



# Microcalorimetric and spectrographic studies on host–guest interactions of $\alpha$ -, $\beta$ -, $\gamma$ - and M $\beta$ -cyclodextrin with resveratrol

Hui Li, Xiangyu Xu, Min Liu, Dezhi Sun\*, Linwei Li\*

College of Chemistry and Chemical Engineering, Liaocheng University, Liaocheng 252059, Shandong Province, People's Republic of China

## ARTICLE INFO

### Article history:

Received 17 May 2010

Received in revised form 8 July 2010

Accepted 9 July 2010

Available online 16 July 2010

### Keywords:

Resveratrol

Cyclodextrin

Isothermal titration calorimetry

Inclusion complex

Microcalorimetry

## ABSTRACT

Thermal effects of inclusion processes of  $\alpha$ -,  $\beta$ -,  $\gamma$ - and M $\beta$ -cyclodextrin with resveratrol (RES) in aqueous solutions were determined by isothermal titration calorimetry (ITC) with nanowatt sensitivity at the temperature of 298.15 K. Standard enthalpy changes, stoichiometry and equilibrium constants of the inclusion complexes were derived from the direct calorimetric data utilizing nonlinear simulation. The thermodynamic parameters were discussed in the light of weak interactions between the host and the guest molecules combining with UV spectral message. The results indicate that all of the complexes formed in the aqueous solutions are in 1:1 stoichiometry. The binding processes of  $\alpha$ -,  $\beta$ - and M $\beta$ -cyclodextrin with the guest are mainly driven by enthalpy, while that of  $\gamma$ -cyclodextrin with the drug is driven by both enthalpy and entropy.

© 2010 Elsevier B.V. All rights reserved.

## 1. Introduction

Resveratrol (3,5,4'-trihydroxy-*trans*-stilbene), a polyphenol, has been found in a wide variety of about 70 plant species, including grapes, mulberries, and peanuts [1], which could protect against a range of illnesses and diseases, e.g., cardiovascular diseases [2–4], cancer [5,6], viral attacks [7], neurodegenerative diseases [8] and ageing [9]. Furthermore, it could increase lifespan [10] and physical activity of diverse species [11,12]. Various studies have also indicated that resveratrol shows several interesting biological effects such as antioxidant [13–15], anti-inflammatory [16] and anti-proliferative activities [17]. However, due to its low water solubility, resveratrol cannot be used for oral administration. This limitation could be overcome by the formation of inclusion complexes of the drug with cyclodextrins [18].

Cyclodextrins (CDs) are the most prominent host molecules used in supramolecular chemistry up to now [19,20], whose molecular structures are shown in Scheme 2 as cyclic oligosaccharides composed of glucopyranose units and can be characterized by a truncated cone structure with hydrophobic interior and hydrophilic exterior [21]. They have been used in many fields such as pharmaceutical industry, food technology, biotechnology, cosmetics and catalysis [22–24]. The most notable feature

of cyclodextrins is their ability to form host–guest inclusion complexes with a very wide range of compounds by molecular complexation, so that they can enhance the solubility and stability of some guest molecule [25–27].

Isothermal titration calorimetry (ITC) is an extremely powerful and highly sensitive technique that is capable of determining the interaction of reacting species in solution and has hitherto been used with great success in the study of interaction between biomolecules in dilute aqueous solutions both from thermodynamic and from kinetics points of view [28]. In a word, it is of significance to study the interactions of CDs with resveratrol in aqueous system using the method of ITC. In the present work, heat effects caused by interactions of several CDs with resveratrol were determined by ITC and thermodynamic parameters were calculated based on the calorimetric data and were discussed according to the weak interactions between the host and the guest molecules. In order to verify the results acquired from ITC, UV spectra were also determined and analyzed.

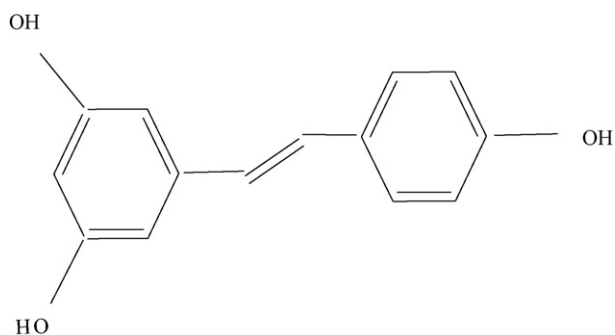
## 2. Experimental

### 2.1. Materials

Resveratrol was purchased from Xi'an Sino-Herb Biotechnology Company and the stated purity was better than 98%, which was dried under reduced pressure at the temperature of 350 K in order to remove the impurity (mainly ethanol). The molecular structure of resveratrol is provided in Scheme 1.  $\alpha$ - and  $\gamma$ -CD were obtained from Aldrich Company and were also

\* Corresponding authors. Tel.: +86 635 8230614; fax: +86 635 8239196.

