. *N*-(4'-Methyl-*o*-nitrophenyl)phthalimide (4b). This product was isolated as yellow needles in 91% yield; mp= 181 °C (lit.,³² mp=187 °C); IR (KBr) v 3082, 1716, 1534, 1383 cm⁻¹; ¹H NMR (DMSO d₆) δ 2.50 (s, 3H, CH₃), 7.67 (d, 1H, 1H_{benzene}, *J*=7.6 Hz), 7.77 (d, 1H, 1H_{benzene}, *J*=7.6 Hz), 7.77 (d, 1H, 1H_{benzene}, *J*=7.6 Hz), 7.94–8.08 (m, 2H, 1H_{benzene} and 4H_{phthalimide}); ¹³C NMR (DMSO d₆) δ 19.8 (CH₃), 121.4 (Cq), 123.4 (2CH), 125.0 (CH), 130.3 (CH), 130.6 (Cq), 134.6 (CH), 134.8 (2CH), 140.6 (Cq), 144.6 (2Cq), 165.8 (2CO); Anal. Calcd for C₁₅H₁₀N₂O₄ (282.25): C, 63.83; H, 3.57; N, 9.93. Found: C, 63.66; H, 3.28; N, 9.87.

4.3. General procedure for synthesis of imides (4c,d) and (10a–c)

A mixture of α , β -diamine **4c**, **4d**, **9a**, **9b**, or **9c** (10 mmol), phthalic anhydride (1) (1.48 g, 10 mmol) and triethylamine (0.5 mL, 3.6 mmol) in toluene (50 mL) was refluxed with a Dean–stark apparatus for 12 h. The reaction mixture was

cooled, then concentrated under reduced pressure. The residue was dissolved into dichloromethane, washed with 5% hydrochloric acid solution then with a 5% sodium hydrogenocarbonate solution. The organic layer was dried over magnesium sulfate, concentrated under vacuo, and recrystallization of the residue gave the expected imides 4c,d or 10a-c in good yields.

4.3.1. *N*-(*o*-*Tert*-butoxycarbonylamidophenyl)phthalimide (4c). This product was isolated as white crystals in 91% yield; mp=269 °C (decomposition); IR (KBr) v 3417, 3034, 2990, 1731, 1714 cm⁻¹; ¹H NMR (CDCl₃) δ 1.58 (s, 9H, 3CH₃), 7.12–7.53 (m, 4H, 4H_{benzene}), 7.61–7.82 (m, 2H, 2H_{phthalimide}), 7.90–7.98 (m, 2H, 2H_{phthalimide}), 8.15 (s broad, 1H, NH); ¹³C NMR (CDCl₃) δ 28.9 (3CH₃), 80.1 (Cq), 123.4 (Cq), 124.8 (2CH), 125.6 (CH), 127.3 (CH), 129.6 (Cq), 131.1 (CH), 142.3 (CH), 134.4 (2CH), 140.6 (2Cq), 164.8 (CO), 167.2 (2CO); Anal. Calcd for C₁₉H₁₈N₂O₄ (338.36): C, 67.44; H, 5.36; N, 8.28. Found: C, 67.18; H, 5.22; N, 8.31.

4.3.2. *N*-(*o*-Acetamidophenyl)phthalimide (4d). This product was isolated as white crystals in 91% yield; mp = 200 °C; IR (KBr) v 3244, 3038, 1720, 1657 cm⁻¹; ¹H NMR (CDCl₃) δ 2.01 (s, 3H, COCH₃), 7.24–7.31 (m, 2H, 1H_{benzene} and NH, exchangeable with D₂O), 7.36–7.58 (m, 2H, 2H_{benzene}), 7.77–7.86 (m, 3H, 1H_{benzene} and 2H_{phthalimide}), 7.89–7.96 (m, 2H, 2H_{phthalimide}); ¹³C NMR (CDCl₃) δ 24.3 (CH₃), 124.0 (Cq), 124.2 (2CH), 125.9 (CH), 126.0 (CH), 128.6 (CH), 129.7 (CH), 131.8 (Cq), 133.9 (2Cq), 134.8 (2CH), 167.4 (2CO), 168.6 (CO); Anal. Calcd for C₁₆H₁₂N₂O₃ (280.28): C, 68.56; H, 4.32; N, 9.99. Found: C, 68.39; H, 4.25; N, 10.05.