E-mail addresses: [sundezhisdz@163.com](mailto:sundezhisdz@163.com), [dzsun@eyou.com](mailto:dzsun@eyou.com) (D. Sun), [lilinwei@lcu.edu.cn](mailto:lilinwei@lcu.edu.cn) (L. Li).

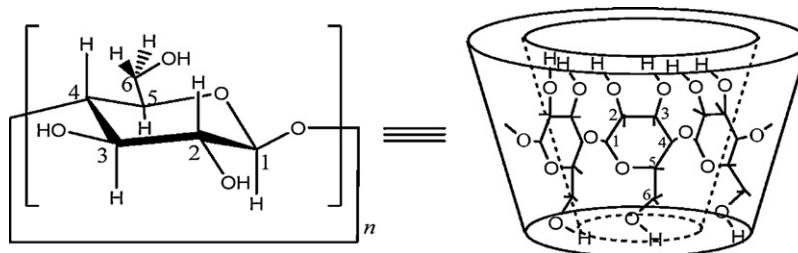


Scheme 1. Molecular structure of resveratrol.

$\geq 98\%$  pure.  $\beta$ - and  $M\beta$ -CD were purchased from Shanghai Chemical Reagent Company and purified twice by recrystallization in redistilled water. All of the CD samples were dried under reduced pressure at 358 K for 24 h prior to use. Double distilled water used in the experiment was prepared in the presence of basic potassium permanganate. Mass of each sample was weighed using a Mettler Toledo AG 135 analytical balance with a precision of  $\pm 0.01$  mg.

## 2.2. Microcalorimetric measurement

Titration calorimetric measurement was performed by a 201 nanowatt scale isothermal titration microcalorimeter supported by Thermal Activity Monitor TAM 2277 (Thermometric, Sweden), which was controlled by Digitam 4.1 software. This instrument can be electrically calibrated with a precision better than  $\pm 1\%$  that was verified by measuring the dilution enthalpy of a concentrated sucrose solution with the literature method [29]. In the experiment, the reaction cell and reference cell of the calorimeter were initially loaded with 500  $\mu\text{L}$  resveratrol solution at  $5.21 \times 10^{-4}$  mol  $\text{L}^{-1}$  and 750  $\mu\text{L}$  pure water, respectively; the titrant solution was injected into the stirred sample vessel in 30 aliquots of 12  $\mu\text{L}$  using a Hamilton syringe controlled by a 612 Lund Pump. The concentrations of  $\alpha$ -,  $\beta$ -,  $\gamma$ - and  $M\beta$ -cyclodextrins as titrants were, respectively,  $1.46 \times 10^{-3}$ ,  $1.43 \times 10^{-3}$ ,  $1.40 \times 10^{-3}$  and  $8.42 \times 10^{-4}$  mol  $\text{L}^{-1}$ . The interval between two injections was 40 min, which was sufficiently long for the signal to return to the baseline. The system was stirred at 30 rpm with a gold propeller. All experiments were performed at the temperature of  $298.15 \pm 0.01$  K and were started after the baseline became stable so that the heat produced by stir can be automatically deducted. To deduct the dilution heats of resveratrol and CD solution, titration experiments were performed for each of CD solution titrated into the solvent (pure water) and the solvent into resveratrol solution, respectively. All of the dilution heats were found to be negligible.



$$[n = 6 (\alpha\text{-CD}), n = 7 (\beta\text{-CD}), n = 8 (\gamma\text{-CD})]$$

Scheme 2. Molecular structure of CDs.

## 2.3. UV-vis spectrum determination

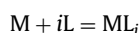
An Hp8453 UV-vis spectrophotometer (USA) was used for absorbance determination of solutions. The temperature of the sample was controlled at  $298.0 \pm 0.1$  K. Four kinds of CDs with the concentration of  $4.00 \times 10^{-4}$  mol  $\text{L}^{-1}$  were added into the resveratrol solution in which the concentration of resveratrol was always at  $4.34 \times 10^{-5}$  mol  $\text{L}^{-1}$ .