4.3.3. *N*-(*o*-Phenylaminophenyl)phthalimide (10a). This product was isolated as a yellow solid in 76% yield; mp= 200 °C (ethanol); IR (KBr) ν 3320, 3006, 1710 cm⁻¹; ¹H NMR (CDCl₃) δ 6.04 (s, 1H, NH, exchangeable with D₂O), 7.08–7.27 (m, 4H, 4H_{benzene}), 7.37–7.80 (m, 5H, 5H_{benzene}), 8.01–8.12 (m, 2H, 2H_{phthalimide}), 8.17–8.26 (m, 2H, 2H_{phthalimide}); ¹³C NMR (CDCl₃) δ 118.5 (2CH), 121.4 (CH), 121.7 (CH), 123.3 (Cq), 123.9 (CH), 124.2 (2CH), 129.6 (2CH), 129.7 (CH), 130.2 (CH), 132.3 (2Cq), 134.8 (2CH), 140.8 (Cq), 143.4 (Cq), 167.7 (2CO); Anal. Calcd for C₂₀H₁₄N₂O₂ (314.34): C, 76.42; H, 4.49; N, 8.91. Found: C, 76.09; H, 4.25; N, 8.71.

4.3.4. *N*-(*o*-Phenylaminoethyl)phthalimide (10b). This product was isolated as a white solid in 85% yield; mp= 100 °C (ethanol); IR (KBr) ν 3345, 3010, 2989, 1712; ¹H NMR (CDCl₃) δ 3.42 (t, 2H, CH₂, *J*=6.3 Hz), 3.96 (t, 2H, CH₂, *J*=6.3 Hz), 6.57–6.72 (m, 3H, 3H_{benzene}), 7.08–7.19 (m, 2H, 2H_{benzene}), 7.66–7.75 (m, 2H, 2H_{phthalimide}), 7.77–7.87 (m, 2H, 2H_{phthalimide}); ¹³C NMR (CDCl₃) δ 37.7 (CH₂), 43.3 (CH₂), 112.9 (2CH), 117.9 (CH), 123.7 (2CH), 129.6 (2CH), 132.3 (2Cq), 134.4 (2CH), 147.9 (Cq), 169 (2CO); Anal. Calcd for C₁₆H₁₄N₂O₂ (266.29): C, 72.16; H, 5.30; N, 10.52. Found: C, 72.02; H, 5.15; N, 10.41.

4.3.5. *N*-(2-(5'-Nitropyridin-2'-ylamino)ethyl)phthalimide (10c). This product was isolated as a white solid in 86% yield; mp=191 °C (ethanol); IR (KBr) ν 3242, 3005, 2950, 1710, 1503, 1396; ¹H NMR (DMSO d₆) δ 3.37–3.76 (m, 4H, 2CH₂), 6.42 (d, 1H, H_{pyridine}, J=8.6 Hz), 7.79–7.85 (m, 2H, 2H_{phthalimide}), 8.04 (d, 1H, H_{pyridine}, J=8.6 Hz), 8.12–8.24 (m, 2H, 2H_{phthalimide}), 8.71 (s, 1H, H_{pyridine}), 8.90 (t, 1H, NH, J=2.4 Hz, exchangeable with D₂O); ¹³C NMR (DMSO d₆) δ 36.9 (CH₂), 38.5 (CH₂), 108.5 (CH), 122.6 (2CH), 131.4 (CH), 131.6 (Cq), 133.9 (2CH), 134.4 (2Cq), 146.1 (CH), 161.4 (Cq), 167.8 (2CO); Anal. Calcd for C₁₅H₁₂N₄O₄ (312.09): C, 57.69; H, 3.87; N, 17.94. Found: C, 57.51; H, 3.66; N, 17.63.

4.4. General procedure for reduction of imides (4a–d) and (10a–c)

To a mixture of 5 mmol of imide 4a (4b, 4c, 4d, 10a, 10b or 10c) in dry methanol (40 mL) at -5 to 0 °C was added sodium borohydride (283-565 mg, 7.5-15 mmol) by portions during 5 min. To this mixture was added 5 drops of ethanolic hydrochloric acid solution (prepared by addition of nine drops of concentrated hydrochloric acid into 9 mL of dry ethanol) at regular intervals of 10 min. The reaction was monitored by TLC using CH₂Cl₂ as eluent (CH₂Cl₂/MeOH (9/1) in the case of 10c). After the end of the reaction (45 min to 1 h), the excess of sodium borohydride was decomposed by careful addition of cold water (15 mL) and 10% hydrochloric acid until pH 4. Sodium hydrogen carbonate was added and the solvent was evaporated. The resulting residue was triturated with water and dichloromethane and the organic layer was separated, washed with water, brine, dried and concentrated in vacuo. The resulting product was purified by chromatography on silica gel column or by recrystallization to give in appreciable yield **5b**, **6c**, **6d**, **8a**, **8'a**, **11a**, **11b** or **11c**, respectively.