## 3. Results and discussion

### 3.1. Thermodynamic data analysis

#### 3.1.1. Analysis process

To evaluate the standard enthalpy of formation for inclusion complex of drug with cyclodextrin in aqueous medium,  $\Delta H_i^\circ$ , apparent equilibrium constants  $\beta_i$  for overall reactions have been defined as follows:



$$\beta_i = \frac{[ML_i]}{[M][L]^i} \quad \Delta H_i^\circ \quad (1)$$

where  $i = 1, 2, 3$ . In Eq. (1), M or L can represent either guest (Resveratrol) or host (CD), so there are five possible reaction models. The total concentrations of L and M can be obtained according to the following formulas:

$$[M]_0 = [M] \left( 1 + \sum \beta_i [L]^i \right) \quad (2)$$

$$[L]_0 = [L] \left( 1 + [M] \sum i \beta_i [L]^{(i-1)} \right) \quad (3)$$

The relationship between the overall equilibrium constants,  $\beta_i$ , and the stepwise equilibrium constants,  $K_i$ , is given by

$$\beta_i = \prod K_i \quad (4)$$

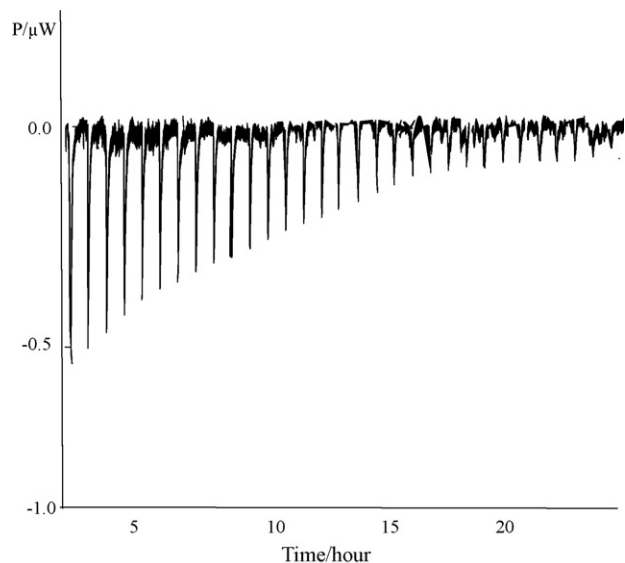
$\Delta H_i^\circ$  and  $\beta_i$  (or  $K_i$ ) can be obtained by the multi-nonlinear regression equation analysis with the Ligand Binding program of the Digitam 4.1 software. By comparing the congruousness between the simulation curve and the experiment point, the most reasonable result can be obtained as final result. Furthermore, the standard changes of Gibbs free energy ( $\Delta G^\circ$ ) and entropy effect ( $T\Delta S^\circ$ ) of the combination can be derived using the following thermodynamic formulas [30]:

$$\Delta G_i^\circ = -RT \ln \beta_i \quad (5)$$

$$\Delta G_i^\circ = \Delta H_i^\circ - T\Delta S_i^\circ$$

**Table 1**  
Thermodynamic parameters of the complexation of host ( $\alpha$ -,  $\beta$ -,  $\gamma$ -, M $\beta$ -CD) with guest (RES) in aqueous solution at 298.15 K.

Host	Reaction model	$\beta_i$ (L/mol)	$\Delta H^\circ$ (kJ/mol)	$\Delta G^\circ$ (kJ/mol)	$T\Delta S^\circ$ (kJ/mol)
$\alpha$ -CD	M + L = ML	877.24	-27.18	-16.80	-10.38
$\beta$ -CD	M + L = ML	952.29	-26.16	-17.00	-9.16
$\gamma$ -CD	M + L = ML	$4.74 \times 10^4$	-1.50	-26.69	25.19
M $\beta$ -CD	M + L = ML	3054.10	-20.36	-19.89	-0.47

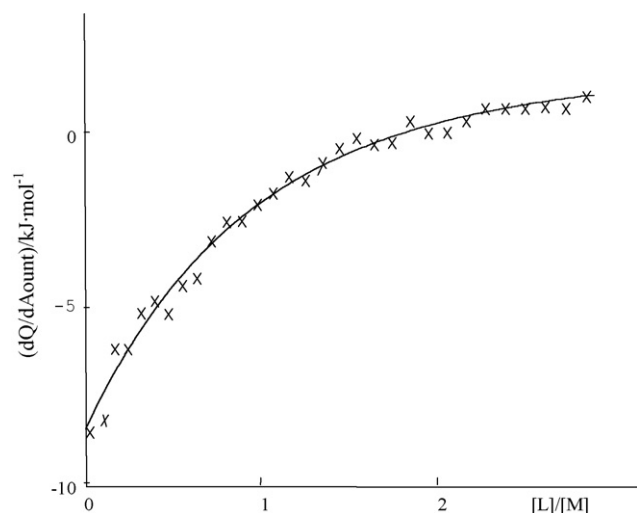


**Fig. 1.** Variation of heat-flow/electrical power as a function of time, titrant:  $\beta$ -CD ( $1.43 \times 10^{-3} \text{ mol L}^{-1}$ ); titrand: RES ( $5.21 \times 10^{-4} \text{ mol L}^{-1}$ ).

The relationship between the enthalpy of formation ( $\Delta H_i^\circ$ ) and the apparent equilibrium constant ( $\beta_i$ ) can be shown by Eq. (6).

$$\Delta H_i^\circ = -RT \ln \beta_i + T\Delta S^\circ \quad (6)$$

The thermodynamic parameters for the resveratrol complexes with several CDs are listed in Table 1, and a typical calorimetric titration plot and a typical fitting curve of the binding complex system are shown in Fig. 1 and Fig. 2, respectively.



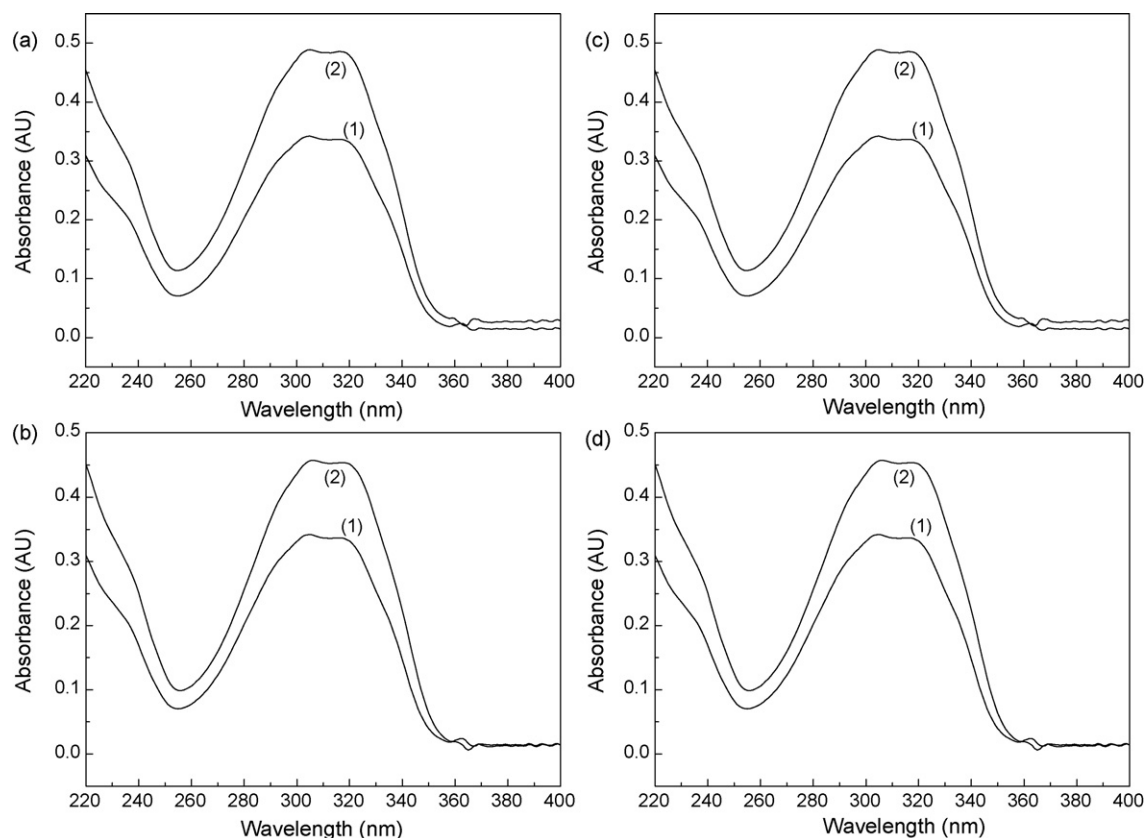
**Fig. 2.** Rate of change in the heat caused by the host-guest interaction versus the ratio of concentrations  $[L]/[M]$ . L and M, respectively, represent  $\beta$ -CD and RES, the points are gotten from experiment and the line is the result of calculated. The concentrations of the titrant and the titrand are, respectively, the same as those in Fig. 1.