4.4.1. 3-Hydroxy-2,3-dihydro-2-(*o*-nitrophenyl)isoind**ol-1-one** (5a). This product was not isolated but react immediately in situ to give the cyclised product **8**'a and/or **8**a.

4.4.2. 6-Oxoisoindolo[2,1-*a*]benzimidazole (8a). This product was obtained in a range of 45-61% yield and have same characteristics to that reported in literature (lit., ⁹ mp > 290 °C, 39% yield).

4.4.3. 10b-Methoxy-10b,11-dihydrobenzo[4,5]imidazo[2,1-*a***]isoindol-6-one (8'a). This product was isolated as orange crystals in 39% yield after chromatography on silica gel column using a mixture of dichloromethane/ cyclohexane (7:3) as eluent; mp=128 °C; IR (KBr) v 3246, 3023, 2958, 1718 cm⁻¹; ¹H NMR (CDCl₃) \delta 3.80 (s, 3H, OCH₃), 7.16 (t, 1H, 1H_{isoindole},** *J***=7.8, 7.0 Hz), 7.46–7.57 (m, 3H, 3H_{benzene}), 7.65 (t, 1H, 1H_{isoindole},** *J***=7.8, 7.0 Hz), 7.91 (d, 1H, 1H_{benzene},** *J***=7.0 Hz), 8.17 (d, 1H, 1H_{isoindole},** *J***=7.8 Hz), 8.84 (d, 1H, 1H_{isoindole},** *J***=7.8 Hz), 10.45 (s, 3H, NH, exchangeable with D₂O); ¹³C NMR (CDCl₃) \delta 52.8 (OCH₃), 122.6 (CH), 123.8 (CH), 125.9 (CH), 127.3 (CH), 129.2 (Cq), 130.6 (CH), 130.7 (CH), 132.6 (CH), 134.7 (Cq), 136.3 (CH), 136.8 (Cq), 137.9 (Cq), 166.8 (CO), 167.9 (Cq); Anal. Calcd for C₁₅H₁₂N₂O₂ (252.27): C, 71.42; H, 4.79; N, 11.10. Found: C, 71.25; H, 4.64; N, 11.01.**

4.4.4. 3-Hydroxy-2,3-dihydro-2-(4'-methyl-*o*-nitrophen-yl)isoindol-1-one (5b). This product was isolated as a yellow solid in 65% yield; mp=158 °C; IR (KBr) v 3351, 3082, 1690, 1532, 1383 cm⁻¹; ¹H NMR (CDCl₃) δ 2.43 (s, 3H, CH₃), 3.59 (d, 1H, OH, J=11.7 Hz, exchangeable with D₂O), 6.17 (d, 1H, CH, J=11.7 Hz), 7.35–7.55 (m, 3H, 1H_{benzene} and 2H_{isoindole}), 7.56–7.75 (m, 3H, 1H_{benzene} and 2H_{isoindole}), 7.84 (d, 1H, 1H_{benzene}, J=10.6 Hz); ¹³C NMR (CDCl₃) δ 20.9 (CH₃), 83.9 (CH), 123.8 (CH), 124.6 (CH), 125.9 (CH), 127.5 (Cq), 130.2 (CH), 130.6 (CH), 130.8 (Cq), 134.0 (CH), 135.4 (CH), 139.7 (Cq), 145.4 (Cq), 146.7 (Cq), 166.9 (CO); Anal. Calcd for C₁₅H₁₂N₂O₄ (284.27): C, 63.38; H, 4.25; N, 9.85. Found: C, 63.21; H, 4.08; N, 9.71.