### 3.1.2. Thermodynamic parameters

As shown by the data in Table 1, the four host-guest complexes are all in 1:1 stoichiometry (M + L = ML). The cause might be that the concentrations of CD and resveratrol are rather low in the solution, so there is little opportunity for the host and the guest to form complexes in other stoichiometry. From the values of  $\beta_i$  ( $=\beta_1$ ), it could be observed that the inclusion complex of  $\gamma$ -CD with the drug is much more stable than those of other CDs with the drug. This phenomenon can be attributed to the different inner sizes of the hydrophobic cavity of the CDs. The molecular dimension of  $\gamma$ -CD is the largest, so it might be just right for the guest molecule entering into the cavity of  $\gamma$ -CD molecule.

It can be seen from Table 1 that the standard formation enthalpies ( $\Delta H^\circ$ ) of all the CD complexes are negative, i.e., the heat effect is beneficial to the formation of host-guest supramolecular structure. The negative heat effects include the energy released by transferring of water molecules from the holes of completely hydrated CD molecules into the bulk aqueous phase and that caused by hydrophobic interaction between the CD and the drug molecules [31,32]. Assuming that the volume of a water molecule is  $\sim 30 \text{ \AA}^3$ , the number of water molecules that per molecule of  $\alpha$ -,  $\beta$ -, and  $\gamma$ -CD can accommodate are 6, 9 and 14, respectively [33,34]. It was proposed [35] that they are not able to develop a full hydrogen bonded network inside the cavity, probably due to the high curvature of the inside of the cavity. When the water molecules are released, the hydrogen bonds reform, which leads to a release of heat.  $\Delta H^\circ$  is less negative for the inclusion compounds formed by the drug with  $\beta$ -CD and M $\beta$ -CD than that formed by the drug with  $\alpha$ -CD and much negative than that with  $\gamma$ -CD. This phenomenon may be caused by the incomplete intramolecular hydrogen-bond belt in  $\alpha$ -CD molecule, since one glucopyranose unit is in a distorted position [36]. Consequently, the molecular conformation of  $\alpha$ -CD can alter to combine with the drug molecule more tightly. So, binding between the interior of  $\alpha$ -CD molecule with the hydrophobic part of the drug molecule releases a larger amount of energy. For  $\beta$ -CD or M $\beta$ -CD molecule, they have smaller curvature inside their cavity, to which the water molecules can better adapt. All water molecules are expected to be expelled upon complexation of both  $\beta$ -CD and M $\beta$ -CD with the guest. However, the minimum of the binding enthalpies of  $\gamma$ -CD with drug can be attributed to the retention of the water in the inclusion complex cavity allowing for a 'looseness' of the guest molecule fits into the  $\gamma$ -CD cavity. In a word, the foregoing trend can be well understood by considering all of the factors together.

The entropy effect ( $T\Delta S^\circ$ ) of  $\alpha$ -,  $\beta$ -,  $\gamma$ - and M $\beta$ -CD-guest complexes are  $-10.38$ ,  $-9.61$ ,  $25.19$  and  $-0.47 \text{ kJ mol}^{-1}$ , respectively, which are all negative values except for  $\gamma$ -CD-drug complex. In order to explain this tendency, structural factors governing the entropy effect should be taken into account. As mentioned above, the drug molecule possesses hydrophobic groups, benzene ring and hydrocarbyl chain, which can drive water molecules from the molecular cavity of CD to the bulk solvent, this transport of water molecules can cause increase in entropy. Furthermore, collapse of the iceberg structure formed by water molecules around each hydrophobic group also causes the entropy increase when the hydrophobic groups combine with the CD molecule. If only these two entropy-increasing factors were considered, formation of CD



**Fig. 3.** Influence of CD on the UV-vis spectrum of RES, (a):  $\alpha$ -CD; (b):  $\beta$ -CD; (c):  $\gamma$ -CD; (d): M $\beta$ -CD. Curve (1): aqueous solution of RES at  $4.34 \times 10^{-5} \text{ mol L}^{-1}$ ; Curve (2): aqueous solution of RES at  $4.34 \times 10^{-5} \text{ mol L}^{-1}$  in coexistence with CD at  $4.00 \times 10^{-4} \text{ mol L}^{-1}$ .

complex with the drug would be an entropy-increasing process. However, there is an important factor causing entropy decrease, that is the directly hydrophobic interaction of molecular hole of CD with the included part of the drug molecule, which makes negative contribution to entropy. From the values of  $T\Delta S^\circ$ , we can deduce that the former influence surpass the later one for the interaction of  $\gamma$ -CD with the drug, while the later influence become dominant for the interactions of  $\alpha$ -,  $\beta$ - and M $\beta$ -CD with the drug. The values of  $T\Delta S^\circ$  also indicate that the order of the contribution of entropy to the formation of inclusion complex is that:  $\gamma$ -CD–drug > M $\beta$ -CD–drug >  $\beta$ -CD–drug >  $\alpha$ -CD–drug, which is opposite to the tendency of the enthalpy change. This tendency might be mainly decided by the transfer of water molecules from molecular cavity of CD to the bulk medium. The larger the inner cavity of CD molecule is, the more the water molecules can be driven from the host cavity.

The values of standard Gibbs free energy changes ( $\Delta G^\circ$ ), which are decided by enthalpy changes as well as entropy changes, indicate that all of the inclusion complexes can spontaneously form in aqueous solution. Combining with the values of  $\Delta H^\circ$  and  $T\Delta S^\circ$ , we can deduce that the complexation processes of  $\alpha$ -,  $\beta$ - and M $\beta$ -CD with the drug were shown to be enthalpy favored, while the formation of  $\gamma$ -CD–drug was synergistically driven by enthalpy and entropy.

### 3.2. Ultraviolet absorption spectra

Fig. 3(a)–(d), respectively, shows the UV spectra of resveratrol in the presence of  $\alpha$ -,  $\beta$ -,  $\gamma$ - and M $\beta$ -cyclodextrin. From these figures we can see that the typical absorption peaks of resveratrol center at 306 nm, which is in good accordance with that reported in the literature [18]. When the drug is in coexistence

**Table 2**

The changes in absorbance at the peak wavelength ( $\Delta A$ ) caused by the CDs at the concentration about 10 times of the guest concentration, the concentrations of resveratrol and each of the CDs are  $4.34 \times 10^{-5}$  and  $4.00 \times 10^{-4} \text{ mol L}^{-1}$ , respectively.

Hosts	$\alpha$ -CD	$\beta$ -CD	$\gamma$ -CD	M $\beta$ -CD
$\Delta A$	0.160	0.123	0.147	0.116

with a CD, the absorption peak becomes higher at almost the same wavelength, this indicate that the host–guest inclusion complex is formed [20]. There is nearly no influence of each of the CDs on the peak wavelength of the drug (Fig. 3), which means that the interaction between the host and the guest is quite weak. The changes in absorbance at the peak wavelength ( $\Delta A$ ) caused by the CDs at the concentration about 10 times of the guest concentration are shown in Table 2. The value of  $\Delta A$  corresponding to the formation of the  $\alpha$ -CD complex with the drug is the largest, showing that its inner surface is most near to the hydrophobic part of the guest, in other words, association force between  $\alpha$ -CD and resveratrol is the strongest. This is in accordance with the largest negative formation enthalpy value (Table 1).

### 4. Conclusions

Calorimetric measurement for the interactions of  $\alpha$ -,  $\beta$ -,  $\gamma$ - and M $\beta$ -CD with resveratrol at 298.15 K have shown that all of the stoichiometry of the host–guest complex is in 1:1. The complexation processes of  $\alpha$ -,  $\beta$ - and M $\beta$ -CD–drug are predominantly enthalpy favored. While  $\gamma$ -CD–drug is mainly enthalpy and entropy driven. The UV–vis spectra can be regarded as evidence for the formation of inclusion complexes.

## Acknowledgements

The authors are grateful to the National Natural Science Foundation of China (No. 20773059) for financial support.