4.4.5. 3-Hydroxy-2,3-dihydro-2-(*o-tert*-butoxycarbonylamidophenyl)isoindol-1-one (6c). This product was isolated as a white solid in 85% yield; mp=188 °C; IR (KBr) v 3446, 3280, 3010, 2970, 1736, 1663 cm⁻¹; ¹H NMR (DMSO d₆) δ 1.39 (s, 9H, 3CH₃), 6.23 (d, 1H, OH, J=7.1 Hz, exchangeable with D₂O), 6.77 (d, 1H, CH, J= 7.1 Hz), 7.14 (t, 1H, 1H_{benzene}, J=7.8, 7.0 Hz), 7.34 (t, 2H, 1H_{benzene} and 1H_{isoindole}, J=7.8, 7.8 Hz), 7.58–7.88 (m, 5H, 2H_{benzene} and 3H_{isoindole}), 8.35 (s, 1H, NH, exchangeable with D₂O); ¹³C NMR (DMSO d₆) δ 28.1 (3CH₃), 79.5 (Cq), 83.6 (CH), 122.2 (CH), 122.9 (CH), 123.5 (CH), 123.7 (CH), 127.5 (Cq), 138.0 (CH), 129.2 (CH), 129.6 (CH), 131.4 (Cq), 132.6 (Cq), 136.6 (Cq), 145.8 (Cq), 152.9 (CO), 165.8 (CO); Anal. Calcd for C₁₉H₂₀N₂O₄ (340.37): C, 67.05; H, 5.92; N, 8.23. Found: C, 67.12; H, 5.81; N, 8.09.

4.4.6. 3-Hydroxy-2,3-dihydro-2-(*o*-acetamidophenyl)isoindol-1-one (6d). This product was isolated as a white solid in 89% yield; mp=185 °C (ethanol); IR (KBr) v 3439, 3247, 3006, 2923, 1689, 1673 cm⁻¹; ¹H NMR (DMSO d₆) δ 1.98 (s, 9H, CH₃), 6.17 (s, 1H, CH), 6.65 (s broad, 1H, OH, exchangeable with D₂O), 7.11–7.34 (m, 3H, 3H_{benzene}), 7.46–7.56 (m, 1H, 1H_{benzene}), 7.60–7.62 (m, 2H, 2H_{isoindole}), 7.79 (d, 1H, 1H_{isoindole}, *J*=7.8 Hz), 8.03 (d, 1H, 1H_{isoindole}), *J*=7.8 Hz),_{isoindole}), 8.48 (s, 1H, NH, exchangeable with D₂O); ¹³C NMR (DMSO d₆) δ 24.3 (CH₃), 84.6 (CH), 123.5 (2CH), 124.1 (CH), 125.0 (CH), 127.7 (CH), 128.4 (CH), 129.7 (CH), 131.0 (Cq), 132.7 (CH), 135.8 (Cq), 141.3 (Cq), 144.9 (Cq), 170.1 (CO), 174.9 (CO); Anal. Calcd for C₁₆H₁₄N₂O₃ (282.29): C, 68,07; H, 5,00; N, 9,92. Found: C, 67,98; H, 4,81; N, 9,84.

4.4.7. 3-Hydroxy-2,3-dihydro-2-(*o*-phenylaminophenyl)-1*H*-isoindol-1-one (11a). This product was isolated as a yellow solid in 80% after recrystallization from ethanol; mp = 73 °C; IR (KBr) ν 3381, 3333, 3038, 1703 cm⁻¹; ¹H NMR (CDCl₃) δ 6.22 (s, 1H, CH), 7.17–7.36 (m, 9H, 9H_{benzene}), 6.65–7.64 (m, 12H, 9H_{benzene} and 3H_{isoindole}), 7.80–7.89 (m, 1H, 1H_{isoindole}); ¹³C NMR (CDCl₃) δ 84.5 (CH), 118.5 (2CH), 120.5 (CH), 121.2 (CH), 122.5 (CH), 123.8 (CH), 124.2 (CH), 127.0 (Cq), 128.3 (CH), 129.1 (CH), 129.6 (2CH), 130.5 (CH), 131.3 (Cq), 133.1 (CH), 141.5 (Cq), 143.6 (Cq), 144.3 (Cq), 167.2 (CO); Anal. Calcd for C₂₀H₁₆N₂O₂ (316.35): C, 75.93; H, 5.10; N, 8.86. Found: C, 75.80; H, 5.02; N, 8.65.