## References

- [1] P. Saiko, A. Szakmary, W. Jaeger, T. Szekeres, Resveratrol and its analogs: defense against cancer, coronary disease and neurodegenerative maladies or just a fad? *Mutat. Res.* 658 (2008) 68–94.
- [2] D. Colin, A. Lancon, D. Delmas, G. Lizard, J. Abrossinow, E. Kahn, B. Jannin, N. Latruffe, Antiproliferative activities of resveratrol and related compounds in human hepatocyte derived HepG2 cells are associated with biochemical cell disturbance revealed by fluorescence analyses, *Biochimie* 90 (2008) 1674–1684.
- [3] D. Delmas, B. Jannin, N. Latruffe, Resveratrol: preventing properties against vascular alterations and ageing, *Mol. Nutr. Food Res.* 49 (2005) 377–395.
- [4] B. Olas, B. Wachowicz, Resveratrol, A phenolic antioxidant with effects on blood platelet functions, *Platelets* 16 (2005) 251–260.
- [5] C.S. Yang, J.M. Landau, M.T. Huang, H.L. Newmark, Inhibition of carcinogenesis by dietary polyphenolic compounds, *Annu. Rev. Nutr.* 21 (2001) 381–406.
- [6] M. Nielsen, R.J. Ruch, O. Vang, Resveratrol reverses tumor-promoter-induced inhibition of gap-junctional intercellular communication, *Biochem. Biophys. Res. Commun.* 275 (2000) 804–808.
- [7] V. Krishnan, S.L. Zeichner, Host cell gene expression during human immunodeficiency virus type 1 latency and reactivation and effects of targeting genes that are differentially expressed in viral latency, *J. Virol.* 78 (2004) 9458–9473.
- [8] J.A. Parker, M. Arango, S. Abderrahmane, E. Lambert, C. Tourette, H. Catoire, C. Neri, Resveratrol rescues mutant polyglutamine cytotoxicity in nematode and mammalian neurons, *Nat. Genet.* 37 (2005) 349–350.
- [9] C.A. de la Lastra, I. Villegas, Resveratrol as an anti-inflammatory and anti-aging agent: mechanisms and clinical implications, *Mol. Nutr. Food Res.* 49 (2005) 405–430.
- [10] K.T. Howitz, K.J. Bitterman, H.Y. Cohen, D.W. Lamming, S. Lavu, J.G. Wood, R.E. Zipkin, P. Chung, A. Kisielewski, L.L. Zhang, B. Scherer, D.A. Sinclair, Small molecule activators of sirtuins extend *Saccharomyces cerevisiae* lifespan, *Nature* 425 (2003) 191–196.
- [11] J.A. Baur, K.J. Pearson, N.L. Price, H.A. Jamieson, C. Lerin, A. Kalra, V.V. Prabhu, J.S. Allard, G. Lopez-Lluch, K. Lewis, P.J. Pistell, S. Poosala, K.G. Becker, O. Boss, D. Gwinn, M. Wang, S. Ramaswamy, K.W. Fishbein, R.G. Spencer, E.G. Lakatta, D. Le Couteur, R.J. Shaw, P. Navas, P. Puigserver, D.K. Ingram, R. de Cabo, D.A. Sinclair, Resveratrol improves health and survival of mice on a high-calorie diet, *Nature* 444 (2006) 337–342.
- [12] M. Lagouge, C. Argmann, Z. Gerhart-Hines, H. Meziane, C. Lerin, F. Daussin, N. Messadeq, J. Milne, P. Lambert, P. Elliott, B. Geny, M. Laakso, P. Puigserver, J. Auwerx, Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1 $\alpha$ , *Cell* 127 (2006) 1109–1122.
- [13] M. Sengottavelan, K. Deeptha, N. Nalini, Resveratrol ameliorates DNA damage, prooxidant and antioxidant imbalance in 1,2-dimethylhydrazine induced rat colon carcinogenesis, *Chem.-Biol. Interact.* 181 (2009) 193–201.
- [14] S. Fabris, F. Momo, G. Ravagnan, R. Stevanato, Antioxidant properties of resveratrol and piceid on lipid peroxidation in micelles and monolamellar liposomes, *Biophys. Chem.* 135 (2008) 76–83.
- [15] B. Fauconneau, P. Waffo-Teguo, F. Huguët, L. Barrier, A. Decendit, J.M. Merillon, Comparative study of radical scavenger and antioxidant properties of phenolic compounds from *Vitis vinifera* cell cultures using in vitro tests, *Life Sci.* 61 (1997) 2103–2110.
- [16] X.H. Shu, H. Li, Z. Sun, M.L. Wu, J.X. Ma, J.M. Wang, Q. Wang, Y. Sun, Y.S. Fu, X.Y. Chen, Q.Y. Kong, J. Liu, Identification of metabolic pattern and bioactive form of resveratrol in human medulloblastoma cells, *Biochem. Pharmacol.* 79 (2010) 1516–1525.
- [17] B. Billack, V. Radkar, C. Adiabouah, In vitro evaluation of the cytotoxic and antiproliferative properties of resveratrol and several of its analogs, *Cell Mol. Biol. Lett.* 13 (2008) 553–569.