4.4.8. 3-Hydroxy-2,3-dihydro-2-(*o*-**phenylaminoethyl**)-**1***H***-isoindol-1-one (11b).** This product was isolated as a white solid in 88% after recrystallization from ethanol; mp=76 °C; IR (KBr) ν 3363, 3009, 2988, 1709 cm⁻¹; ¹H NMR (CDCl₃) δ 3.35 (t, 2H, CH₂, *J*=5.5 Hz), 3.51–3.67 (m, 1H, CH₂), 3.72–3.88 (m, 1H, CH₂), 4.09 (s large, 1H, NH, exchangeable with D₂O), 5.78 (s, 1H, CH), 6,58 (d, 2H, J=7.8 Hz), 6,69 (t, 1H, J=7.1 Hz), 7.12 (t, 2H, J=7.8 Hz), 7.39–7.57 (m, 3H, 3H_{isoindole}), 7.66 (d, 1H, 1H_{isoindole}, J=7.1 Hz); ¹³C NMR (CDCl₃) δ 40.3 (CH₂), 43.4 (CH₂), 83 (CH), 113.5 (2CH), 118.3 (CH), 123.5 (2CH), 129.6 (2CH), 130.1 (CH), 131.5 (Cq), 132.7 (CH), 144.1 (Cq), 147.9 (Cq), 168.6 (CO); Anal. Calcd for C₁₆H₁₆N₂O₂ (268.31): C, 75.93; H, 5.10; N, 8.86. Found: C, 75.80; H, 5.02; N, 8.65.

4.4.9. 3-Hydroxy-2,3-dihydro-2-[2-(5'-nitropyridin-2'-ylamino)ethyl]-1*H***-isoindol-1-one (11c). This product was isolated as a yellow solid in 82% after recrystallization from dry ethanol; mp=120 °C (decomposition); IR (KBr) \nu 3467, 3307, 3093, 2935, 1704, 1504, 1337 cm⁻¹; ¹H NMR (CDCl₃, this product is no stable in the solution) \delta 3.08 (s broad, 1H, NH, exchangeable with D₂O), 3.28–3.71 (m, 3H, CH₂–CH₂), 4.13–4.18 (m, 1H, CH₂–CH₂), 3.08 (s broad, 1H, OH, exchangeable with D₂O), 5.83 (s, 1H, CH), 6.37 (d, 1H, 1H_{pyridine},** *J***=9.4 Hz), 7.29–7.43 (m, 1H, 1H_{benzene}), 7.47–7.57 (m, 3H, 3H_{benzene}), 7.99 (d, 1H, 1H_{pyridine},** *J***=9.4 Hz), 8.58 (s, 1H, 1H_{pyridine}); Anal. Calcd for C₁₅H₁₄N₄O₄ (314.30): C, 57.32; H, 4.49; N, 17.83. Found: C, 57.18; H, 4.27; N, 17.65.**

4.5. Procedure for the reductive cyclization of nitrohydroxylactam 5b

To a solution of 6 mmol of nitro-hydroxylactam **5b** in dry methanol (40 mL) at -5 to 0 °C was added sodium borohydride (679 mg, 18 mmol) by portions during 10 min. To this mixture was added 5 drops of ethanolic hydrochloric acid solution (as prepared above) at regular intervals of 5 min. The reaction was monitored by TLC using CH₂Cl₂ as eluent. After the end of the reaction (2 h), the excess of sodium borohydride was decomposed by careful addition of cold water (15 mL) and 10% hydrochloric or H₂SO₄ acid until pH 4. After sodium hydrogen carbonate was added and the solvent was evaporated. The resulting residue was triturated with water and dichloromethane and the organic layer was separated, washed with water, brine, dried and concentrated in vacuo.

4.5.1. 2-Methyl-6-oxoisoindolo[2,1-*a*]benzimidazole (8b). This product was obtained as a white powder in 63% after recrystallization from absolute ethanol; mp=238 °C; IR (KBr) ν 3025, 2952, 1719, 1640 cm⁻¹; ¹H NMR (CDCl₃) δ 2.47 (s, 3H, CH₃), 7.1 (d, 1H, 1H_{benzene}, *J*=8.1 Hz), 7.41 (s, 1H, 1H_{benzene}), 7.49 (d, 1H, 1H_{benzene}, *J*=8.1 Hz), 7.57–7.94 (m, 4H, 4H_{isoindole}); ¹³C NMR (CDCl₃) δ 22.8 (CH₃), 115.8 (CH), 116.6 (CH), 125.4 (CH), 131.2 (Cq), 131.4 (2CH), 131.7 (CH), 132.7 (Cq), 133.3 (CH), 134.6 (Cq), 138.4 (Cq), 139.6 (Cq), 152.4 (Cq), 170.4 (CO); Anal. Calcd for C₁₅H₁₀N₂O (234.26): C, 76.91; H, 4.30; N, 11.96. Found: C, 76.81; H, 4.09; N, 12.05.

4.6. General procedure for acid cyclization of hydroxylactams (6c,b) and (11a–c)

The procedure is general and concerns the use of neat or weak TFA, AcOH or catalytic PTSA at rt or reflux without or with a solvent as CH_2Cl_2 or toluene. For more details see Table 2. A stirred solution of 6 mmol of amino-hydroxylactams **6c,d** (or hydroxy-lactams **11a–c**) in 10 mL of appropriate acid or catalytic amount (in the case of PTSA) was left to react at room temperature or reflux in the presence or not of the solvent (For details see Table 2). After the required time, the solvent was evaporated in vacuo, and the residue was diluted with dichloromethane (10 mL) and treated with a saturated solution of hydrogenocarbonate. After separation, the organic layer was washed with water, brine, dried and concentrated in vacuo. The resulting residue was purified either by recrystallisation from ethanol or through a silica gel column.

4.6.1. 6-Oxoisoindolo[2,1-*a*]benzimidazole (8c). This product is identical to 8a described above.

4.6.2. 11-Acetyl-10b,11-dihydrobenzo[4,5]imidazo[2,1*a*]isoindol-6-one (8d). This product was isolated as a white-yellow crystals after chromatography on silica gel column using a mixture of chloroform/cyclohaxane (2:3) as eluent in 91% yield; mp=168 °C; IR (KBr) v 3021, 2943, 1732, 1667 cm⁻¹; ¹H NMR (CDCl₃) δ 2.52 (s, 3H, CH₃), 6.93 (s, 1H, CH), 7.05–7.19 (m, 3H, 3H_{benzene}), 7.49–7.64 (m, 2H, 1H_{benzene} and 1H_{isoindole}), 7.66–7.75 (m, 1H, 1H_{isoindole}), 7.85 (d, 1H, 1H_{isoindole}, *J*=7.0 Hz), 8.25 (d, 1H, 1H_{isoindole}, *J*=7.0 Hz); ¹³C NMR (CDCl₃) δ 25.3 (CH₃), 79.4 (CH), 113.1 (CH), 117.2 (CH), 124.1 (CH), 124.5 (CH), 124.9 (CH), 128.3 (CH), 130.3 (CH), 132.1 (Cq), 132.8 (Cq), 133.7 (CH), 136.0 (Cq), 145.0 (Cq), 169.3 (CO), 171.6 (CO); Anal. Calcd for C₁₆H₁₂N₂O₂ (264.28): C, 72.72; H, 4.58; N, 10.60. Found: C, 72.59; H, 4.32; N, 10.28.

4.6.3. 11-Phenyl-6(10*bH***)-oxoisoindolo[2,1-***a***]benzimidazole (12a).** This product was obtained as white crystals in 79% after recrystallization from ethanol; mp=208–210 °C (decomposition); IR (KBr) ν 3015, 1709 cm⁻¹; ¹H NMR (CDCl₃) δ 7.03–7.12 (m, 1H, H benzene), 7.24 (s, 1H, CH), 7.26–7.42 (m, 2H, 2H_{benzene}), 7.64–8.03 (m, 9H, 9H_{benzene}), 8.28–8.37 (m, 1H, 1H_{benzene}); ¹³C NMR (CDCl₃) δ 83.3 (CH), 108.6 (CH), 116.9 (CH), 120.4 (CH), 124.2 (CH), 124.8 (2CH), 125.3 (CH), 125.6 (CH), 126.4 (CH), 130.2 (2CH), 130.4 (CH), 131.5 (Cq), 133.1 (CH), 133.9 (Cq), 141.9 (Cq), 144.1 (Cq), 146.3 (Cq), 170.7 (CO); Anal. Calcd for C₂₀H₁₄N₂O (298.34): C, 80.52; H, 4.73; N, 9.39. Found: C, 80.42; H, 4.59; N, 9.19.