- [18] V. Bertacche, N. Lorenzi, D. Nava, E. Pini, C. Sinico, Host–guest interaction study of resveratrol with natural and modified cyclodextrins, *J. Incl. Phenom. Macro.* 55 (2006) 279–287.
- [19] R. Villalonga, R. Cao, A. Frago, Supramolecular chemistry of cyclodextrins in enzyme technology, *Chem. Rev.* 107 (2007) 3088–3116.
- [20] S.H. Gellman, Introduction: molecular recognition, *Chem. Rev.* 97 (1997) 1231–1734.
- [21] N. Li, L. Xu, Thermal analysis of  $\beta$ -cyclodextrin/Berberine chloride inclusion compounds, *Thermochim. Acta* 499 (2010) 166–170.
- [22] A. Avci, S. Dönmez, A novel thermophilic anaerobic bacteria producing cyclodextrin glycosyltransferase, *Process Biochem.* 44 (2009) 36–42.
- [23] A. Gunaratne, H. Corke, Effect of hydroxypropyl  $\beta$ -cyclodextrin on physical properties and transition parameters of amylose–lipid complexes of native and acetylated starches, *Food Chem.* 108 (2008) 14–22.
- [24] F.B. Sousa, A.M.L. Denadai, I.S. Lula, J.F. Lopes, H.F.D. Santos, W.B.D. Almeida, R.D. Sinisterra, Supramolecular complex of fluoxetine with  $\beta$ -cyclodextrin: an experimental and theoretical study, *Int. J. Pharm.* 353 (2008) 160–169.
- [25] M.B. Jesus, L.M.A. Pinto, L.F. Fraceto, Y. Takahata, A.C.S. Lino, C. Jaime, E. Paula, Theoretical and experimental study of a praziquantel and  $\beta$ -cyclodextrin inclusion complex using molecular mechanic calculations and  $^1\text{H}$ -nuclear magnetic resonance, *J. Pharm. Biomed. Anal.* 41 (2006) 1428–1432.
- [26] H.R. Zhang, G. Chen, L. Wang, L. Ding, Y. Tian, W.Q. Jin, H.Q. Zhang, Study on the inclusion complexes of cyclodextrin and sulfonated azo dyes by electrospray ionization mass spectrometry, *Int. J. Mass Spectrom.* 252 (2006) 1–10.
- [27] A.C. Illapakurthy, C.M. Wyandt, S.P. Stodghill, Isothermal titration calorimetry method for determination of cyclodextrin complexation thermodynamics between artemisinin and naproxen under varying environmental conditions, *Eur. J. Pharm. Biopharm.* 59 (2005) 325–332.
- [28] I. Jelesarov, H.R. Bosshard, Isothermal titration calorimetry and differential scanning calorimetry as complementary tools to investigate the energetics of biomolecular recognition, *J. Mol. Recognit.* 12 (1999) 3–18.
- [29] G.Y. Bai, Y.J. Wang, H.K. Yan, Thermodynamics of interaction between cationic Gemini surfactants and hydrophobically modified polymers in aqueous solutions, *J. Phys. Chem. B* 106 (2002) 2153–2159.
- [30] V. Karlson, E.B. Heggset, M. Sölle, The use of isothermal titration calorimetry to determine the thermodynamics of metal ion binding to low-cost sorbents, *Thermochim. Acta* 501 (2010) 119–121.
- [31] D.Z. Sun, L. Li, X.M. Qiu, F. Liu, B.L. Yin, Isothermal titration calorimetry and  $^1\text{H}$  NMR studies on host–guest interaction of paeonol and two of its isomers with  $\beta$ -cyclodextrin, *Int. J. Pharm.* 316 (2006) 7–13.
- [32] Q. Zhao, X.Y. Xu, X.J. Sun, M. Liu, D.Z. Sun, L.W. Li, A calorimetric study on interactions of colchicine with human serum albumin, *J. Mol. Struct.* 931 (2009) 31–34.
- [33] V.J. Smith, N.M. Rougier, R.H. de Rossi, M.R. Caira, E.I. Buján, M.A. Fernández, S.A. Bourne, Investigation of the inclusion of the herbicide metobromuron in native cyclodextrins by powder X-ray diffraction and isothermal titration calorimetry, *Carbohydr. Res.* 344 (2009) 2388–2393.
- [34] M. Nilsson, A.J.M. Valente, G. Olofsson, O. Soderman, M. Bonini, Thermodynamic and kinetic characterization of host–guest association between bolaform surfactants and  $\alpha$ - and  $\beta$ -cyclodextrins, *J. Phys. Chem. B* 112 (2008) 11310–11316.
- [35] T. Loftsson, M. Masson, M.E. Brewster, Self-association of cyclodextrins and cyclodextrin complexes, *Pharm. Sci.* 93 (2004) 1091–1099.
- [36] J. Szejtli, Introduction and general overview of cyclodextrin chemistry, *Chem. Rev.* 98 (1998) 1743–1754.