4.6.4. 5-Phenyl-4*b***,5,6,7-tetrahydroimidazo**[**2,1***-a*]**iso-indol-9-one** (**12b**). This product was obtained as a white powder in 82% after recrystallization from absolute ethanol; mp = 143 °C; IR (KBr) ν 3005, 2992, 1711 cm⁻¹; ¹H NMR (CDCl₃) δ 3.39–3.68 (m, 2H, CH₂), 3.77–3.92 (m, 1H, CH₂), 4.26–4.39 (m, 1H, CH₂), 6.12 (s, 1H, CH), 6.87–7.28 (m, 3H, 3H_{benzene}), 7.29–7.41 (m, 2H, 2H_{benzene}), 7.44–7.54 (m, 2H, 2H_{isoindole}), 7.71–7.86 (m, 2H, 2H_{isoindole}); ¹³C NMR (CDCl₃) δ 42.4 (CH₂), 53.3 (CH₂), 75.7 (CH), 114.7 (2CH), 119.3 (CH), 124.6 (2CH), 129.7 (2CH), 130.0 (CH), 132.9 (CH), 133.3 (Cq), 145.7 (Cq), 146.8 (Cq), 173.1 (CO); Anal. Calcd for C₁₆H₁₄N₂O (250.30): C, 76.78; H, 5.64; N, 11.19. Found: C, 76.84; H, 5.48; N, 11.27.

4.6.5. 2-(1-Benzyl-1*H*-benzimidazol-2-yl)-3-hydroxy-2,3dihydroisoindol-1-one (12c). This product was obtained as a yellow orange powder in 85% after recrystallization from absolute ethanol; mp=234 °C; IR (KBr) ν 3079, 2963, 1699, 1513, 1327 cm⁻¹; ¹H NMR (DMSO d₆) δ 3.50–3.82 (m, 2H, CH₂), 3.87–3.99 (m, 1H, CH₂), 4.33 (dd, 1H, CH₂, J=6.4, 10.7 Hz), 6.61 (s, 1H, CH), 6.78 (d, 1H, 1H_{pyridine}, J=9.1 Hz), 7.56–7.82 (m, 3H, 3H_{isoindole}), 8.30–8.44 (m, 2H, 2H_{isoindole} and 1H_{pyridine}), 9.26 (d, 1H, 1H_{pyridine}, J=2.7 Hz); ¹³C NMR (DMSO d₆) δ 42.8 (CH₂), 50.4 (CH₂), 76.2 (CH), 108.6 (CH), 124.3 (CH), 128.2 (CH), 131.2 (CH), 133.2 (Cq), 134.0 (2CH), 136.6 (Cq), 145.3 (Cq), 146.8 (CH), 159.5 (Cq), 173.1 (Cq); Anal. Calcd for C₁₅H₁₂N₄O₃ (296.28): C, 60.81; H, 4.08; N, 18.91. Found: C, 60.70; H, 4.10; N, 18.69.

4.6.6. 5,15*b***-Dihydro-11***H***-isoindolo**[**2,1***-d*]**dibenzo**[*b*,*f*]-[**1,4**]**diazepin-11-one** (**13a**). This product was obtained as a yellow powder in 81–91% after recrystallization from absolute ethanol; mp=248 °C; IR (KBr) ν 3346, 3013, 1709 cm⁻¹; ¹H NMR (CDCl₃) δ 6.13 (s, 1H, CH), 6.61– 6.83 (m, 3H, 3H_{benzene}), 6.88–7.16 (m, 4H, 4H_{benzene}), 7.32 (s, 1H, NH, exchangeable with D₂O), 7.42–7.68 (m, 1H, 1H_{isoindole}), 8.2 (m, 1H, 1H_{isoindole}); ¹³C NMR (CDCl₃) δ 63.3 (CH), 118.9 (CH), 119.3 (CH), 119.5 (CH), 120.1 (CH), 122.7 (CH), 124.2 (CH), 124.4 (CH), 125.3 (CH), 126.1 (Cq), 126.2 (CH), 127.2 (Cq), 128.6 (CH), 129.1 (CH), 131.1 (Cq), 133.1 (Cq), 135.2 (Cq), 142.0 (Cq), 144.1 (Cq), 166.9 (CO); Anal. Calcd for C₂₀H₁₄N₂O (298.34): C, 80.52; H, 4.73; N, 9.39. Found: C, 80.41; H, 4.59; N, 9.21.

4.6.7. 5,6,7,13b-Tetrahydroisoindolo[2,1-d][1,4]benzodiazepin-9-one (13b). This product was obtained as a white solid in 89-93% after chromatography over silica gel column using a mixture of CH₂Cl₂/hexane (9.5/0.5) as eluent and recrystallization from absolute ethanol; mp= 139 °C (lit.,²⁹ mp=135–137 °C); IR (KBr) ν 3336, 3013, 2986, 1709 cm⁻¹; ¹H NMR (CDCl₃) δ 3.03–3.19 (m, 1H, CH₂), 3.49-3.68 (m, 2H, CH₂), 4.0 (s br, 1H, NH, exchangeable with D₂O), 4.25-4.39 (m, 1H, CH₂), 5.73 (s, 1H, CH), 6.77-6.92 (m, 2H, 2H_{benzene}), 7.03 (d, 1H, 1H_{benzene}, J=7.1 Hz), 7.16-7.25 (t, 1H, 1H_{benzene}, J= 7.1 Hz), 7.47–7.68 (m, 3H, 3H_{isoindole}), 7.94 (d, 1H, 1H_{isoindole}, J=7.2, Hz); ¹³C NMR (CDCl₃) δ 44.3 (CH₂), 47.5 (CH₂), 62.5 (CH), 120.7 (CH), 121.8 (CH), 124.6 (CH), 125.4 (CH), 127.3 (Cq), 127.7 (CH), 128.9 (CH), 129.3 (CH), 131.2 (CH), 133.5 (Cq), 143.1 (Cq), 150.0 (Cq), 168.6 (CO); Anal. Calcd for C₁₆H₁₄N₂O (250.30): C, 76.78; H, 5.64; N, 11.19. Found: C, 76.59; H, 5.61; N, 11.08.

4.7. General procedure for acidic transformation of isoindoloimidazoles (12a,b) into isoindolobenzodiazepines (13a,b)

4.7.1. Products 13a and 12b. All chemicals and physicals characteristics of these products are identical to that described above in Sections 4.6.6 and 4.6.7, respectively.

4.8. General procedure for keto acids/diamines cyclodehydration into isoindoloimidazole derivatives (8a,b,e), and (12a-c)

To a solution of 2-carboxybenz-aldehyde (2, 1.5 g, 10 mmol) in anhydrous toluene with or without addition of PTSA (50 mg), was added 10 mmol of diamine 7a, 7b, 7e, 9a, 9b or 9c. The reaction mixture was refluxed for 12 h with a Dean–Stark apparatus. After cooling and removal of the solvent in vacuo, the residue was purified by

recrystallization from ethanol to yield the expected isoindolobenzimidazoles **8a** (82%), **8b** (54%), **8e** (51%) or **12a** (91%), or isoindoloimidazoles **12b** (82%) or **12c** (96%), respectively, as crystals.

4.8.1. Products 8a,b and 12a–c. All chemicals and physicals characteristics of these products **8a,b** and **12a–c** are identical to that reported above.

4.8.2. 3-Methoxycarbonyl-6-oxoisoindolo[**2**,**1**-*a*]**benzimidazole** (**8e**). This product was isolated as white crystals in 51% yield; mp=269 °C; IR (KBr) ν 3009, 1709, 1698, 1635 cm⁻¹; ¹H NMR (DMSO d₆) δ 3.95 (s, 3H, CH₃), 7.64–7.98 (m, 6H, 3H_{benzene} and 3H_{isoindole}), 8.12 (s, 1H, H_{isoindole}); ¹³C NMR (DMSO d₆) δ 53.0 (CH₃), 115.7 (CH), 118.0 (CH), 124.1 (CH and Cq), 130.5 (CH), 131.1 (CH and Cq), 131.4 (CH), 132.1 (CH), 134.0 (Cq), 140.1 (Cq), 143.3 (Cq), 155.1 (Cq), 167.7 (CO), 169.4 (CO); Anal. Calcd for C₁₆H₁₀N₂O₃ (278.27): C, 69.06; H, 3.62; N, 10.07. Found: C, 68.98; H, 3.51; N, 10.01.

Acknowledgements

We acknowledge the Johnson and Johnson Pharmaceutical Research and Development, Division of Janssen–Cilag, for scientific and financial support of this program. We thank also the Region of 'Haute Normandie' for a 'Regional-Industrial Graduate Fellowship' (BRI), 1998–2001, attributed to A. Cul. We also thank C. Meyer, from J&J PRD, a division of Janssen-Cilag, for the AM1 calculations.

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